

inclass09

Quarto

Quarto enables you to weave together content and executable code into a finished document. To learn more about Quarto see <https://quarto.org>.

Running Code

When you click the **Render** button a document will be generated that includes both content and the output of embedded code. You can embed code like this:

```
1 + 1
```

```
[1] 2
```

You can add options to executable code like this

```
[1] 4
```

The `echo: false` option disables the printing of code (only output is displayed).

```
db<- read.csv("Data Export Summary.csv")
db
```

	Molecular.Type	X.ray	EM	NMR	Multiple.methods	Neutron	Other
1	Protein (only)	154,766	10,155	12,187	191	72	32
2	Protein/Oligosaccharide	9,083	1,802	32	7	1	0
3	Protein/NA	8,110	3,176	283	6	0	0
4	Nucleic acid (only)	2,664	94	1,450	12	2	1
5	Other	163	9	32	0	0	0

6	Oligosaccharide (only)	11	0	6	1	0	4
	Total						
1	177,403						
2	10,925						
3	11,575						
4	4,223						
5	204						
6	22						

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
sum_xray <- sum(as.numeric( gsub(",", "", db$X.ray) ))
sum_em <- sum(as.numeric( gsub(",", "", db$EM) ))
```

How can we make this into a function?

```
#x is a column from a matrix that is specified, in the format : matrix$columnname
#function substitutes comma for an empty string, makes it into a number, and sums the value
sum_comma<- function(x) {
  return(sum(as.numeric( gsub(",", "", x) )))
}
```

Now we can answer Q1

```
#for xray
sum_comma(db$X.ray)/sum_comma(db$Total)
```

```
[1] 0.8553721
```

```
#for EM
round(sum_comma(db$EM)/sum_comma(db$Total), 2 )
```

```
[1] 0.07
```

Q2: What proportion of structures in the PDB are protein?

```
round(sum_comma(db$Total[1])/sum_comma(db$Total), 2)
```

```
[1] 0.87
```

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB? Q6. Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain and the critical water (we recommend “Ball & Stick” for these side-chains). Add this figure to your Quarto document.



Figure 1: HIV-PR structure from MERK with a bound drug

Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

Because the resolution is too low to see all three atoms. You need a sub 1 Angstrom resolution to see them.

Q5: There is a critical “conserved” water molecule in the binding site. Can you

identify this water molecule? What residue number does this water molecule have

HOH308

#Section 3: Workigim with structures in R

We can use the `bio3d` package to read and perfrom bioinformatics calculations on PDG structures

```
library(bio3d)

pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
pdb
```

Call: `read.pdb(file = "1hsg")`

```
Total Models#: 1
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 172 (residues: 128)
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]
```

Protein sequence:

```
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF
```

```
+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call
```

```
attributes(pdb)
```

```
$names
[1] "atom"    "xyz"     "seqres"  "helix"   "sheet"   "calpha"  "remark"  "call"
```

```
$class
[1] "pdb" "sse"
```

```
head(pdb$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40

	segid	elesy	charge
1	<NA>	N	<NA>
2	<NA>	C	<NA>
3	<NA>	C	<NA>
4	<NA>	O	<NA>
5	<NA>	C	<NA>
6	<NA>	C	<NA>

read an ADK structure

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file
PDB has ALT records, taking A only, rm.alt=TRUE

```
adk
```

```
Call: read.pdb(file = "6s36")
```

```
Total Models#: 1
Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)

Protein Atoms#: 1654 (residues/Calpha atoms#: 214)
```

Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 244 (residues: 244)

Non-protein/nucleic resid values: [CL (3), HOH (238), MG (2), NA (1)]

Protein sequence:

```
MRILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAAVKSGSELGKQAKDIMDAGKLV  
DELVIALVKERIAQEDCRNGFLDGFRTIPQADAMKEAGINVDYVLEFDVPDELIVDKI  
VGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG  
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
```

+ attr: atom, xyz, seqres, helix, sheet,
calpha, remark, call

Q7: How many amino acid residues are there in this pdb object? 198

Q8: Name one of the two non-protein residues? HOH, MK1

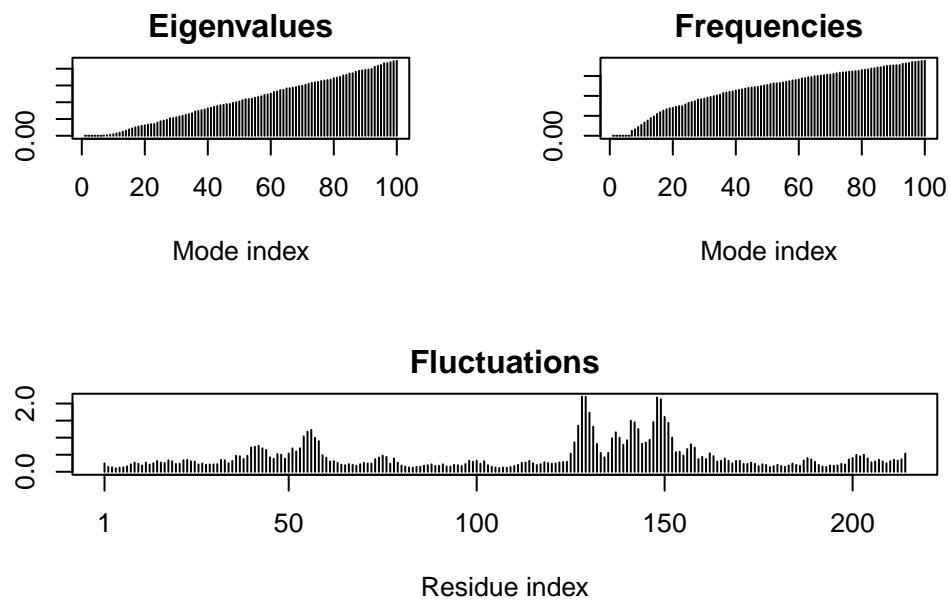
Q9: How many protein chains are in this structure? 2

Perfrom a predcition of felxibility with a technique called NMA (normal mode analysis)

```
#Perform a flexibility prediction  
m <-nma(adk)
```

```
Building Hessian...      Done in 0.064 seconds.  
Diagonalizing Hessian... Done in 0.633 seconds.
```

```
plot(m)
```



Write out a “movie” (aka trajectory) if tge motion forr viewing in MOL star

```
mktrj(m, file="adk_m7.pdb")
```