Create MRexp and Phyloseq Objects

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Generating MR experiment and phyloseq objects for the three pipelines.

Metadata

```
meta_df <- sampleSheet %>%
     filter(barcode_lab == "JHU", seq_lab == "JHU") %>%
     select(biosample_id, titration, t_fctr, pcr_16S_plate, pos) %>%
     unite(id, pcr_16S_plate, pos, sep = "-",remove = FALSE) %>%
     as.data.frame()
## PCR replicate information
half1 <- paste0(rep(c("A","B","C","D","E","F","G","H"), each = 6), 1:6)
meta_df <- meta_df %>%
               mutate(pcr_half = if_else(pos %in% half1, "1","2"),
                                  pcr_rep = paste0(pcr_16S_plate,":",pcr_half))
rownames(meta_df) <- meta_df$id</pre>
glimpse(meta_df)
## Observations: 192
## Variables: 8
## $ biosample_id <chr> "E01JH0004", "E01JH0004", "E01JH0004", "E01JH000...
## $ t_fctr
                                                <fctr> 20, 1, 2, 3, 4, 5, 10, 15, 0, 20, 1, 2, 3, 4, 5...
## $ id
                                               <chr> "1-A1", "1-B1", "1-C1", "1-D1", "1-E1", "1-F1", ...
## $ pos <chr> "A1", "B1", "C1", "D1", "E1", "F1", "G1", "H1", ...
## $ pcr_half
                                             <chr> "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1", "1:1"
```

Mothur

Mothur Phyloseq Object

```
mothur_ps
## phyloseq-class experiment-level object
                OTU Table:
                               [ 38358 taxa and 192 samples ]
## otu table()
## sample_data() Sample Data:
                                    [ 192 samples by 8 sample variables ]
                 Taxonomy Table: [ 38358 taxa by 6 taxonomic ranks ]
## tax table()
Mothur MRexperiment Object
mothur_mrexp <- phyloseq_to_metagenomeSeq(mothur_ps)</pre>
mothur_mrexp
## MRexperiment (storageMode: environment)
## assayData: 38358 features, 192 samples
     element names: counts
## protocolData: none
## phenoData
     sampleNames: 1-A1 1-A10 ... 2-H9 (192 total)
     varLabels: biosample_id titration ... pcr_rep (8 total)
##
    varMetadata: labelDescription
## featureData
   featureNames: Otu00001 Otu00002 ... Otu38358 (38358 total)
##
##
    fvarLabels: OTUname Rank1 ... Rank6 (7 total)
## fvarMetadata: labelDescription
## experimentData: use 'experimentData(object)'
## Annotation:
Save Mothur Objects
saveRDS(mothur_ps, "mothur/mothur_ps.rds")
saveRDS(mothur_mrexp, "mothur/mothur_mrexp.rds")
QIIME
QIIME Phyloseq Object
# qiime_dir <- "qiime/otus_uc_fast_no_chimera/"</pre>
qiime_dir <- "qiime/otus_uc_fast/"</pre>
biom_file <- file.path(qiime_dir, "otu_table_mc2_w_tax_no_pynast_failures.biom")</pre>
seq_file <- file.path(qiime_dir, "rep_set.fna")</pre>
tree_file <- file.path(qiime_dir, "rep_set.tre")</pre>
qiime_ps <- phyloseq::import_biom(BIOMfilename = biom_file)</pre>
## Warning in strsplit(msg, "\n"): input string 1 is invalid in this locale
#sample_names(qiime_ps) <- sample_names(qiime_ps) %>% str_replace("centroid=","")
## Adding sample data
phyloseq::sample_data(qiime_ps) <- meta_df</pre>
```

```
qiime_ps
## phyloseq-class experiment-level object
## otu_table() OTU Table: [ 11385 taxa and 192 samples ]
## sample_data() Sample Data: [ 192 samples by 8 sample variables ]
## tax_table() Taxonomy Table: [ 11385 taxa by 7 taxonomic ranks ]
QIIME MRexperiment Object
qiime_mrexp <- phyloseq_to_metagenomeSeq(qiime_ps)</pre>
qiime mrexp
## MRexperiment (storageMode: environment)
## assayData: 11385 features, 192 samples
    element names: counts
## protocolData: none
## phenoData
     sampleNames: 2-B5 2-E12 ... 2-D12 (192 total)
##
     varLabels: biosample_id titration ... pcr_rep (8 total)
##
     varMetadata: labelDescription
## featureData
##
    featureNames: 4333897 1036749 ... New.CleanUp.ReferenceOTU19782
##
        (11385 total)
##
    fvarLabels: OTUname Rank1 ... Rank7 (8 total)
## fvarMetadata: labelDescription
## experimentData: use 'experimentData(object)'
## Annotation:
Save QIIME Objects
saveRDS(qiime_ps, "qiime/qiime_ps.rds")
saveRDS(qiime_mrexp, "qiime/qiime_mrexp.rds")
```

DADA2 Phyloseq Object

```
seqtab <- readRDS("dada2/seqtab_nochim.rds")
otu_tbl <- otu_table(seqtab, taxa_are_rows=FALSE)

## Rep sequences
sv_seqs <- colnames(otu_tbl)
names(sv_seqs) <- paste0("SV",1:ncol(otu_tbl))

## Rename features
colnames(otu_tbl) <- paste0("SV",1:ncol(otu_tbl))

taxa <- readRDS("dada2/taxa.rds")
rownames(taxa) <- names(sv_seqs)[match(sv_seqs, rownames(taxa))]

dada_ps <- phyloseq(otu_tbl,</pre>
```

```
sample_data(meta_df),
                    tax_table(taxa))
## Removing O entry samples
dada_samples <- sample_names(dada_ps)</pre>
dada_nonzero_sample <- dada_samples[phyloseq::sample_sums(dada_ps) != 0]</pre>
dada_ps <- phyloseq::prune_samples(dada_nonzero_sample, dada_ps)</pre>
dada_ps
## phyloseq-class experiment-level object
## otu_table()
                OTU Table:
                                   [ 3144 taxa and 191 samples ]
## sample_data() Sample Data: [ 191 samples by 8 sample variables ]
                 Taxonomy Table: [ 3144 taxa by 6 taxonomic ranks ]
## tax_table()
DADA2 MRexperiment Object
dada_mrexp <- phyloseq_to_metagenomeSeq(dada_ps)</pre>
dada_mrexp
## MRexperiment (storageMode: environment)
## assayData: 3144 features, 191 samples
     element names: counts
## protocolData: none
## phenoData
##
    sampleNames: 1-A1 1-A10 ... 2-H9 (191 total)
##
     varLabels: biosample_id titration ... pcr_rep (8 total)
##
    varMetadata: labelDescription
## featureData
   featureNames: SV1 SV2 ... SV3144 (3144 total)
##
## fvarLabels: OTUname Rank1 ... Rank6 (7 total)
## fvarMetadata: labelDescription
## experimentData: use 'experimentData(object)'
## Annotation:
Save DADA2 Objects
saveRDS(dada_ps, "dada2/dada_ps.rds")
saveRDS(dada_mrexp, "dada2/dada_mrexp.rds")
DNAStringSet(sv_seqs) %>% writeXStringSet("dada2/sv_seqs.rds")
```

Session information

```
s_info <- devtools::session_info()
print(s_info$platform)

## setting value
## version R version 3.3.3 (2017-03-06)
## system x86_64, darwin15.6.0
## ui unknown</pre>
```

package	version	date	source
bbmle	1.0.18	2016-02-11	CRAN (R 3.3.2)
Biobase	2.34.0	2016 - 11 - 07	Bioconductor
BiocGenerics	0.20.0	2016 - 11 - 07	Bioconductor
BiocParallel	1.8.1	2016 - 11 - 07	Bioconductor
Biostrings	2.42.1	2016-12-19	Bioconductor
DESeq	1.26.0	2016 - 11 - 28	Bioconductor
DESeq2	1.15.28	2017-02-02	bioc (readonly/DESeq2@125913)
dplyr	0.5.0	2016-06-24	CRAN (R 3.3.2)
edgeR	3.16.5	2017-02-02	Bioconductor
forcats	0.2.0	2017-01-23	CRAN (R 3.3.2)
foreach	1.4.3	2015-10-13	CRAN (R 3.3.1)
GenomeInfoDb	1.10.3	2017 - 03 - 28	Bioconductor
GenomicAlignments	1.10.1	2017 - 03 - 28	Bioconductor
GenomicRanges	1.26.4	2017 - 03 - 28	Bioconductor
ggplot2	2.2.1	2016-12-30	CRAN (R 3.3.2)
glmnet	2.0 - 5	2016-03-17	CRAN (R 3.3.1)
IRanges	2.8.2	2017-03-28	Bioconductor
knitr	1.15.1	2016-11-22	CRAN (R 3.3.2)
lattice	0.20 - 34	2016-09-06	CRAN (R 3.3.3)
limma	3.30.13	2017-03-28	Bioconductor
locfit	1.5 - 9.1	2013-04-20	CRAN (R 3.3.1)
Matrix	1.2 - 8	2017-01-20	CRAN (R 3.3.3)
metagenomeSeq	1.16.0	2016-11-07	Bioconductor
modelr	0.1.0	2016-08-31	cran (@0.1.0)
permute	0.9 - 4	2016-09-09	CRAN (R 3.3.1)
phyloseq	1.19.1	2017-01-04	Bioconductor
ProjectTemplate	0.7	2016-08-11	CRAN (R 3.3.1)
purrr	0.2.2	2016-06-18	CRAN (R 3.3.1)
RColorBrewer	1.1-2	2014-12-07	CRAN (R 3.3.1)
readr	1.1.0	2017-03-22	CRAN (R 3.3.2)
readxl	0.1.1	2016-03-28	cran (@0.1.1)
Rqc	1.8.0	2016-11-07	Bioconductor
Rsamtools	1.26.1	2016-11-07	Bioconductor
S4Vectors	0.12.2	2017-03-28	Bioconductor
sads	0.3.1	2016-05-13	CRAN (R 3.3.2)
savR	1.12.0	2016-11-07	Bioconductor
ShortRead	1.32.1	2017-03-28	Bioconductor
stringr	1.2.0	2017-02-18	CRAN (R 3.3.2)
SummarizedExperiment	1.4.0	2016-11-07	Bioconductor
tibble	1.2	2016-08-26	CRAN (R 3.3.1)
tidyr	0.6.1	2017-01-10	CRAN (R 3.3.2)
tidyverse	1.1.1	2017-01-27	CRAN (R 3.3.2)
vegan	2.4-2	2017-01-17	CRAN (R 3.3.2)
XVector	0.14.1	2017-03-28	Bioconductor