



Carnegie
Mellon
University

CB Computational
Biology
Department

Predicting the evolutionary stability of synonymous mutations

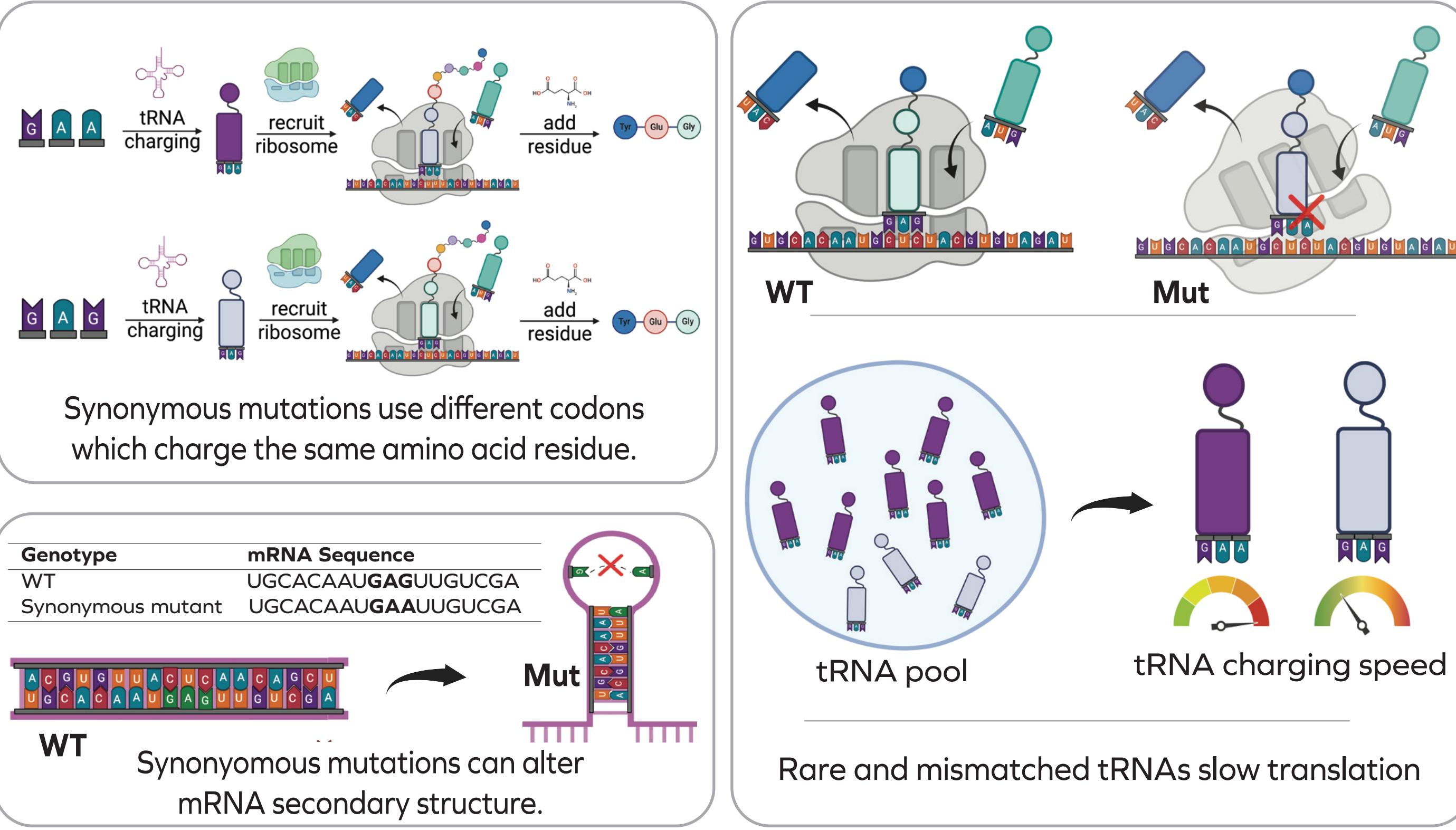
Seh Na Mellick

Ray and Stephanie Lane Computational Biology Department
Carnegie Mellon University

cb

Synonymous ≠ silent

Synonymous mutations are traditionally viewed as neutral because they do not alter protein sequences. However, they can still be deleterious through effects on expression, mRNA structure, or regulation.



Dataset

To predict the evolutionary fitness effects of synonymous mutations, we used a dataset from Shen et al. 2022 [1] which includes:

- (i) ~8,000 single SNPs over 150bp coding regions in 22 nonessential *Saccharomyces cerevisiae* genes
- (ii) Relative fitness measurements for each mutation in a pooled competition assay
- (iii) Wild-type and mutant codons for each mutation

Genomic, structural, and conservation features

Codon usage bias

$$CAI = \left(\prod_{i=1}^L w_i \right)^{1/L}$$

$$w_i = \frac{RSCU_i}{\max(RSCU_{syn})}$$

$$L = \# \text{ codons in gene}$$

$RSCU_i$: Relative Synonymous Codon Usage of codon i

CAI [2] quantifies how well a gene's codon usage matches the preferred codons of highly expressed genes in the organism.

Secondary mRNA structure

$$MFE = \min_{\text{all foldings}} \Delta G_{\text{structure}}$$

$$\Delta G_{\text{structure}} : \text{sum of energy contributions from secondary structures}$$

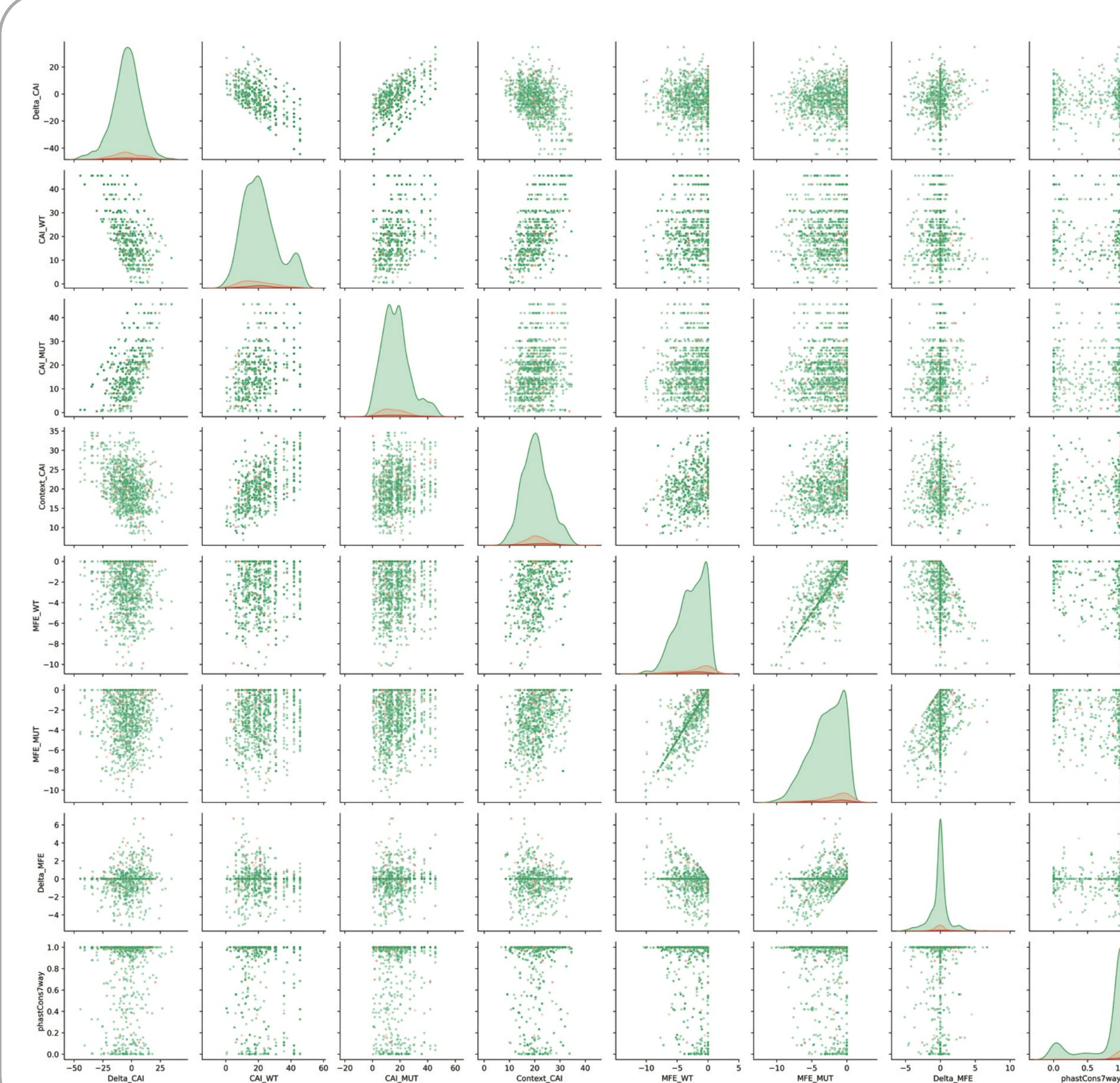
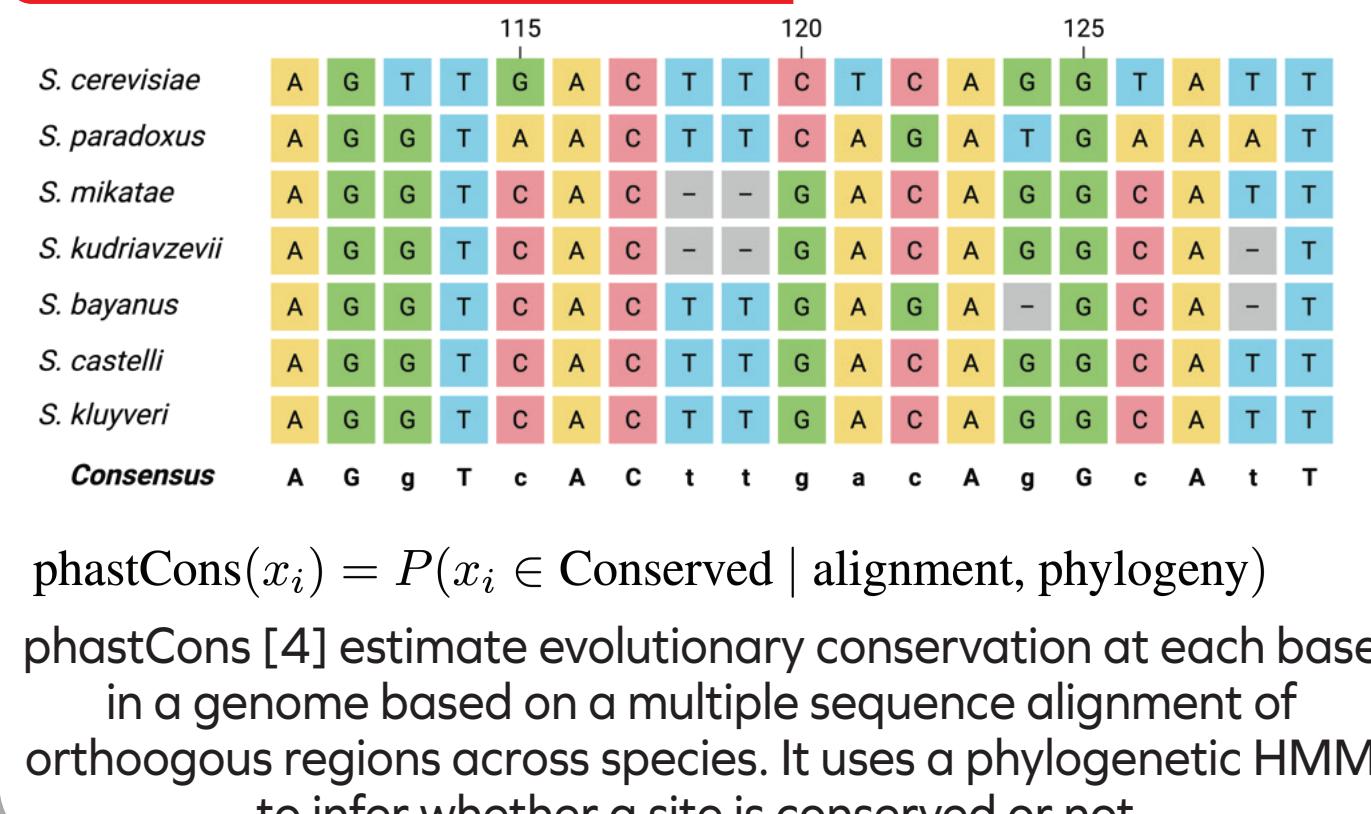
MFE [3] represents the predicted stability of an mRNA's secondary structure, which can affect translation efficiency and the rate of mRNA decay

Objectives

We compiled a curated dataset of SNPs from *S. cerevisiae* coding regions, annotated with codon usage bias metrics, mRNA folding energies, and conservation scores. Each mutation is associated with an experimentally measured fitness value from DMS experiments [1].

Our objective was twofold:
(i) to identify molecular features which best distinguish deleterious from neutral synonymous mutations, and (ii) to classify mutations into evolutionary fitness effect categories—strongly deleterious, mildly deleterious, or neutral.

Evolutionary conservation



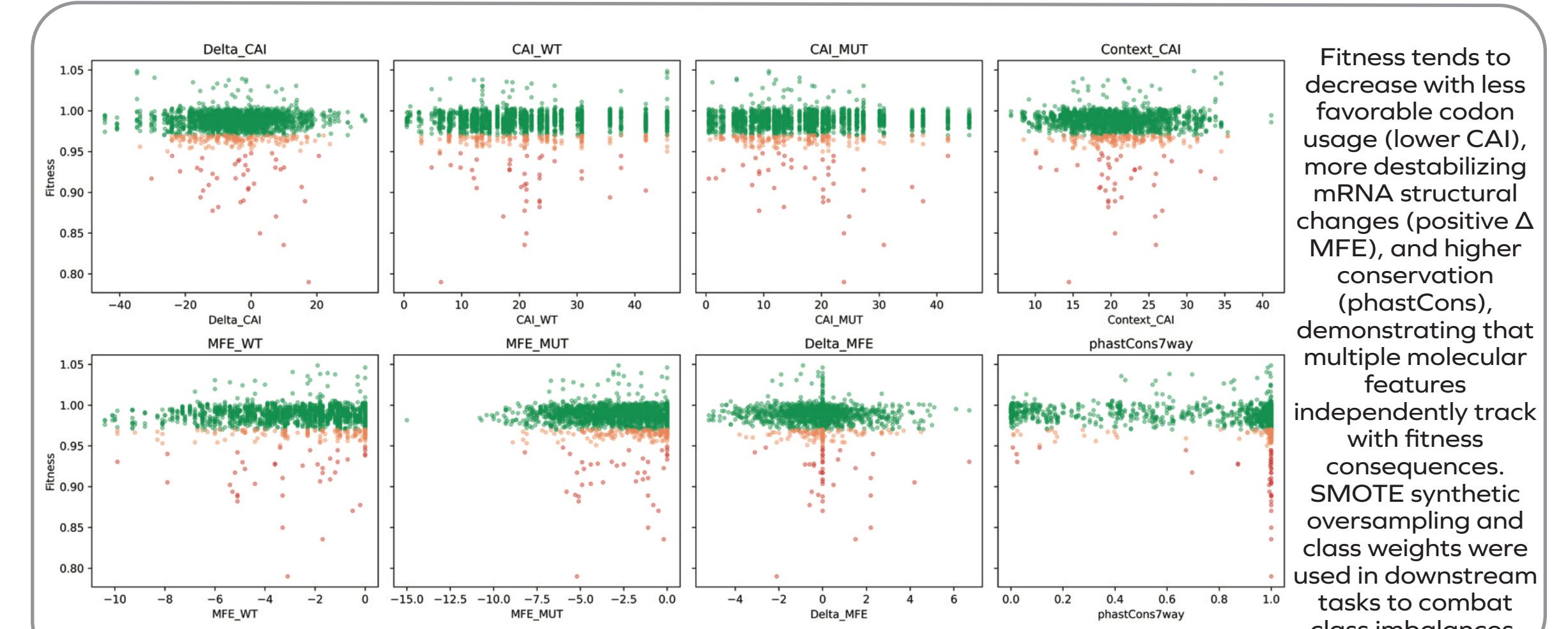
Fitness Class

- Strongly Deleterious
- Mildly Deleterious
- Neutral

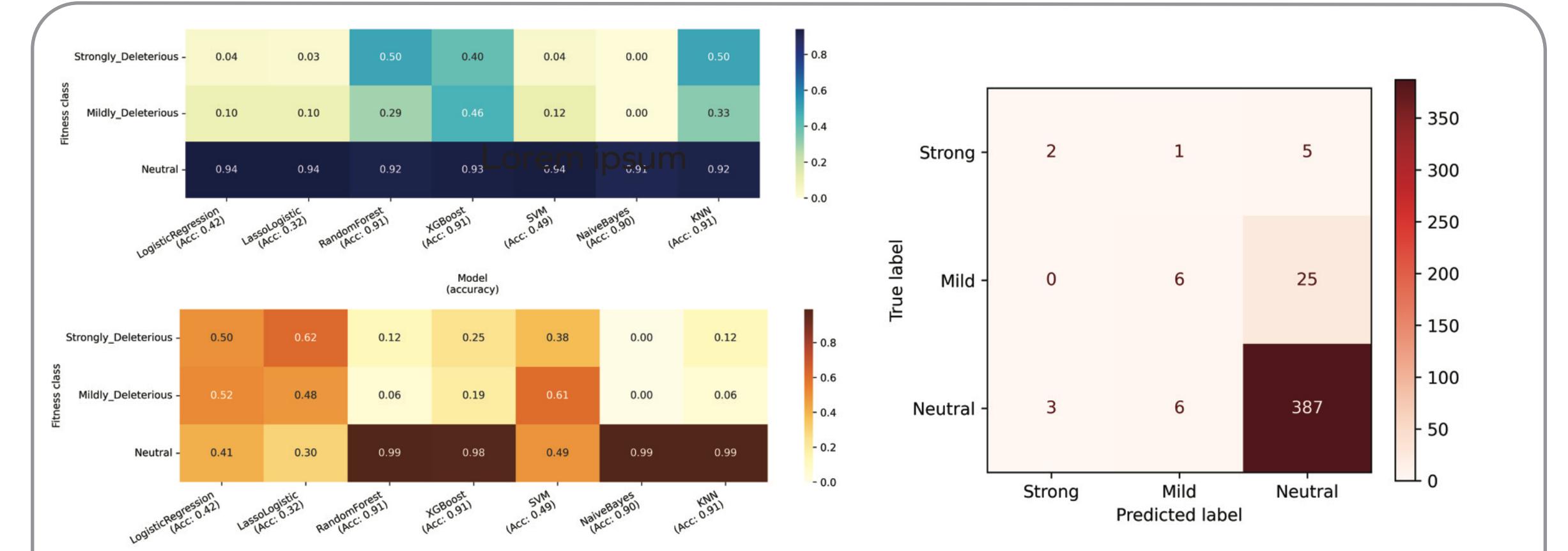
Pairwise relationships between genomic, structural, and evolutionary features of synonymous mutations, colored by fitness class.

Strongly deleterious mutations (red) show reduced CAI and more negative ΔMFE relative to neutral mutations, while phastCons appear positively associated with higher fitness. Overall, this suggests correlated shifts across multiple feature types.

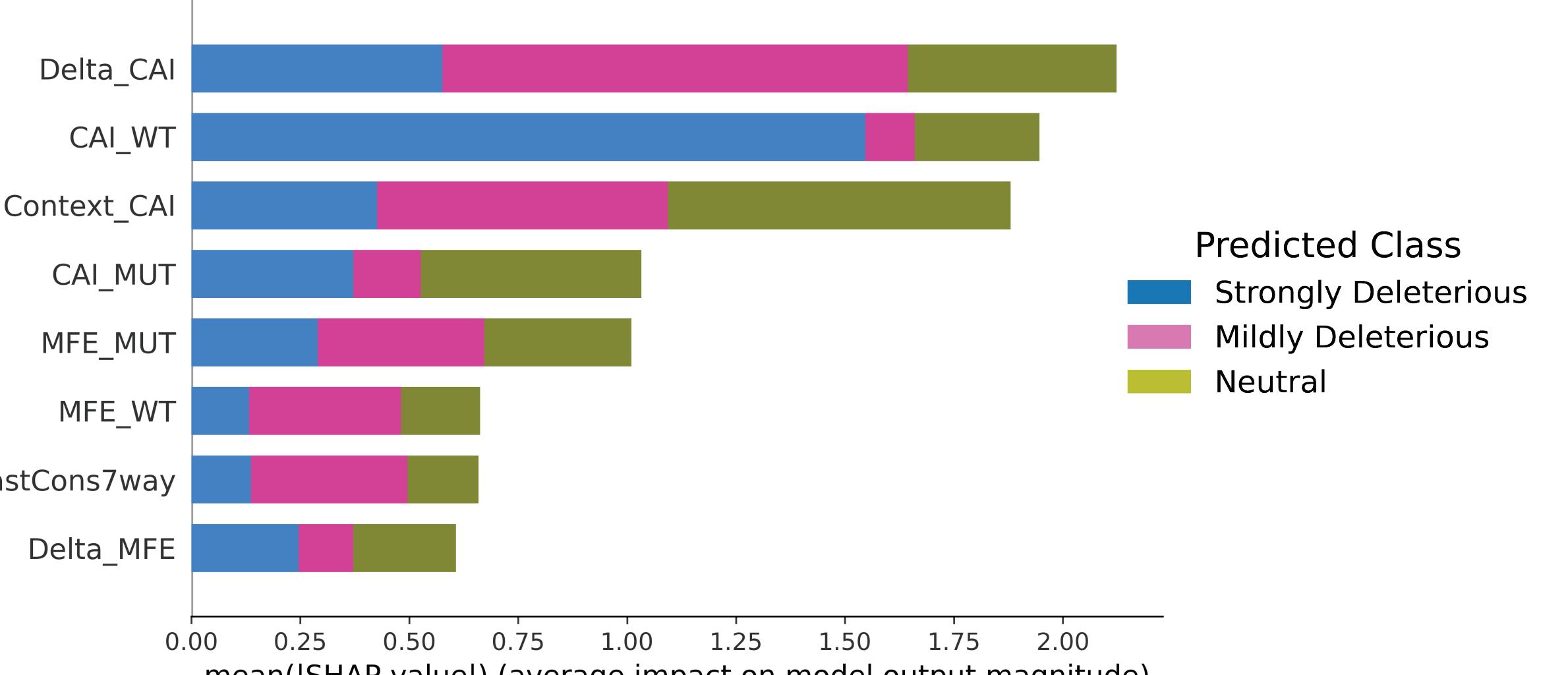
Predicting fitness effects



Fitness tends to decrease with less favorable codon usage (lower CAI), more destabilizing mRNA structural changes (positive ΔMFE), and higher conservation (phastCons), demonstrating that multiple molecular features independently track with fitness consequences. SMOTE synthetic oversampling and class weights were used in downstream tasks to combat class imbalances.



Feature importance for classification



Discussion

- (i) Our work supports the hypothesis that synonymous mutations can produce substantial fitness effects through codon usage, mRNA structural stability, and evolutionary conservation, despite not altering the protein sequence. This further challenges the common assumption that synonymous mutations are functionally silent.
- (ii) Our models predict neutral mutations well but struggle with predicting strongly deleterious synonymous mutations.
- (iii) Future work could use additional molecular data like ribosome profiling or RNA stability data to better capture post-transcriptional effects and refine predictive resolution for borderline fitness classes.
- (iv) The DMS dataset is inherently imbalanced, with most synonymous mutations being neutral and thus limiting classification performance. Future work could explore alternative sampling strategies or loss functions beyond class weighting and SMOTE.

References

1. Shen, X., et al. (2022). Synonymous mutations in representative yeast genes are mostly strongly nonneutral. *Nature*, 606(7915):725-31.
2. Puigbo, P., et al., (2008). Caical: a combined set of tools to assess codon usage adaptation. *Biology Direct*, 3(38).
3. Lorenz, R., et al., (2011). ViennaRNA package 2.0.. *Algorithms for Molecular Biology*, 6(1):1-14.
4. Siepel, A., et al., (2005). Evolutionarily conserved elements in vertebrate, insect, worm, and yeast genomes. *Genome Research*, 15(8):1034-50.
5. Lundberg, S., and Lee, S., (2017). A unified approach to interpreting model predictions. *Advances in Neural Information Processing Systems*, 30.