Homework 4 - Answer Key

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1. Perform a Simple Linear Regression for:

- OUTCOME variable cesd: "Center for Epidemiological Studies-Depression (CESD) total score Baseline"
- PREDICTOR variable indtot: "Inventory of Drug Use Consequences (InDue) total score Baseline"
- decide if you want to transform either variable cesd or indtot and if so, what transformation you applied and why you can also decide not to transform (i.e. tradeoffs between model fit and interpretability of your results) discuss your reasoning.

1. Answer

Here is the code and output after running a simple linear regression using lm() function.

```
library(tidyverse)
library(haven)

helpdat <- haven::read_spss("helpmkh.sav")

# create subset
# select indtot, cesd and racegrp

h1 <- helpdat %>%
    select(indtot, cesd, racegrp)

# run simple linear regression
# using the lm
# save the results in the fit1 object
fit1 <- lm(cesd ~ indtot, data=h1)</pre>
```

Model summary

Coefficients:

```
# look at a summary() of the model
summary(fit1)

##
## Call:
## lm(formula = cesd ~ indtot, data = h1)
##
## Residuals:
## Min 1Q Median 3Q Max
## -34.356 -7.658 0.644 8.057 30.674
##
```

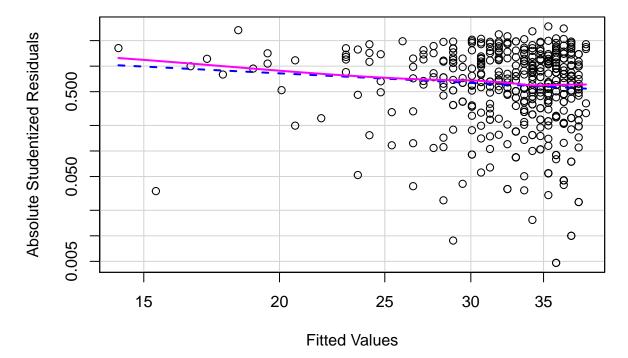
```
##
               Estimate Std. Error t value Pr(>|t|)
## (Intercept) 11.86597
                           2.82788
                                     4.196 3.27e-05 ***
  indtot
                0.58725
                           0.07761
                                     7.566 2.18e-13 ***
##
                     '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
## Signif. codes:
##
## Residual standard error: 11.8 on 451 degrees of freedom
## Multiple R-squared: 0.1126, Adjusted R-squared: 0.1107
## F-statistic: 57.25 on 1 and 451 DF, p-value: 2.176e-13
```

Possible Transformation

I probably would not do a transformation since the residuals look fairly normal (see diagnostic plots below), but you could run the spreadLevelPlot() function from the car package to see if a power transformation is suggested. This suggests a power transformation of 1.639 which could be rounded up to 2. This is optional and not needed for this data.

```
library(car)
# look at the spreadLevelPlot
# this also provides a suggestion of
# possible power transformation
car::spreadLevelPlot(fit1)
```

Spread-Level Plot for fit1



```
##
## Suggested power transformation: 1.63886
```

Plus the gvlma() function from the gvlma package can be run to check model assumptions, which also all look ok.

```
# global test of linear model assumptions
# install gvlma package
library(gvlma)
gvmodel <- gvlma::gvlma(fit1)
summary(gvmodel)</pre>
```

```
##
## Call:
## lm(formula = cesd ~ indtot, data = h1)
##
## Residuals:
##
                10 Median
                                3Q
      Min
                                       Max
  -34.356 -7.658
                     0.644
                             8.057
                                    30.674
##
## Coefficients:
##
              Estimate Std. Error t value Pr(>|t|)
## (Intercept) 11.86597
                           2.82788
                                     4.196 3.27e-05 ***
                                     7.566 2.18e-13 ***
                0.58725
                           0.07761
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 11.8 on 451 degrees of freedom
## Multiple R-squared: 0.1126, Adjusted R-squared: 0.1107
## F-statistic: 57.25 on 1 and 451 DF, p-value: 2.176e-13
##
##
## ASSESSMENT OF THE LINEAR MODEL ASSUMPTIONS
## USING THE GLOBAL TEST ON 4 DEGREES-OF-FREEDOM:
## Level of Significance = 0.05
##
## Call:
   gvlma::gvlma(x = fit1)
##
                                                     Decision
                        Value p-value
## Global Stat
                     6.65298 0.1554 Assumptions acceptable.
                     2.17129 0.1406 Assumptions acceptable.
## Skewness
## Kurtosis
                     2.89772 0.0887 Assumptions acceptable.
## Link Function
                     1.52631 0.2167 Assumptions acceptable.
## Heteroscedasticity 0.05766 0.8102 Assumptions acceptable.
```

2. Perform regression diagnostics:

- check the normality of the residuals (histogram and Q-Q plots)
- check for linearity is there any systematic relationship between the residuals and the predicted (or fitted) values?
- homoscedasticity plot of standardized residuals versus fitted values this is known as a "Scale-Location" graph.
- check for outliers and data points with high leverage or influence: outliers are often identified with standardized residuals > 3 (or <-3) and influential observations are often identified using Cook's D

2. Answer

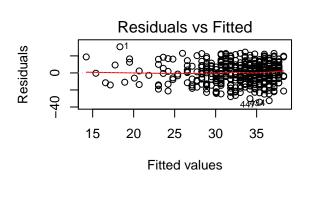
A good set of 4 diagnostic plots can be obtained using the plot() function for the fitted model output.

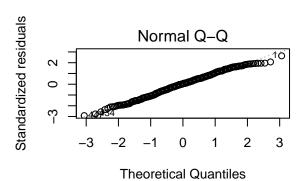
Diagnostic plots

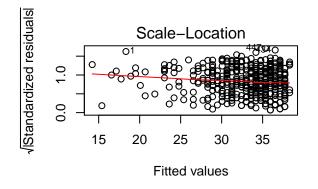
These 4 diagnostic plots show:

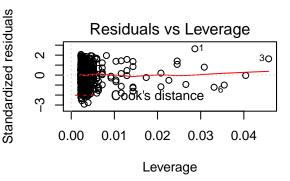
- residuals vs fitted values the line here is flat and shows no obvious trend, but the data do cluster on the higher end of the fitted values than the lower end, indicating some skewness
- normal Q-Q plot of the residuals this plot looks fairly linear indicating a close to normal distribution
- scale-location plot there is a slight trend downwards, but this slope is minor and in general the variability looks pretty consistent across all of the fitted values (no obvious heteroscedasticity)
- the last plot of Cook's distance does highlight a few possible outliers cases 1, 3 and 6, in the but these appear to be minor as they are not obvious in the Q-Q plot nor in the histogram

```
# get diagnostic plots
par(mfrow=c(2,2))
plot(fit1)
```









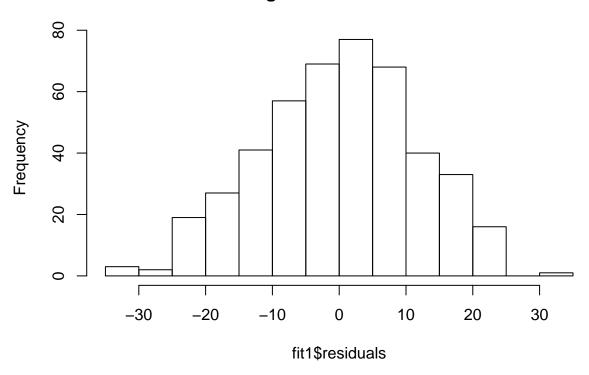
```
# reset par
par(mfrow=c(1,1))
```

Histogram of the Residuals

The histogram of the residuals look normal. No skewness and no obvious outliers.

```
# histogram of the residuals
hist(fit1$residuals)
```

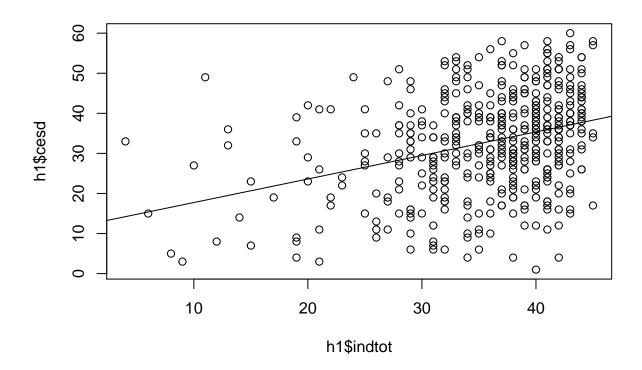
Histogram of fit1\$residuals



Scatterplot of Data with Fitted Line

There is quite a bit of scatter in the plot with a lot of variability in both indtot and cesd, so a linear fit line looks as good as any - there is no obvious curvature to the data.

```
plot(h1$indtot, h1$cesd)
abline(lm(h1$cesd ~ h1$indtot))
```



3. Provide a summary of the regression results.

- provide a **FIGURE** of the model, in this case a scatterplot with the fitted line overlaid and 95% confidence intervals if you can
- Make a TABLE presenting the fitted regression model (coefficients and tests of significance for those coefficients)
- describe the variance explained by the model (based on r2)
- describe the model itself based on the y-intercept and slope terms
- note any limitations or issues with the model fit or interpretation of the model

Model fit with 95% confidence intervals for fitted line

```
# using a ggplot2 approach
ggplot(h1, aes(indtot, cesd)) +
  geom_point() +
  stat_smooth(method = lm)
```

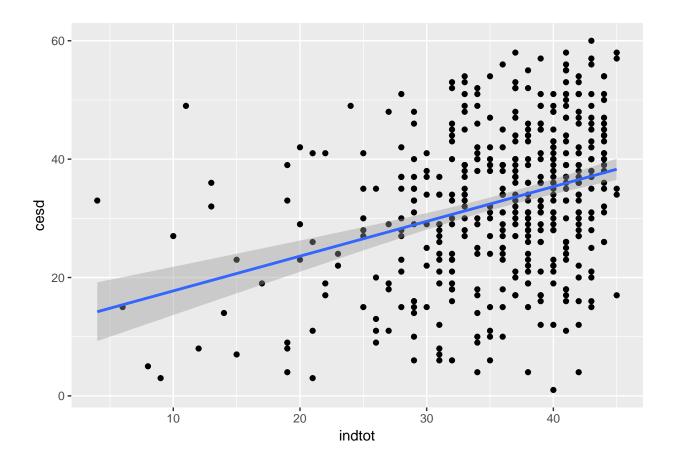


Table of the model fit

```
library(knitr)
library(xtable)
print(xtable(summary(fit1)), type = "html")
```

Estimate

Std. Error

t value

 $\Pr(>|t|)$

(Intercept)

11.8660

2.8279

4.20

0.0000

ind to t

0.5873

0.0776

7.57

0.0000

Variance explained

The variablity explained by this model is 11.0676446%, which is the adjusted r2 captured as summary(fit1)\$adj.r.squared.

Describe the model

For indtot scores equal to 0, the model estimates that a subject would have a cesd score of 11.8659738 given the y-intercept. For each 1 point increase in indtot scores, the cesd score will increase on average by 0.5872545 based on the slope estimate.

Model fit and Any Other issues

It is worth noting that given the wide variability in both the indtot and cesd a linear fit line indicates a weak positive correlation between these 2 variables, but the residuals do show wide variation about the best fitted line, indicating that the linear trend is weak at best.

4. Perform a One-way ANOVA for:

- OUTCOME variable cesd: "Center for Epidemiological Studies-Depression (CESD) total score Baseline"
- GROUP variable racegrp: "Racial Group of Respondent"
- options you can use either an ANOVA or GLM modeling approach
- if the GROUP variable is significant, also perform *post hoc* tests use some kind of pairwise error rate adjustment (i.e. bonferroni, sidak, Tukey's HSD, etc) be sure to report which one you used and why

ANOVA Model Results with dummy coding

```
# one-way ANOVA
# we can use the lm() function
# it does "dummy" coding on the fly
# run racegrp as either the character
# type or as a factor - either will work
fit2.lm <- lm(cesd ~ racegrp, data=h1)
summary(fit2.lm)</pre>
```

```
##
## Call:
## lm(formula = cesd ~ racegrp, data = h1)
##
## Residuals:
##
        Min
                  1Q
                       Median
                                     3Q
                                             Max
                       0.8199
## -31.3012 -8.3012
                                 8.6400 27.8199
##
## Coefficients:
```

```
##
                  Estimate Std. Error t value Pr(>|t|)
                               0.8468 35.642 < 2e-16 ***
## (Intercept)
                   30.1801
## racegrphispanic
                               1.9346
                    4.1799
                                       2.161
                                                0.0313 *
                                                0.0252 *
## racegrpother
                    5.7430
                                        2.246
                               2.5565
## racegrpwhite
                    5.1211
                               1.2761
                                      4.013 7.02e-05 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 12.3 on 449 degrees of freedom
## Multiple R-squared: 0.04042,
                                   Adjusted R-squared:
## F-statistic: 6.304 on 3 and 449 DF, p-value: 0.0003396
```

ANOVA results for group effect overall

```
# the aov() function
# gives the global test for the "group" effect
fit2.aov <- aov(cesd ~ racegrp, data=h1)</pre>
summary(fit2.aov)
##
                Df Sum Sq Mean Sq F value Pr(>F)
## racegrp
                     2861
                            953.7
                                    6.304 0.00034 ***
## Residuals
               449
                   67927
                            151.3
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
```

Yes, racegrp is significant - there are significant differences between the 4 races.

Post hoc tests

Since racegrp was significant, let's run all of the pairwise comparisons. The code below will show the options for adjusting the error-rate due to multiple pairwise comparisons including: Bonferroni, Holm, and Tukey HSD.

```
# post hoc tests
# Tukey HSD
TukeyHSD(fit2.aov)
```

```
Tukey multiple comparisons of means
##
##
       95% family-wise confidence level
##
## Fit: aov(formula = cesd ~ racegrp, data = h1)
##
## $racegrp
##
                        diff
                                   lwr
                                             upr
## hispanic-black 4.1799052 -0.8088317 9.168642 0.1359780
## other-black
                  5.7429821 -0.8494072 12.335372 0.1125093
## white-black
                  5.1211100 1.8305336 8.411686 0.0004071
## other-hispanic 1.5630769 -6.1057988 9.231953 0.9528809
## white-hispanic 0.9412048 -4.1754298 6.057839 0.9647000
## white-other
                 -0.6218721 -7.3115695 6.067825 0.9951504
```

```
# using Bonferroni error-rate correction
pairwise.t.test(h1$cesd, h1$racegrp, p.adj = "bonf")
##
   Pairwise comparisons using t tests with pooled SD
##
## data: h1$cesd and h1$racegrp
##
##
                    hispanic other
            black
## hispanic 0.18753 -
## other
            0.15097 1.00000 -
## white
           0.00042 1.00000 1.00000
##
## P value adjustment method: bonferroni
# using the Holm error-rate correction
pairwise.t.test(h1$cesd, h1$racegrp, p.adj = "holm")
##
   Pairwise comparisons using t tests with pooled SD
##
##
## data: h1$cesd and h1$racegrp
##
           black
                    hispanic other
## hispanic 0.12581 -
           0.12581 1.00000 -
## other
## white
            0.00042 1.00000 1.00000
##
## P value adjustment method: holm
```

These pairwise comparisons show that there are significant differences between white-black, but none of the other pairwise comparisons were significant.

5. Perform model diagnostics:

- homoscedasticity look at a test for equal variance (Levene's test or Bartlett's test or equivalent).
- if this test of equal variances fails, you may want to report a modified F-test (e.g. Welch's test)

Test of Equal Variances

```
# barlett's test for homogenity of variances
# note: put the formula back in
bartlett.test(cesd ~ racegrp, data=h1)

##
## Bartlett test of homogeneity of variances
##
## data: cesd by racegrp
## Bartlett's K-squared = 3.4367, df = 3, p-value = 0.3291
```

This was not significant, so we can report the usual F-statistic test for racegrp rather than the "robust" Welch's F-test.

6. Present a summary of the ANOVA results.

- Make a **FIGURE** of the group mean differences either an error-bar plot or a series of boxplots one for each group to show the group differences in the outcome
- Make a **TABLE** presenting the ANOVA results
- describe the model results was the GROUP (racegrp) significant?
- If GROUP is significant, what did the post hoc tests reveal?

Plot of Means and 95% Confidence Intervals

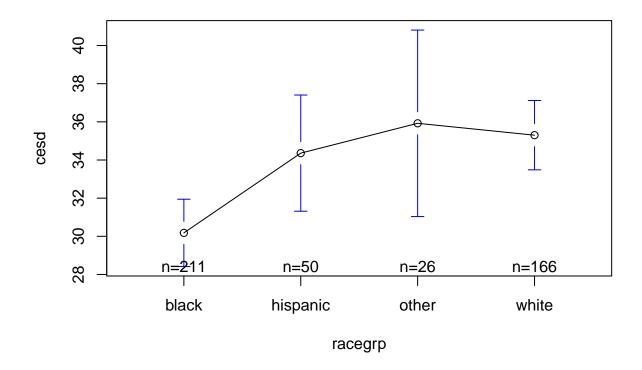


Table of the ANOVA results - overall group effect

racegrp was significant - no significant diff

```
print(xtable(summary(fit2.aov)),
    type = "html")
```

Df

 $\operatorname{Sum}\,\operatorname{Sq}$

Mean Sq

F value

Pr(>F)

racegrp

3

2861.03

953.68

6.30

0.0003

Residuals

449

67927.46

151.29