Create MRexp and Phyloseq Objects

Nate Olson

2017-03-14

Generating MRexperiment and phyloseq objects for the three pipelines.

Metadata

```
meta_df <- sampleSheet %>%
  filter(barcode_lab == "JHU", seq_lab == "JHU") %>%
  select(biosample_id, titration, pcr_16S_plate, pos) %>%
  unite(id, pcr_16S_plate, pos, sep = "-",remove = TRUE)  %>%
  as.data.frame()

rownames(meta_df) <- meta_df$id

glimpse(meta_df)
```

```
## Observations: 192
## Variables: 3
## $ biosample_id <chr> "E01JH0004", "E01JH0004", "E01JH0004", "E01JH0004...
## $ titration    <dbl> 20, 0, 1, 2, 3, 4, 5, 10, 15, 20, 0, 1, 2, 3, 4, ...
## $ id           <chr> "1-F6", "1-A1", "1-B1", "1-C1", "1-D1", "1-E1", "...
```

Mothur

Mothur Phyloseq Object

```
shared_file <- "mothur/mgtst.trim.contigs.good.unique.good.filter.unique.precluster.pick.opti_mcc.unique

constax_file <- "mothur/mgtst.trim.contigs.good.unique.good.filter.unique.precluster.pick.opti_mcc.uniqu

mothur_ps <- phyloseq::import_mothur(mothur_shared_file = shared_file,
                      mothur_constaxonomy_file = constax_file)

## Adding sample data
phyloseq::sample_data(mothur_ps) <- meta_df

mothur_ps
```

```
## phyloseq-class experiment-level object
## otu_table()   OTU Table:         [ 38358 taxa and 192 samples ]
## sample_data() Sample Data:       [ 192 samples by 3 sample variables ]
## tax_table()   Taxonomy Table:    [ 38358 taxa by 6 taxonomic ranks ]
```

Mothur MRexperiment Object

```
mothur_mrexp <- phyloseq_to_metagenomeSeq(mothur_ps)
```

```
mothur_mrexp
```

```
## MRexperiment (storageMode: environment)
## assayData: 38358 features, 192 samples
##   element names: counts
## protocolData: none
## phenoData
##   sampleNames: 1-A1 1-A10 ... 2-H9 (192 total)
##   varLabels: biosample_id titration id
##   varMetadata: labelDescription
## featureData
##   featureNames: Otu00001 Otu00002 ... Otu38358 (38358 total)
##   fvarLabels: OTUname Rank1 ... Rank6 (7 total)
##   fvarMetadata: labelDescription
## experimentData: use 'experimentData(object)'
## Annotation:
```

Save Mothur Objects

```
saveRDS(mothur_ps, "mothur/mothur_ps.rds")
saveRDS(mothur_mrexp, "mothur/mothur_mrexp.rds")
```

QIIME

QIIME Phyloseq Object

```
qiime_dir <- "qiime/otus_uc_fast_no_chimera/"
biom_file <- file.path(qiime_dir, "otu_table_mc2_w_tax_no_pynast_failures.biom")

qiime_ps <- phyloseq::import_biom(BIOMfilename = biom_file)
```

```
## Warning in strsplit(msg, "\n"): input string 1 is invalid in this locale
```

```
sample_names(qiime_ps) <- sample_names(qiime_ps) %>% str_replace("centroid=","")
```

```
## Adding sample data
phyloseq::sample_data(qiime_ps) <- meta_df
```

```
qiime_ps
```

```
## phyloseq-class experiment-level object
## otu_table()   OTU Table:        [ 2886 taxa and 189 samples ]
## sample_data() Sample Data:      [ 189 samples by 3 sample variables ]
## tax_table()   Taxonomy Table:   [ 2886 taxa by 7 taxonomic ranks ]
```

NOTE The three samples removed by the QIIME pipeline are no template controls.

```
meta_df %>% rownames_to_column() %>%
  filter(!(rowname %in% sample_names(qiime_ps)))
```

```
##   rowname biosample_id titration   id
## 1    1-A6          NTC        NA  1-A6
```

```
## 2   1-A12          NTC         NA 1-A12
## 3   1-D12          NTC         NA 1-D12
```

QIIME MRexperiment Object

```
qiime_mrexp <- phyloseq_to_metagenomeSeq(qiime_ps)
```

```
qiime_mrexp
```

```
## MRexperiment (storageMode: environment)
## assayData: 2886 features, 189 samples
##   element names: counts
## protocolData: none
## phenoData
##   sampleNames: 1-E3 2-G3 ... 1-H6 (189 total)
##   varLabels: biosample_id titration id
##   varMetadata: labelDescription
## featureData
##   featureNames: 3943182 130864 ... New.CleanUp.ReferenceOTU15362
##     (2886 total)
##   fvarLabels: OTUname Rank1 ... Rank7 (8 total)
##   fvarMetadata: labelDescription
## experimentData: use 'experimentData(object)'
## Annotation:
```

Save Mothur Objects

```
saveRDS(qiime_ps, "qiime/qiime_ps.rds")
saveRDS(qiime_mrexp, "qiime/qiime_mrexp.rds")
```

DADA2 Phyloseq Object

```
seqtab <- readRDS("dada2/seqtab_nochim.rds")
otu_tbl <- otu_table(seqtab, taxa_are_rows=FALSE)

## Rep sequences
sv_seqs <- colnames(otu_tbl)
names(sv_seqs) <- paste0("SV",1:ncol(otu_tbl))

## Rename features
colnames(otu_tbl) <- paste0("SV",1:ncol(otu_tbl))

taxa <- readRDS("dada2/taxa.rds")
rownames(taxa) <- names(sv_seqs)[match(sv_seqs, rownames(taxa))]

dada_ps <- phyloseq(otu_tbl,
                    sample_data(meta_df),
                    tax_table(taxa))

## Removing 0 entry samples
dada_samples <- sample_names(dada_ps)
```

```
dada_nonzero_sample <- dada_samples[phyloseq::sample_sums(dada_ps) != 0]
dada_ps <- phyloseq::prune_samples(dada_nonzero_sample, dada_ps)
```

```
dada_ps
```

```
## phyloseq-class experiment-level object
## otu_table()   OTU Table:         [ 3144 taxa and 191 samples ]
## sample_data() Sample Data:       [ 191 samples by 3 sample variables ]
## tax_table()   Taxonomy Table:    [ 3144 taxa by 6 taxonomic ranks ]
```

DADA2 MRexperiment Object

```
dada_mrexp <- phyloseq_to_metagenomeSeq(dada_ps)
```

```
dada_mrexp
```

```
## MRexperiment (storageMode: environment)
## assayData: 3144 features, 191 samples
##   element names: counts
## protocolData: none
## phenoData
##   sampleNames: 1-A1 1-A10 ... 2-H9 (191 total)
##   varLabels: biosample_id titration id
##   varMetadata: labelDescription
## featureData
##   featureNames: SV1 SV2 ... SV3144 (3144 total)
##   fvarLabels: OTUname Kingdom ... Genus (7 total)
##   fvarMetadata: labelDescription
## experimentData: use 'experimentData(object)'
## Annotation:
```

Save DADA2 Objects

```
saveRDS(dada_ps, "dada2/dada_ps.rds")
saveRDS(dada_mrexp, "dada2/dada_mrexp.rds")
DNAStringSet(sv_seqs) %>% writeXStringSet("dada2/sv_seqs.rds")
```

Session information

```
s_info <- devtools::session_info()
print(s_info$platform)
```

```
##  setting  value
##  version  R version 3.3.2 (2016-10-31)
##  system   x86_64, darwin15.6.0
##  ui       unknown
##  language (EN)
##  collate  en_US.UTF-8
##  tz       America/New_York
##  date     2017-03-14
```

```r
s_info$packages %>% filter(`*` == "*") %>% select(-`*`) %>%
    knitr::kable()
```

package	version	date	source
bbmle	1.0.18	2016-02-11	CRAN (R 3.3.2)
Biobase	2.34.0	2016-11-07	Bioconductor
BiocGenerics	0.20.0	2016-11-07	Bioconductor
BiocParallel	1.8.1	2016-11-07	Bioconductor
Biostrings	2.42.1	2016-12-19	Bioconductor
DESeq	1.26.0	2016-11-28	Bioconductor
DESeq2	1.15.28	2017-02-02	bioc (readonly/DESeq2@125913)
dplyr	0.5.0	2016-06-24	CRAN (R 3.3.2)
edgeR	3.16.5	2017-02-02	Bioconductor
forcats	0.2.0	2017-01-23	CRAN (R 3.3.2)
foreach	1.4.3	2015-10-13	CRAN (R 3.3.1)
GenomeInfoDb	1.10.2	2017-01-04	Bioconductor
GenomicAlignments	1.10.0	2016-11-07	Bioconductor
GenomicRanges	1.26.2	2017-01-04	Bioconductor
ggplot2	2.2.1	2016-12-30	CRAN (R 3.3.2)
glmnet	2.0-5	2016-03-17	CRAN (R 3.3.1)
IRanges	2.8.1	2016-11-18	Bioconductor
knitr	1.15.1	2016-11-22	CRAN (R 3.3.2)
lattice	0.20-34	2016-09-06	CRAN (R 3.3.2)
limma	3.30.9	2017-02-02	Bioconductor
locfit	1.5-9.1	2013-04-20	CRAN (R 3.3.1)
Matrix	1.2-8	2017-01-20	CRAN (R 3.3.2)
metagenomeSeq	1.16.0	2016-11-07	Bioconductor
modelr	0.1.0	2016-08-31	cran (@0.1.0)
permute	0.9-4	2016-09-09	CRAN (R 3.3.1)
phyloseq	1.19.1	2017-01-04	Bioconductor
ProjectTemplate	0.7	2016-08-11	CRAN (R 3.3.1)
purrr	0.2.2	2016-06-18	CRAN (R 3.3.1)
RColorBrewer	1.1-2	2014-12-07	CRAN (R 3.3.1)
readr	1.0.0	2016-08-03	CRAN (R 3.3.1)
readxl	0.1.1	2016-03-28	cran (@0.1.1)
Rqc	1.8.0	2016-11-07	Bioconductor
Rsamtools	1.26.1	2016-11-07	Bioconductor
S4Vectors	0.12.1	2016-12-19	Bioconductor
sads	0.3.1	2016-05-13	CRAN (R 3.3.2)
savR	1.12.0	2016-11-07	Bioconductor
ShortRead	1.32.0	2016-11-07	Bioconductor
stringr	1.1.0	2016-08-19	CRAN (R 3.3.1)
SummarizedExperiment	1.4.0	2016-11-07	Bioconductor
tibble	1.2	2016-08-26	CRAN (R 3.3.1)
tidyr	0.6.1	2017-01-10	CRAN (R 3.3.2)
tidyverse	1.1.1	2017-01-27	CRAN (R 3.3.2)
vegan	2.4-2	2017-01-17	CRAN (R 3.3.2)
XVector	0.14.0	2016-11-07	Bioconductor