

# Investigating the coherency between logical relationships and gene expressions for each connected gene pair in the signaling networks

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## Introduction

Study the correlation between the gene expression profiles at both mRNA and protein level is common. Several studies have shown that mRNA and protein expression are poorly correlated. There are some reasons such as miRNA activity on mRNA transcripts or post-translational modifications which cause the variation in amounts of mRNAs and active/inactive Proteins. However, the association between mRNA and protein of housekeeping genes which is not mostly related to signaling processes has been also reported. Now, another major question is the association between mRNA or protein level with the logical relations inside the wiring diagram of the signaling networks. Considering the sign of interactions between components, does the gene expression or protein amount help the signal to be transduced or not? In two simple elements, there are eight possible states (figure 1). Four of these states conform to the direction of the signal transduction, i.e., if the interaction is activation, both of two related components are up-regulated or down-regulated; and if the interaction is inhibitory, the expression of two associated components is inverse (Panel A). The other four possible states are against to signal transduction's direction (panel B). Analyzing these two different clusters of states in various biological conditions could be an interesting subject for the discussion. Figure 2 is the flowchart which depicts the different steps to reach the data needed to analyze the coherency between the gene pairs in KEGG signaling pathways.

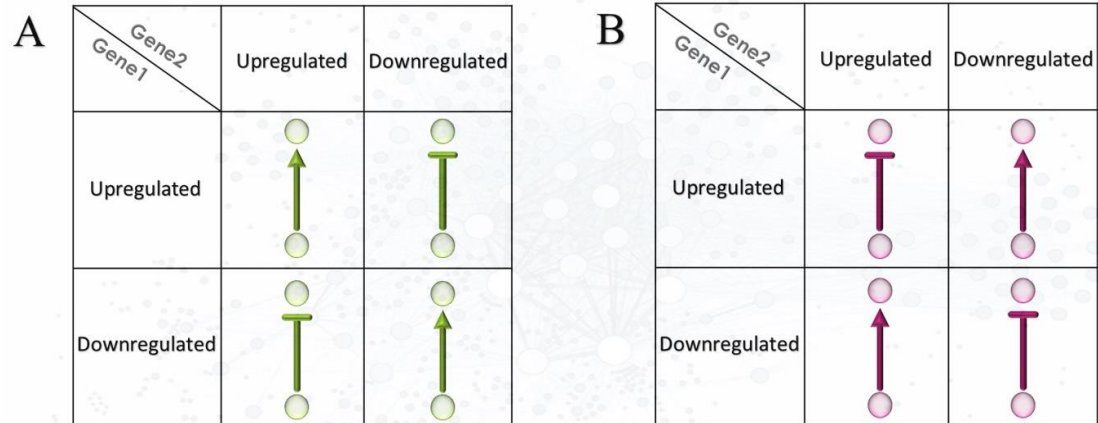
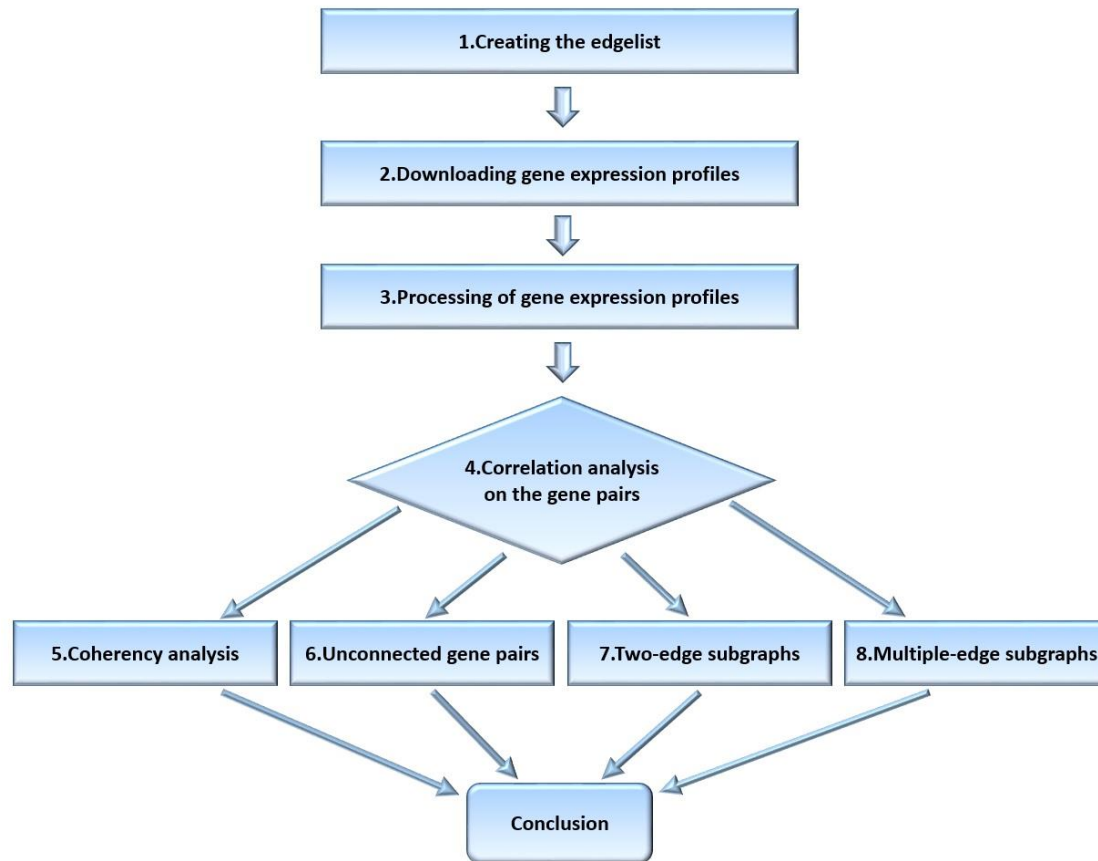


Figure1

## The Procedure



*Figure2: Different steps to analyze the coherency between the gene pairs*

## 1. Creating an edgelist from KEGG database

An edgelist was created from all human KEGG pathways using “KEGGgraph” package.

### 1.1. Downloading KEGG pathways

All the human signaling pathways were downloaded in form of KGML files from KEGG database [KEGG Pathways](#). There were 206 human pathways on July 24th, 2017.

An example of these pathways is: [Ras Signaling Pathway](#)

## 1.2.Importing pathways into R in form of graphs

Downloaded pathways were imported into R using the combination of “parseKGML2Graph” and “lapply” functions. The “parseKGML2Graph” is a function which creates a class of graph (graphNEL) from each pathway and is suitable for networks with few edges and high nodes. All 206 graphs were stored in KGMLGraphs object.

```
library(KEGGgraph)

#Define path
kgfiles<-list.files("F:/Projects/Project Coherency/KEGG GEO/kegg
xml",full.names=T)

#Read kgml files
KGMLGraphs=lapply(kgfiles,parseKGML2Graph,genesOnly=F)
length(KGMLGraphs)

## [1] 206

head(KGMLGraphs)

## [[1]]
## A graphNEL graph with directed edges
## Number of Nodes = 94
## Number of Edges = 247
##
## [[2]]
## A graphNEL graph with directed edges
## Number of Nodes = 115
## Number of Edges = 311
##
## [[3]]
## A graphNEL graph with directed edges
## Number of Nodes = 64
## Number of Edges = 18
##
## [[4]]
## A graphNEL graph with directed edges
## Number of Nodes = 84
## Number of Edges = 79
##
## [[5]]
## A graphNEL graph with directed edges
## Number of Nodes = 93
## Number of Edges = 282
##
## [[6]]
## A graphNEL graph with directed edges
## Number of Nodes = 267
## Number of Edges = 856
```

### 1.3.Extracting edge information

Edge information is derived from each pathway using “getKEGGedgeData” function, and they are stored in **l** object.

```
l = lapply(KGMLGraphs,getKEGGedgeData)
length(l)

## [1] 206
```

### 1.4. Constructing the edgelist

The following codes construct the edgelist from all 206 signaling pathways and remove the non-informative information. The loop contains two “if” conditions to exclude edges without complete data. Note that, nodes (e.g., hsa:1950) are genes and Subtype is the relationship between genes, e.g., activation, inhibition and so on. A large list called **listedgelist** is created, and each element contains an edgelist.

```
# an example of complete informative edge:
l[[1]][[1]]

##    KEGG Edge (Type: PPrel):
## -----
## [ Entry 1 ID ]: hsa:1950
## [ Entry 2 ID ]: hsa:1956
## [ Subtype ]:
## [ Subtype name ]: activation
## [ Subtype value ]: -->
## -----

# First exclusive condition: There is no edge between nodes. For example:
KGMLGraphs[[26]]

## A graphNEL graph with directed edges
## Number of Nodes = 133
## Number of Edges = 0

# Second exclusive condition: There is no information about the edge. For
example:
l[[10]][[251]]

##    KEGG Edge (Type: PCrel):
## -----
## [ Entry 1 ID ]: hsa:6543
## [ Entry 2 ID ]: cpd:C01330
## [ Subtype ]:
## -----
```

```

a = NULL
class(a)

## [1] "NULL"

b = list()

listedgelists = list()

for(i in 1:length(l)){
  if(is.list(l[[i]]) & length(l[[i]]) == 0){
    listedgelists[[i]] = c(0,0,0)
  } else
  {
    d = matrix(0,length(l[[i]]),3)
    d = as.data.frame(d)
    for(j in 1:length(l[[i]])){
      d[j,1] = l[[i]][[j]]@entry1ID
      d[j,2] = l[[i]][[j]]@entry2ID
      c1 = l[[i]][[j]]@subtype$subtype

      if(class(c1) == class(a)){
        d[j,3] = "none"
      } else
      {
        d[j,3] = l[[i]][[j]]@subtype$subtype@name
      }
    }

    listedgelists[[i]] = d
  }
}

```

listedgelists object contains all 206 edgelists.

## 1.5.Merging all edgelists

All the edgelists are attached together and a large edgelist is created.

```

table = do.call("rbind", listedgelists)
dim(table)

## [1] 68757      3

```

Duplicated rows are omitted as below.

```
table1 = table[!duplicated(table),]
```

## 1.6. Preprocessing the edgelist

In the following code, “activation” and “inhibition” interactions are separated from each other.

```
idx1 = which(table1[,3] == "activation")
idx2 = which(table1[,3] == "inhibition")
idx = c(idx1,idx2)
table2 = table1[idx,]
dim(table2)
## [1] 28870      3
```

After that, only edges which are KEGG Ids starting with “hsa” are selected.

```
idx3 = grep("^hsa" , table2[,1])
idx4 = grep("^hsa" , table2[,2])
idx5 = intersect(idx3,idx4)
table3 = table2[idx5,]
dim(table3)
## [1] 26490      3
```

```
sum(!grepl("^hsa" , table3[,1]))
## [1] 0
sum(!grepl("^hsa" , table3[,2]))
## [1] 0
```

Next, all the KEGG Ids are changed into gene Ids.

```

kgid1 = as.character(table3[,1])
kgid2 = as.character(table3[,2])
geneid1 <- translateKEGGID2GeneID(kgid1)
geneid2 <- translateKEGGID2GeneID(kgid2)

any(is.na(geneid1))
## [1] FALSE

any(is.na(geneid2))
## [1] FALSE

```

Gene Ids are mapped to gene symbols using “org.Hs.eg.db” package which contains annotation for the human genome.

```

require(org.Hs.eg.db)

genesymbol1 <- sapply(mget(geneid1, org.Hs.egSYMBOL,
                           ifnotfound=NA), "[", 1)
length(genesymbol1)
## [1] 26490

any(is.na(genesymbol1))
## [1] TRUE

genesymbol2 <- sapply(mget(geneid2, org.Hs.egSYMBOL,
                           ifnotfound=NA), "[", 1)
length(genesymbol2)
## [1] 26490

any(is.na(genesymbol2))
## [1] TRUE

```

## 1.7. Final edgelist

So far, there are three columns in the edgelist. The first two columns contain interacting genes and the last one shows interaction type: activation or inhibition

```
edgelist = table3
```



```

edgelist[,1] = genesymbol1

edgelist[,2] = genesymbol2

dim(edgelist)

## [1] 26490      3

```

Finally, an ID column is added to the edgelist.

```

id1 = paste0("E0000" , 1:9 )
id2 = paste0("E000" , 10:99)
id3 = paste0("E00" , 100:999)
id4 = paste0("E0" , 1000:9999)
id5 = paste0("E" , 10000:length(edgelist[,1]))
ID = c(id1,id2,id3,id4,id5)
edgelist = cbind(ID,edgelist)
colnames(edgelist) = c("ID" , "Gene1" , "Gene2" , "Interaction type")
edgelist = apply(edgelist , 2 , as.character)
edgelist = as.data.frame(edgelist , stringsAsFactors = F)
class(edgelist)

## [1] "data.frame"

```

```

head(edgelist)

##      ID Gene1 Gene2 Interaction type
## 1 E00001   EGF  EGFR      activation
## 2 E00002  TGFA  EGFR      activation
## 3 E00003   HGF   MET      activation
## 4 E00004   MET ERBB3      activation
## 5 E00005  IGF1 IGF1R      activation
## 6 E00006 VEGFA   KDR      activation

```

## 2. Downloading all up or down gene expression profiles

All the genes are stored at **edgelistGenes** object.

```

edgelistGenes = unique(c(edgelist[,2],edgelist[,3]))

length(edgelistGenes)

```

There were 3187 unique genes in the KEGG edgelist. We downloaded all the human up-down gene expression profiles for these genes from GEO database (<https://www.ncbi.nlm.nih.gov/geoprofiles/>). This database contains gene expression profiles derived from curated GEO DataSets and presents them as a chart that displays the expression level of one gene across all Samples within a DataSet. Due to the limitation in NCBI query rules (importing less than 200 gene names in the query and downloading 500 gene profiles in each step), we downloaded 423 text files, each contains nearly 500 gene profiles. A query example for some of the genes is shown here: [GEO Profile](#).

A query example:

```
("Homo sapiens"[Organism] OR Homo sapiens[All Fields]) AND ("PPIA"[Gene Symbol]
OR"GRB2"[Gene Symbol] OR"RAC1"[Gene Symbol] OR"AP2B1"[Gene Symbol]
OR"GNAI1"[Gene Symbol] OR"PPP3R1"[Gene Symbol] OR"YWHAZ"[Gene Symbol] AND "up
down genes"[filter])
```

## 2.1.Merging all gene expression profiles into one file

All the downloaded gene profiles are merged into Geneprofiles.txt as below (figure3):

- 1) all the text files are placed in a folder in drive C.
  - 2) In the Windows7 command line shell, the following codes were run.
- ```
. cd C: Geneprofiles (Enter)
. dir (Enter) (Shows all the files in the directory)
. copy *.txt Geneprofiles.txt (Enter)
```

```

C:\Windows\system32\cmd.exe
Microsoft Windows [Version 6.1.7601]
Copyright (c) 2009 Microsoft Corporation. All rights reserved.

C:\Users\DiamonD>cd C:\Geneprofiles

C:\Geneprofiles>dir
Volume in drive C has no label.
Volume Serial Number is E0B8-A5E6

Directory of C:\Geneprofiles

08/19/2017  09:53 AM    <DIR>          .
08/19/2017  09:53 AM    <DIR>          ..
07/27/2017  01:58 PM             1,945,733 profile_data (1).txt
07/27/2017  02:00 PM             1,704,632 profile_data (2).txt
07/27/2017  02:01 PM             1,997,962 profile_data (3).txt
07/27/2017  02:01 PM             1,820,277 profile_data (4).txt
07/27/2017  02:02 PM             1,868,848 profile_data (5).txt
               5 File(s)              9,337,452 bytes
               2 Dir(s)  12,688,789,504 bytes free

C:\Geneprofiles>copy *.txt Mergedfiles.txt

```

Figure3

## 2.2.Importing Geneprofiles file into R

library(readxl)

Geneprofiles = read\_excel("Geneprofiles.xlsx", col\_names = F)

dim(Geneprofiles)

Geneprofiles = as.data.frame(Geneprofiles)

|    | X_1              | X_2                    | X_3                    | X_4                    | X_5                                         | X_6                                                     | X_7                      |
|----|------------------|------------------------|------------------------|------------------------|---------------------------------------------|---------------------------------------------------------|--------------------------|
| 1  | GDS53            | N/A                    | N/A                    | N/A                    | N/A                                         | N/A                                                     | N/A                      |
| 2  | ID_REF           | GSM575                 | GSM576                 | GSM577                 | GSM578                                      | GSM579                                                  | Gene title               |
| 3  | cell type        | CD34+ from bone marrow | CD34+ from bone marrow | CD34+ from bone marrow | CD34+ from G-CSF-mobilized peripheral blood | CD34+ from G-CSF-mobilized peripheral blood             | N/A                      |
| 4  | M22877_at        | 4419                   | 4528.3                 | 4732.00000000000004    | 2309.30000000000002                         | 1757.6                                                  | cytochrome c, somatic    |
| 5  | M27068_s_at      | 2204.9                 | 2417                   | 2456.0999999999998     | 5088.2                                      | 4615.3                                                  | fibroblast growth factor |
| 6  | N/A              | N/A                    | N/A                    | N/A                    | N/A                                         | N/A                                                     | N/A                      |
| 7  | GDS157           | N/A                    | N/A                    | N/A                    | N/A                                         | N/A                                                     | N/A                      |
| 8  | ID_REF           | GSM2289                | GSM2294                | GSM2299                | GSM2304                                     | GSM2309                                                 | GSM2313                  |
| 9  | metabolism       | insulin resistant      | insulin resistant      | insulin resistant      | insulin resistant                           | insulin resistant                                       | insulin sensitive        |
| 10 | U90907_at        | 1443[A]                | 1520[A]                | 1676                   | 518                                         | 764                                                     | 507[A]                   |
| 11 | N/A              | N/A                    | N/A                    | N/A                    | N/A                                         | N/A                                                     | N/A                      |
| 12 | GDS160           | N/A                    | N/A                    | N/A                    | N/A                                         | N/A                                                     | N/A                      |
| 13 | ID_REF           | GSM2284                | GSM2315                | GSM2320                | GSM2325                                     | GSM2330                                                 | GSM2286                  |
| 14 | metabolism       | insulin sensitive      | insulin sensitive      | insulin sensitive      | insulin sensitive                           | insulin sensitive                                       | insulin resistant        |
| 15 | RC_AA018907_s_at | null                   | 127                    | 291                    | 223                                         | 186                                                     | 68[A]                    |
| 16 | N/A              | N/A                    | N/A                    | N/A                    | N/A                                         | N/A                                                     | N/A                      |
| 17 | GDS161           | N/A                    | N/A                    | N/A                    | N/A                                         | N/A                                                     | N/A                      |
| 18 | ID_REF           | GSM2287                | GSM2292                | GSM2297                | GSM2302                                     | GSM2307                                                 | GSM2311                  |
| 19 | metabolism       | insulin resistant      | insulin resistant      | insulin resistant      | insulin resistant                           | insulin resistant                                       | insulin sensitive        |
| 20 | RC_T10824_s_at   | 1803[A]                | 703[A]                 | 376[A]                 | 61[A]                                       | null                                                    | 219[A]                   |
| 21 | N/A              | N/A                    | N/A                    | N/A                    | N/A                                         | N/A                                                     | N/A                      |
| 22 | GDS162           | N/A                    | N/A                    | N/A                    | N/A                                         | N/A                                                     | N/A                      |
| 23 | ID_REF           | GSM2288                | GSM2293                | GSM2298                | GSM2303                                     | GSM2308                                                 | GSM2312                  |
| 24 | metabolism       | insulin resistant      | insulin resistant      | insulin resistant      | insulin resistant                           | insulin resistant                                       | insulin sensitive        |
| 25 | RC_AA279937_at   | null                   | null                   | 357[A]                 | 504[A]                                      | 376[A]                                                  | 80[A]                    |
| 26 | N/A              | N/A                    | N/A                    | N/A                    | N/A                                         | N/A                                                     | N/A                      |
| 27 | GDS215           | N/A                    | N/A                    | N/A                    | N/A                                         | N/A                                                     | N/A                      |
| 28 | ID_REF           | GSM4363                | GSM4364                | GSM4367                | GSM4368                                     |                                                         | Gene title               |
| 29 | specimen         | PLGC-(136-129)         | PLGC-(136-129)         | PSGA-(133-188)         | PSGA-(133-188)                              |                                                         | Gene symbol              |
| 30 | 40704_at         | 73.855000000000004     | 70.414000000000001     | 137.28700000000001     | 137.94                                      | phosphatidylinositol-4,5-bisphosphate 3-kinase catal... | PIK3CA                   |

Showing 1 to 31 of 558,097 entries

### 3.Processing gene expression profiles

#### 3.1.Creating a large list called listGeneprofiles which contains each part of Geneprofiles dataframe in one of its elements

The following codes are to create a large list called **listGeneprofiles**. each element of the list contains one row for the GDS name, one row for ID\_REF, GSMs, Gene ids and Gene symbol, and multiple rows for the expression values.

An empty list called listGeneprofiles is created. To extract the information of each GDS part, rows indexes containing "GDS" are found. A "loop" is used to exert some operations on each GDS part. v1 is a vector which contains the row after GDS i (the row which start with ID\_REF). "index" is a vector holding all the elements of v1 which start with GSM letters. "l" shows how many GSMs are present in row idx[i] + 1. "d" is a data frame as the same dimension as each GDS part. The number of rows in matrix d, is equal to The number of rows starting from GDS[i] to GDS[i+1] minus two ("length((idx[i]):(idx[i+1] - 2))"). The number of columns in object d, is equal to the sum of column one as well as all the GSM columns and two columns after GSM columns (length(c(1,index,index[l]+c(1,2))))). Whenever d is completed for GDS i, extra rows are deleted. In other words, row one and rows which contain expression values (have "at" or numbers at the end) are kept.

```
listGeneprofiles = list()
idx = grep("^GDS" , Geneprofiles[,1])
for(i in 1:(length(idx)-1)){

  v1 = data[idx[i]+1,]
  index = grep("^GSM" , v1)
  l = length(index)

  d = as.data.frame(matrix(0,length((idx[i]):(idx[i+1] - 2)) ,
length(c(1,index,index[l]+c(1,2)))))
  for(j in 1:length((idx[i]):(idx[i+1] - 2))) {
    d[j,] = Geneprofiles[idx[i]+j-1,c(1,index,index[l]+c(2,3))]
  }

  colnames(d) = d[2,]
  idxxx = grep(paste(paste(c(0:9,'_at'),"$",sep=""),collapse="|"),d[,1])
  listGeneprofiles[[i]] = d[idxxx,]

  #print(i)
}

length(listGeneprofiles)

## [1] 73460
```

listGeneprofiles contains 73460 elements. each element has a similar structure as below.

```
listGeneprofiles[[1]]
```

|      | ID_REF      | GSM575  | GSM576 | GSM577             | GSM578             | GSM579 |
|------|-------------|---------|--------|--------------------|--------------------|--------|
| ## 1 | GDS53       | <NA>    | <NA>   | <NA>               | <NA>               | <NA>   |
| ## 4 | M22877_at   | 4419    | 4528.3 | 4732.6000000000004 | 2399.3000000000002 | 1757.6 |
| ## 5 | M27968_s_at | 2204.9  | 2417   | 2456.6999999999998 | 5088.2             | 4615.3 |
| ##   | Gene symbol | Gene ID |        |                    |                    |        |
| ## 1 | <NA>        | <NA>    |        |                    |                    |        |
| ## 4 | CYCS        | 54205   |        |                    |                    |        |
| ## 5 | FGF2        | 2247    |        |                    |                    |        |

```
listGeneprofiles[[5777]]
```

|                    | ID_REF             | GSM136055          | GSM136056          | GSM136057          |
|--------------------|--------------------|--------------------|--------------------|--------------------|
| ## 1               | GDS2494            | <NA>               | <NA>               | <NA>               |
| ## 5               | 200065_s_at        | 3630.35            | 3685.9             | 3662.81            |
| ## 6               | 202512_s_at        | 48.658799999999999 | 49.640799999999999 | 50.71              |
| ## 7               | 221497_x_at        | 162.16499999999999 | 128.107            | 138.99700000000001 |
| ##                 | GSM136064          | GSM136065          | GSM136066          |                    |
| GSM136058          |                    |                    |                    |                    |
| ## 1               | <NA>               | <NA>               | <NA>               |                    |
| <NA>               |                    |                    |                    |                    |
| ## 5               | 3622.76            | 3423.19            | 3399.93            |                    |
| 3083.94            |                    |                    |                    |                    |
| ## 6               | 72.458799999999997 | 55.823999999999998 | 71.829800000000006 |                    |
| 38.535200000000003 |                    |                    |                    |                    |
| ## 7               | 90.802700000000002 | 82.813800000000001 | 78.484999999999999 |                    |
| 142.18299999999999 |                    |                    |                    |                    |
| ##                 | GSM136059          | GSM136060          | Gene symbol        | Gene ID            |
| ## 1               | <NA>               | <NA>               | <NA>               | <NA>               |
| ## 5               | 3125.62            | 3177.01            | MIR3620///ARF1     | 100500810///375    |
| ## 6               | 38.714700000000001 | 43.994900000000001 | ATG5               | 9474               |
| ## 7               | 156.648            | 143.34800000000001 | EGLN1              | 54583              |

### 3.2.Creating a list called preListGDS which contains each GDS (dataset) in one of its elements

The following codes show how many GDSes (datasets) does exist in the “listGeneprofiles” object.

```
gds = c()
for(i in 1:length(listGeneprofiles)){
  gds[i] = listGeneprofiles[[i]][1,1]
```

```

}
gds1 = unique(gds)

length(gds)

## [1] 73460

```

Same datasets are merged as below.

```

preListGDS = list()
for(i in 1:length(gds)){
  idx = which(gds1[i]==gds)
  preListGDS[[i]] = do.call("rbind",listGeneprofiles[idx])
}
length(preListGDS)

## [1] 1693

# There are 1693 datasets

```

| ID_REF         | GSM2289 | GSM2294 | GSM2299 | GSM2304 | GSM2309 | GSM2313 | GSM2318 | GSM2323 | GSM2328 | GSM2333 | Gene symbol | Gene ID |
|----------------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|-------------|---------|
| 1 GDS157       | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA          | NA      |
| 4 U09007_at    | 1443[A] | 1520[A] | 1676    | 518     | 764     | 507[A]  | 910[A]  | 551     | 494     | 503     | PIK3R3      | 8503    |
| 11 GDS157      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA          | NA      |
| 41 U11821_s_at | null    | 115[A]  | 93[A]   | null    | 19[A]   | 5[A]    | null    | null    | 11[A]   | null    | FASLG       | 356     |
| 5 X57025_at    | 1       | 21[A]   | 21[A]   | 125     | 140     | 142[A]  | null    | 115     | 150     | 108     | IGF1        | 3479    |
| 12 GDS157      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA          | NA      |
| 42 K03021_at   | null    | 25[A]   | 76[A]   | 102[A]  | 46[A]   | 289[A]  | null    | 188[A]  | null    | 22[A]   | PLAT        | 5327    |
| 51 M74088_s_at | null    | null    | 164[A]  | 93[A]   | null    | null    | null    | 21[A]   | 21[A]   | 20[A]   | APC         | 324     |
| 13 GDS157      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA          | NA      |
| 43 L13266_s_at | 292[A]  | 589[A]  | 787[A]  | 227[A]  | null    | 305[A]  | 408[A]  | 71[A]   | 56[A]   | 77[A]   | GRIN1       | 2902    |
| 14 GDS157      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA          | NA      |
| 44 L06132_at   | 2496    | 1957    | 2742    | 5133    | 5697    | 7767    | 1813    | 6071    | 6205    | 6147    | VDAC1       | 7416    |
| 52 X09101_at   | 1532    | 1604    | 1215    | 525     | 746     | 1137    | 1330    | 570     | 574[A]  | 575[A]  | ESR2        | 2100    |
| 15 GDS157      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA          | NA      |
| 45 M22382_at   | 836     | 403     | 968     | 1096    | 1262    | 1313    | 601     | 1275    | 2084    | 1431    | HSPD1       | 3329    |
| 16 GDS157      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA          | NA      |
| 46 M62994_at   | 178[A]  | 131[A]  | 244[A]  | 5[A]    | 84[A]   | null    | 52[A]   | 39[A]   | 42[A]   | 39[A]   | FLNB        | 2317    |
| 17 GDS157      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA          | NA      |
| 47 U58087_at   | 280[A]  | 240[A]  | null    | 434     | 113     | 576[A]  | 541[A]  | 348     | 331     | 310     | CUL1        | 8454    |
| 18 GDS157      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA          | NA      |
| 48 L07590_at   | 2224    | 2198    | 2117    | 1392    | 1585    | 2171    | 1439    | 1726    | 1371    | 1608    | PPP2R3A     | 5523    |
| 53 L13740_at   | 1266[A] | 1232[A] | 1386    | 705     | 415[A]  | 890[A]  | 1296[A] | 497     | 516     | 473     | NR4A1       | 3164    |
| 19 GDS157      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA          | NA      |
| 49 X02317_at   | 2229    | 2280    | 2542    | 1757    | 2269    | 2099    | 1902    | 1900    | 2128    | 1975    | SOD1        | 6647    |
| 110 GDS157     | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA          | NA      |
| 410 D79990_at  | 3[A]    | 30[A]   | null    | 22[A]   | 10[A]   | null    | 197[A]  | 38[A]   | 82[A]   | 71[A]   | RASSF2      | 9770    |
| 54 U35459_at   | 336[A]  | 437[A]  | 403[A]  | 70[A]   | 237[A]  | 424[A]  | 197[A]  | 118[A]  | 89[A]   | 140[A]  | SERPINB10   | 5273    |
| 111 GDS157     | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA          | NA      |
| 411 Z29574_at  | 306[A]  | 248[A]  | 76[A]   | 94[A]   | 116[A]  | 196     | 61[A]   | 65[A]   | 55[A]   | 54[A]   | TNFRSF17    | 608     |
| 112 GDS157     | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA      | NA          | NA      |

Showing 1 to 31 of 79 entries

Figure5

Some datasets does not contain gene symbols or gene ids. We delete them as below.

```
v = c()
for(i in 1:length(preListGDS)){
  v[i]="Gene symbol" %in% colnames(preListGDS[[i]]) & "Gene ID" %in%
colnames(preListGDS[[i]])
}

index = which(!v)
index

## [1] 933 1062 1229 1332

l = preListGDS[index]

c(l[[1]][1,1],l[[2]][1,1],l[[3]][1,1],l[[4]][1,1])

## [1] "GDS2771" "GDS3795" "GDS3268" "GDS4206"

colnames(l[[2]])

## [1] "ID_REF" "GSM483301" "GSM483302" "GSM483303" "GSM483305"
"GSM483307"
## [7] "GSM483312" "GSM483313" "GSM483317" "GSM483318" "GSM483319"
"GSM483322"
## [13] "GSM483327" "GSM483328" "GSM483330" "GSM483332" "GSM483333"
"GSM483336"
## [19] "GSM483337" "GSM483339" "GSM483351" "GSM483352" "GSM483354"
"GSM483358"
## [25] "GSM483384" "GSM483386" "GSM483388" "GSM483390" "GSM483391"
"GSM483396"
## [31] "GSM483399" "GSM483400" "GSM483401" "GSM483412" "GSM483418"
"GSM483420"
## [37] "GSM483421" "GSM483426" "GSM483428" "GSM483431" "GSM483436"
"GSM483442"
## [43] "GSM483443" "GSM483444" "GSM483447" "GSM483448" "GSM483450"
"GSM483455"
## [49] "GSM483458" "GSM483461" "GSM483462" "GSM483464" "GSM483466"
"GSM483468"
## [55] "GSM483476" "GSM483477" "GSM483300" "GSM483308" "GSM483310"
"GSM483311"
## [61] "GSM483323" "GSM483338" "GSM483353" "GSM483361" "GSM483363"
"GSM483364"
## [67] "GSM483366" "GSM483368" "GSM483371" "GSM483372" "GSM483373"
"GSM483374"
## [73] "GSM483379" "GSM483380" "GSM483381" "GSM483389" "GSM483404"
"GSM483405"
## [79] "GSM483410" "GSM483411" "GSM483413" "GSM483416" "GSM483417"
"GSM483419"
## [85] "GSM483427" "GSM483433" "GSM483434" "GSM483445" "GSM483459"
"GSM483465"
```

```
## [91] "GSM483470" "GSM483473" "GSM483478" "GSM483304" "GSM483315"
"GSM483320"
## [97] "GSM483325" "GSM483329" "GSM483331" "GSM483334" "GSM483341"
"GSM483343"
## [103] "GSM483344" "GSM483347" "GSM483348" "GSM483349" "GSM483350"
"GSM483356"
## [109] "GSM483362" "GSM483365" "GSM483367" "GSM483369" "GSM483370"
"GSM483375"
## [115] "GSM483376" "GSM483377" "GSM483378" "GSM483385" "GSM483402"
"GSM483403"
## [121] "GSM483406" "GSM483407" "GSM483408" "GSM483414" "GSM483415"
"GSM483424"
## [127] "GSM483437" "GSM483439" "GSM483440" "GSM483446" "GSM483449"
"GSM483454"
## [133] "GSM483456" "GSM483460" "GSM483463" "GSM483471" "GSM483297"
"GSM483298"
## [139] "GSM483299" "GSM483306" "GSM483309" "GSM483314" "GSM483316"
"GSM483321"
## [145] "GSM483324" "GSM483326" "GSM483335" "GSM483340" "GSM483342"
"GSM483345"
## [151] "GSM483346" "GSM483355" "GSM483357" "GSM483359" "GSM483360"
"GSM483382"
## [157] "GSM483383" "GSM483387" "GSM483392" "GSM483393" "GSM483394"
"GSM483395"
## [163] "GSM483397" "GSM483398" "GSM483409" "GSM483422" "GSM483423"
"GSM483425"
## [169] "GSM483429" "GSM483430" "GSM483432" "GSM483435" "GSM483438"
"GSM483441"
## [175] "GSM483451" "GSM483452" "GSM483453" "GSM483457" "GSM483467"
"GSM483469"
## [181] "GSM483472" "GSM483474" "GSM483475" "GSM483479" "GSM483480"
"GSM483481"
## [187] "GSM483482" "GSM483483" "GSM483484" "GSM483485" "GSM483486"
"GSM483487"
## [193] "GSM483488" "GSM483489" "0" "0"
```

```
preListGDS = preListGDS[-index]
length(preListGDS)

## [1] 1689
```

GDS rows are deleted from each element of preListGDS as below.

```
for(i in 1:length(preListGDS)){
  preListGDS[[i]] = preListGDS[[i]][!grepl("^GDS",preListGDS[[i]],[1]),-1]
}
```



### 3.3.Editing preListGDS object into ListGDS

Datasets in **preListGDS** are required to be edited; so, a “for loop” along with 4 conditions are created to edit preListGDS. edited preListGDS is called **ListGDS**. In the gene symbol column of most of GDSes, there are multiple gene symbols for one value. In that case, the gene symbol which is in the edgelist is selected.

```
ListGDS = preListGDS

# The first condition determines if there is a need for editing

# The second condition is for when there are multiple gene symbols for one
value and
# more than one of them is mapped to the edgelist genes.

# The third condition is for when there are multiple gene symbols for one
value,
# but none of them mapped to the gene set "Gene"

# The fourth condition is for when there are multiple gene symbols for one
value and
# only one of them is mapped to the gene set "Gene"

for(i in 1:length(ListGDS)){

  l1 = length(ListGDS[[i]][1,]) - 1

  a = ListGDS[[i]][,l1]

  a = strsplit(a,"///",fixed = T)

  l = unlist(lapply(a,length))

  idxx = which(l>1)

  if(length(idxx>0)){
    a1 = a[idxx]
    for(j in 1:length(a1)){
      idxx1 = which(a1[[j]] %in% edgelistGenes)
      if(length(idxx1>1)){
        ListGDS[[i]][,l1][idxx[j]] = a[idxx][[j]][idxx1[1]]
      } else if(length(idxx1)==0){
        ListGDS[[i]][,l1][idxx[j]] = a[idxx][[j]][1]
      } else {
        ListGDS[[i]][,l1][idxx[j]] = a[idxx][[j]][idxx1]
      }
    }
  }
}
```

```

    }
}

# Deleting regular [A] characters next to the some values
# changing class of values to numeric
# changing order of columns

ListGDS[[i]] = cbind(ListGDS[[i]][,l1,drop=F],ListGDS[[i]][,1:(l1-
1),drop=F])
ListGDS[[i]] = as.data.frame(ListGDS[[i]],stringsAsFactors=F)

for(j in 2:l1){
  ListGDS[[i]][,j] = sub("\\[A]", "", ListGDS[[i]][,j])
  ListGDS[[i]][,j] = as.numeric(ListGDS[[i]][,j])
}

#print(i)
}

```

```
head(ListGDS[[1]] , 20)
```

```
##      Gene symbol  GSM575  GSM576  GSM577  GSM578  GSM579
## 4          CYCS  4419.0  4528.3  4732.6  2399.3  1757.6
## 5          FGF2  2204.9  2417.0  2456.7  5088.2  4615.3
## 41         GAS6   765.9   562.4   667.5   364.8   395.6
## 51         GRB2   226.2   212.6   198.2    10.3    34.3
## 42        IL2RB   367.8   508.4   328.2    57.9    34.3
## 52          F3   573.5   607.5   356.9    79.9    21.6
## 6        CASP10  1765.1  1400.3  1843.8   660.6   479.1
## 7         PTPRM   676.4   622.7   611.0   309.4   298.9
## 43        IQGAP1   508.0   482.1   480.6   323.9   328.9
## 44        SREBF1  8303.8  9411.2  8260.5  3804.7  4592.9
## 53         FGD1  2637.9  2035.4  1447.3  5435.8  4638.0
## 61         RHO   6781.8  6509.0  7001.6 10833.1 11141.8
## 45         THRA 21144.2 45869.4 52175.2  9517.2  6752.6
## 46        PLA2R1   721.7   375.6   462.5  5271.3  6598.9
## 47        RAB7A  4117.5  4042.4  2926.3 10662.4 11609.5
## 48        SDC2   1846.2  1794.0  2030.2   656.4   776.9
## 54          C5   167.4    41.9    33.5   326.9   295.8
## 49        PPP2R5A 6597.4  7377.2  8328.5  3951.5  3474.7
## 410        IRF9   445.3   315.8   283.7  2369.0  3024.7
## 411       SERPINB10 570.1   478.5   465.0   123.3   109.5

```

```
head(preListGDS[[1]] , 20)
```

```
##          GSM575          GSM576          GSM577
GSM578
## 4          4419          4528.3 4732.6000000000004
2399.3000000000002
## 5          2204.9          2417 2456.6999999999998

```

|                     |                    |                     |                    |
|---------------------|--------------------|---------------------|--------------------|
| 5088.2              |                    |                     |                    |
| ## 41               | 765.9              | 562.4               | 667.5              |
| 364.8               |                    |                     |                    |
| ## 51               | 226.2[A]           | 212.6[A]            | 198.2[A]           |
| 10.3[A]             |                    |                     |                    |
| ## 42               | 367.8              | 508.4               | 328.2              |
| 57.9                |                    |                     |                    |
| ## 52               | 573.5              | 607.5               | 356.9              |
| 79.9[A]             |                    |                     |                    |
| ## 6                | 1765.1             | 1400.3              | 1843.8             |
| 660.6[A]            |                    |                     |                    |
| ## 7                | 676.4              | 622.70000000000005  | 611                |
| 309.4[A]            |                    |                     |                    |
| ## 43               | 508[A]             | 482.1[A]            | 480.6[A]           |
| 323.9[A]            |                    |                     |                    |
| ## 44               | 8303.7999999999993 | 9411.20000000000007 | 8260.5             |
| 3804.7              |                    |                     |                    |
| ## 53               | 2637.9             | 2035.4              | 1447.3             |
| 5435.8              |                    |                     |                    |
| ## 61               | 6781.8             | 6509                | 7001.6             |
| 10833.1             |                    |                     |                    |
| ## 45               | 21144.2            | 45869.4             | 52175.199999999997 |
| 9517.20000000000007 |                    |                     |                    |
| ## 46               | 721.7              | 375.6               | 462.5              |
| 5271.3              |                    |                     |                    |
| ## 47               | 4117.5             | 4042.4              | 2926.3             |
| 10662.4             |                    |                     |                    |
| ## 48               | 1846.2             | 1794                | 2030.2             |
| 656.4               |                    |                     |                    |
| ## 54               | 167.4[A]           | 41.9[A]             | 33.5[A]            |
| 326.89999999999998  |                    |                     |                    |
| ## 49               | 6597.4             | 7377.2              | 8328.5             |
| 3951.5              |                    |                     |                    |
| ## 410              | 445.3              | 315.8               | 283.7              |
| 2369                |                    |                     |                    |
| ## 411              | 570.1              | 478.5               | 465                |
| 123.3[A]            |                    |                     |                    |
| ##                  | GSM579             | Gene symbol         | Gene ID            |
| ## 4                | 1757.6             | CYCS                | 54205              |
| ## 5                | 4615.3             | FGF2                | 2247               |
| ## 41               | 395.6              | GAS6                | 2621               |
| ## 51               | 34.3[A]            | GRB2                | 2885               |
| ## 42               | 34.3[A]            | IL2RB               | 3560               |
| ## 52               | 21.6               | F3                  | 2152               |
| ## 6                | 479.1[A]           | CASP10              | 843                |
| ## 7                | 298.89999999999998 | PTPRM               | 5797               |
| ## 43               | 328.9              | IQGAP1              | 8826               |
| ## 44               | 4592.8999999999996 | SREBF1              | 6720               |
| ## 53               | 4638               | FGD1                | 2245               |
| ## 61               | 11141.8            | RHO                 | 6010               |

```
## 45          6752.6      THRA      7067
## 46          6598.9      PLA2R1    22925
## 47          11609.5     RAB7A     7879
## 48           776.9      SDC2     6383
## 54          295.8[A]      C5       727
## 49          3474.7     PPP2R5A    5525
## 410         3024.7      IRF9     10379
## 411         109.5     SERPINB10    5273

class(ListGDS[[1]][,4])

## [1] "numeric"

# We keep gene symbols and delete columns related to gene Ids

dim(ListGDS[[1]])

## [1] 49  6

dim(preListGDS[[1]])

## [1] 49  7
```

The following code, shows the indices in GDSes containing repetitious GSMs.

```
indexx = list()
for(i in 1:length(ListGDS)){

  logic=c()

  for(j in (1:length(ListGDS))[-i]){
    logic[j]=any(colnames(ListGDS[[i]])[-1] %in% colnames(ListGDS[[j]]))
  }
  index = which(logic)
  if(length(index) > 0){
    indexx[i] = index
  }
  print(i)
}

index.of.GDSes.having.repetitious.GSMs = which(unlist(lapply(indexx ,
length)) > 0)
```

In the code below, repetitious GSMs are deleted in different GDSes.

```
for(i in 1:length(ListGDS)){

  logic=c()
```

```

for(j in (1:length(ListGDS))[-i]){
  logic[j]=any(colnames(ListGDS[[i]])[-1] %in% colnames(ListGDS[[j]]))
}
index = which(logic)
if(length(index) > 0){
  for(z in 1:length(index)){
    logicc = colnames(ListGDS[[index[z]]]) %in% colnames(ListGDS[[i]])[-1]
    indexx = which(logicc)
    ListGDS[[index[z]]]=ListGDS[[index[z]][, -indexx]
  }
}
# print(i)
}

```

Indices in ListGDS for which there is no GSM left are omitted.

```

index = which(unlist(lapply(lapply(lapply(ListGDS , dim) , function(x) x[2])
, function(x) length(x) == 0)))
ListGDS = ListGDS[-index]

# ALL the datasets have more than two GSMs
which(unlist(lapply(lapply(lapply(ListGDS , dim) , function(x) x[2]),
function(x) x < 2)))
## integer(0)

```

Indices in ListGDS for which there is no gene left are removed.

```

index = which(unlist(lapply(lapply(lapply(ListGDS , dim) , function(x) x[1])
, function(x) x < 2)))
ListGDS = ListGDS[-index]

```

All the gene names that are in the ListGDS object, are stored in an object called **profilesGenes**.

```

profilesGenes = c()

for(i in 1:length(ListGDS)){
  profilesGenes = c(profilesGenes,ListGDS[[i]][,1])
}

profilesGenes = unique(profilesGenes)

any(is.na(profilesGenes))

profilesGenes = profilesGenes[-which(is.na(profilesGenes))]

```

```
length(profilesGenes)
```

```
## [1] 3207
```

The first row in the KEGG edgelist contains “EGF” gene. Check how many datasets contain this gene.

```
which(profilesGenes == "EGF")
```

```
## [1] 1679
```

```
logic=c()  
for(i in 1:length(ListGDS)){  
  logic[i] = profilesGenes[1679] %in% ListGDS[[i]][,1]  
}  
idxng1679 = which(logic)  
length(which(logic))
```

```
## [1] 44
```

```
head(ListGDS[[idxng1679[20]]],10)
```

| ##    | Gene symbol | GSM1200171 | GSM1200172 | GSM1200173 | GSM1200174 |
|-------|-------------|------------|------------|------------|------------|
| ## 4  | PRKACB      | 3762.2700  | 3588.7100  | 5354.8300  | 5143.6300  |
| ## 5  | FAS         | 204.5730   | 195.6560   | 507.2090   | 473.8230   |
| ## 6  | FAS         | 178.3230   | 169.3150   | 401.8230   | 366.3830   |
| ## 7  | EGF         | 72.6697    | 66.5497    | 22.1397    | 22.2993    |
| ## 8  | CYCS        | 8361.6400  | 8924.8400  | 5821.4400  | 6205.0000  |
| ## 9  | HGF         | 36.7687    | 40.6940    | 85.0087    | 88.0442    |
| ## 10 | HGF         | 51.4297    | 47.0476    | 83.9657    | 83.3615    |
| ## 11 | FAS         | 107.4120   | 103.9990   | 317.9370   | 290.5730   |
| ## 12 | FAS         | 60.8529    | 55.7002    | 161.3550   | 140.7340   |
| ## 13 | CYCS        | 40.3911    | 42.7323    | 20.5259    | 17.2582    |

```
length(profilesGenes) # 3207 is the number of genes extracted from all the  
gene profiles
```

```
## [1] 3207
```

```
length(edgelistGenes) # 3187 is the number of genes at the edgelist
```

```
## [1] 3187
```

common genes between gene profiles and edgelist are stored in **Genes** object.

```
sum(profilesGenes %in% edgelistGenes)
```

```
Genes = profilesGenes[which(profilesGenes %in% edgelistGenes)]
```

```
length(Genes)
## [1] 3047
# There are 3047 common genes between the gene profiles and the edgelist
```

In the following code all the GSM names are stored at **gsms** object.

```
list3=list()
for(i in 1:(length(ListGDS))){
  index = grep("^GSM" , colnames(ListGDS[[i]]))
  list3[[i]] = colnames(ListGDS[[i]])[index]
}
length(list3)

gsm = unlist(list3)

gsms = unique(gsm)

length(gsms)
## [1] 40903
```

### 3.4. Constructing an expression matrix called Exprtable which contains all the gene values amongst the GSMs

First of all, an NA matrix called **Exprtable** is created which has genes in row and GSMs in column. for each gene name, a sublist is created by a loop. For gene one, we check which GDSes of ListGDS object contain it. Then, we store these GDSes at a **sublist** object. After that, for each element in the sublist, the rows indicating the gene i are stored at **idx** object. If there is just one row, we put that row in Exprtable object. Otherwise, the row with larger IQR is located in Exprtable expression matrix.

```
Exprtable = matrix(NA,length(Genes),length(gsms))

Exprtable = as.data.frame(Exprtable)

rownames(Exprtable) = Genes

colnames(Exprtable) = gsms

dim(Exprtable)
## [1] 3047 40903
```

```

for(i in 1:length(Genes)){

  logic = c()
  for(j in 1:length(ListGDS)){
    logic[j] = Genes[i] %in% ListGDS[[j]][,1]
  }
  sublist = ListGDS[logic]

  for(j in 1:length(sublist)){

    gsm = colnames(sublist[[j]])[-1]

    idxx = which(sublist[[j]][,1] == Genes[i])

    if(length(idxx) == 1){
      Exprtable[Genes[i],gsm] = sublist[[j]][idxx,-1]
    } else {
      d = sublist[[j]][idxx,-1]
      IQRs = c()
      for(z in 1:length(d[,1])){
        a=as.numeric(na.omit(as.numeric(d[z,])))
        IQRs[z] = quantile(a, 0.75 ) - quantile(a, 0.25)
        Exprtable[Genes[i],gsm] = d[which.max(IQRs),]
      }
    }
  }
  #print(i)
}

sum(is.na(Exprtable))
## [1] 120876590

sum(!(is.na(Exprtable)))
## [1] 3754851

```

### 3.5.designing a list called preSignalingNet1 which contains expression values for each edge

In this step, both **Exprtable** and **edgelist** are used. For each edge in the edgelist, a data frame is created. First of all, an empty list called preSignalongNet is created. source gene of row i of the edgelist is stored at gene1 object. Target gene of the same row is stored at gene2 object. The source and target gene rows in Exprtable are stored at a and b objects



respectively. If there exist more than two non-NA GSMs in both source and target genes, these values are saved as a dataframe in element *i* of `preSignalingNet1` object. If not, we put "empty" in `preSignalingNet1[i]`.

```
preSignalingNet1 = list()
for(i in 1:length(edgelist[,1])){
  gene1 = edgelist[,2:3][i,1]
  gene2 = edgelist[,2:3][i,2]

  idx1 = which(gene1 == rownames(Exprtable))
  idx2 = which(gene2 == rownames(Exprtable))
  a = Exprtable[idx1,]
  b = Exprtable[idx2,]
  l1 = !is.na(a)
  l2 = !is.na(b)

  if(length(intersect(which(l1),which(l2))) > 0){
    d = rbind(a,b)
    idx = intersect(which(l1),which(l2))
    preSignalingNet1[[i]] = d[,idx]
    names(preSignalingNet1)[i] = edgelist[i,1]
  } else {
    preSignalingNet1[[i]] = "empty"
    names(preSignalingNet1)[i] = edgelist[i,1]
  }
  #print(i)
}
```

The information for the edge 10000 is depicted as follow. THBS4 is the source gene and CD47 is the target gene.

```
edgelist[10000,]

##           ID Gene1 Gene2 Interaction type
## 10000 E10000 THBS4  CD47          activation

preSignalingNet1[[10000]]

##           GSM183695 GSM185526 GSM185527 GSM185528 GSM185529 GSM185530
GSM185531
## THBS4      86.5063    80.9516   13.80250    285.017    310.7550    39.8476
521.398
## CD47       20.8250    33.1526    3.27527     11.985     32.0414    24.6110
3.422
##           GSM185532 GSM185533 GSM185534 GSM185535 GSM185536 GSM47867 GSM47868
## THBS4 254.00900 686.64900 525.22700 404.1640 733.72700 33.9 36.0
## CD47 7.69714 4.67268 7.08067 29.6316 9.12542 90.9 26.7
##           GSM47700 GSM47862 GSM47869 GSM47863 GSM47864 GSM47870 GSM47871
GSM47865
```

```
## THBS4      23.2      31.1      22.2      6.3      6.1      91.4      58.3
61.7
## CD47       38.5      50.4      74.3      75.0      44.1      4.8      22.8
4.3
##          GSM47866 GSM112271 GSM112272 GSM112273 GSM112274 GSM112275 GSM112276
## THBS4      56.3     -0.7179  -0.1888  -0.7167  -0.6063  -0.9728  -0.8966
## CD47       1.7      0.4725  0.3915  0.0521  0.1217  -0.5224  -0.4888
##          GSM112277 GSM112278 GSM112279 GSM112280 GSM112281 GSM112282
GSM112283
## THBS4     -1.1997  -2.0078  -0.9423  -0.6307  -1.0251  -1.0449  -
1.2765
## CD47      -0.8899  -0.9997  0.0867  0.1363  -0.5541  -0.5307  -
1.0612
##          GSM112284 GSM112285 GSM112286 GSM112287 GSM112288 GSM112289
GSM112290
## THBS4     -1.3115  0.4004  0.2038  0.1565  0.3431  -0.5992  -
0.6471
## CD47      -1.1395  0.6933  0.6035  0.2097  0.2109  0.0369
0.0715
##          GSM112293 GSM112294 GSM112295 GSM112296 GSM112297 GSM112298
## THBS4      0.1637  0.1986  -0.8115  -0.6926  -0.8596  -0.8864
## CD47      0.4232  0.4921  -0.0582  -0.2459  -0.5638  -0.7999
```

The Object **idx.NO.gsms** has the indices of the edges having less than three GSMS.

```
idx.NO.gsms = which(preSignalingNet1 %in% "empty" )
length(idx.NO.gsms)
## [1] 10650
```

### 3.7.Creating preSignalingNet2

All the values in each preSignalingNet1 element are separated based on the source of datasets and they are stored at **preSignalingNet2** object. Furthermore, for some indices of preSignalingNet1, source and target genes are the same and they have exactly the same values. These indices are stored at **index.of.not.matching.preSignalingNet.and.ListGDS** object and they are assigned empty in preSignalingNet2 object. **‘These indices are related to loops in the edgelist.’**

```
preSignalingNet2 = preSignalingNet1
index.of.not.matching.preSignalingNet.and.ListGDS=c()
for(i in (1:length(preSignalingNet1))[-idx.NO.gsms]){
```

```

# Here the index object get indices of all the GDSes which have created
signalingnet[[i]] and have the source and target genes
logic=c()
for(j in 1:length(ListGDS)){
  logic[j]=any(colnames(preSignalingNet1[[i]]) %in% colnames(ListGDS[[j]]))
}
& all(rownames(preSignalingNet1[[i]]) %in% ListGDS[[j]][,1])
}
index = which(logic)

if(length(index) > 1){
  coexpr = ListGDS[index]
  coExpr = list()
  colNAMES = lapply(coexpr,colnames)
  for(z in 1:length(coexpr)){
    coExpr[[z]]=preSignalingNet1[[i]][ , colNAMES[[z]][which( colNAMES[[z]]
%in% colnames(preSignalingNet1[[i]]))]]
  }
  preSignalingNet2[[i]] = coExpr
} else if(length(index) == 1){
  preSignalingNet2[[i]] = preSignalingNet1[[i]]
} else {
  preSignalingNet2[[i]] = "empty"
  index.of.not.matching.preSignalingNet.and.ListGDS[i] = i
}
}

index.of.not.matching.preSignalingNet.and.ListGDS =
which(!is.na(unlist(index.of.not.matching.preSignalingNet.and.ListGDS)))

```

```

index.of.not.matching.preSignalingNet.and.ListGDS

## [1] 213 5245 5258 5271 5284 5297 5310 5323 5336 5349 5362
5375
## [13] 5388 8711 8715 8719 10057 10060 10077 10080 10081 10082 10083
10084
## [25] 10134 16235 17298 18206 18445 18447 18574 21348 21352 21356 25757

edgelist[index.of.not.matching.preSignalingNet.and.ListGDS[1:4],]

## ID Gene1 Gene2 Interaction type
## 213 E00213 HBEGF HBEGF activation
## 5245 E05245 ANAPC10 ANAPC10 activation
## 5258 E05258 CDC26 CDC26 activation
## 5271 E05271 ANAPC13 ANAPC13 activation

```

```
preSignalingNet1[[index.of.not.matching.preSignalingNet.and.ListGDS[2]]][,1:6]
##          GSM1305903 GSM1305905 GSM1305907 GSM1305909 GSM1305902 GSM1305904
## ANAPC10         49.98      52.17      56.98      54.39      33.34      27.26
## ANAPC101        49.98      52.17      56.98      54.39      33.34      27.26
```

```
Eligibles = (1:length(preSignalingNet2))[-
c(index.of.not.matching.preSignalingNet.and.ListGDS,idx.NO.gsms)]
length(Eligibles)
# There are 15805 edges for down stream analysis
```

Correlation analysis was performed on each edge (gene pair) after preprocessing step. Samples having value for the gene pairs, may come from different datasets and they should be separated and analyzed independently. Figure 6 represents the effect of this preprocessing on a gene pairs in the dataset. In panel A, because all the values are related to multiple datasets in preSignalingNet1[[4]], we can see a highly data dispersion. However, in the preSignalingNe2[[4]] all of these values are separated into 14 datasets. in panel B, six of these separated datasets are plotted.

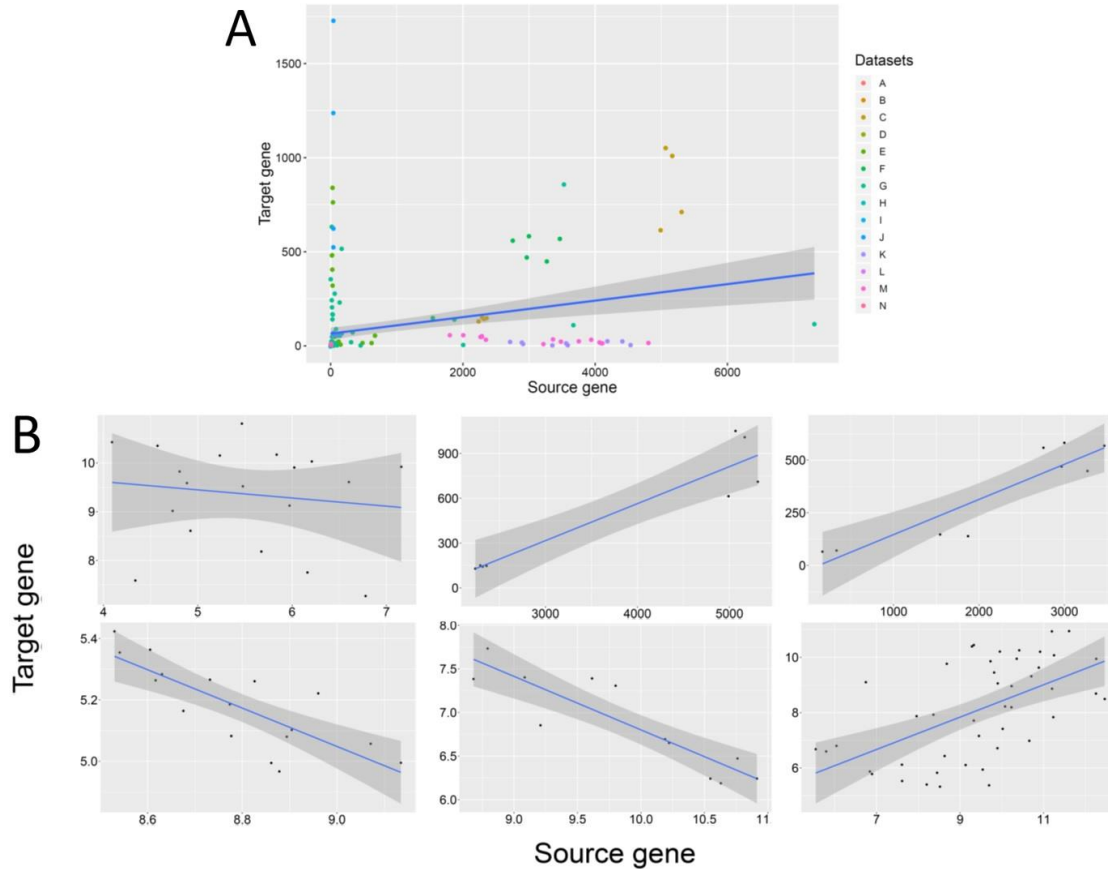


Figure 6

```
## null device
##          1
```

### Getting rid of outliers

Values 1.5 times more than IQR are considered as outliers and are taken out.

```
for(i in Eligibles){
  if(class(preSignalingNet2[[i]])=="list"){
    for(j in 1:length(preSignalingNet2[[i]]))
      outliers1 = boxplot.stats(as.numeric(preSignalingNet2[[i]][[j]][1]))$out
      outliers2 = boxplot.stats(as.numeric(preSignalingNet2[[i]][[j]][2]))$out

    index1 = which(as.numeric(preSignalingNet2[[i]][[j]][2,]) %in% outliers1)
    index2 = which(as.numeric(preSignalingNet2[[i]][[j]][2,]) %in% outliers2)
```

```

index = unique(c(index1,index2))
if(length(index) > 0){
preSignalingNet2[[i]][[j]] = preSignalingNet2[[i]][[j]][,-index]
}

} else {
outliers1 = boxplot.stats(as.numeric(preSignalingNet2[[i]][1,]))$out
outliers2 = boxplot.stats(as.numeric(preSignalingNet2[[i]][2,]))$out

index1 = which(as.numeric(preSignalingNet2[[i]][1,]) %in% outliers1)
index2 = which(as.numeric(preSignalingNet2[[i]][2,]) %in% outliers2)
index = unique(c(index1,index2))
if(length(index) > 0){
  preSignalingNet2[[i]] = preSignalingNet2[[i]][,-index]}
}
}

```

The following code proves that all the elements in preSignalingNet2 which is in the class dataframe, have more than two samples. The code must return a TRUE value.

```

CLASS = lapply(preSignalingNet2 , class)
all(lapply(lapply(preSignalingNet2[which(CLASS == "data.frame")] , dim),
function(x) x[2]) > 2)
## [1] FALSE

```

The following code presents the number of datasets which have two or fewer datasets.

```

sum(unlist(lapply(preSignalingNet2[which(CLASS == "list")] , function(x)
lapply(x,function(x) dim(x)[2] <= 2))))
## [1] 43

```

The following code shows the indices in preSignalingNet2 in the class list having datasets with less than three samples.

```

index = which(unlist(lapply(preSignalingNet2[which(CLASS == "list")] ,
function(x) any(unlist(lapply(x,function(x) dim(x)[2] < 3))))))
index
## E00598 E00868 E00877 E01073 E01081 E01430 E01439 E04464 E04490 E06633
E06640
##      393      550      556      683      689      914      921      2509      2527      3398
3405
## E06648 E06655 E06657 E06665 E06672 E06858 E06865 E06873 E06880 E06882

```

```

E06890
## 3413 3420 3422 3430 3436 3569 3576 3583 3589 3590
3596
## E06897 E07488 E07495 E07503 E07510 E07512 E07520 E07527 E09571 E09777
E09842
## 3602 4081 4087 4091 4096 4098 4103 4109 4999 5144
5193
## E13198 E16423 E16428 E17413 E17743 E18042 E18711 E25851 E25910 E25914
## 6901 7581 7586 8053 8216 8389 8787 11007 11050 11054

```

```

preSignalingNet2[which(CLASS == "list")][[index[1]]]

## [[1]]
##      GSM4312 GSM4317
## TGFB3 296.045 109.721
## TGFB2 24.545 136.243
##
## [[2]]
##      GSM7621 GSM7622 GSM7623 GSM7624 GSM6681 GSM6682 GSM6683 GSM6684
## TGFB3 848.063 827.429 3403.71 2631.38 739.542 606.972 3023.23 2892.95
## TGFB2 5545.760 5872.560 8368.38 7563.18 5431.500 5991.290 7298.31 6956.94
##      GSM6685 GSM6686 GSM6687 GSM6688
## TGFB3 715.008 824.526 3184.55 2850.80
## TGFB2 4540.660 4318.670 7407.48 6698.04

```

Ineligible datasets are omitted from the preSignalingNet2 through the following code. Ineligible datasets are those which contain less than three samples.

```

for(i in 1:length(index)){
  idx = which(unlist(lapply(lapply(preSignalingNet2[which(CLASS ==
"list")][index][[i]] , dim) , function(x) x[2]<= 2)))
  preSignalingNet2[which(CLASS == "list")][index][[i]] =
preSignalingNet2[which(CLASS == "list")][index][[i]][-idx]
}

```

the following code is to check whether there is any element of preSignalingNet2 in class list with the length of one.

```

index = which(lapply(preSignalingNet2 , class) == "list" &
lapply(preSignalingNet2 , length) == 1)
index

## E00598
##      598

preSignalingNet2[[index]] = as.data.frame(preSignalingNet2[[index]])

```

Because some changes may be imposed on some elements, another checking is done to make sure all the datasets have more than two GSMs.

```
CLASS = lapply(preSignalingNet2 , class)
all(lapply(lapply(preSignalingNet2[which(CLASS == "data.frame")] , dim),
function(x) x[2]) > 2)

## [1] FALSE

all(unlist(lapply(preSignalingNet2[which(CLASS == "list")] , function(x)
any(unlist(lapply(x,function(x) dim(x)[2] >= 3))))))

## [1] TRUE
```

The elements which contain less than three GSMs are assigned empty.

```
index = which(lapply(lapply(preSignalingNet2[which(CLASS == "data.frame")] ,
dim), function(x) x[2]) < 3)
index

## E12586
## 2460

if(length(index)>0){
preSignalingNet2[which(CLASS == "data.frame")][[index]]
preSignalingNet2[which(CLASS == "data.frame")][[index]] = "empty"
}
```

Eligible edges are updated.

```
Eligibles = which(lapply(preSignalingNet2 , class) == "list" |
lapply(preSignalingNet2 , class) == "data.frame")
length(Eligibles)

## [1] 15804
```



## 4. Correlation analysis for each edge

### 4.1. Creating a large list called SignalingNet which contains expression values and correlation information for each edge

The next step is to compute the correlation between the two interacting genes. Pearson, Spearman and Kendall correlation tests are calculated for each edge and a list called **SignalingNet** is created.

```
SignalingNet = preSignalingNet2

for(i in Eligibles){

  if(class(SignalingNet[[i]])=="list"){
    corAnalysis = list()
    dim = list()
    for(j in 1:length(SignalingNet[[i]])){

      p = cor.test(as.numeric(SignalingNet[[i]][[j]][1,]) ,
as.numeric(SignalingNet[[i]][[j]][2,]) , method = "pearson")
      s = cor.test(as.numeric(SignalingNet[[i]][[j]][1,]) ,
as.numeric(SignalingNet[[i]][[j]][2,]) , method = "spearman")
      k = cor.test(as.numeric(SignalingNet[[i]][[j]][1,]) ,
as.numeric(SignalingNet[[i]][[j]][2,]) , method = "kendall")
      pearson = as.data.frame(cbind(pearsoncor = unname(p$estimate) , pearsonpval
= p$p.value))
      spearman = as.data.frame(cbind(spearmancor = unname(s$estimate) ,
spearmanpval = s$p.value))
      kendall = as.data.frame(cbind(kendallcor = unname(k$estimate) , kendallpval
= k$p.value))
      corAnalysis[[j]] = list(pearson = pearson,spearman = spearman,kendall =
kendall)
      dim[[j]] = dim(SignalingNet[[i]][[j]])
    }

    SignalingNet[[i]] = list(Source = unname(edgeList[i,2]) , Target =
unname(edgeList[i,3]) , length = length(SignalingNet[[i]]) ,dim = dim ,
coExpr = SignalingNet[[i]] ,
corAnalysis = corAnalysis , Interactiontype =
unname(edgeList[i,4]) , Coherency = "empty")

  } else {
    p = cor.test(as.numeric(SignalingNet[[i]][1,]) ,
as.numeric(SignalingNet[[i]][2,]) , method = "pearson")
    s = cor.test(as.numeric(SignalingNet[[i]][1,]) ,
as.numeric(SignalingNet[[i]][2,]) , method = "spearman")
    k = cor.test(as.numeric(SignalingNet[[i]][1,]) ,
as.numeric(SignalingNet[[i]][2,]) , method = "kendall")
    pearson = as.data.frame(cbind(pearsoncor = unname(p$estimate) ,
```

```

pearsonpval = p$p.value))
  spearman = as.data.frame(cbind(spearmancor = unname(s$estimate) ,
spearmanpval = s$p.value))
  kendall = as.data.frame(cbind(kendallcor = unname(k$estimate) ,
kendallpval = k$p.value))

  SignalingNet[[i]] = list(Source = unname(edgelist[i,2]) , Target =
unname(edgelist[i,3]) , length = 1 ,dim = dim(SignalingNet[[i]]) , coExpr =
SignalingNet[[i]] ,
                        corAnalysis = list(pearson = pearson,spearman
= spearman,kendall = kendall) ,
                        Interactiontype = unname(edgelist[i,4]) ,
Coherency = "empty")
}

}

for(i in (1:length(SignalingNet))[-Eligibles]){
  SignalingNet[[i]] = list(Source = unname(edgelist[i,2]) , Target =
unname(edgelist[i,3]) , length = 0 ,dim = 0 , coExpr = NA , corAnalysis = NA
, Interactiontype = unname(edgelist[i,4]) , Coherency = "empty")
}

```

### An example of eligible edge:

```
SignalingNet[[70]]
```

```

## $Source
## [1] "PDGFRB"
##
## $Target
## [1] "PLCG2"
##
## $length
## [1] 4
##
## $dim
## $dim[[1]]
## [1] 2 8
##
## $dim[[2]]
## [1] 2 8
##
## $dim[[3]]
## [1] 2 10
##

```

```

## $dim[[4]]
## [1] 2 30
##
##
## $coExpr
## $coExpr[[1]]
##      GSM490979 GSM490980 GSM490981 GSM490982 GSM490983 GSM490984
GSM490985
## PDGFRB      8.13064      8.35757      7.80769      8.03344      9.01956      8.54397
9.12372
## PLCG2       6.11117      6.29475      6.50859      6.23847      5.81704      5.82973
5.92684
##      GSM490986
## PDGFRB      9.15383
## PLCG2       6.08239
##
## $coExpr[[2]]
##      GSM244647 GSM244649 GSM244651 GSM244653 GSM244648 GSM244650
GSM244652
## PDGFRB      36.1238      39.6828      6.70644      13.8024      21.5159      34.3139
11.9610
## PLCG2      360.3680      373.0510      85.62970      68.4027      428.7340      424.1310
59.5258
##      GSM244654
## PDGFRB      12.7781
## PLCG2      104.2860
##
## $coExpr[[3]]
##      GSM102789 GSM102785 GSM102787 GSM102790 GSM102786 GSM102788
GSM102681
## PDGFRB      176.298      124.255      107.709      352.027      239.746      95.4943
1298.46
## PLCG2       565.279      259.747      554.512      1922.380      365.583      2091.8100
1613.89
##      GSM102783 GSM102782 GSM102784
## PDGFRB       780.864      951.725      846.224
## PLCG2      1324.060      1713.440      1369.790
##
## $coExpr[[4]]
##      GSM1143676 GSM1143677 GSM1143678 GSM1143679 GSM1143680 GSM1143681
## PDGFRB       7.31532      7.23638      7.54621      7.48674      7.67071      7.35978
## PLCG2       7.30048      7.22943      7.15666      6.98803      7.30929      6.96447
##      GSM1143682 GSM1143683 GSM1143684 GSM1143685 GSM1143686 GSM1143687
## PDGFRB       7.54044      7.39747      7.56700      7.40331      7.44638      7.82191
## PLCG2       7.11817      7.32129      7.08797      6.95786      7.06090      7.56741
##      GSM1143688 GSM1143689 GSM1143690 GSM1143691 GSM1143692 GSM1143693
## PDGFRB       7.54010      7.59168      7.78653      7.47773      7.25271      7.41038
## PLCG2       7.36197      7.49527      7.60299      7.37531      7.32986      7.18851
##      GSM1143694 GSM1143695 GSM1143696 GSM1143697 GSM1143698 GSM1143699
## PDGFRB       7.41699      7.27599      7.48022      7.45763      7.52638      7.78671

```

```

## PLCG2      7.19516      7.25258      7.05503      7.03926      7.35168      7.10651
##          GSM1143700 GSM1143701 GSM1143702 GSM1143703 GSM1143704 GSM1143705
## PDGFRB      7.52223      7.06359      7.76968      7.32825      7.41585      7.43055
## PLCG2      7.10267      7.24621      7.28328      7.25294      7.31378      7.46169
##
##
## $corAnalysis
## $corAnalysis[[1]]
## $corAnalysis[[1]]$pearson
##   pearsoncor pearsonpval adjusted.pearsonpval
## 1 -0.7402235  0.03573155          0.0790027
##
## $corAnalysis[[1]]$spearman
##   spearmancor spearmanpval
## 1 -0.7142857  0.05758929
##
## $corAnalysis[[1]]$kendall
##   kendallcor kendallpval
## 1      -0.5    0.1086806
##
##
## $corAnalysis[[2]]
## $corAnalysis[[2]]$pearson
##   pearsoncor pearsonpval adjusted.pearsonpval
## 1  0.8483993  0.007750156          0.02304932
##
## $corAnalysis[[2]]$spearman
##   spearmancor spearmanpval
## 1  0.6666667  0.08308532
##
## $corAnalysis[[2]]$kendall
##   kendallcor kendallpval
## 1  0.4285714    0.178869
##
##
## $corAnalysis[[3]]
## $corAnalysis[[3]]$pearson
##   pearsoncor pearsonpval adjusted.pearsonpval
## 1  0.4567883  0.1844641          0.2898644
##
## $corAnalysis[[3]]$spearman
##   spearmancor spearmanpval
## 1  0.2606061  0.4696753
##
## $corAnalysis[[3]]$kendall
##   kendallcor kendallpval
## 1  0.2444444    0.3807198
##
##
## $corAnalysis[[4]]

```

```
## $corAnalysis[[4]]$pearson
##   pearsoncor pearsonpval adjusted.pearsonpval
## 1  0.2661583   0.1551258             0.2530683
##
## $corAnalysis[[4]]$spearman
##   spearmancor spearmanpval
## 1   0.1764182   0.3495161
##
## $corAnalysis[[4]]$kendall
##   kendallcor kendallpval
## 1  0.1310345   0.3206742
##
##
##
## $Interactiontype
## [1] "activation"
##
## $Coherency
## [1] "empty"
```

#### An example of ineligible edge:

```
SignalingNet[[43]]
```

```
## $Source
## [1] "IGF1R"
##
## $Target
## [1] "PLCG1"
##
## $length
## [1] 0
##
## $dim
## [1] 0
##
## $coExpr
## [1] NA
##
## $corAnalysis
## [1] NA
##
## $Interactiontype
## [1] "activation"
##
## $Coherency
## [1] "empty"
```

Using the following code, all the Pearson, Spearman and Kendall correlation coefficients and p-values are saved in the following objects.

```
pearsonpvalues = c()
spearmanpvalues = c()
kendallpvalues = c()
pearsoncors = c()
spearmancors = c()
kendallcors = c()

for(i in Eligibles){
  if(SignalingNet[[i]]$length > 1){
    for(j in 1:length(SignalingNet[[i]]$corAnalysis)){

      a = SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsonpval
      pearsonpvalues = c(pearsonpvalues,a)
      b = SignalingNet[[i]]$corAnalysis[[j]]$spearman$spearmanpval
      spearmanpvalues = c(spearmanpvalues,b)
      c = SignalingNet[[i]]$corAnalysis[[j]]$kendall$kendallpval
      kendallpvalues = c(kendallpvalues,c)
      d = SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor
      pearsoncors = c(pearsoncors,d)
      e = SignalingNet[[i]]$corAnalysis[[j]]$spearman$spearmancor
      spearmancors = c(spearmancors,e)
      f = SignalingNet[[i]]$corAnalysis[[j]]$kendall$kendallcor
      kendallcors = c(kendallcors,f)

    }
  }else {

    a = SignalingNet[[i]]$corAnalysis$pearson$pearsonpval
    pearsonpvalues = c(pearsonpvalues,a)
    b = SignalingNet[[i]]$corAnalysis$spearman$spearmanpval
    spearmanpvalues = c(spearmanpvalues,b)
    c = SignalingNet[[i]]$corAnalysis$kendall$kendallpval
    kendallpvalues = c(kendallpvalues,c)
    d = SignalingNet[[i]]$corAnalysis$pearson$pearsoncor
    pearsoncors = c(pearsoncors,d)
    e = SignalingNet[[i]]$corAnalysis$spearman$spearmancor
    spearmancors = c(spearmancors,e)
    f = SignalingNet[[i]]$corAnalysis$kendall$kendallcor
    kendallcors = c(kendallcors,f)
  }
}

length(pearsonpvalues)
```

```
## [1] 76897
length(spearmanpvalues)
## [1] 76897
length(kendallpvalues)
## [1] 76897
```

Using the codes below, p-values are adjusted using “fdr” method.

```
adjusted.pearsonpvalues = p.adjust(pearsonpvalues , method = "fdr")
all(lapply(SignalingNet[Eligibles] , function(x) x$length) > 0)
## [1] TRUE
```

Adjusted p-values are added to the **SignalingNet** object. After completing this operation, adjusted.pearsonpvalues object must be NULL.

```
for(i in Eligibles){
  l = SignalingNet[[i]]$length
  if(l > 1){
    for(j in 1:l){
      SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval =
        adjusted.pearsonpvalues[j]
    }
    adjusted.pearsonpvalues = adjusted.pearsonpvalues[-1:-1]
  }else{
    SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval =
      adjusted.pearsonpvalues[1]
    adjusted.pearsonpvalues = adjusted.pearsonpvalues[-1]
  }
}
```

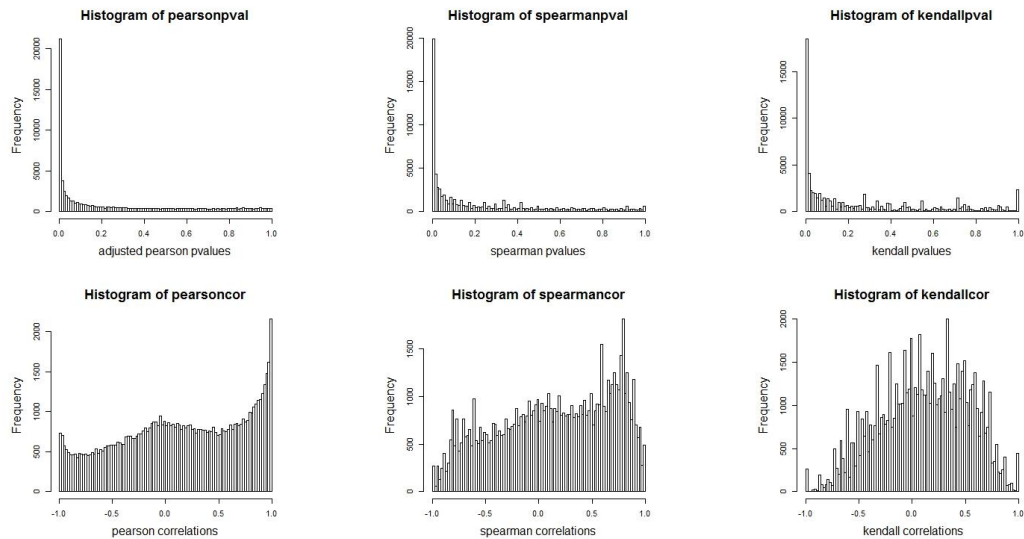


Figure7

In the histograms, most of the p-values are under 0.1. The number of positive coefficients are larger than negative coefficients. so, histograms of coefficients are left-skewed.

The code show that how many edges have a p-value less than a specified value.

```
cdfpearsonpval = c()
length(pearsonpvalues)

## [1] 76897

pvalue = seq(0.01 , 0.99 , by = 0.01)
for(i in 1:99){
  cdfpearsonpval[i] = sum(na.omit(pearsonpvalues < pvalue[i]))
}
cdfpearsonpval = cdfpearsonpval/length(pearsonpvalues)
cdfpearsonpval = rbind(pvalue,cdfpearsonpval)
cdfpearsonpval[,1:5]

##           [,1]      [,2]      [,3]      [,4]      [,5]
## pvalue      0.0100000 0.0200000 0.0300000 0.0400000 0.0500000
## cdfpearsonpval 0.3521854 0.4038779 0.4370782 0.4620987 0.4842452

# For example in the codes below, 35 percent of edges had a pvalue less than 0.01
```

```
cdfspearmanpval = c()
pvalue = seq(0.01 , 0.99 , by = 0.01)
for(i in 1:99){
  cdfspearmanpval[i] = sum(na.omit(spearmanpvalues < pvalue[i]))
}
```



```

}
cdfspearmanpval = cdfspearmanpval/length(spearmanpvalues)
cdfspearmanpval = rbind(pvalue,cdfspearmanpval)
cdfspearmanpval[,1:5]

##           [,1]      [,2]      [,3]      [,4]      [,5]
## pvalue      0.010000 0.0200000 0.0300000 0.0400000 0.050000
## cdfspearmanpval 0.259477 0.3160461 0.3524975 0.3870892 0.410094

cdfkendallpval = c()
pvalue = seq(0.01 , 0.99 , by = 0.01)
for(i in 1:99){
  cdfkendallpval[i] = sum(na.omit(kendallpvalues < pvalue[i]))
}
cdfkendallpval = cdfkendallpval/length(kendallpvalues)
cdfkendallpval = rbind(pvalue,cdfkendallpval)
cdfkendallpval[,1:5]

##           [,1]      [,2]      [,3]      [,4]      [,5]
## pvalue      0.0100000 0.0200000 0.0300000 0.0400000 0.0500000
## cdfkendallpval 0.2406726 0.2939256 0.3237578 0.3505208 0.3763086

```

```

cdfpearsoncorr = c()
length(pearsoncors)

## [1] 76897

positiveCorr = seq(0 , 0.9 , by = 0.1)
for(i in 1:10){
  cdfpearsoncorr[i] = sum(na.omit(pearsoncors > positiveCorr[i]))
}
cdfpearsoncorr = cdfpearsoncorr/length(pearsoncors)
cdfpearsoncorr = rbind(positiveCorr,cdfpearsoncorr)
# about 10% of edges have pearson corr larger than 0.9
cdfpearsoncorr

##           [,1]      [,2]      [,3]      [,4]      [,5]      [,6]
## positiveCorr 0.0000000 0.1000000 0.2000000 0.3000000 0.4000000 0.5000000
## cdfpearsoncorr 0.5919607 0.5378103 0.4845313 0.4329558 0.3831749 0.3341223
##           [,7]      [,8]      [,9]      [,10]
## positiveCorr 0.6000000 0.7000000 0.8000000 0.9000000
## cdfpearsoncorr 0.2848486 0.2310753 0.1724905 0.1018245

```

```

cdfpearsoncorr1 = c()
length(pearsoncors)

## [1] 76897

```

```

negativeCorr = seq(0 , -0.9 , by = -0.1)
for(i in 1:10){
  cdfpearsoncorr1[i] = sum(na.omit(pearsoncors < negativeCorr[i]))
}
cdfpearsoncorr1 = cdfpearsoncorr1/length(pearsoncors)
cdfpearsoncorr1 = rbind(negativeCorr,cdfpearsoncorr1)
# about 3.9% of edges have pearson corr less than -0.9
cdfpearsoncorr1

##                [,1]      [,2]      [,3]      [,4]      [,5]
[,6]
## negativeCorr      0.0000000 -0.1000000 -0.2000000 -0.3000000 -0.4000000 -
0.5000000
## cdfpearsoncorr1 0.4064528  0.3499096  0.2969817  0.2505169  0.2071992
0.168212
##                [,7]      [,8]      [,9]      [,10]
## negativeCorr     -0.6000000 -0.7000000 -0.8000000 -0.9000000
## cdfpearsoncorr1 0.1320208  0.09957476  0.06936551  0.03939035

```

```

cdfspearmancorr = c()
positiveCorr = seq(0 , 0.9 , by = 0.1)
for(i in 1:10){
  cdfspearmancorr[i] = sum(na.omit(spearmancors > positiveCorr[i]))
}
cdfspearmancorr = cdfspearmancorr/length(spearmancors)
cdfspearmancorr = rbind(positiveCorr,cdfspearmancorr)
# about 3.5% of edges have spearman corr larger than 0.9
cdfspearmancorr

##                [,1]      [,2]      [,3]      [,4]      [,5]
[,6]
## positiveCorr      0.0000000 0.1000000 0.2000000 0.3000000 0.4000000
0.5000000
## cdfspearmancorr 0.6002185 0.5422188 0.4857667 0.4329688 0.3782202
0.3209748
##                [,7]      [,8]      [,9]      [,10]
## positiveCorr      0.6000000 0.7000000 0.8000000 0.9000000
## cdfspearmancorr 0.2557057 0.1897083 0.1024227 0.03524195

```

```

cdfspearmancorr1 = c()
negativeCorr = seq(0 , -0.9 , by = -0.1)
for(i in 1:10){
  cdfspearmancorr1[i] = sum(na.omit(spearmancors < negativeCorr[i]))
}
cdfspearmancorr1 = cdfspearmancorr1/length(spearmancors)
cdfspearmancorr1 = rbind(negativeCorr,cdfspearmancorr1)

```

*# about 1.2% of edges have spearman core less than -0.9*

cdfspearmancorr1

```
##           [,1]      [,2]      [,3]      [,4]      [,5]
[,6]
## negativeCorr      0.000000 -0.100000 -0.200000 -0.300000 -0.400000 -
0.500000
## cdfspearmancorr1 0.39523  0.339155  0.2878396  0.2420901  0.2003329
0.1601883
##           [,7]      [,8]      [,9]      [,10]
## negativeCorr      -0.6000000 -0.7000000 -0.8000000 -0.9000000
## cdfspearmancorr1  0.1183401  0.08026321  0.03743969  0.01204208
```

```
cdfkendallcorr = c()
```

```
positiveCorr = seq(0 , 0.9 , by = 0.1)
```

```
for(i in 1:10){
```

```
  cdfkendallcorr[i] = sum(na.omit(kendallcors > positiveCorr[i]))
```

```
}
```

```
cdfkendallcorr = cdfkendallcorr/length(kendallcors)
```

```
cdfkendallcorr = rbind(positiveCorr,cdfkendallcorr)
```

*# about 0.8% of edges have kendall corr larger than 0.9*

cdfkendallcorr

```
##           [,1]      [,2]      [,3]      [,4]      [,5]      [,6]
## positiveCorr      0.000000 0.1000000 0.2000000 0.3000000 0.400000 0.5000000
## cdfkendallcorr 0.595966 0.5151306 0.4342172 0.3588176 0.277228 0.1983823
##           [,7]      [,8]      [,9]      [,10]
## positiveCorr      0.6000000 0.7000000 0.8000000 0.9000000
## cdfkendallcorr 0.1231387 0.06464491 0.02395412 0.008517888
```

```
cdfkendallcorr1 = c()
```

```
negativeCorr = seq(0 , -0.9 , by = -0.1)
```

```
for(i in 1:10){
```

```
  cdfkendallcorr1[i] = sum(na.omit(kendallcors < negativeCorr[i]))
```

```
}
```

```
cdfkendallcorr1 = cdfkendallcorr1/length(kendallcors)
```

```
cdfkendallcorr1 = rbind(negativeCorr,cdfkendallcorr1)
```

*# about 0.32% of edges have kendall core less than -0.9*

cdfkendallcorr1

```
##           [,1]      [,2]      [,3]      [,4]      [,5]
## negativeCorr      0.0000000 -0.1000000 -0.2000000 -0.3000000 -0.4000000
## cdfkendallcorr1 0.3903793  0.3138874  0.2426232  0.1856639  0.1293549
##           [,6]      [,7]      [,8]      [,9]
##           [,10]
## negativeCorr      -0.50000000 -0.60000000 -0.70000000 -0.80000000 -
0.900000000
```

```
## cdfkendallcorr1  0.08337126  0.04706295  0.02379807  0.00923313
0.004200424
```

Here we create a function for our database “SignalingNet” to check the number of its elements having less than n GSMs among eligible elements.

```
GSMnumber = function(x){logic = c()

for(i in Eligibles){
  if(SignalingNet[[i]]$length > 1){
    logic[i] = dim(do.call("cbind" , SignalingNet[[i]]$coExpr))[2] < x
  }else{logic[i] = dim(SignalingNet[[i]]$coExpr)[2] < x}
}
length(which(logic))
}

# to show the number of edges that have less than 10 GSMs:
GSMnumber(10)

## [1] 1567
```

## 5.One edge Subgraphs

### 5.1.Getting the number of Activation and Inhibition edges having p-values and correlations of interest.

In this section the number of edges having the specific p-value and correlation are computed. The first condition determines if SignalingNet[[i]]\$coranalysis has one element or multiple elements. If there is more than one element, all the elements should be non-NA. Then we check all the elements in SignalingNet[[i]] to have p-value < 0.05 and correlation > 0 . If all the conditions are met, the index of the edge is stored at **index.pval.cor1** object.

```
index.pval.cor1 = c()

for(i in Eligibles){
  logic =c()
  if(SignalingNet[[i]]$length > 1){

    logic1=c()
    for(j in 1:SignalingNet[[i]]$length){
      logic1[j] =
!any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))
    }
  }
}
```

```

    if(all(logic1)){
      for(j in 1:SignalingNet[[i]]$length){
        logic[j] = SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval
        < 0.05 & SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor > 0
      }
      if(all(logic)) {
        index.pval.cor1 = c(index.pval.cor1,i)
      }
    } else { if(!any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na)))){
      a = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
      SignalingNet[[i]]$corAnalysis$pearson$pearsoncor > 0
      if(a){
        index.pval.cor1 = c(index.pval.cor1,i)
      }
    }
  }
}

```

The first condition determines if there are more than one GDS for each edge. The second condition tells we need edges for which all the Pearson p-values are NA or > 0.05. If all the conditions are met, the index of the edge is stored at **index.pval.cor.NA** object.

```

index.pval.cor.NA = c()

for(i in Eligibles){
  logic = c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){

      logic[j]=any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))
      | SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval > 0.05
    }
    if(all(logic)){ index.pval.cor.NA = c(index.pval.cor.NA,i) }

  }else{ if(any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na))) |
  SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval > 0.05){
    index.pval.cor.NA = c(index.pval.cor.NA,i)}
  }
}

```

The first condition determines whether SignalingNet[[i]]\$coranalysis has one element or multiple elements. If there are several elements, all of them should be non-NA. Then we check that all elements in SignalingNet[[i]] to have the p-value < 0.05 or correlation < 0. If all the conditions are met, the index of the edge will be stored at **index.pval.cor2** object.

```
index.pval.cor2 = c()

for(i in Eligibles){
  logic=c()
  if(SignalingNet[[i]]$length > 1){

    logic1=c()
    for(j in 1:SignalingNet[[i]]$length){
      logic1[j] =
!any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))
    }
    if(all(logic1)){

      for(j in 1:SignalingNet[[i]]$length){

        logic[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor < 0
      }
      if(all(logic)) {
        index.pval.cor2 = c(index.pval.cor2,i)
      }
    } else { if(!any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na)))){
      a = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor < 0
      if(a){
        index.pval.cor2 = c(index.pval.cor2,i)
      }
    }
  }
}
```

## 5.2.Determining coherency of edges

If an edge is activation, and p-value < 0.05 and Pearson correlation > 0, or If an edge is inhibition, and p-value < 0.05 and pearson correlation < 0, the edge is coherent. If edge is activation, and p-value < 0.05 and pearson correlation < 0, or If edge is inhibition, and p-value < 0.05 and Pearson correlation > 0, the edge is incoherent. If p-value > 0.05 or p-value == NA, the edge is NA.

```
logic = edgelist[index.pval.cor1,4] == "activation"  
length(which(logic))
```

```
## [1] 896
```

```
idx.Act1 = which(logic)
```

```
logic = edgelist[index.pval.cor1,4] == "inhibition"  
length(which(logic))
```

```
## [1] 243
```

```
idx.Inh1 = which(logic)
```

```
for(i in index.pval.cor1){  
  if(SignalingNet[[i]][[7]] == "activation"){  
    SignalingNet[[i]][[8]] = "Coherent"  
  } else {SignalingNet[[i]][[8]] = "Incoherent"}  
}
```

```
logic = edgelist[index.pval.cor2,4] == "activation"  
length(which(logic))
```

```
## [1] 457
```

```
idx.Act2 = which(logic)
```

```
logic = edgelist[index.pval.cor2,4] == "inhibition"  
length(which(logic))
```

```
## [1] 130
```

```
idx.Inh2 = which(logic)
```

```
for(i in index.pval.cor2){  
  if(SignalingNet[[i]][[7]] == "activation"){  
    SignalingNet[[i]][[8]] = "Incoherent"  
  } else {SignalingNet[[i]][[8]] = "Coherent"}  
}
```

```

logic = edgelist[index.pval.cor.NA,4] == "activation"
length(which(logic))

## [1] 4602

idx.Act.NA = which(logic)

logic = edgelist[index.pval.cor.NA,4] == "inhibition"
length(which(logic))

## [1] 1246

idx.Inh.NA = which(logic)

```

```

for(i in index.pval.cor.NA){
  SignalingNet[[i]][[8]] = NA
}

```

```

number.of.coherent.edges=0
number.of.incoherent.edges=0
number.of.NA.edges=0

for(i in c(index.pval.cor1,index.pval.cor2)){
  if(SignalingNet[[i]][[8]] == "Coherent"){number.of.coherent.edges =
number.of.coherent.edges + 1
  }else if(SignalingNet[[i]][[8]] == "Incoherent"){number.of.incoherent.edges
= number.of.incoherent.edges + 1
  }
}

for(i in index.pval.cor.NA){
  if(is.na(SignalingNet[[i]][[8]])){number.of.NA.edges = number.of.NA.edges
+ 1}
}

number.of.coherent.edges

## [1] 1026

number.of.incoherent.edges

## [1] 700

number.of.NA.edges

## [1] 5848

```



In the following code, some eligible edges which are heterogeneous are stored at the object called **index.heterogeneous**. Heterogeneous edges are those for which there are multiple datasets with dissimilar p-values and correlations (Table 2).

```
index.heterogeneous = Eligibles[!(Eligibles %in% index)]

length(index.heterogeneous )

## [1] 15803

length(c(index.pval.cor1 , index.pval.cor2 , index.pval.cor.NA ,
index.heterogeneous ))

## [1] 23377

length(unique(c(index.pval.cor1 , index.pval.cor2 , index.pval.cor.NA ,
index.heterogeneous )))

## [1] 15805

length(Eligibles)

## [1] 15804

logic = edgelist[index.heterogeneous,4] == "activation"
length(which(logic))

## [1] 12693

logic = edgelist[index.heterogeneous,4] == "inhibition"
length(which(logic))

## [1] 3110
```

## 6. Unconnected gene pairs

For all the four independent analyses, we randomly select 1000 unconnected gene pairs through adjacency matrix self-multiplication. After that, correlation analysis is done on these gene pairs. To do that, a graph is created from the eligible edges in the edgelist, and a non-weighted adjacency matrix is created from the giant component of the graph. For more information refer to the section 8. If an entry between two genes in an adjacency matrix stays zero in all the multiplications, there is no path which could connect these two genes.

```
matpower2 = AdjMatrix %**% AdjMatrix
matpower3 = matpower2 %**% AdjMatrix
matpower4 = matpower3 %**% AdjMatrix
matpower5 = matpower4 %**% AdjMatrix
matpower6 = matpower5 %**% AdjMatrix
matpower7 = matpower6 %**% AdjMatrix
matpower8 = matpower7 %**% AdjMatrix
matpower9 = matpower8 %**% AdjMatrix
matpower10 = matpower9 %**% AdjMatrix
matpower11 = matpower10 %**% AdjMatrix
matpower12 = matpower11 %**% AdjMatrix
matpower13 = matpower12 %**% AdjMatrix
matpower14 = matpower13 %**% AdjMatrix
matpower15 = matpower14 %**% AdjMatrix
matpower16 = matpower15 %**% AdjMatrix
matpower17 = matpower16 %**% AdjMatrix
matpower18 = matpower17 %**% AdjMatrix
matpower19 = matpower18 %**% AdjMatrix
matpower20 = matpower19 %**% AdjMatrix
```

```
matpower21 = matpower20 %% AdjMatrix
matpower22 = matpower21 %% AdjMatrix
matpower23 = matpower22 %% AdjMatrix
matpower24 = matpower23 %% AdjMatrix
matpower25 = matpower24 %% AdjMatrix
matpower26 = matpower25 %% AdjMatrix
matpower27 = matpower26 %% AdjMatrix
matpower28 = matpower27 %% AdjMatrix
matpower29 = matpower28 %% AdjMatrix
matpower30 = matpower29 %% AdjMatrix
matpower31 = matpower30 %% AdjMatrix
matpower32 = matpower31 %% AdjMatrix
matpower33 = matpower32 %% AdjMatrix
matpower34 = matpower33 %% AdjMatrix
matpower35 = matpower34 %% AdjMatrix
matpower36 = matpower35 %% AdjMatrix
matpower37 = matpower36 %% AdjMatrix
matpower38 = matpower37 %% AdjMatrix
matpower39 = matpower38 %% AdjMatrix
matpower40 = matpower39 %% AdjMatrix
```

```
rn = rownames(AdjMatrix)
cn = colnames(AdjMatrix)
lzeros = list()
```

In the following code, gene pairs for which the value is zero in all 40 non-weighted adjacency matrices are selected.

```
for(i in 1:length(matpower40[1,])){
  a = cbind(1,2)
  for(j in 1:length(matpower40[,1])){
    if(matpower2[i,j] == 0 & matpower3[i,j] == 0 & matpower4[i,j] == 0 &
matpower5[i,j] == 0 &
      matpower6[i,j] == 0 & matpower7[i,j] == 0 & matpower8[i,j] == 0 &
matpower9[i,j] == 0 &
      matpower10[i,j] == 0 & matpower11[i,j] == 0 & matpower12[i,j] == 0 &
matpower13[i,j] == 0 &
      matpower14[i,j] == 0 & matpower15[i,j] == 0 & matpower16[i,j] == 0 &
matpower17[i,j] == 0 &
      matpower18[i,j] == 0 & matpower19[i,j] == 0 & matpower20[i,j] == 0 &
matpower21[i,j] == 0 &
      matpower22[i,j] == 0 & matpower23[i,j] == 0 & matpower24[i,j] == 0 &
matpower25[i,j] == 0 &
      matpower26[i,j] == 0 & matpower27[i,j] == 0 & matpower28[i,j] == 0 &
matpower29[i,j] == 0 &
      matpower30[i,j] == 0 & matpower31[i,j] == 0 & matpower32[i,j] == 0 &
matpower33[i,j] == 0 &
      matpower34[i,j] == 0 & matpower35[i,j] == 0 & matpower36[i,j] == 0 &
matpower37[i,j] == 0 &
      matpower38[i,j] == 0 & matpower39[i,j] == 0 & matpower40[i,j] == 0 &
AdjMatrix[i,j] == 0 ){

      a = rbind(cbind(rn[i],cn[j]),a)
    }
  }

  lzeros[[i]] = a
  print(i)
}

unconnected.edgelist = do.call("rbind" , lzeros)
index = which(unconnected.edgelist[,1]=="1")
unconnected.edgelist=unconnected.edgelist[-index,]
```

There should be no common edges between the main edgelist and unconnected-gene-pair edgelist. because matpower40 adjacency matrix was driven from the giant component of the graph which was created from eligible-edge edgelist, there may exist some common edges between unconnected.edgelist and the eligible-edge edgelist. The reason why this happens is that some eligible edges are not included in the giant component. These edges

should be omitted from the `unconnected.edgelist`. After that, we randomly selected 2000 edges from this `edgelist`.

```
e = edgelist[Eligibles,2:3]

index.of.rows.in.x.that.are.in.y <- function(x,y)
{
  x.vec <- apply(x, 1, paste, collapse = "")
  y.vec <- apply(y, 1, paste, collapse = "")
  index = x.vec %in% y.vec
  return(which(index))
}
Index = index.of.rows.in.x.that.are.in.y(unconnected.edgelist,e)
Index

# unconnected.edgelist = unconnected.edgelist[-Index,]

short.unconnected.edgelist = unconnected.edgelist[seq(1,4000000,2000),]
```

The loop-form gene pairs are removed through the following code.

```
#
logic=c()
for(i in 1:2000){
  logic[i] = short.unconnected.edgelist[i,1]==short.unconnected.edgelist[i,2]
}
idxloop = which(logic)
idxloop
short.unconnected.edgelist[idxloop,]
short.unconnected.edgelist = short.unconnected.edgelist[-idxloop,]
```

All the gene names of each dataset are stored as an element of **list.gene** object.

```
list.genes = lapply(ListGDS , function(x) unname(x[,1]))
```

Here, a large list called **pre.unconnected.SignalingNet1** is created. Each element of this list is representative of each gene pair in **short.unconnected.edgelist**, and contains gene expression profiles for that gene pair. Some gene pairs are in multiple datasets and some of them are in one dataset. So, some elements in `pre.unconnected.SignalingNet1` contain a list of multiple datasets and some of them contain just one dataset. To construct this object the following instruction is applied using **ListGDS** object:

The first condition is to see which datasets contain gene pairs in edge i in the short.unconnected.edgelist. The second condition tells if there are more than one datasets which satisfy the previous condition. This condition separates each element of pre.unconnected.SignalingNet1 into multiple datasets. The third condition tells that in dataset j, just two rows are related to the gene pairs. If the third condition is not satisfied, then between the similar-gene-named rows, we select the one with the largest IQR.

```
pre.unconnected.SignalingNet1 = list()
```

```
for(i in 1:length(short.unconnected.edgelist[,1])){
  index = which(unlist(lapply(list.genes , function(x)
all(short.unconnected.edgelist[i,] %in% x))))
  if(length(index) > 0){

    small.list = list()
    for(j in 1:length(index)){
      d = ListGDS[[index[j]]][which(list.genes[[index[j]]] %in%
short.unconnected.edgelist[i,]) , ]

      if(length(d[,1])==2){
        rownames(d) = d[,1]
        d = d[,-1]
        small.list[[j]] = d

      }else{
        n = d[,1]
        n1 = unique(n)

        index1 = which(n %in% n1[1])
        index2 = which(n %in% n1[2])

        if(length(index1)>1){
          d1 = d[index1,]

          IQRs = c()
          for(z in 1:length(d1[,1])){
            a=as.numeric(na.omit(as.numeric(d1[z,-1])))
            IQRs[z] = quantile(a, 0.75 ) - quantile(a, 0.25)
          }
          row.one = d1[which.max(IQRs),]

        }else{
          row.one = d[which(d[,1] == n1[1]),]
        }

      }

      if(length(index2)>1){
```

```

d2 = d[index2,]

IQRs = c()
for(z in 1:length(d2[,1])){
  a=as.numeric(na.omit(as.numeric(d2[z,-1])))
  IQRs[z] = quantile(a, 0.75 ) - quantile(a, 0.25)
}
row.two = d2[which.max(IQRs),]

}else{
  row.two = d[which(d[,1] == n1[2]),]
}

dNEW = rbind(row.one,row.two)
rownames(dNEW) = dNEW[,1]
dNEW = dNEW[,-1]
small.list[[j]] = dNEW

}
}

if(j > 1){
pre.unconnected.SignalingNet1[[i]] = small.list
}else{ pre.unconnected.SignalingNet1[[i]] = small.list[[1]]}

}
#print(i)
}

```

Some elements in pre.unconnected.SignalingNet1 are a list of multiple datasets and are in class list, some elements are a dataset and are in class dataframe, and some elements of are NULL. All the elements of this object containing datasets are stored in **pre.unconnected.SignalingNet2** object.

```

CLASS = lapply(pre.unconnected.SignalingNet1 , class)
index = which(CLASS == "list" | CLASS == "data.frame")

pre.unconnected.SignalingNet2 = pre.unconnected.SignalingNet1[index]

```

### Getting rid of outliers

```

for(i in 1:length(pre.unconnected.SignalingNet2)){

```

```

if(class(pre.unconnected.SignalingNet2[[i]])=="list"){
  for(j in 1:length(pre.unconnected.SignalingNet2[[i]]))
    outliers1 =
boxplot.stats(as.numeric(pre.unconnected.SignalingNet2[[i]][[j]][1,]))$out
    outliers2 =
boxplot.stats(as.numeric(pre.unconnected.SignalingNet2[[i]][[j]][2,]))$out

    index1 = which(as.numeric(pre.unconnected.SignalingNet2[[i]][[j]][2,])
%in% outliers1)
    index2 = which(as.numeric(pre.unconnected.SignalingNet2[[i]][[j]][2,])
%in% outliers2)
    index = unique(c(index1,index2))
    if(length(index) > 0){
      pre.unconnected.SignalingNet2[[i]][[j]] =
pre.unconnected.SignalingNet2[[i]][[j]][,-index]
    }

  } else {
    outliers1 =
boxplot.stats(as.numeric(pre.unconnected.SignalingNet2[[i]][1,]))$out
    outliers2 =
boxplot.stats(as.numeric(pre.unconnected.SignalingNet2[[i]][2,]))$out

    index1 = which(as.numeric(pre.unconnected.SignalingNet2[[i]][1,]) %in%
outliers1)
    index2 = which(as.numeric(pre.unconnected.SignalingNet2[[i]][2,]) %in%
outliers2)
    index = unique(c(index1,index2))
    if(length(index) > 0){
      pre.unconnected.SignalingNet2[[i]] =
pre.unconnected.SignalingNet2[[i]][,-index]}

  }
}

```

```

CLASS = lapply(pre.unconnected.SignalingNet2 , class)

```

*# The following code proves that all the elements in pre.unconnected.SignalingNet2 which is data frame, having more than two samples.*

```

all(lapply(lapply(pre.unconnected.SignalingNet2[which(CLASS == "data.frame")],
dim), function(x) x[2]) > 2)

```



```

lapply(pre.unconnected.SignalingNet2[which(CLASS == "list")] , function(x)
lapply(x,dim))

# The following code present the number of datasets with less than three
samples.
sum(unlist(lapply(pre.unconnected.SignalingNet2[which(CLASS == "list")] ,
function(x) lapply(x,function(x) dim(x)[2] <= 2))))

# The following code shows the indices of pre.unconnected.SignalingNet2 which
are in the class list having datasets with less than three samples.
index = which(!(unlist(lapply(pre.unconnected.SignalingNet2[which(CLASS ==
"list")] , function(x) all(unlist(lapply(x,function(x) dim(x)[2] > 2)))))))
index
# 92 216 491 582

# We deleted the ineligible datasets from the pre.unconnected.SignalingNet2
through the following code
for(i in 1:length(index)){
  idx = which(unlist(lapply(lapply(pre.unconnected.SignalingNet2[which(CLASS
== "list")][index][[i]] , dim) , function(x) x[2]<= 2)))
  pre.unconnected.SignalingNet2[which(CLASS == "list")][index][[i]] =
pre.unconnected.SignalingNet2[which(CLASS == "list")][index][[i]][-idx]
}

# Through the following code we check whether or not there is any element in
pre.unconnected.SignalingNet2 in the class list having the length of one
(have just one dataset)
any(lapply(pre.unconnected.SignalingNet2[which(CLASS == "list")] , length) ==
1)

```

## Correlation analysis

A new list called **unconnected.SignalingNet** is created by the following code involving the correlation results.

```

unconnected.SignalingNet = pre.unconnected.SignalingNet2

for(i in (1:length(unconnected.SignalingNet))){

```

```

if(class(unconnected.SignalingNet[[i]])=="list"){
  corAnalysis = list()
  dim = list()
  for(j in 1:length(unconnected.SignalingNet[[i]])){

    p = cor.test(as.numeric(unconnected.SignalingNet[[i]][[j]][1,]) ,
as.numeric(unconnected.SignalingNet[[i]][[j]][2,]) , method = "pearson")
    s = cor.test(as.numeric(unconnected.SignalingNet[[i]][[j]][1,]) ,
as.numeric(unconnected.SignalingNet[[i]][[j]][2,]) , method = "spearman")
    k = cor.test(as.numeric(unconnected.SignalingNet[[i]][[j]][1,]) ,
as.numeric(unconnected.SignalingNet[[i]][[j]][2,]) , method = "kendall")
    pearson = as.data.frame(cbind(pearsoncor = unname(p$estimate) ,
pearsonpval = p$p.value))
    spearman = as.data.frame(cbind(spearmancor = unname(s$estimate) ,
spearmanpval = s$p.value))
    kendall = as.data.frame(cbind(kendallcor = unname(k$estimate) ,
kendallpval = k$p.value))
    corAnalysis[[j]] = list(pearson = pearson,spearman = spearman,kendall =
kendall)
    dim[[j]] = dim(unconnected.SignalingNet[[i]][[j]])
  }

  unconnected.SignalingNet[[i]] = list(Source =
rownames(unconnected.SignalingNet[[i]][[1]])[1] , Target =
rownames(unconnected.SignalingNet[[i]][[1]])[2] , length =
length(unconnected.SignalingNet[[i]]) ,dim = dim , coExpr =
unconnected.SignalingNet[[i]] ,
corAnalysis = corAnalysis , Interactiontype =
"None" , Coherency = "empty")

} else {
  p = cor.test(as.numeric(unconnected.SignalingNet[[i]][1,]) ,
as.numeric(unconnected.SignalingNet[[i]][2,]) , method = "pearson")
  s = cor.test(as.numeric(unconnected.SignalingNet[[i]][1,]) ,
as.numeric(unconnected.SignalingNet[[i]][2,]) , method = "spearman")
  k = cor.test(as.numeric(unconnected.SignalingNet[[i]][1,]) ,
as.numeric(unconnected.SignalingNet[[i]][2,]) , method = "kendall")
  pearson = as.data.frame(cbind(pearsoncor = unname(p$estimate) ,
pearsonpval = p$p.value))
  spearman = as.data.frame(cbind(spearmancor = unname(s$estimate) ,
spearmanpval = s$p.value))
  kendall = as.data.frame(cbind(kendallcor = unname(k$estimate) ,
kendallpval = k$p.value))

  unconnected.SignalingNet[[i]] = list(Source =
unconnected.SignalingNet[[i]][1] , Target =
unconnected.SignalingNet[[i]][2] , length = 1 ,dim =

```

```

dim(unconnected.SignalingNet[[i]]) , coExpr = unconnected.SignalingNet[[i]]
,
corAnalysis = list(pearson = pearson,spearman =
spearman,kendall = kendall) ,
Interactiontype = "None" , Coherency = "empty")
}
#print(i)
}

```

*# The first 1000 elements of unconnected.SignalingNet object are selected.*  
unconnected.SignalingNet = unconnected.SignalingNet[1:1000]

```
unconnected.SignalingNet[[1]]
```

```

## $Source
## [1] "EGFR"
##
## $Target
## [1] "GDF7"
##
## $length
## [1] 3
##
## $dim
## $dim[[1]]
## [1] 2 6
##
## $dim[[2]]
## [1] 2 10
##
## $dim[[3]]
## [1] 2 12
##
##
## $coExpr
## $coExpr[[1]]
##      GSM148690 GSM148691 GSM148692 GSM148687 GSM148688 GSM148689
## EGFR    7.43996   8.09821   7.97677   6.60359   6.57079   6.6735
## GDF7    9.69006   9.43124   9.50077   7.13693   7.91709   7.9194
##
## $coExpr[[2]]
##      GSM15785 GSM15790 GSM15787 GSM15791 GSM15788 GSM15792 GSM15795
## EGFR 1452.4300 1280.6000 1701.4000 1416.290 1551.1200 1384.5200 1779.4400
## GDF7  68.7106  59.7835  67.5011  58.521  62.3464  57.2744  69.3889
##      GSM15786 GSM15789 GSM15794
## EGFR 1379.5800 1724.8500 1130.1800
## GDF7  57.7585  67.9647  58.8681
##
## $coExpr[[3]]

```

```

##      GSM3911  GSM3913 GSM3909  GSM3912 GSM3914 GSM3910  GSM3918  GSM3915
## EGFR 264.1880 355.9830 265.477 471.5480 377.299 288.368 271.8510 263.7640
## GDF7 63.4734 28.6645 143.890 59.4679 15.271 126.904 77.6652 57.4021
##      GSM3916  GSM3919 GSM3920  GSM3917
## EGFR 126.383 1752.890 577.190 1759.9300
## GDF7 187.364 197.137 90.366 49.2624
##
##
## $corAnalysis
## $corAnalysis[[1]]
## $corAnalysis[[1]]$pearson
## pearsoncor pearsonpval adjusted.pearsonpval
## 1 0.889057 0.01777975 0.05270126
##
## $corAnalysis[[1]]$spearman
## spearmancor spearmanpval
## 1 0.7142857 0.1361111
##
## $corAnalysis[[1]]$kendall
## kendallcor kendallpval
## 1 0.4666667 0.2722222
##
##
## $corAnalysis[[2]]
## $corAnalysis[[2]]$pearson
## pearsoncor pearsonpval adjusted.pearsonpval
## 1 0.784558 0.00720065 0.02588214
##
## $corAnalysis[[2]]$spearman
## spearmancor spearmanpval
## 1 0.7333333 0.02116648
##
## $corAnalysis[[2]]$kendall
## kendallcor kendallpval
## 1 0.5555556 0.02860946
##
##
## $corAnalysis[[3]]
## $corAnalysis[[3]]$pearson
## pearsoncor pearsonpval adjusted.pearsonpval
## 1 0.1628474 0.6130792 0.7266001
##
## $corAnalysis[[3]]$spearman
## spearmancor spearmanpval
## 1 -0.1818182 0.5729582
##
## $corAnalysis[[3]]$kendall
## kendallcor kendallpval
## 1 -0.1212121 0.6383613
##

```

```
##
##
## $Interactiontype
## [1] "None"
##
## $Coherency
## [1] "empty"
```

All the p-values and correlations are stored at these objects.

```
pearsonpvalues = c()
spearmanpvalues = c()
kendallpvalues = c()
pearsoncors = c()
spearmancors = c()
kendallcors = c()

for(i in 1:1000){
  if(unconnected.SignalingNet[[i]]$length > 1){
    for(j in 1:length(unconnected.SignalingNet[[i]]$corAnalysis)){

      a = unconnected.SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsonpval
      pearsonpvalues = c(pearsonpvalues,a)
      b =
unconnected.SignalingNet[[i]]$corAnalysis[[j]]$spearman$spearmanpval
      spearmanpvalues = c(spearmanpvalues,b)
      c = unconnected.SignalingNet[[i]]$corAnalysis[[j]]$kendall$kendallpval
      kendallpvalues = c(kendallpvalues,c)
      d = unconnected.SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor
      pearsoncors = c(pearsoncors,d)
      e = unconnected.SignalingNet[[i]]$corAnalysis[[j]]$spearman$spearmancor
      spearmancors = c(spearmancors,e)
      f = unconnected.SignalingNet[[i]]$corAnalysis[[j]]$kendall$kendallcor
      kendallcors = c(kendallcors,f)

    }
  }else {

    a = unconnected.SignalingNet[[i]]$corAnalysis$pearson$pearsonpval
    pearsonpvalues = c(pearsonpvalues,a)
    b = unconnected.SignalingNet[[i]]$corAnalysis$spearman$spearmanpval
    spearmanpvalues = c(spearmanpvalues,b)
    c = unconnected.SignalingNet[[i]]$corAnalysis$kendall$kendallpval
    kendallpvalues = c(kendallpvalues,c)
    d = unconnected.SignalingNet[[i]]$corAnalysis$pearson$pearsoncor
    pearsoncors = c(pearsoncors,d)
```

```

    e = unconnected.SignalingNet[[i]]$corAnalysis$spearman$spearmancor
    spearmancors = c(spearmancors,e)
    f = unconnected.SignalingNet[[i]]$corAnalysis$kendall$kendallcor
    kendallcors = c(kendallcors,f)
  }

  #print(i)
}

```

All the adjusted Pearson p-values are stored at the **adjusted.pearsonpvalues** object.

```

adjusted.pearsonpvalues = p.adjust(pearsonpvalues , method = "fdr")
adjusted.pearsonpvalues

all(lapply(unconnected.SignalingNet , function(x) x$length) > 0)

```

Adjusted Pearson p-values are added to the unconnected.SignalingNet object. After completing the process, adjusted.pearsonpvalues object should be empty or NULL.

```

for(i in 1:1000){
  l = unconnected.SignalingNet[[i]]$length
  if(l > 1){
    for(j in 1:l){

unconnected.SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval =
adjusted.pearsonpvalues[j]
    }
    adjusted.pearsonpvalues = adjusted.pearsonpvalues[-1:-1]
  }else{
    unconnected.SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval =
adjusted.pearsonpvalues[1]
    adjusted.pearsonpvalues = adjusted.pearsonpvalues[-1]
  }
}

```

```

adjusted.pearsonpvalues

## numeric(0)

```

The next codes are for getting the number of edges which are engaged in unconnected gene pairs (Table 2). The indices of gene pairs having p-values > 0.05 or p-values == NA are stored at **index.pval.cor.NA** object.

```

index.pval.cor.NA = c()

for(i in 1:1000){
  logic = c()
  if(unconnected.SignalingNet[[i]]$length > 1){

    for(j in 1:unconnected.SignalingNet[[i]]$length){

      logic[j]=any(unlist(lapply(unconnected.SignalingNet[[i]]$corAnalysis[[j]],is.
na))) |
      unconnected.SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval >
0.05
    }
    if(all(logic)){ index.pval.cor.NA = c(index.pval.cor.NA,i) }

  }else{
    if(any(unlist(lapply(unconnected.SignalingNet[[i]]$corAnalysis,is.na)))) |
    unconnected.SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval >
0.05){
      index.pval.cor.NA = c(index.pval.cor.NA,i)}
    }
  }
}

length(index.pval.cor.NA)

## [1] 437

```

The indices of gene pairs which have p-values < 0.05 and correlation > 0 are stored at **index.pval.cor1** object.

```

index.pval.cor1 = c()

for(i in 1:1000){
  logic =c()
  if(unconnected.SignalingNet[[i]]$length > 1){

    logic1=c()
    for(j in 1:unconnected.SignalingNet[[i]]$length){
      logic1[j] =
!any(unlist(lapply(unconnected.SignalingNet[[i]]$corAnalysis[[j]],is.na)))
    }
    if(all(logic1)){

      for(j in 1:unconnected.SignalingNet[[i]]$length){

        logic[j] =

```

```

unconnected.SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval <
0.05 & unconnected.SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor > 0
    }
    if(all(logic)) {
      index.pval.cor1 = c(index.pval.cor1,i)
    }
  } else {
if(!any(unlist(lapply(unconnected.SignalingNet[[i]]$corAnalysis,is.na)))){
  a =
unconnected.SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05
& unconnected.SignalingNet[[i]]$corAnalysis$pearson$pearsoncor > 0
  if(a){
    index.pval.cor1 = c(index.pval.cor1,i)
  }
}
}
}

length(index.pval.cor1)
## [1] 59

```

The indices of gene pairs which have p-values < 0.05 and correlation < 0 are stored at **index.pval.cor1** object.

```

index.pval.cor2 = c()

for(i in 1:1000){
  logic =c()
  if(unconnected.SignalingNet[[i]]$length > 1){

    logic1=c()
    for(j in 1:unconnected.SignalingNet[[i]]$length){
      logic1[j] =
!any(unlist(lapply(unconnected.SignalingNet[[i]]$corAnalysis[[j]],is.na)))
    }
    if(all(logic1)){

      for(j in 1:unconnected.SignalingNet[[i]]$length){

        logic[j] =
unconnected.SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval <
0.05 & unconnected.SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor < 0
      }
      if(all(logic)) {

```



```

        index.pval.cor2 = c(index.pval.cor2,i)
      }
    } else {
  if(!any(unlist(lapply(unconnected.SignalingNet[[i]]$corAnalysis,is.na)))){
    a =
unconnected.SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05
& unconnected.SignalingNet[[i]]$corAnalysis$pearson$pearsoncor < 0
    if(a){
      index.pval.cor2 = c(index.pval.cor2,i)
    }
  }
}
}

length(index.pval.cor2)
## [1] 33

```

```

1000 - 392 - 52 - 51
## [1] 505

```

## 7.Two Edge Subgraphs

```
load("Two Edge Subgraphs KEGG GEO.RData")
```

The following code shows if there is any loop among the eligible edges.

```

logic=c()
for(i in Eligibles){
  logic[i] = edgelist[i,2]==edgelist[i,3]
}
idxloop = which(logic)
edgelist[idxloop,]

## [1] ID          Gene1          Gene2          Interaction type
## <0 rows> (or 0-length row.names)

# There is no loop

```

Edges which has been participated in two-edge subgraphs are saved at **twoEdges** edgelist. the indices of these edges are stored at **idxTWOedges** object.

```

idxTWOedges = list()
for(i in Eligibles){
  a = edgelist[i,2]
  b = edgelist[i,3]

  index = which(edgelist[,2] == b & edgelist[,3] == a)
  if(length(index) > 0){idxTWOedges[[i]] = c(i,index)}
}
idxTWOedges = unlist(idxTWOedges)

length(idxTWOedges)
twoEdges=edgelist[idxTWOedges,]
sum(duplicated(twoEdges))
sum(duplicated(edgelist[2:4]))

twoEdges = twoEdges[!duplicated(twoEdges),]
idxTWOedges = rownames(twoEdges)
idxTWOedges = as.numeric(idxTWOedges)

```

```

length(idxTWOedges)

## [1] 314

tail(twoEdges)

##          ID  Gene1  Gene2 Interaction type idxTWOedges
## 20272 E20272 CDKN1B  CCND2      inhibition      20272
## 25769 E25769 CCND2  CDKN1B      inhibition      25769
## 20273 E20273 CDKN1B  CCND3      inhibition      20273
## 25770 E25770 CCND3  CDKN1B      inhibition      25770
## 21710 E21710 CSNK1E  WWTR1      inhibition      21710
## 21726 E21726 WWTR1  CSNK1E      inhibition      21726

```

```
head(twoEdges)
```

```
##           ID Gene1 Gene2 Interaction type idxTW0edges
## 21      E00021  EGFR   SRC      activation         21
## 16566 E16566   SRC   EGFR      activation        16566
## 264     E00264  PTK2  PIK3CA      activation         264
## 4387   E04387 PIK3CA  PTK2      activation        4387
## 265     E00265  PTK2  PIK3CB      activation         265
## 4388   E04388 PIK3CB  PTK2      activation        4388
```

Two-edge subgraphs are stored at **two.edge.subgraphs** object. This table is a **bisection edgelist**, and it contains columns which determine the indices of edges that has been participated in two edge subgraphs. Each row of the table presents a two-edge subgraph (Figure8).

```
twoEdges[idxTW0edges] = idxTW0edges
```

```
which(duplicated(edgelist[,2:4]))
```

```
index = seq(1 , length(idxTW0edges) , by = 2 )
```

```
two.edge.subgraphs = cbind(twoEdges[index,] , twoEdges[index + 1 ,])
```

| ID  | Gene1  | Gene2 | Interaction type | idxTW0edges | ID  | Gene1  | Gene2  | Interaction type | idxTW0edges |
|-----|--------|-------|------------------|-------------|-----|--------|--------|------------------|-------------|
| 21  | E00021 | EGFR  | SRC              | activation  | 21  | E16566 | SRC    | EGFR             | activation  |
| 264 | E00264 | PTK2  | PIK3CA           | activation  | 264 | E04387 | PIK3CA | PTK2             | activation  |
| 265 | E00265 | PTK2  | PIK3CB           | activation  | 265 | E04388 | PIK3CB | PTK2             | activation  |
| 266 | E00266 | PTK2  | PIK3CD           | activation  | 266 | E04389 | PIK3CD | PTK2             | activation  |
| 267 | E00267 | PTK2  | PIK3R1           | activation  | 267 | E04390 | PIK3R1 | PTK2             | activation  |
| 268 | E00268 | PTK2  | PIK3R2           | activation  | 268 | E04391 | PIK3R2 | PTK2             | activation  |
| 269 | E00269 | PTK2  | PIK3R3           | activation  | 269 | E04392 | PIK3R3 | PTK2             | activation  |
| 341 | E00341 | SRC   | ESR1             | activation  | 341 | E16581 | ESR1   | SRC              | activation  |
| 342 | E00342 | SRC   | ESR2             | activation  | 342 | E16582 | ESR2   | SRC              | activation  |
| 628 | E00628 | RAC1  | PAK1             | activation  | 628 | E08977 | PAK1   | RAC1             | activation  |
| 629 | E00629 | RAC2  | PAK1             | activation  | 629 | E08984 | PAK1   | RAC2             | activation  |
| 630 | E00630 | RAC3  | PAK1             | activation  | 630 | E08991 | PAK1   | RAC3             | activation  |
| 631 | E00631 | CDC42 | PAK1             | activation  | 631 | E08867 | PAK1   | CDC42            | activation  |
| 632 | E00632 | RAC1  | PAK2             | activation  | 632 | E08978 | PAK2   | RAC1             | activation  |
| 633 | E00633 | RAC2  | PAK2             | activation  | 633 | E08985 | PAK2   | RAC2             | activation  |

Figure8: The bisection edgelist

DNFBL-two-edge subgrphs are stored at the bisection edgelist called **DNFBL**, DPFBL1-two-edge subgrphs are stored at the bisection edgelist called **DPFBL1** and DPFBL2-two-edge subgrphs are stored at the bisection edgelist called **DPFBL2** (Tables 1 and 2).

```

idxDNFBL.1 = which(two.edge.subgraphs[,4]=="activation" &
two.edge.subgraphs[,9]=="inhibition")
DNFBL.1 = two.edge.subgraphs[idxDNFBL.1,]

length(DNFBL.1[,1])

## [1] 20

head(DNFBL.1)

##          ID   Gene1   Gene2 Interaction type idxTwOedges      ID
Gene1
## 1741 E01741   RAC1    RHOA      activation      1741 E25247
RHOA
## 1742 E01742   RAC2    RHOA      activation      1742 E25248
RHOA
## 1743 E01743   RAC3    RHOA      activation      1743 E25249
RHOA
## 2900 E02900  PRKACB   PPP1R1B      activation      2900 E25642
PPP1R1B
## 3107 E03107 TNFSF11  TNFRSF11B      activation      3107 E21675
TNFRSF11B
## 4885 E04885   INSR    PTPN11      activation      4885 E25571
PTPN11
##          Gene2 Interaction type idxTwOedges
## 1741    RAC1      inhibition      25247
## 1742    RAC2      inhibition      25248
## 1743    RAC3      inhibition      25249
## 2900  PRKACB      inhibition      25642
## 3107 TNFSF11      inhibition      21675
## 4885   INSR      inhibition      25571

idxDNFBL.2 = which(two.edge.subgraphs[,4]=="inhibition" &
two.edge.subgraphs[,9]=="activation")
DNFBL.2 = two.edge.subgraphs[idxDNFBL.2,]

DNFBL = rbind(DNFBL.1 , DNFBL.2)
idxDNFBL = c(idxDNFBL.1,idxDNFBL.2)

```

```

idxDPFBL1 = which(two.edge.subgraphs[,4]=="activation" &
two.edge.subgraphs[,9]=="activation")
DPFBL1 = two.edge.subgraphs[idxDPFBL1,]

length(DPFBL1[,1])

## [1] 125

head(DPFBL1)

```

```
##          ID Gene1  Gene2 Interaction type idxTW0edges      ID  Gene1 Gene2
## 21  E00021  EGFR    SRC      activation        21 E16566    SRC  EGFR
## 264 E00264  PTK2  PIK3CA      activation      264 E04387  PIK3CA  PTK2
## 265 E00265  PTK2  PIK3CB      activation      265 E04388  PIK3CB  PTK2
## 266 E00266  PTK2  PIK3CD      activation      266 E04389  PIK3CD  PTK2
## 267 E00267  PTK2  PIK3R1      activation      267 E04390  PIK3R1  PTK2
## 268 E00268  PTK2  PIK3R2      activation      268 E04391  PIK3R2  PTK2
##      Interaction type idxTW0edges
## 21      activation      16566
## 264      activation      4387
## 265      activation      4388
## 266      activation      4389
## 267      activation      4390
## 268      activation      4391
```

```
idxDPFBL2 = which(two.edge.subgraphs[,4]=="inhibition" &
two.edge.subgraphs[,9]=="inhibition")
DPFBL2 = two.edge.subgraphs[idxDPFBL2,]
```

```
length(DPFBL2[,1])
```

```
## [1] 12
```

```
head(DPFBL2)
```

```
##          ID Gene1  Gene2 Interaction type idxTW0edges      ID  Gene1
Gene2
## 21354 E21354  GLI1    GLI3      inhibition      21354 E21350  GLI3
GLI1
## 19192 E19192  BCL2    BAX      inhibition      19192 E26437  BAX
BCL2
## 19202 E19202  BCL2    BAD      inhibition      19202 E19216  BAD
BCL2
## 19217 E19217  BCL2L1  BAK1      inhibition      19217 E25805  BAK1
BCL2L1
## 19841 E19841  CDK2    CDKN1B      inhibition      19841 E19850  CDKN1B
CDK2
## 19842 E19842  CDK2    CDKN1C      inhibition      19842 E19851  CDKN1C
CDK2
##      Interaction type idxTW0edges
## 21354      inhibition      21350
## 19192      inhibition      26437
## 19202      inhibition      19216
## 19217      inhibition      25805
## 19841      inhibition      19850
## 19842      inhibition      19851
```

## 7.1.Computing the number of edges constructing in three kinds of two-edge subgraphs

### DNFBL

```
index = c(DNFBL[,5],DNFBL[,10])

DNFBL.pval.cor1 = 0
DNFBL.pval.cor.NA = 0
DNFBL.pval.cor2 = 0

for(i in index){
  logic=c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){
      logic[j]=
!any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))
    }
    if(all(logic)){
      logic1 = c()
      logic2 = c()
      for(j in 1:SignalingNet[[i]]$length){

        logic1[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor > 0
        logic2[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor < 0
      }
      if(all(logic1)) {DNFBL.pval.cor1=DNFBL.pval.cor1+1}
      if(all(logic2)) {DNFBL.pval.cor2=DNFBL.pval.cor2+1}
    }
  } else{
    if(!any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na)))){
      a = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor > 0
      b = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor < 0
      if(a){DNFBL.pval.cor1=DNFBL.pval.cor1+1}
      if(b){DNFBL.pval.cor2=DNFBL.pval.cor2+1}
    }
  }
}
```

```

    }
}

for(i in index){
  logic = c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){

      logic[j]=any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))
| SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval > 0.05
    }
    if(all(logic)){ DNFBFL.pval.cor.NA = DNFBFL.pval.cor.NA + 1 }

  }else{ if(any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na))) |
SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval > 0.05){
    index.pval.cor.NA = DNFBFL.pval.cor.NA + 1}
  }
}

DNFBFL.pval.cor1
## [1] 0

DNFBFL.pval.cor.NA
## [1] 10

DNFBFL.pval.cor2
## [1] 3

length(DNFBFL[,1]) * 2 - c(DNFBFL.pval.cor1 + DNFBFL.pval.cor.NA +
DNFBFL.pval.cor2)
## [1] 27

```

## DPFBL1

```
index = c(DPFBL1[,5] , DPFBL1[,10])
```

```

DPFBL1.pval.cor1 = 0
DPFBL1.pval.cor.NA = 0
DPFBL1.pval.cor2 = 0

```

```

for(i in index){
  logic = c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){
      logic[j]=(!class(SignalingNet[[i]]$corAnalysis[[j]]) == "logical" |
any(is.na(unlist(SignalingNet[[i]]$corAnalysis[[j]]))))
    }
    if(all(logic)){
      logic1 = c()
      logic2 = c()
      for(j in 1:SignalingNet[[i]]$length){

        logic1[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor > 0
        logic2[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor < 0
      }
      if(all(logic1)) {DPFBL1.pval.cor1=DPFBL1.pval.cor1+1}
      if(all(logic2)) {DPFBL1.pval.cor2=DPFBL1.pval.cor2+1}
    }
  } else if(SignalingNet[[i]]$length == 1) {
    if(!class(SignalingNet[[i]]$corAnalysis) == "logical" |
any(is.na(unlist(SignalingNet[[i]]$corAnalysis)))){
      a = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor > 0
      b = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor < 0
      if(a){DPFBL1.pval.cor1=DPFBL1.pval.cor1+1}
      if(b){DPFBL1.pval.cor2=DPFBL1.pval.cor2+1}
    }
  }
}

```

```

for(i in index){
  logic = c()
  if(SignalingNet[[i]]$length > 1){
    for(j in 1:SignalingNet[[i]]$length){
      logic[j]=!class(SignalingNet[[i]]$corAnalysis[[j]]) == "logical"
    }
  }
}

```



```

    if(all(logic)){
      logic1 =c()
      for(j in 1:SignalingNet[[i]]$length){
        logic1[j]=
is.na(SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval) |
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval > 0.05
      }
      if(all(logic1)){DPFBL1.pval.cor.NA = DPFBL1.pval.cor.NA+1}
    }
  } else {
    if(!class(SignalingNet[[i]]$corAnalysis) == "logical"){
      a = is.na(SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval) |
SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval > 0.05
      if(a){DPFBL1.pval.cor.NA=DPFBL1.pval.cor.NA+1}
    }
  }
}
DPFBL1.pval.cor1
## [1] 34
DPFBL1.pval.cor.NA
## [1] 85
DPFBL1.pval.cor2
## [1] 6
length(DPFBL1[,1]) * 2 - c(DPFBL1.pval.cor1 + DPFBL1.pval.cor.NA +
DPFBL1.pval.cor2)
## [1] 125

```

## DPFBL2

```
index = c(DPFBL2[,5] , DPFBL2[,10])
```

```

DPFBL2.pval.cor1 = 0
DPFBL2.pval.cor.NA = 0
DPFBL2.pval.cor2 = 0

```

```

for(i in index){
  logic =c()

```

```

if(SignalingNet[[i]]$length > 1){
  for(j in 1:SignalingNet[[i]]$length){
    logic[j]=(!class(SignalingNet[[i]]$corAnalysis[[j]]) == "logical" |
any(is.na(unlist(SignalingNet[[i]]$corAnalysis[[j]]))))))
  }
  if(all(logic)){
    logic1 = c()
    logic2 = c()
    for(j in 1:SignalingNet[[i]]$length){
      logic1[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor > 0
      logic2[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor < 0
    }
    if(all(logic1)) {DPFBL2.pval.cor1=DPFBL2.pval.cor1+1}
    if(all(logic2)) {DPFBL2.pval.cor2=DPFBL2.pval.cor2+1}
  }
} else if(SignalingNet[[i]]$length == 1) {
  if(!class(SignalingNet[[i]]$corAnalysis) == "logical" |
any(is.na(unlist(SignalingNet[[i]]$corAnalysis))))){
    a = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor > 0
    b = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor < 0
    if(a){DPFBL2.pval.cor1=DPFBL2.pval.cor1+1}
    if(b){DPFBL2.pval.cor2=DPFBL2.pval.cor2+1}
  }
}
}

for(i in index){
  logic =c()
  if(SignalingNet[[i]]$length > 1){
    for(j in 1:SignalingNet[[i]]$length){
      logic[j]=!class(SignalingNet[[i]]$corAnalysis[[j]]) == "logical"
    }
    if(all(logic)){
      logic1 =c()
      for(j in 1:SignalingNet[[i]]$length){
        logic1[j]=
is.na(SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval) |

```

```

SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval > 0.05
    }
    if(all(logic1)){DPFBL2.pval.cor.NA = DPFBL2.pval.cor.NA+1}
  }

} else {
  if(!class(SignalingNet[[i]]$corAnalysis) == "logical"){
    a = is.na(SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval) |
SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval > 0.05
    if(a){DPFBL2.pval.cor.NA=DPFBL2.pval.cor.NA+1}
  }
}
}

DPFBL2.pval.cor1
## [1] 0

DPFBL2.pval.cor.NA
## [1] 10

DPFBL2.pval.cor2
## [1] 0

length(DPFBL2[,1]) * 2 - (DPFBL2.pval.cor1 + DPFBL2.pval.cor.NA +
DPFBL2.pval.cor2)
## [1] 14

```

## 8. Multiple Edge subgraphs

To compute the number of edges which participate in multiple-edge subgraphs, an edgelist is created from eligible edges called **shortEdgelist**.

```
library(igraph)

shortEdgelist = edgelist[Eligibles,]

shortEdgelist[which(shortEdgelist[,4] == "activation"),4] = 1
shortEdgelist[which(shortEdgelist[,4] == "inhibition"),4] = -1
```

A weighted graph was created from shortEdgelist object, and -1 was assigned to inhibitions and 1 was assigned to activations as weights. Then, the largest component of the eligible edges in the graph was extracted called **weighted.giant.component**. We changed this object into an adjacency matrix called **WadjaMatrix**. The same processes were done for non-weighted adjacency matrix and an object called **none.weighted.giant.component** was created. Then, **AdjaMatrix** object was created from that object. The diameter of the two giant components was computed and using matrix self-multiplication, the number of edges which are engaged in 8 kinds of multiple-edge subgraphs was computed (Table1 and Table2).

If the edge i in the edgelist is activation and the entity between the gene pair in one of the weighted matrices equals to their unweighted counterparts while the entities are not zero or NA, this edge is engaged in MFFL1 or MPFBL1 subgraphs.

If the edge i in the edgelist is inhibition and the entity between the gene pair in one of the weighted matrices equals to their unweighted counterparts while the entities are not zero or NA, this edge is engaged in MNFFL2 or MNFBL2 subgraphs.

If the edge i in the edgelist is activation and the entity between the gene pair in one of the weighted matrices does not equal to their unweighted counterparts while the entities are not zero or NA, this edge is engaged in MNFBL1 or MNFFL1 subgraphs.

If the edge i in the edgelist is inhibition and the entity between the gene pair in one of the weighted matrices does not equal to their unweighted counterparts while the entities are not zero or NA, this edge is engaged in MFFL2 or MPFBL2 subgraphs.

```
w = as.numeric(shortEdgelist[,4])
shortEdgelist = as.matrix(shortEdgelist)

g = graph_from_edgelist(shortEdgelist[,2:3] , directed = T)
E(g)$weights = w
```

```
shortEdgelistGenes = unique(c(shortEdgelist[,2] , c(shortEdgelist[,3])))

c= components(g)
```

```

weighted.giant.component = induced.subgraph(g , which(c$membership == 1))
weighted.giant.component

## IGRAPH e173162 DN-- 2549 15718 --
## + attr: name (v/c), weights (e/n)
## + edges from e173162 (vertex names):
## [1] EGF ->EGFR TGFA ->EGFR HGF ->MET MET ->ERBB3 IGF1 ->IGF1R
## [6] VEGFA->KDR PDGFA->PDGFRA PDGFB->PDGFRA PDGFC->PDGFRA PDGFD->PDGFRA
## [11] PDGFA->PDGFRB PDGFB->PDGFRB PDGFC->PDGFRB PDGFD->PDGFRB FGF2 ->FGFR3
## [16] FGF2 ->FGFR2 GAS6 ->AXL IL6 ->IL6R EGFR ->JAK1 EGFR ->JAK2
## [21] EGFR ->SRC EGFR ->GAB1 EGFR ->PLCG1 EGFR ->PLCG2 EGFR ->SHC2
## [26] EGFR ->SHC4 EGFR ->SHC3 EGFR ->SHC1 MET ->JAK1 MET ->JAK2
## [31] MET ->SRC MET ->GAB1 MET ->PLCG1 MET ->PLCG2 MET ->SHC2
## [36] MET ->SHC4 MET ->SHC3 MET ->SHC1 IGF1R->JAK1 IGF1R->JAK2
## + ... omitted several edges

is.connected(weighted.giant.component)

## [1] TRUE

WadjMatrix = as_adjacency_matrix(weighted.giant.component , attr = "weights")

WadjMatrix = as.matrix(WadjMatrix)

diameter(weighted.giant.component)

## [1] 17

```

```

Wmatpower2 = WadjMatrix %**% WadjMatrix

Wmatpower3 = Wmatpower2 %**% WadjMatrix

Wmatpower4 = Wmatpower3 %**% WadjMatrix

Wmatpower5 = Wmatpower4 %**% WadjMatrix

Wmatpower6 = Wmatpower5 %**% WadjMatrix

Wmatpower7 = Wmatpower6 %**% WadjMatrix

Wmatpower8 = Wmatpower7 %**% WadjMatrix

Wmatpower9 = Wmatpower8 %**% WadjMatrix

Wmatpower10 = Wmatpower9 %**% WadjMatrix

```

```

Wmatpower11 = Wmatpower10 %% WadjMatrix
Wmatpower12 = Wmatpower11 %% WadjMatrix
Wmatpower13 = Wmatpower12 %% WadjMatrix
Wmatpower14 = Wmatpower13 %% WadjMatrix
Wmatpower15 = Wmatpower14 %% WadjMatrix
Wmatpower16 = Wmatpower15 %% WadjMatrix
Wmatpower17 = Wmatpower16 %% WadjMatrix

```

```

g1 = graph_from_edgelist(shortEdgelist[,2:3] , directed = T)

c= components(g1)

none.weighted.giant.component = induced.subgraph(g1 , which(c$membership == 1))

none.weighted.giant.component

## IGRAPH 4d382e2 DN-- 2549 15718 --
## + attr: name (v/c)
## + edges from 4d382e2 (vertex names):
## [1] EGF ->EGFR TGFA ->EGFR HGF ->MET MET ->ERBB3 IGF1 ->IGF1R
## [6] VEGFA->KDR PDGFA->PDGFRA PDGFB->PDGFRA PDGFC->PDGFRA PDGFD->PDGFRA
## [11] PDGFA->PDGFRB PDGFB->PDGFRB PDGFC->PDGFRB PDGFD->PDGFRB FGF2 ->FGFR3
## [16] FGF2 ->FGFR2 GAS6 ->AXL IL6 ->IL6R EGFR ->JAK1 EGFR ->JAK2
## [21] EGFR ->SRC EGFR ->GAB1 EGFR ->PLCG1 EGFR ->PLCG2 EGFR ->SHC2
## [26] EGFR ->SHC4 EGFR ->SHC3 EGFR ->SHC1 MET ->JAK1 MET ->JAK2
## [31] MET ->SRC MET ->GAB1 MET ->PLCG1 MET ->PLCG2 MET ->SHC2
## [36] MET ->SHC4 MET ->SHC3 MET ->SHC1 IGF1R->JAK1 IGF1R->JAK2
## + ... omitted several edges

is.connected(none.weighted.giant.component)

## [1] TRUE

AdjMatrix = as_adjacency_matrix(none.weighted.giant.component)

AdjMatrix = as.matrix(AdjMatrix)

diameter(none.weighted.giant.component)

## [1] 17

```

```

matpower2 = AdjMatrix %**% AdjMatrix
matpower3 = matpower2 %**% AdjMatrix
matpower4 = matpower3 %**% AdjMatrix
matpower5 = matpower4 %**% AdjMatrix
matpower6 = matpower5 %**% AdjMatrix
matpower7 = matpower6 %**% AdjMatrix
matpower8 = matpower7 %**% AdjMatrix
matpower9 = matpower8 %**% AdjMatrix
matpower10 = matpower9 %**% AdjMatrix
matpower11 = matpower10 %**% AdjMatrix
matpower12 = matpower11 %**% AdjMatrix
matpower13 = matpower12 %**% AdjMatrix
matpower14 = matpower13 %**% AdjMatrix
matpower15 = matpower14 %**% AdjMatrix
matpower16 = matpower15 %**% AdjMatrix
matpower17 = matpower16 %**% AdjMatrix

```

**listAdj1** object is a large list containing matrices as the same dimension as the adjacency matrices. This list contains 16 matrices, and Each matrix is representative of one of the powered matrices.

```

Apower = matrix(0 , 2549 , 2549)
colnames(Apower) = colnames(AdjMatrix)
rownames(Apower) = rownames(AdjMatrix)
listAdj1 = list(Apower2 = Apower,
Apower3 = Apower,
Apower4 = Apower,
Apower5 = Apower,
Apower6 = Apower,

```

```

Apower7 = Apower,
Apower8 = Apower,
Apower9 = Apower,
Apower10 = Apower,
Apower11 = Apower,
Apower12 = Apower,
Apower13 = Apower,
Apower14 = Apower,
Apower15 = Apower,
Apower16 = Apower,
Apower17 = Apower
)

```

Before computing the number of edges which participate in multiple-edge subgraphs, the following algorithms are required to be applied: If  $WmatpowerX[i,j] == matpowerX[i,j]$ , we put the value of  $WmatpowerX[i,j]$  at the  $ApowerX[i,j]$  in **listAdj1** list. Otherwise, we put NA. After that, the number of edges which take part in “MPFBL1”, “MNFBL2”, “MFFL1” and “MNFFL2” multiple-edge subgraphs are computed. For the other multiple-edge subgraphs called “MNFBL1”, “MPFBL2”, “MFFL2” and “MNFFL1”, If  $WmatpowerX[i,j] != matpowerX[i,j]$ , we put the value of  $WmatpowerX[i,j]$  at the  $ApowerX[i,j]$  in **listAdj2** list. Otherwise, we put NA.

```

for(i in 1:length(AdjMatrix[,1])){

for(j in 1:length(AdjMatrix[1,])){

if(Wmatpower2[i,j] == matpower2[i,j]){
  listAdj1$Apower2[i,j] = Wmatpower2[i,j]
} else { listAdj1$Apower2[i,j] = NA }

if(Wmatpower3[i,j] == matpower3[i,j]){
  listAdj1$Apower3[i,j] = Wmatpower3[i,j]
} else { listAdj1$Apower3[i,j] = NA }

if(Wmatpower4[i,j] == matpower4[i,j]){
  listAdj1$Apower4[i,j] = Wmatpower4[i,j]
} else { listAdj1$Apower4[i,j] = NA }

if(Wmatpower5[i,j] == matpower5[i,j]){
  listAdj1$Apower5[i,j] = Wmatpower5[i,j]
} else { listAdj1$Apower5[i,j] = NA }

if(Wmatpower6[i,j] == matpower6[i,j]){

```



```
listAdj1$Apower6[i,j] = Wmatpower6[i,j]
} else { listAdj1$Apower6[i,j] = NA }

if(Wmatpower7[i,j] == matpower7[i,j]){
  listAdj1$Apower7[i,j] = Wmatpower7[i,j]
} else { listAdj1$Apower7[i,j] = NA }

if(Wmatpower8[i,j] == matpower8[i,j]){
  listAdj1$Apower8[i,j] = Wmatpower8[i,j]
} else { listAdj1$Apower8[i,j] = NA }

if(Wmatpower9[i,j] == matpower9[i,j]){
  listAdj1$Apower9[i,j] = Wmatpower9[i,j]
} else { listAdj1$Apower9[i,j] = NA }

if(Wmatpower10[i,j] == matpower10[i,j]){
  listAdj1$Apower10[i,j] = Wmatpower10[i,j]
} else { listAdj1$Apower10[i,j] = NA }

if(Wmatpower11[i,j] == matpower11[i,j]){
  listAdj1$Apower11[i,j] = Wmatpower11[i,j]
} else { listAdj1$Apower11[i,j] = NA }

if(Wmatpower12[i,j] == matpower12[i,j]){
  listAdj1$Apower12[i,j] = Wmatpower12[i,j]
} else { listAdj1$Apower12[i,j] = NA }

if(Wmatpower13[i,j] == matpower13[i,j]){
  listAdj1$Apower13[i,j] = Wmatpower13[i,j]
} else { listAdj1$Apower13[i,j] = NA }

if(Wmatpower14[i,j] == matpower14[i,j]){
  listAdj1$Apower14[i,j] = Wmatpower14[i,j]
} else { listAdj1$Apower14[i,j] = NA }

if(Wmatpower15[i,j] == matpower15[i,j]){
  listAdj1$Apower15[i,j] = Wmatpower15[i,j]
} else { listAdj1$Apower15[i,j] = NA }
```

```

if(Wmatpower16[i,j] == matpower16[i,j]){
  listAdj1$Apower16[i,j] = Wmatpower16[i,j]
} else { listAdj1$Apower16[i,j] = NA }

if(Wmatpower17[i,j] == matpower17[i,j]){
  listAdj1$Apower17[i,j] = Wmatpower17[i,j]
} else { listAdj1$Apower17[i,j] = NA }

}
# print(i)
}

```

```

Apower = matrix(0 , 2549 , 2549)
colnames(Apower) = colnames(AdjMatrix)
rownames(Apower) = rownames(AdjMatrix)
listAdj2 = list(Apower2 = Apower,
               Apower3 = Apower,
               Apower4 = Apower,
               Apower5 = Apower,
               Apower6 = Apower,
               Apower7 = Apower,
               Apower8 = Apower,
               Apower9 = Apower,
               Apower10 = Apower,
               Apower11 = Apower,
               Apower12 = Apower,
               Apower13 = Apower,
               Apower14 = Apower,
               Apower15 = Apower,
               Apower16 = Apower,
               Apower17 = Apower
)

for(i in 1:length(AdjMatrix[,1])){

  for(j in 1:length(AdjMatrix[1,])){

    if(Wmatpower2[i,j] != matpower2[i,j]){
      listAdj2$Apower2[i,j] = Wmatpower2[i,j]
    } else { listAdj2$Apower2[i,j] = NA }

    if(Wmatpower3[i,j] != matpower3[i,j]){
      listAdj2$Apower3[i,j] = Wmatpower3[i,j]
    } else { listAdj2$Apower3[i,j] = NA }

  }

}

```

```
if(Wmatpower4[i,j] != matpower4[i,j]){
  listAdj2$Apower4[i,j] = Wmatpower4[i,j]
} else { listAdj2$Apower4[i,j] = NA }

if(Wmatpower5[i,j] != matpower5[i,j]){
  listAdj2$Apower5[i,j] = Wmatpower5[i,j]
} else { listAdj2$Apower5[i,j] = NA }

if(Wmatpower6[i,j] != matpower6[i,j]){
  listAdj2$Apower6[i,j] = Wmatpower6[i,j]
} else { listAdj2$Apower6[i,j] = NA }

if(Wmatpower7[i,j] != matpower7[i,j]){
  listAdj2$Apower7[i,j] = Wmatpower7[i,j]
} else { listAdj2$Apower7[i,j] = NA }

if(Wmatpower8[i,j] != matpower8[i,j]){
  listAdj2$Apower8[i,j] = Wmatpower8[i,j]
} else { listAdj2$Apower8[i,j] = NA }

if(Wmatpower9[i,j] != matpower9[i,j]){
  listAdj2$Apower9[i,j] = Wmatpower9[i,j]
} else { listAdj2$Apower9[i,j] = NA }

if(Wmatpower10[i,j] != matpower10[i,j]){
  listAdj2$Apower10[i,j] = Wmatpower10[i,j]
} else { listAdj2$Apower10[i,j] = NA }

if(Wmatpower11[i,j] != matpower11[i,j]){
  listAdj2$Apower11[i,j] = Wmatpower11[i,j]
} else { listAdj2$Apower11[i,j] = NA }

if(Wmatpower12[i,j] != matpower12[i,j]){
  listAdj2$Apower12[i,j] = Wmatpower12[i,j]
} else { listAdj2$Apower12[i,j] = NA }

if(Wmatpower13[i,j] != matpower13[i,j]){
  listAdj2$Apower13[i,j] = Wmatpower13[i,j]
```

```

    } else { listAdj2$Apower13[i,j] = NA }

    if(Wmatpower14[i,j] != matpower14[i,j]){
      listAdj2$Apower14[i,j] = Wmatpower14[i,j]
    } else { listAdj2$Apower14[i,j] = NA }

    if(Wmatpower15[i,j] != matpower15[i,j]){
      listAdj2$Apower15[i,j] = Wmatpower15[i,j]
    } else { listAdj2$Apower15[i,j] = NA }

    if(Wmatpower16[i,j] != matpower16[i,j]){
      listAdj2$Apower16[i,j] = Wmatpower16[i,j]
    } else { listAdj2$Apower16[i,j] = NA }

    if(Wmatpower17[i,j] != matpower17[i,j]){
      listAdj2$Apower17[i,j] = Wmatpower17[i,j]
    } else { listAdj2$Apower17[i,j] = NA }

  }
}

```

Using the following code, the indices of the giant component edges in the **KEGG edgelist** are found, and they are stored at **commonIndex** object.

```

m = as_edgelist(weighted.giant.component)
m = as.data.frame(m , stringsAsFactors = F)
e = edgelist[,2:3]

index.of.rows.in.x.that.are.in.y <- function(x,y)
{
  x.vec <- apply(x, 1, paste, collapse = "")
  y.vec <- apply(y, 1, paste, collapse = "")
  index = x.vec %in% y.vec
  return(which(index))
}
commonIndex = index.of.rows.in.x.that.are.in.y(e,m)

```

## 8.1.Computing the number of edges participating in MPFBL1, MNFBL2, MFFL1 and MNFFL2 multiple-edge subgraphs

The indices of edges taking part in MFFL1 subgraph are saved at **idxMFFL1** object through the following code.

```
MFFL1 = list()

idxMFFL1 = list()

for(i in commonIndex){

  if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower2[edgeList[i,2],edgeList[i,3]]) |
  listAdj1$Apower2[edgeList[i,2],edgeList[i,3]] == 0)){
    MFFL1[i]=listAdj1$Apower2[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower3[edgeList[i,2],edgeList[i,3]]) |
  listAdj1$Apower3[edgeList[i,2],edgeList[i,3]] == 0)){
    MFFL1[i]=listAdj1$Apower3[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower4[edgeList[i,2],edgeList[i,3]]) |
  listAdj1$Apower4[edgeList[i,2],edgeList[i,3]] == 0)){
    MFFL1[i]=listAdj1$Apower4[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower5[edgeList[i,2],edgeList[i,3]]) |
  listAdj1$Apower5[edgeList[i,2],edgeList[i,3]] == 0)){
    MFFL1[i]=listAdj1$Apower5[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower6[edgeList[i,2],edgeList[i,3]]) |
  listAdj1$Apower6[edgeList[i,2],edgeList[i,3]] == 0)){
    MFFL1[i]=listAdj1$Apower6[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower7[edgeList[i,2],edgeList[i,3]]) |
  listAdj1$Apower7[edgeList[i,2],edgeList[i,3]] == 0)){
    MFFL1[i]=listAdj1$Apower7[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower8[edgeList[i,2],edgeList[i,3]]) |
  listAdj1$Apower8[edgeList[i,2],edgeList[i,3]] == 0)){
    MFFL1[i]=listAdj1$Apower8[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower9[edgeList[i,2],edgeList[i,3]]) |
  listAdj1$Apower9[edgeList[i,2],edgeList[i,3]] == 0)){
    MFFL1[i]=listAdj1$Apower9[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower10[edgeList[i,2],edgeList[i,3]]) |
  listAdj1$Apower10[edgeList[i,2],edgeList[i,3]] == 0)){
    MFFL1[i]=listAdj1$Apower10[edgeList[i,2],edgeList[i,3]]
  }
```

```

} else if(edgelist[i,4] == "1" &
!(is.na(listAdj1$Apower11[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower11[edgelist[i,2],edgelist[i,3]] == 0)){
  MFFL1[i]=listAdj1$Apower11[edgelist[i,2],edgelist[i,3]]
} else if(edgelist[i,4] == "1" &
!(is.na(listAdj1$Apower12[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower12[edgelist[i,2],edgelist[i,3]] == 0)){
  MFFL1[i]=listAdj1$Apower12[edgelist[i,2],edgelist[i,3]]
} else if(edgelist[i,4] == "1" &
!(is.na(listAdj1$Apower13[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower13[edgelist[i,2],edgelist[i,3]] == 0)){
  MFFL1[i]=listAdj1$Apower13[edgelist[i,2],edgelist[i,3]]
} else if(edgelist[i,4] == "1" &
!(is.na(listAdj1$Apower14[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower14[edgelist[i,2],edgelist[i,3]] == 0)){
  MFFL1[i]=listAdj1$Apower14[edgelist[i,2],edgelist[i,3]]
} else if(edgelist[i,4] == "1" &
!(is.na(listAdj1$Apower15[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower15[edgelist[i,2],edgelist[i,3]] == 0)){
  MFFL1[i]=listAdj1$Apower15[edgelist[i,2],edgelist[i,3]]
} else if(edgelist[i,4] == "1" &
!(is.na(listAdj1$Apower16[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower16[edgelist[i,2],edgelist[i,3]] == 0)){
  MFFL1[i]=listAdj1$Apower16[edgelist[i,2],edgelist[i,3]]
} else if(edgelist[i,4] == "1" &
!(is.na(listAdj1$Apower17[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower17[edgelist[i,2],edgelist[i,3]] == 0)){
  MFFL1[i]=listAdj1$Apower17[edgelist[i,2],edgelist[i,3]]
}

if( (edgelist[i,4] == "1" &
!(is.na(listAdj1$Apower2[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower2[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj1$Apower3[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower3[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj1$Apower4[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower4[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj1$Apower5[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower5[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj1$Apower6[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower6[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj1$Apower7[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower7[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj1$Apower8[edgelist[i,2],edgelist[i,3]]) |

```

```

listAdj1$Apower8[edgelist[i,2],edgelist[i,3]] == 0)) |
  (edgelist[i,4] == "1" &
  !(is.na(listAdj1$Apower9[edgelist[i,2],edgelist[i,3]])) |
  listAdj1$Apower9[edgelist[i,2],edgelist[i,3]] == 0)) |
  (edgelist[i,4] == "1" &
  !(is.na(listAdj1$Apower10[edgelist[i,2],edgelist[i,3]])) |
  listAdj1$Apower10[edgelist[i,2],edgelist[i,3]] == 0)) |
  (edgelist[i,4] == "1" &
  !(is.na(listAdj1$Apower11[edgelist[i,2],edgelist[i,3]])) |
  listAdj1$Apower11[edgelist[i,2],edgelist[i,3]] == 0)) |
  (edgelist[i,4] == "1" &
  !(is.na(listAdj1$Apower12[edgelist[i,2],edgelist[i,3]])) |
  listAdj1$Apower12[edgelist[i,2],edgelist[i,3]] == 0)) |
  (edgelist[i,4] == "1" &
  !(is.na(listAdj1$Apower13[edgelist[i,2],edgelist[i,3]])) |
  listAdj1$Apower13[edgelist[i,2],edgelist[i,3]] == 0)) |
  (edgelist[i,4] == "1" &
  !(is.na(listAdj1$Apower14[edgelist[i,2],edgelist[i,3]])) |
  listAdj1$Apower14[edgelist[i,2],edgelist[i,3]] == 0)) |
  (edgelist[i,4] == "1" &
  !(is.na(listAdj1$Apower15[edgelist[i,2],edgelist[i,3]])) |
  listAdj1$Apower15[edgelist[i,2],edgelist[i,3]] == 0)) |
  (edgelist[i,4] == "1" &
  !(is.na(listAdj1$Apower16[edgelist[i,2],edgelist[i,3]])) |
  listAdj1$Apower16[edgelist[i,2],edgelist[i,3]] == 0)) |
  (edgelist[i,4] == "1" &
  !(is.na(listAdj1$Apower17[edgelist[i,2],edgelist[i,3]])) |
  listAdj1$Apower17[edgelist[i,2],edgelist[i,3]] == 0))
){idxMFFL1[i] = i}

}

MFFL1 = unlist(MFFL1)

idxMFFL1 = unlist(idxMFFL1)

length(idxMFFL1)

## [1] 8416

```

The indices of edges taking part in MPFBL1 subgraph are saved at **idxMPFBL1** object through the following code.

```

MPFBL1 = list()

idxMPFBL1 = list()

```

```

for(i in commonIndex){

  if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower2[edgeList[i,3],edgeList[i,2]])) |
  listAdj1$Apower2[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL1[i]=listAdj1$Apower2[edgeList[i,3],edgeList[i,2]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower3[edgeList[i,3],edgeList[i,2]])) |
  listAdj1$Apower3[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL1[i]=listAdj1$Apower3[edgeList[i,3],edgeList[i,2]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower4[edgeList[i,3],edgeList[i,2]])) |
  listAdj1$Apower4[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL1[i]=listAdj1$Apower4[edgeList[i,3],edgeList[i,2]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower5[edgeList[i,3],edgeList[i,2]])) |
  listAdj1$Apower5[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL1[i]=listAdj1$Apower5[edgeList[i,3],edgeList[i,2]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower6[edgeList[i,3],edgeList[i,2]])) |
  listAdj1$Apower6[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL1[i]=listAdj1$Apower6[edgeList[i,3],edgeList[i,2]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower7[edgeList[i,3],edgeList[i,2]])) |
  listAdj1$Apower7[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL1[i]=listAdj1$Apower7[edgeList[i,3],edgeList[i,2]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower8[edgeList[i,3],edgeList[i,2]])) |
  listAdj1$Apower8[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL1[i]=listAdj1$Apower8[edgeList[i,3],edgeList[i,2]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower9[edgeList[i,3],edgeList[i,2]])) |
  listAdj1$Apower9[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL1[i]=listAdj1$Apower9[edgeList[i,3],edgeList[i,2]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower10[edgeList[i,3],edgeList[i,2]])) |
  listAdj1$Apower10[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL1[i]=listAdj1$Apower10[edgeList[i,3],edgeList[i,2]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower11[edgeList[i,3],edgeList[i,2]])) |
  listAdj1$Apower11[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL1[i]=listAdj1$Apower11[edgeList[i,3],edgeList[i,2]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower12[edgeList[i,3],edgeList[i,2]])) |
  listAdj1$Apower12[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL1[i]=listAdj1$Apower12[edgeList[i,3],edgeList[i,2]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj1$Apower13[edgeList[i,3],edgeList[i,2]])) |
  listAdj1$Apower13[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL1[i]=listAdj1$Apower13[edgeList[i,3],edgeList[i,2]]
  }
}

```



```

    } else if(edge1[i,4] == "1" &
!(is.na(listAdj1$Apower14[edge1[i,3],edge1[i,2]])) |
listAdj1$Apower14[edge1[i,3],edge1[i,2]] == 0)){
    MPFBL1[i]=listAdj1$Apower14[edge1[i,3],edge1[i,2]]
    } else if(edge1[i,4] == "1" &
!(is.na(listAdj1$Apower15[edge1[i,3],edge1[i,2]])) |
listAdj1$Apower15[edge1[i,3],edge1[i,2]] == 0)){
    MPFBL1[i]=listAdj1$Apower15[edge1[i,3],edge1[i,2]]
    } else if(edge1[i,4] == "1" &
!(is.na(listAdj1$Apower16[edge1[i,3],edge1[i,2]])) |
listAdj1$Apower16[edge1[i,3],edge1[i,2]] == 0)){
    MPFBL1[i]=listAdj1$Apower16[edge1[i,3],edge1[i,2]]
    } else if(edge1[i,4] == "1" &
!(is.na(listAdj1$Apower17[edge1[i,3],edge1[i,2]])) |
listAdj1$Apower17[edge1[i,3],edge1[i,2]] == 0)){
    MPFBL1[i]=listAdj1$Apower17[edge1[i,3],edge1[i,2]]
    }

    if( (edge1[i,4] == "1" &
!(is.na(listAdj1$Apower2[edge1[i,3],edge1[i,2]])) |
listAdj1$Apower2[edge1[i,3],edge1[i,2]] == 0)) |
(edge1[i,4] == "1" &
!(is.na(listAdj1$Apower3[edge1[i,3],edge1[i,2]])) |
listAdj1$Apower3[edge1[i,3],edge1[i,2]] == 0)) |
(edge1[i,4] == "1" &
!(is.na(listAdj1$Apower4[edge1[i,3],edge1[i,2]])) |
listAdj1$Apower4[edge1[i,3],edge1[i,2]] == 0)) |
(edge1[i,4] == "1" &
!(is.na(listAdj1$Apower5[edge1[i,3],edge1[i,2]])) |
listAdj1$Apower5[edge1[i,3],edge1[i,2]] == 0)) |
(edge1[i,4] == "1" &
!(is.na(listAdj1$Apower6[edge1[i,3],edge1[i,2]])) |
listAdj1$Apower6[edge1[i,3],edge1[i,2]] == 0)) |
(edge1[i,4] == "1" &
!(is.na(listAdj1$Apower7[edge1[i,3],edge1[i,2]])) |
listAdj1$Apower7[edge1[i,3],edge1[i,2]] == 0)) |
(edge1[i,4] == "1" &
!(is.na(listAdj1$Apower8[edge1[i,3],edge1[i,2]])) |
listAdj1$Apower8[edge1[i,3],edge1[i,2]] == 0)) |
(edge1[i,4] == "1" &
!(is.na(listAdj1$Apower9[edge1[i,3],edge1[i,2]])) |
listAdj1$Apower9[edge1[i,3],edge1[i,2]] == 0)) |
(edge1[i,4] == "1" &
!(is.na(listAdj1$Apower10[edge1[i,3],edge1[i,2]])) |
listAdj1$Apower10[edge1[i,3],edge1[i,2]] == 0)) |
(edge1[i,4] == "1" &
!(is.na(listAdj1$Apower11[edge1[i,3],edge1[i,2]])) |
listAdj1$Apower11[edge1[i,3],edge1[i,2]] == 0)) |
(edge1[i,4] == "1" &
!(is.na(listAdj1$Apower12[edge1[i,3],edge1[i,2]])) |

```

```

listAdj1$Apower12[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "1" &
  !(is.na(listAdj1$Apower13[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower13[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "1" &
  !(is.na(listAdj1$Apower14[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower14[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "1" &
  !(is.na(listAdj1$Apower15[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower15[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "1" &
  !(is.na(listAdj1$Apower16[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower16[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "1" &
  !(is.na(listAdj1$Apower17[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower17[edgelist[i,3],edgelist[i,2]] == 0))
  ){idxMPFBL1[i] = i}

}

MPFBL1 = unlist(MPFBL1)

idxMPFBL1 = unlist(idxMPFBL1)
length(idxMPFBL1)

length(idxMPFBL1)

## [1] 2818

```

The indices of edges taking part in MNFFL2 subgraph are stored at **idxMNFFL2** object through the following code.

```

MNFFL2 = list()

idxMNFFL2 = list()

for(i in commonIndex){
  if(edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower2[edgelist[i,2],edgelist[i,3]])) |
  listAdj1$Apower2[edgelist[i,2],edgelist[i,3]] == 0)){
    MNFFL2[i]=listAdj1$Apower2[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower3[edgelist[i,2],edgelist[i,3]])) |
  listAdj1$Apower3[edgelist[i,2],edgelist[i,3]] == 0)){

```

```

    MNFFL2[i]=listAdj1$Apower3[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower4[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower4[edgelist[i,2],edgelist[i,3]] == 0)){
    MNFFL2[i]=listAdj1$Apower4[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower5[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower5[edgelist[i,2],edgelist[i,3]] == 0)){
    MNFFL2[i]=listAdj1$Apower5[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower6[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower6[edgelist[i,2],edgelist[i,3]] == 0)){
    MNFFL2[i]=listAdj1$Apower6[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower7[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower7[edgelist[i,2],edgelist[i,3]] == 0)){
    MNFFL2[i]=listAdj1$Apower7[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower8[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower8[edgelist[i,2],edgelist[i,3]] == 0)){
    MNFFL2[i]=listAdj1$Apower8[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower9[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower9[edgelist[i,2],edgelist[i,3]] == 0)){
    MNFFL2[i]=listAdj1$Apower9[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower10[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower10[edgelist[i,2],edgelist[i,3]] == 0)){
    MNFFL2[i]=listAdj1$Apower10[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower11[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower11[edgelist[i,2],edgelist[i,3]] == 0)){
    MNFFL2[i]=listAdj1$Apower11[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower12[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower12[edgelist[i,2],edgelist[i,3]] == 0)){
    MNFFL2[i]=listAdj1$Apower12[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower13[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower13[edgelist[i,2],edgelist[i,3]] == 0)){
    MNFFL2[i]=listAdj1$Apower13[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower14[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower14[edgelist[i,2],edgelist[i,3]] == 0)){
    MNFFL2[i]=listAdj1$Apower14[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower15[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower15[edgelist[i,2],edgelist[i,3]] == 0)){
    MNFFL2[i]=listAdj1$Apower15[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &

```

```

!(is.na(listAdj1$Apower16[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower16[edgelist[i,2],edgelist[i,3]] == 0)){
  MNFFL2[i]=listAdj1$Apower16[edgelist[i,2],edgelist[i,3]]
} else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower17[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower17[edgelist[i,2],edgelist[i,3]] == 0)){
  MNFFL2[i]=listAdj1$Apower17[edgelist[i,2],edgelist[i,3]]
}

  if( (edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower2[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower2[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower3[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower3[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower4[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower4[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower5[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower5[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower6[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower6[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower7[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower7[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower8[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower8[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower9[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower9[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower10[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower10[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower11[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower11[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower12[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower12[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower13[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower13[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower14[edgelist[i,2],edgelist[i,3]]) |
listAdj1$Apower14[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower15[edgelist[i,2],edgelist[i,3]]) |

```

```

listAdj1$Apower15[edgelist[i,2],edgelist[i,3]] == 0)) |
  (edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower16[edgelist[i,2],edgelist[i,3]])) |
  listAdj1$Apower16[edgelist[i,2],edgelist[i,3]] == 0)) |
  (edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower17[edgelist[i,2],edgelist[i,3]])) |
  listAdj1$Apower17[edgelist[i,2],edgelist[i,3]] == 0))
  ){idxMNFFL2[i] = i}

}

MNFFL2 = unlist(MNFFL2)

idxMNFFL2 = unlist(idxMNFFL2)

length(idxMNFFL2)

## [1] 659

```

The indices of edges taking part in MNFBL2 subgraph are stored at **idxMNFBL2** object through the following code.

```

MNFBL2 = list()

idxMNFBL2 = list()

for(i in commonIndex){

  if(edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower2[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower2[edgelist[i,3],edgelist[i,2]] == 0)){
    MNFBL2[i]=listAdj1$Apower2[edgelist[i,3],edgelist[i,2]]
  } else if(edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower3[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower3[edgelist[i,3],edgelist[i,2]] == 0)){
    MNFBL2[i]=listAdj1$Apower3[edgelist[i,3],edgelist[i,2]]
  } else if(edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower4[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower4[edgelist[i,3],edgelist[i,2]] == 0)){
    MNFBL2[i]=listAdj1$Apower4[edgelist[i,3],edgelist[i,2]]
  } else if(edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower5[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower5[edgelist[i,3],edgelist[i,2]] == 0)){
    MNFBL2[i]=listAdj1$Apower5[edgelist[i,3],edgelist[i,2]]
  } else if(edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower6[edgelist[i,3],edgelist[i,2]])) |

```

```

listAdj1$Apower6[edgelist[i,3],edgelist[i,2]] == 0)){
  MNFBL2[i]=listAdj1$Apower6[edgelist[i,3],edgelist[i,2]]
} else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower7[edgelist[i,3],edgelist[i,2]])) |
listAdj1$Apower7[edgelist[i,3],edgelist[i,2]] == 0)){
  MNFBL2[i]=listAdj1$Apower7[edgelist[i,3],edgelist[i,2]]
} else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower8[edgelist[i,3],edgelist[i,2]])) |
listAdj1$Apower8[edgelist[i,3],edgelist[i,2]] == 0)){
  MNFBL2[i]=listAdj1$Apower8[edgelist[i,3],edgelist[i,2]]
} else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower9[edgelist[i,3],edgelist[i,2]])) |
listAdj1$Apower9[edgelist[i,3],edgelist[i,2]] == 0)){
  MNFBL2[i]=listAdj1$Apower9[edgelist[i,3],edgelist[i,2]]
} else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower10[edgelist[i,3],edgelist[i,2]])) |
listAdj1$Apower10[edgelist[i,3],edgelist[i,2]] == 0)){
  MNFBL2[i]=listAdj1$Apower10[edgelist[i,3],edgelist[i,2]]
} else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower11[edgelist[i,3],edgelist[i,2]])) |
listAdj1$Apower11[edgelist[i,3],edgelist[i,2]] == 0)){
  MNFBL2[i]=listAdj1$Apower11[edgelist[i,3],edgelist[i,2]]
} else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower12[edgelist[i,3],edgelist[i,2]])) |
listAdj1$Apower12[edgelist[i,3],edgelist[i,2]] == 0)){
  MNFBL2[i]=listAdj1$Apower12[edgelist[i,3],edgelist[i,2]]
} else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower13[edgelist[i,3],edgelist[i,2]])) |
listAdj1$Apower13[edgelist[i,3],edgelist[i,2]] == 0)){
  MNFBL2[i]=listAdj1$Apower13[edgelist[i,3],edgelist[i,2]]
} else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower14[edgelist[i,3],edgelist[i,2]])) |
listAdj1$Apower14[edgelist[i,3],edgelist[i,2]] == 0)){
  MNFBL2[i]=listAdj1$Apower14[edgelist[i,3],edgelist[i,2]]
} else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower15[edgelist[i,3],edgelist[i,2]])) |
listAdj1$Apower15[edgelist[i,3],edgelist[i,2]] == 0)){
  MNFBL2[i]=listAdj1$Apower15[edgelist[i,3],edgelist[i,2]]
} else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower16[edgelist[i,3],edgelist[i,2]])) |
listAdj1$Apower16[edgelist[i,3],edgelist[i,2]] == 0)){
  MNFBL2[i]=listAdj1$Apower16[edgelist[i,3],edgelist[i,2]]
} else if(edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower17[edgelist[i,3],edgelist[i,2]])) |
listAdj1$Apower17[edgelist[i,3],edgelist[i,2]] == 0)){
  MNFBL2[i]=listAdj1$Apower17[edgelist[i,3],edgelist[i,2]]
}

if( (edgelist[i,4] == "-1" &
!(is.na(listAdj1$Apower2[edgelist[i,3],edgelist[i,2]])) |

```

```

listAdj1$Apower2[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower3[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower3[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower4[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower4[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower5[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower5[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower6[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower6[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower7[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower7[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower8[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower8[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower9[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower9[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower10[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower10[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower11[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower11[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower12[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower12[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower13[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower13[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower14[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower14[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower15[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower15[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower16[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower16[edgelist[i,3],edgelist[i,2]] == 0)) |
  (edgelist[i,4] == "-1" &
  !(is.na(listAdj1$Apower17[edgelist[i,3],edgelist[i,2]])) |
  listAdj1$Apower17[edgelist[i,3],edgelist[i,2]] == 0))
){idxMNFB2[i] = i}
}

```



```

MNFBFL2 = unlist(MNFBFL2)

idxMNFBFL2 = unlist(idxMNFBFL2)

length(idxMNFBFL2)

## [1] 430

```

Now it's time to compute the number of edges involved in multiple-edge subgraphs (table 2).

### MFFL1

```

MFFL1.pval.cor1 = 0
MFFL1.pval.cor.NA = 0
MFFL1.pval.cor2 = 0

for(i in idxMFFL1){
  logic = c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){
      logic[j]=
!any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))
    }
    if(all(logic)){
      logic1 = c()
      logic2 = c()
      for(j in 1:SignalingNet[[i]]$length){

        logic1[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor > 0
        logic2[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor < 0
      }
      if(all(logic1)) {MFFL1.pval.cor1=MFFL1.pval.cor1+1}
      if(all(logic2)) {MFFL1.pval.cor2=MFFL1.pval.cor2+1}
    }
  } else{
    if(!any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na)))){
      a = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor > 0
    }
  }
}

```



```

        b = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor < 0
        if(a){MFFL1.pval.cor1=MFFL1.pval.cor1+1}
        if(b){MFFL1.pval.cor2=MFFL1.pval.cor2+1}
    }
}
}

for(i in idxMFFL1){
  logic = c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){

      logic[j]=any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na))) |
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval > 0.05
    }
    if(all(logic)){ MFFL1.pval.cor.NA = MFFL1.pval.cor.NA + 1 }

  }else{ if(any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na))) |
SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval > 0.05){
    index.pval.cor.NA = MFFL1.pval.cor.NA + 1}
  }
}

MFFL1.pval.cor1
## [1] 537

MFFL1.pval.cor.NA
## [1] 3061

MFFL1.pval.cor2
## [1] 270

length(idxMFFL1) - (MFFL1.pval.cor1 + MFFL1.pval.cor.NA + MFFL1.pval.cor2)
## [1] 4548

```

## MNFBL2

```

MNFBL2.pval.cor1 = 0
MNFBL2.pval.cor.NA = 0
MNFBL2.pval.cor2 = 0

```

```

for(i in idxMNFBL2){
  logic = c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){
      logic[j]=
!any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))
    }
    if(all(logic)){
      logic1 = c()
      logic2 = c()
      for(j in 1:SignalingNet[[i]]$length){

        logic1[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor > 0
        logic2[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor < 0
      }
      if(all(logic1)) {MNFBL2.pval.cor1=MNFBL2.pval.cor1+1}
      if(all(logic2)) {MNFBL2.pval.cor2=MNFBL2.pval.cor2+1}
    }
  } else{
    if(!any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na)))){
      a = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor > 0
      b = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor < 0
      if(a){MNFBL2.pval.cor1=MNFBL2.pval.cor1+1}
      if(b){MNFBL2.pval.cor2=MNFBL2.pval.cor2+1}
    }
  }
}

for(i in idxMNFBL2){
  logic = c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){

      logic[j]=any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))
| SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval > 0.05

```

```

    }
    if(all(logic)){ MNFBL2.pval.cor.NA = MNFBL2.pval.cor.NA + 1 }

    }else{ if(any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na))) |
SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval > 0.05){
    index.pval.cor.NA = MNFBL2.pval.cor.NA + 1}
    }
}

MNFBL2.pval.cor1

## [1] 24

MNFBL2.pval.cor.NA

## [1] 181

MNFBL2.pval.cor2

## [1] 13

length(idXMNFBL2) - (MNFBL2.pval.cor1 + MNFBL2.pval.cor.NA +
MNFBL2.pval.cor2)

## [1] 212

```

## MNFFL2

```

MNFFL2.pval.cor1 = 0
MNFFL2.pval.cor.NA = 0
MNFFL2.pval.cor2 = 0

for(i in idxMNFFL2){
  logic = c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){
      logic[j]=
!any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))
    }
    if(all(logic)){
      logic1 = c()
      logic2 = c()
      for(j in 1:SignalingNet[[i]]$length){

        logic1[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &

```

```

SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor > 0
  logic2[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor < 0
  }
  if(all(logic1)) {MNFFL2.pval.cor1=MNFFL2.pval.cor1+1}
  if(all(logic2)) {MNFFL2.pval.cor2=MNFFL2.pval.cor2+1}
  }
} else{
  if(!any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na)))){
    a = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor > 0
    b = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor < 0
    if(a){MNFFL2.pval.cor1=MNFFL2.pval.cor1+1}
    if(b){MNFFL2.pval.cor2=MNFFL2.pval.cor2+1}
  }
}
}

for(i in idxMNFFL2){
  logic = c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){

      logic[j]=any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))
| SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval > 0.05
    }
    if(all(logic)){ MNFFL2.pval.cor.NA = MNFFL2.pval.cor.NA + 1 }

  }else{ if(any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na))))|
SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval > 0.05){
    index.pval.cor.NA = MNFFL2.pval.cor.NA + 1}
  }
}

MNFFL2.pval.cor1
## [1] 43

MNFFL2.pval.cor.NA
## [1] 254

MNFFL2.pval.cor2
## [1] 27

```

```
length(idxMNFFL2) - (MNFFL2.pval.cor1 + MNFFL2.pval.cor.NA +
MNFFL2.pval.cor2)
```

```
## [1] 335
```

## MPFBL1

```
MPFBL1.pval.cor1 = 0
```

```
MPFBL1.pval.cor.NA = 0
```

```
MPFBL1.pval.cor2 = 0
```

```
for(i in idxMPFBL1){
  logic = c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){
      logic[j]=
!any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))
    }
    if(all(logic)){
      logic1 = c()
      logic2 = c()
      for(j in 1:SignalingNet[[i]]$length){

        logic1[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor > 0
        logic2[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor < 0
      }
      if(all(logic1)) {MPFBL1.pval.cor1=MPFBL1.pval.cor1+1}
      if(all(logic2)) {MPFBL1.pval.cor2=MPFBL1.pval.cor2+1}
    }
  } else{
    if(!any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na)))){
      a = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor > 0
      b = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor < 0
      if(a){MPFBL1.pval.cor1=MPFBL1.pval.cor1+1}
      if(b){MPFBL1.pval.cor2=MPFBL1.pval.cor2+1}
    }
  }
}
```

```

for(i in idxMPFBL1){
  logic = c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){

      logic[j]=any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))
| SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval > 0.05
    }
    if(all(logic)){ MPFBL1.pval.cor.NA = MPFBL1.pval.cor.NA + 1 }

  }else{ if(any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na))) |
SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval > 0.05){
    index.pval.cor.NA = MPFBL1.pval.cor.NA + 1}
  }
}

MPFBL1.pval.cor1
## [1] 224

MPFBL1.pval.cor.NA
## [1] 916

MPFBL1.pval.cor2
## [1] 85

length(idxMPFBL1) - (MPFBL1.pval.cor1 + MPFBL1.pval.cor.NA +
MPFBL1.pval.cor2)
## [1] 1593

```

## 7.2.Computing the number of edges participating in MNFBL1, MPFBL2, MFFL2 and MNFFL1 multiple-edge subgraphs

The indices of edges which are involved in MNFBL1 subgraph are stored at **idxMNFBL1** object through the following code.

```
MNFBL1= list()
```

```

idxMNFBL1= list()

for(i in commonIndex){

  if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower2[edgeList[i,2],edgeList[i,3]]) |
listAdj2$Apower2[edgeList[i,2],edgeList[i,3]] == 0)){
    MNFBL1[i]=listAdj2$Apower2[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower3[edgeList[i,2],edgeList[i,3]]) |
listAdj2$Apower3[edgeList[i,2],edgeList[i,3]] == 0)){
    MNFBL1[i]=listAdj2$Apower3[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower4[edgeList[i,2],edgeList[i,3]]) |
listAdj2$Apower4[edgeList[i,2],edgeList[i,3]] == 0)){
    MNFBL1[i]=listAdj2$Apower4[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower5[edgeList[i,2],edgeList[i,3]]) |
listAdj2$Apower5[edgeList[i,2],edgeList[i,3]] == 0)){
    MNFBL1[i]=listAdj2$Apower5[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower6[edgeList[i,2],edgeList[i,3]]) |
listAdj2$Apower6[edgeList[i,2],edgeList[i,3]] == 0)){
    MNFBL1[i]=listAdj2$Apower6[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower7[edgeList[i,2],edgeList[i,3]]) |
listAdj2$Apower7[edgeList[i,2],edgeList[i,3]] == 0)){
    MNFBL1[i]=listAdj2$Apower7[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower8[edgeList[i,2],edgeList[i,3]]) |
listAdj2$Apower8[edgeList[i,2],edgeList[i,3]] == 0)){
    MNFBL1[i]=listAdj2$Apower8[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower9[edgeList[i,2],edgeList[i,3]]) |
listAdj2$Apower9[edgeList[i,2],edgeList[i,3]] == 0)){
    MNFBL1[i]=listAdj2$Apower9[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower10[edgeList[i,2],edgeList[i,3]]) |
listAdj2$Apower10[edgeList[i,2],edgeList[i,3]] == 0)){
    MNFBL1[i]=listAdj2$Apower10[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower11[edgeList[i,2],edgeList[i,3]]) |
listAdj2$Apower11[edgeList[i,2],edgeList[i,3]] == 0)){
    MNFBL1[i]=listAdj2$Apower11[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower12[edgeList[i,2],edgeList[i,3]]) |
listAdj2$Apower12[edgeList[i,2],edgeList[i,3]] == 0)){
    MNFBL1[i]=listAdj2$Apower12[edgeList[i,2],edgeList[i,3]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower13[edgeList[i,2],edgeList[i,3]]) |

```

```

listAdj2$Apower13[edgelist[i,2],edgelist[i,3]] == 0)){
  MNFBL1[i]=listAdj2$Apower13[edgelist[i,2],edgelist[i,3]]
} else if(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower14[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower14[edgelist[i,2],edgelist[i,3]] == 0)){
  MNFBL1[i]=listAdj2$Apower14[edgelist[i,2],edgelist[i,3]]
} else if(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower15[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower15[edgelist[i,2],edgelist[i,3]] == 0)){
  MNFBL1[i]=listAdj2$Apower15[edgelist[i,2],edgelist[i,3]]
} else if(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower16[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower16[edgelist[i,2],edgelist[i,3]] == 0)){
  MNFBL1[i]=listAdj2$Apower16[edgelist[i,2],edgelist[i,3]]
} else if(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower17[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower17[edgelist[i,2],edgelist[i,3]] == 0)){
  MNFBL1[i]=listAdj2$Apower17[edgelist[i,2],edgelist[i,3]]
}

  if( (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower2[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower2[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower3[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower3[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower4[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower4[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower5[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower5[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower6[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower6[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower7[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower7[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower8[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower8[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower9[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower9[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower10[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower10[edgelist[i,2],edgelist[i,3]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower11[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower11[edgelist[i,2],edgelist[i,3]] == 0)) |

```



```

      (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower12[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower12[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower13[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower13[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower14[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower14[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower15[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower15[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower16[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower16[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower17[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower17[edgelist[i,2],edgelist[i,3]] == 0))
    ){idxMNFBL1[i] = i}
}

MNFBL1 = unlist(MNFBL1)

idxMNFBL1 = unlist(idxMNFBL1)

length(idxMNFBL1)

## [1] 11532

```

The indices of edges involved in MPFBL2 subgraph are stored at **idxMPFBL2** object through the following code.

```

MPFBL2= list()

idxMPFBL2 = list()

for(i in commonIndex){
  if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower2[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower2[edgelist[i,3],edgelist[i,2]] == 0)){
    MPFBL2[i]=listAdj2$Apower2[edgelist[i,3],edgelist[i,2]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower3[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower3[edgelist[i,3],edgelist[i,2]] == 0)){
    MPFBL2[i]=listAdj2$Apower3[edgelist[i,3],edgelist[i,2]]
  }
}

```

```

    } else if(edgeList[i,4] == "-1" &
!(is.na(listAdj2$Apower4[edgeList[i,3],edgeList[i,2]])) |
listAdj2$Apower4[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL2[i]=listAdj2$Apower4[edgeList[i,3],edgeList[i,2]]
    } else if(edgeList[i,4] == "-1" &
!(is.na(listAdj2$Apower5[edgeList[i,3],edgeList[i,2]])) |
listAdj2$Apower5[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL2[i]=listAdj2$Apower5[edgeList[i,3],edgeList[i,2]]
    } else if(edgeList[i,4] == "-1" &
!(is.na(listAdj2$Apower6[edgeList[i,3],edgeList[i,2]])) |
listAdj2$Apower6[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL2[i]=listAdj2$Apower6[edgeList[i,3],edgeList[i,2]]
    } else if(edgeList[i,4] == "-1" &
!(is.na(listAdj2$Apower7[edgeList[i,3],edgeList[i,2]])) |
listAdj2$Apower7[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL2[i]=listAdj2$Apower7[edgeList[i,3],edgeList[i,2]]
    } else if(edgeList[i,4] == "-1" &
!(is.na(listAdj2$Apower8[edgeList[i,3],edgeList[i,2]])) |
listAdj2$Apower8[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL2[i]=listAdj2$Apower8[edgeList[i,3],edgeList[i,2]]
    } else if(edgeList[i,4] == "-1" &
!(is.na(listAdj2$Apower9[edgeList[i,3],edgeList[i,2]])) |
listAdj2$Apower9[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL2[i]=listAdj2$Apower9[edgeList[i,3],edgeList[i,2]]
    } else if(edgeList[i,4] == "-1" &
!(is.na(listAdj2$Apower10[edgeList[i,3],edgeList[i,2]])) |
listAdj2$Apower10[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL2[i]=listAdj2$Apower10[edgeList[i,3],edgeList[i,2]]
    } else if(edgeList[i,4] == "-1" &
!(is.na(listAdj2$Apower11[edgeList[i,3],edgeList[i,2]])) |
listAdj2$Apower11[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL2[i]=listAdj2$Apower11[edgeList[i,3],edgeList[i,2]]
    } else if(edgeList[i,4] == "-1" &
!(is.na(listAdj2$Apower12[edgeList[i,3],edgeList[i,2]])) |
listAdj2$Apower12[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL2[i]=listAdj2$Apower12[edgeList[i,3],edgeList[i,2]]
    } else if(edgeList[i,4] == "-1" &
!(is.na(listAdj2$Apower13[edgeList[i,3],edgeList[i,2]])) |
listAdj2$Apower13[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL2[i]=listAdj2$Apower13[edgeList[i,3],edgeList[i,2]]
    } else if(edgeList[i,4] == "-1" &
!(is.na(listAdj2$Apower14[edgeList[i,3],edgeList[i,2]])) |
listAdj2$Apower14[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL2[i]=listAdj2$Apower14[edgeList[i,3],edgeList[i,2]]
    } else if(edgeList[i,4] == "-1" &
!(is.na(listAdj2$Apower15[edgeList[i,3],edgeList[i,2]])) |
listAdj2$Apower15[edgeList[i,3],edgeList[i,2]] == 0)){
    MPFBL2[i]=listAdj2$Apower15[edgeList[i,3],edgeList[i,2]]
    } else if(edgeList[i,4] == "-1" &
!(is.na(listAdj2$Apower16[edgeList[i,3],edgeList[i,2]])) |

```

```

listAdj2$Apower16[edgelist[i,3],edgelist[i,2]] == 0)){
  MPFBL2[i]=listAdj2$Apower16[edgelist[i,3],edgelist[i,2]]
} else if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower17[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower17[edgelist[i,3],edgelist[i,2]] == 0)){
  MPFBL2[i]=listAdj2$Apower17[edgelist[i,3],edgelist[i,2]]
}

  if( (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower2[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower2[edgelist[i,3],edgelist[i,2]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower3[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower3[edgelist[i,3],edgelist[i,2]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower4[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower4[edgelist[i,3],edgelist[i,2]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower5[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower5[edgelist[i,3],edgelist[i,2]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower6[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower6[edgelist[i,3],edgelist[i,2]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower7[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower7[edgelist[i,3],edgelist[i,2]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower8[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower8[edgelist[i,3],edgelist[i,2]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower9[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower9[edgelist[i,3],edgelist[i,2]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower10[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower10[edgelist[i,3],edgelist[i,2]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower11[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower11[edgelist[i,3],edgelist[i,2]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower12[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower12[edgelist[i,3],edgelist[i,2]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower13[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower13[edgelist[i,3],edgelist[i,2]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower14[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower14[edgelist[i,3],edgelist[i,2]] == 0)) |
(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower15[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower15[edgelist[i,3],edgelist[i,2]] == 0)) |

```

```

      (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower16[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower16[edgelist[i,3],edgelist[i,2]] == 0)) |
      (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower17[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower17[edgelist[i,3],edgelist[i,2]] == 0))
    ){idxMPFBL2[i] = i}

}

MPFBL2 = unlist(MPFBL2)

idxMPFBL2 = unlist(idxMPFBL2)

length(idxMPFBL2)

## [1] 973

```

The indices of edges involved in MFFL2 subgraph are stored at **idxMFFL2** object through the following code.

```

MFFL2 = list()

idxMFFL2 = list()

for(i in commonIndex){

  if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower2[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower2[edgelist[i,2],edgelist[i,3]] == 0)){
    MFFL2[i]=listAdj2$Apower2[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower3[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower3[edgelist[i,2],edgelist[i,3]] == 0)){
    MFFL2[i]=listAdj2$Apower3[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower4[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower4[edgelist[i,2],edgelist[i,3]] == 0)){
    MFFL2[i]=listAdj2$Apower4[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower5[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower5[edgelist[i,2],edgelist[i,3]] == 0)){
    MFFL2[i]=listAdj2$Apower5[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower6[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower6[edgelist[i,2],edgelist[i,3]] == 0)){

```

```

    MFFL2[i]=listAdj2$Apower6[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower7[edgelist[i,2],edgelist[i,3]]) |
listAdj2$Apower7[edgelist[i,2],edgelist[i,3]] == 0)){
    MFFL2[i]=listAdj2$Apower7[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower8[edgelist[i,2],edgelist[i,3]]) |
listAdj2$Apower8[edgelist[i,2],edgelist[i,3]] == 0)){
    MFFL2[i]=listAdj2$Apower8[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower9[edgelist[i,2],edgelist[i,3]]) |
listAdj2$Apower9[edgelist[i,2],edgelist[i,3]] == 0)){
    MFFL2[i]=listAdj2$Apower9[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower10[edgelist[i,2],edgelist[i,3]]) |
listAdj2$Apower10[edgelist[i,2],edgelist[i,3]] == 0)){
    MFFL2[i]=listAdj2$Apower10[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower11[edgelist[i,2],edgelist[i,3]]) |
listAdj2$Apower11[edgelist[i,2],edgelist[i,3]] == 0)){
    MFFL2[i]=listAdj2$Apower11[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower12[edgelist[i,2],edgelist[i,3]]) |
listAdj2$Apower12[edgelist[i,2],edgelist[i,3]] == 0)){
    MFFL2[i]=listAdj2$Apower12[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower13[edgelist[i,2],edgelist[i,3]]) |
listAdj2$Apower13[edgelist[i,2],edgelist[i,3]] == 0)){
    MFFL2[i]=listAdj2$Apower13[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower14[edgelist[i,2],edgelist[i,3]]) |
listAdj2$Apower14[edgelist[i,2],edgelist[i,3]] == 0)){
    MFFL2[i]=listAdj2$Apower14[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower15[edgelist[i,2],edgelist[i,3]]) |
listAdj2$Apower15[edgelist[i,2],edgelist[i,3]] == 0)){
    MFFL2[i]=listAdj2$Apower15[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower16[edgelist[i,2],edgelist[i,3]]) |
listAdj2$Apower16[edgelist[i,2],edgelist[i,3]] == 0)){
    MFFL2[i]=listAdj2$Apower16[edgelist[i,2],edgelist[i,3]]
  } else if(edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower17[edgelist[i,2],edgelist[i,3]]) |
listAdj2$Apower17[edgelist[i,2],edgelist[i,3]] == 0)){
    MFFL2[i]=listAdj2$Apower17[edgelist[i,2],edgelist[i,3]]
  }
}

if( (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower2[edgelist[i,2],edgelist[i,3]]) |
listAdj2$Apower2[edgelist[i,2],edgelist[i,3]] == 0)) |

```

```

      (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower3[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower3[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower4[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower4[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower5[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower5[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower6[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower6[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower7[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower7[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower8[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower8[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower9[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower9[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower10[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower10[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower11[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower11[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower12[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower12[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower13[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower13[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower14[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower14[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower15[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower15[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower16[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower16[edgelist[i,2],edgelist[i,3]] == 0)) |
      (edgelist[i,4] == "-1" &
!(is.na(listAdj2$Apower17[edgelist[i,2],edgelist[i,3]])) |
listAdj2$Apower17[edgelist[i,2],edgelist[i,3]] == 0))
    ){idxMFFL2[i] = i}
}

```

```

MFFL2 = unlist(MFFL2)

```

```

idxMFFL2 = unlist(idxMFFL2)

length(idxMFFL2)

## [1] 2608

```

The indices of edges involved in MNFFL1 subgraph are stored at **MNFFL1** object through the following code.

```

MNFFL1 = list()

idxMNFFL1 = list()

for(i in commonIndex){

  if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower2[edgeList[i,3],edgeList[i,2]])) |
  listAdj2$Apower2[edgeList[i,3],edgeList[i,2]] == 0)){
    MNFFL1[i]=listAdj2$Apower2[edgeList[i,3],edgeList[i,2]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower3[edgeList[i,3],edgeList[i,2]])) |
  listAdj2$Apower3[edgeList[i,3],edgeList[i,2]] == 0)){
    MNFFL1[i]=listAdj2$Apower3[edgeList[i,3],edgeList[i,2]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower4[edgeList[i,3],edgeList[i,2]])) |
  listAdj2$Apower4[edgeList[i,3],edgeList[i,2]] == 0)){
    MNFFL1[i]=listAdj2$Apower4[edgeList[i,3],edgeList[i,2]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower5[edgeList[i,3],edgeList[i,2]])) |
  listAdj2$Apower5[edgeList[i,3],edgeList[i,2]] == 0)){
    MNFFL1[i]=listAdj2$Apower5[edgeList[i,3],edgeList[i,2]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower6[edgeList[i,3],edgeList[i,2]])) |
  listAdj2$Apower6[edgeList[i,3],edgeList[i,2]] == 0)){
    MNFFL1[i]=listAdj2$Apower6[edgeList[i,3],edgeList[i,2]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower7[edgeList[i,3],edgeList[i,2]])) |
  listAdj2$Apower7[edgeList[i,3],edgeList[i,2]] == 0)){
    MNFFL1[i]=listAdj2$Apower7[edgeList[i,3],edgeList[i,2]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower8[edgeList[i,3],edgeList[i,2]])) |
  listAdj2$Apower8[edgeList[i,3],edgeList[i,2]] == 0)){
    MNFFL1[i]=listAdj2$Apower8[edgeList[i,3],edgeList[i,2]]
  } else if(edgeList[i,4] == "1" &
  !(is.na(listAdj2$Apower9[edgeList[i,3],edgeList[i,2]])) |
  listAdj2$Apower9[edgeList[i,3],edgeList[i,2]] == 0)){

```



```

    MNFFL1[i]=listAdj2$Apower9[edgelist[i,3],edgelist[i,2]]
  } else if(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower10[edgelist[i,3],edgelist[i,2]]) |
listAdj2$Apower10[edgelist[i,3],edgelist[i,2]] == 0)){
    MNFFL1[i]=listAdj2$Apower10[edgelist[i,3],edgelist[i,2]]
  } else if(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower11[edgelist[i,3],edgelist[i,2]]) |
listAdj2$Apower11[edgelist[i,3],edgelist[i,2]] == 0)){
    MNFFL1[i]=listAdj2$Apower11[edgelist[i,3],edgelist[i,2]]
  } else if(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower12[edgelist[i,3],edgelist[i,2]]) |
listAdj2$Apower12[edgelist[i,3],edgelist[i,2]] == 0)){
    MNFFL1[i]=listAdj2$Apower12[edgelist[i,3],edgelist[i,2]]
  } else if(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower13[edgelist[i,3],edgelist[i,2]]) |
listAdj2$Apower13[edgelist[i,3],edgelist[i,2]] == 0)){
    MNFFL1[i]=listAdj2$Apower13[edgelist[i,3],edgelist[i,2]]
  } else if(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower14[edgelist[i,3],edgelist[i,2]]) |
listAdj2$Apower14[edgelist[i,3],edgelist[i,2]] == 0)){
    MNFFL1[i]=listAdj2$Apower14[edgelist[i,3],edgelist[i,2]]
  } else if(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower15[edgelist[i,3],edgelist[i,2]]) |
listAdj2$Apower15[edgelist[i,3],edgelist[i,2]] == 0)){
    MNFFL1[i]=listAdj2$Apower15[edgelist[i,3],edgelist[i,2]]
  } else if(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower16[edgelist[i,3],edgelist[i,2]]) |
listAdj2$Apower16[edgelist[i,3],edgelist[i,2]] == 0)){
    MNFFL1[i]=listAdj2$Apower16[edgelist[i,3],edgelist[i,2]]
  } else if(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower17[edgelist[i,3],edgelist[i,2]]) |
listAdj2$Apower17[edgelist[i,3],edgelist[i,2]] == 0)){
    MNFFL1[i]=listAdj2$Apower17[edgelist[i,3],edgelist[i,2]]
  }

  if( (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower2[edgelist[i,3],edgelist[i,2]]) |
listAdj2$Apower2[edgelist[i,3],edgelist[i,2]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower3[edgelist[i,3],edgelist[i,2]]) |
listAdj2$Apower3[edgelist[i,3],edgelist[i,2]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower4[edgelist[i,3],edgelist[i,2]]) |
listAdj2$Apower4[edgelist[i,3],edgelist[i,2]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower5[edgelist[i,3],edgelist[i,2]]) |
listAdj2$Apower5[edgelist[i,3],edgelist[i,2]] == 0)) |
(edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower6[edgelist[i,3],edgelist[i,2]]) |
listAdj2$Apower6[edgelist[i,3],edgelist[i,2]] == 0)) |

```



```

      (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower7[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower7[edgelist[i,3],edgelist[i,2]] == 0)) |
      (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower8[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower8[edgelist[i,3],edgelist[i,2]] == 0)) |
      (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower9[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower9[edgelist[i,3],edgelist[i,2]] == 0)) |
      (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower10[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower10[edgelist[i,3],edgelist[i,2]] == 0)) |
      (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower11[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower11[edgelist[i,3],edgelist[i,2]] == 0)) |
      (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower12[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower12[edgelist[i,3],edgelist[i,2]] == 0)) |
      (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower13[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower13[edgelist[i,3],edgelist[i,2]] == 0)) |
      (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower14[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower14[edgelist[i,3],edgelist[i,2]] == 0)) |
      (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower15[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower15[edgelist[i,3],edgelist[i,2]] == 0)) |
      (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower16[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower16[edgelist[i,3],edgelist[i,2]] == 0)) |
      (edgelist[i,4] == "1" &
!(is.na(listAdj2$Apower17[edgelist[i,3],edgelist[i,2]])) |
listAdj2$Apower17[edgelist[i,3],edgelist[i,2]] == 0))
    ){idxMNFFL1[i] = i}
}

MNFFL1 = unlist(MNFFL1)

idxMNFFL1 = unlist(idxMNFFL1)

length(idxMNFFL1)

## [1] 5267

```

Through the following code, the number of edges which are engaged in the multiple-edge subgraphs are computed.

## MFFL2

```
MFFL2.pval.cor1 = 0
MFFL2.pval.cor.NA = 0
MFFL2.pval.cor2 = 0

for(i in idxMFFL2){
  logic = c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){
      logic[j]=
!any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))
    }
    if(all(logic)){
      logic1 = c()
      logic2 = c()
      for(j in 1:SignalingNet[[i]]$length){

        logic1[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor > 0
        logic2[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor < 0
      }
      if(all(logic1)) {MFFL2.pval.cor1=MFFL2.pval.cor1+1}
      if(all(logic2)) {MFFL2.pval.cor2=MFFL2.pval.cor2+1}
    }
  } else{
    if(!any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na)))){
      a = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor > 0
      b = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor < 0
      if(a){MFFL2.pval.cor1=MFFL2.pval.cor1+1}
      if(b){MFFL2.pval.cor2=MFFL2.pval.cor2+1}
    }
  }
}
```

```

for(i in idxMFFL2){
  logic = c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){

      logic[j]=any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na))) |
      SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval > 0.05
    }
    if(all(logic)){ MFFL2.pval.cor.NA = MFFL2.pval.cor.NA + 1 }

  }else{ if(any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na))) |
  SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval > 0.05){
    index.pval.cor.NA = MFFL2.pval.cor.NA + 1}
  }
}

MFFL2.pval.cor1

## [1] 200

MFFL2.pval.cor.NA

## [1] 1057

MFFL2.pval.cor2

## [1] 106

length(idxMFFL2) - (MFFL2.pval.cor1 + MFFL2.pval.cor.NA + MFFL2.pval.cor2)

## [1] 1245

```

## MNFBL1

```

MNFBL1.pval.cor1 = 0
MNFBL1.pval.cor.NA = 0
MNFBL1.pval.cor2 = 0

```

```

for(i in idxMNFBL1){
  logic =c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){
      logic[j]=
!any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))

```

```

    }
    if(all(logic)){
      logic1 = c()
      logic2 = c()
      for(j in 1:SignalingNet[[i]]$length){

        logic1[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor > 0
        logic2[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor < 0
      }
      if(all(logic1)) {MNFB1.pval.cor1=MNFB1.pval.cor1+1}
      if(all(logic2)) {MNFB1.pval.cor2=MNFB1.pval.cor2+1}
    }
  } else{
    if(!any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na)))){
      a = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor > 0
      b = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor < 0
      if(a){MNFB1.pval.cor1=MNFB1.pval.cor1+1}
      if(b){MNFB1.pval.cor2=MNFB1.pval.cor2+1}
    }
  }
}

for(i in idxMNFB1){
  logic = c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){

      logic[j]=any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))
| SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval > 0.05
    }
    if(all(logic)){ MNFB1.pval.cor.NA = MNFB1.pval.cor.NA + 1 }

  }else{ if(any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na))) |
SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval > 0.05){
    index.pval.cor.NA = MNFB1.pval.cor.NA + 1}
  }
}

MNFB1.pval.cor1

```

```
## [1] 773

MNFBFL1.pval.cor.NA

## [1] 4205

MNFBFL1.pval.cor2

## [1] 423

length(idxMNFBFL1) - (MNFBFL1.pval.cor1 + MNFBFL1.pval.cor.NA +
MNFBFL1.pval.cor2)

## [1] 6131
```

## MNFFL1

```
MNFFL1.pval.cor1 = 0
MNFFL1.pval.cor.NA = 0
MNFFL1.pval.cor2 = 0

for(i in idxMNFFL1){
  logic = c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){
      logic[j]=
!any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))
    }
    if(all(logic)){
      logic1 = c()
      logic2 = c()
      for(j in 1:SignalingNet[[i]]$length){

        logic1[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor > 0
        logic2[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor < 0
      }
      if(all(logic1)) {MNFFL1.pval.cor1=MNFFL1.pval.cor1+1}
      if(all(logic2)) {MNFFL1.pval.cor2=MNFFL1.pval.cor2+1}
    }
  } else{
    if(!any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na)))){
      a = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
```

```

SignalingNet[[i]]$corAnalysis$pearson$pearsoncor > 0
  b = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor < 0
  if(a){MNFFL1.pval.cor1=MNFFL1.pval.cor1+1}
  if(b){MNFFL1.pval.cor2=MNFFL1.pval.cor2+1}
}
}
}

```

```

for(i in idxMNFFL1){
  logic = c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){

      logic[j]=any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))
| SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval > 0.05
    }
    if(all(logic)){ MNFFL1.pval.cor.NA = MNFFL1.pval.cor.NA + 1 }

  }else{ if(any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na))) |
SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval > 0.05){
    index.pval.cor.NA = MNFFL1.pval.cor.NA + 1}
  }
}

```

MNFFL1.pval.cor1

## [1] 393

MNFFL1.pval.cor.NA

## [1] 1820

MNFFL1.pval.cor2

## [1] 193

length(idxMNFFL1) - (MNFFL1.pval.cor1 + MNFFL1.pval.cor.NA +  
MNFFL1.pval.cor2)

## [1] 2861

## MPFBL2

```
MPFBL2.pval.cor1 = 0
MPFBL2.pval.cor.NA = 0
MPFBL2.pval.cor2 = 0
```

```
for(i in idxMPFBL2){
  logic = c()
  if(SignalingNet[[i]]$length > 1){

    for(j in 1:SignalingNet[[i]]$length){
      logic[j]=
!any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))
    }
    if(all(logic)){
      logic1 = c()
      logic2 = c()
      for(j in 1:SignalingNet[[i]]$length){

        logic1[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor > 0
        logic2[j] =
SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis[[j]]$pearson$pearsoncor < 0
      }
      if(all(logic1)) {MPFBL2.pval.cor1=MPFBL2.pval.cor1+1}
      if(all(logic2)) {MPFBL2.pval.cor2=MPFBL2.pval.cor2+1}
    }
  } else{
    if(!any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na)))){
      a = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor > 0
      b = SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval < 0.05 &
SignalingNet[[i]]$corAnalysis$pearson$pearsoncor < 0
      if(a){MPFBL2.pval.cor1=MPFBL2.pval.cor1+1}
      if(b){MPFBL2.pval.cor2=MPFBL2.pval.cor2+1}
    }
  }
}
```

```
for(i in idxMPFBL2){
  logic = c()
  if(SignalingNet[[i]]$length > 1){
```

```

    for(j in 1:SignalingNet[[i]]$length){
      logic[j]=any(unlist(lapply(SignalingNet[[i]]$corAnalysis[[j]],is.na)))
| SignalingNet[[i]]$corAnalysis[[j]]$pearson$adjusted.pearsonpval > 0.05
    }
    if(all(logic)){ MPFBL2.pval.cor.NA = MPFBL2.pval.cor.NA + 1 }

    }else{ if(any(unlist(lapply(SignalingNet[[i]]$corAnalysis,is.na))) |
SignalingNet[[i]]$corAnalysis$pearson$adjusted.pearsonpval > 0.05){
      index.pval.cor.NA = MPFBL2.pval.cor.NA + 1}
    }
  }
}

MPFBL2.pval.cor1

## [1] 45

MPFBL2.pval.cor.NA

## [1] 355

MPFBL2.pval.cor2

## [1] 34

length(idxMPFBL2) - (MPFBL2.pval.cor1 + MPFBL2.pval.cor.NA +
MPFBL2.pval.cor2)

## [1] 539

```







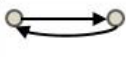

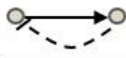
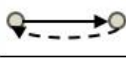


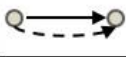
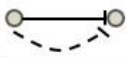
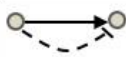
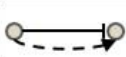
| Simple Subgraphs                                                                    |                                      |              |        |          |
|-------------------------------------------------------------------------------------|--------------------------------------|--------------|--------|----------|
| Structures                                                                          | Names                                | Abbreviation | KEGG   | OmniPath |
|    | Unconnected Gene Pairs               | UGP          | —      | —        |
|    | Activation                           | Act          | 19,170 | 11,437   |
|    | Inhibition                           | Inh          | 7,320  | 3,607    |
| Complex Subgraphs                                                                   |                                      |              |        |          |
|    | Dual Negative Feedback Loop          | DNFBL        | 37     | 107      |
|    | Dual Positive Feedback Loop1         | DPFBL1       | 186    | 321      |
|    | Dual Positive Feedback Loop2         | DPFBL2       | 14     | 49       |
|    | Multiple Negative Feedback Loop1     | MNFBL1       | 17,712 | 10,585   |
|    | Multiple Positive Feedback Loop1     | MPFBL1       | 3,731  | 2,379    |
|    | Multiple Negative Feedback Loop2     | MNFBL2       | 2,417  | 728      |
|   | Multiple Positive Feedback Loop2     | MPFBL2       | 3,232  | 1,949    |
|  | Multiple Feed-Forward Loop1          | MFFL1        | 12,869 | 6,248    |
|  | Multiple Feed-Forward Loop2          | MFFL2        | 6,618  | 3,383    |
|  | Multiple Negative Feed Forward Loop1 | MNFFL1       | 8,918  | 5,628    |
|  | Multiple Negative Feed-Forward Loop2 | MNFFL2       | 2,925  | 637      |

Table1

## Results

| Number of KEGG edges and GEO             |             |                       |                      |                       |                     |
|------------------------------------------|-------------|-----------------------|----------------------|-----------------------|---------------------|
| Simple Subgraphs                         |             |                       |                      |                       |                     |
|                                          | Abreviation | Pval < 0.05 & cor > 0 | Pval > 0.05 or == na | Pval < 0.05 & cor < 0 | Heterogeneous edges |
| Randomly-selected unconnected gene pairs | UGP         | 52                    | 392                  | 51                    | 505                 |
| Activation                               | Act         | 896                   | 4602                 | 457                   | 12693               |
| Inhibition                               | Inh         | 243                   | 1246                 | 130                   | 3110                |
| Complex Subgraphs                        |             |                       |                      |                       |                     |
|                                          | Abreviation | Pval < 0.05 & cor > 0 | Pval > 0.05 or == na | Pval < 0.05 & cor < 0 | Heterogeneous edges |
| Dual negative feedback loop              | DNFBL       | 0                     | 8                    | 3                     | 29                  |
| Dual positive feedback loop1             | DPFBL1      | 34                    | 85                   | 6                     | 125                 |
| Dual positive feedback loop2             | DPFBL2      | 0                     | 10                   | 0                     | 14                  |
| Multiple negative feedback loop1         | MNFBL1      | 773                   | 1851                 | 423                   | 8496                |
| Multiple positive feedback loop1         | MPFBL1      | 224                   | 426                  | 85                    | 2087                |
| Multiple negative feedback loop2         | MNFBL2      | 24                    | 83                   | 13                    | 310                 |
| Multiple positive feedback loop2         | MPFBL2      | 45                    | 160                  | 34                    | 735                 |
| Multiple feed-forward loop1              | MFFL1       | 537                   | 1384                 | 270                   | 6234                |
| Multiple feed-forward loop2              | MFFL2       | 200                   | 457                  | 106                   | 1846                |
| Multiple negative feed forward loop1     | MNFFL1      | 393                   | 801                  | 193                   | 3884                |
| Multiple negative feed-forward loop2     | MNFFL2      | 43                    | 108                  | 27                    | 481                 |

*Table2*

| Ratio of KEGG edges and GEO              |             |                       |                      |                       |                     |
|------------------------------------------|-------------|-----------------------|----------------------|-----------------------|---------------------|
| Simple Subgraphs                         |             |                       |                      |                       |                     |
|                                          | Abreviation | Pval < 0.05 & cor > 0 | Pval > 0.05 or == na | Pval < 0.05 & cor < 0 | Heterogeneous edges |
| Randomly-selected unconnected gene pairs | UGP         | 5.20%                 | 39.20%               | 5.10%                 | 50.50%              |
| Activation                               | Act         | 4.80%                 | 24.68%               | 2.45%                 | 68.07%              |
| Inhibition                               | Inh         | 5.14%                 | 26.35%               | 2.75%                 | 65.76%              |
| Complex Subgraphs                        |             |                       |                      |                       |                     |
|                                          | Abreviation | Pval < 0.05 & cor > 0 | Pval > 0.05 or == na | Pval < 0.05 & cor < 0 | Heterogeneous edges |
| Dual negative feedback loop              | DNFBL       | 0.00%                 | 20.00%               | 7.50%                 | 72.50%              |
| Dual positive feedback loop1             | DPFBL1      | 13.60%                | 34.00%               | 2.40%                 | 50.00%              |
| Dual positive feedback loop2             | DPFBL2      | 0.00%                 | 41.67%               | 0.00%                 | 58.33%              |
| Multiple negative feedback loop1         | MNFBL1      | 6.70%                 | 16.04%               | 3.66%                 | 73.60%              |
| Multiple positive feedback loop1         | MPFBL1      | 7.94%                 | 15.10%               | 3.01%                 | 73.95%              |
| Multiple negative feedback loop2         | MNFBL2      | 5.58%                 | 19.30%               | 3.02%                 | 72.09%              |
| Multiple positive feedback loop2         | MPFBL2      | 4.62%                 | 16.43%               | 3.49%                 | 75.46%              |
| Multiple feed-forward loop1              | MFFL1       | 6.37%                 | 16.43%               | 3.20%                 | 73.99%              |
| Multiple feed-forward loop2              | MFFL2       | 7.67%                 | 17.52%               | 4.06%                 | 70.76%              |
| Multiple negative feed forward loop1     | MNFFL1      | 7.46%                 | 15.20%               | 3.66%                 | 73.69%              |
| Multiple negative feed-forward loop2     | MNFFL2      | 6.53%                 | 16.39%               | 4.10%                 | 72.99%              |

*Table3*