

# Package ‘BioDataome’

November 30, 2017

**Title** Functions used to preprocess datasets stored in BioDataome

**Version** 0.0.0.9000

**Description** BioDataome package contains all the functions used to download, preprocess and annotate gene expression and methylation microarray data from Gene Expression Omnibus, as well as RNASeq data from recount.

**Depends** R (>= 3.4.2), foreach, SCAN.UPC

**Imports** GEOquery, Biobase, RCurl, Rfast, SCAN.UPC, XML, doParallel, parallel, rentrez

**License** What license is it under?

**Encoding** UTF-8

**LazyData** true

**RoxygenNote** 6.0.1

**VignetteBuilder** knitr

**Suggests** BiocStyle, knitr, rmarkdown

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compareDsetList	<i>Column wise comparison of a dataset to a list of datasets.</i>
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### Description

This function finds all datasets of the list y for which dataset x shares samples. The datasets are in the form variables (probes) x samples. The number of variables (probes) in both datasets should be the same.

### Usage

```
compareDsetList(x, y)
```

### Arguments

x	the path to a normalized dataset x
y	a character vector of all paths to datasets to compare

### Value

a character vector of all datasets for which dataset x shares at least one sample, separated by ;

### Examples

Let us assume we want to compare normalized gene expression dataset GSE86013 with datasets: GSE86015, GSE9008, GSE9119. x and y can be either local paths where .Rda normalized data are stored or links to the csv files in BioDataome.

First example runs with datasets stored in .csv in BioDataome.

Since these datasets are large we propose to use fread from package data.table to read datasets faster

```
install.packages("data.table")
library("data.table")
x<-"http://dataome.mensxmachina.org/data/Homo%20sapiens/GPL570/GSE86013.csv"
y<-c("GSE86015.csv", "GSE9008.csv", "GSE9119.csv")
y<-paste0("http://dataome.mensxmachina.org/data/Homo%20sapiens/GPL570/", y)
commonGSEs<-compareDsetList(x,y)
```

---

compareDsets	<i>Column wise comparison of two datasets This function finds how many samples are shared between two datasets. The datasets are in the form variables (probes) x samples. The number of variables (probes) in both datasets should be the same</i>
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### Description

Column wise comparison of two datasets This function finds how many samples are shared between two datasets. The datasets are in the form variables (probes) x samples. The number of variables (probes) in both datasets should be the same

**Usage**

```
compareDsets(d1, d2)
```

**Arguments**

d1                    a numeric matrix of a dataset  
d2                    a numeric matrix of a dataset

**Value**

the number of equal samples

**Examples**

```
Let us assume we want to compare two normalized gene expression datasets from the same platform
d1<-get(load(url("http://dataome.mensxmachina.org/data/Homo%20sapiens/GPL570/GSE86013.Rda")))
d2<-get(load(url("http://dataome.mensxmachina.org/data/Homo%20sapiens/GPL570/GSE86015.Rda")))
commons<-compareDsets(d1,d2)
```

---

controlSamples

*Discover control samples from phenotype data in GEO*

---

**Description**

This function discovers control samples from the series matrix found in GEO. It searches for specific keywords that are often used to denote controls in specific columns of the series matrices.

**Usage**

```
controlSamples(d)
```

**Arguments**

d                    a data frame with the contents of series matrix

**Value**

a data frame of GEO sample ids (i.e. GSM60555) and their class.

**Examples**

```
phenos<-downloadPhenotypePlatform("GSE11761","GPL570")
controls<-controlSamples(phenos)
```

---

curateGSE	<i>Run all steps to download, preprocess and annotate a GEO dataset</i>
-----------	---

---

### Description

Given a GSE id this function downloads, preprocesses, annotates a study and also creates the sample phenotype metadata.

### Usage

```
curateGSE(x, y, z = getwd(), keepRaw = FALSE)
```

### Arguments

x	a GSE series ID
y	a GEO technology id (GPL)
z	the path to save the downloaded files. By default this value is set to the working directory.

### Value

a list with three components:

- "metadata" a data frame of the metadata with sample phenotype information
- "dataNorm" a numeric matrix of the preprocessed data with variables (probes) as rows and samples as columns

### Examples

```
curateGSE("GSE11761", "GPL570", getwd())
```

---

curateRecountRNASeq	<i>Run all steps to download, preprocess and annotate an RNASeq dataset from Recount</i>
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---

### Description

Run all steps to download, preprocess and annotate an RNASeq dataset from Recount

### Usage

```
curateRecountRNASeq(x, y)
```

### Arguments

x	a recount dataset ID
y	the path to write the output

**Value**

writes in the given path two data frames, the preprocessed data and the metadata file with phenotype information

**Examples**

```
curatedRecount<-curateRecountRNASeq("SRP032775",getwd())
```

---

`diseasetoChildrenNodes`

*Map a Disease Ontology (D-O) term to the first children nodes in D-O*

---

**Description**

This function uses internal look up data to map a disease to its first children node.

**Usage**

```
diseasetoChildrenNodes(x)
```

**Arguments**

x                      a disease in D-O terms

**Value**

the first children node of x disease

**Examples**

```
DOChild<-diseasetoChildrenNodes("vesiculitis")
```

---

`diseasetoParentNodes`

*Map a Disease Ontology (D-O) term to the parent nodes in D-O*

---

**Description**

This function uses internal look up data to map a disease to its parent node.

**Usage**

```
diseasetoParentNodes(x)
```

**Arguments**

x                      a disease in D-O terms

**Value**

the parent node of x disease

**Examples**

```
DOParent<-diseaseToParentNodes("vesiculitis")
```

---

downloadPhenotype	<i>Download series matrices from GEO for a given study</i>
-------------------	--

---

**Description**

This function downloads all series matrices related to a given GEO Series (GSE) and saves them in a list. The same GSE study (i.e. GSE11761) may be related to more than one platforms (i.e GPL570 and GPL1261). The length of the output list is the number of the related platforms.

**Usage**

```
downloadPhenotype(x)
```

**Arguments**

x	a GEO Series id (GSE)
---	-----------------------

**Value**

a list of series matrices related to the given study

**Examples**

```
downloadPhenotype("GSE11761")
```

---

downloadPhenotypePlatform	<i>Download series matrix from GEO for a given study and platform</i>
---------------------------	---

---

**Description**

This function downloads returns the series matrix related to a given GEO Series (GSE)

**Usage**

```
downloadPhenotypePlatform(x, y)
```

**Arguments**

x	a GEO Series id (GSE)
y	a GEO platform id (GPL)

**Value**

a data frame with the contents of the series matrix found in GEO

**Examples**

```
downloadPhenotypePlatform("GSE11761", "GPL570")
```

---

downloadRaw	<i>Download raw files from GEO for a given study</i>
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---

**Description**

Download raw files from GEO for a given study

**Usage**

```
downloadRaw(x, y = getwd())
```

**Arguments**

x	a GEO Series id (GSE)
y	the path to save the downloaded files. By default this value is set to the working directory

**Value**

downloadRaw creates a directory in the given path with the GSE name and saves there the compressed RAW files.

**Examples**

Download the raw files of GSE11761 study and save them in a directory named GSE11761 in the working directory

```
downloadRaw("GSE11761",getwd())
```

---

downloadRecount	<i>Download gene-level RangedSummarizedExperiment data from Recount</i>
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---

**Description**

This function downloads the RangedSummarizedExperiment object with the data summarized at the gene level from Recount (<https://jhubiostatistics.shinyapps.io/recount/>)

**Usage**

```
downloadRecount(x)
```

**Arguments**

x	a recount dataset ID
y	the destination path for the downloaded RangedSummarizedExperiment object

**Value**

RangedSummarizedExperiment object for the given study

**Examples**

```
downloadRecount("SRP032775",getwd())
```

---

entrezIDtoGSE

*Find all GSE ids for a given Entrez query*


---

**Description**

Find all GSE ids for a given Entrez query

**Usage**

```
entrezIDtoGSE(x)
```

**Arguments**

x                      an esearch object as a result of an entrez\_search query

**Value**

a matrix the first column of which is the GSE id and the second the entrezID

**Examples**

```
query GEO for all Homo sapiens studies with sample size between 200-300, measured with GPL570 and provide CEL
r_search <- entrez_search(db="gds", term="Homo sapiens[ORGN] AND CEL[SFIL] AND gp1570[ACCN] AND 200:300[Number]
entrezIDtoGSE(r_search)
```

---

GSEmetadata

*Create sample phenotype metadata of a GEO dataset including control discovery*


---

**Description**

Given a GSE id this function downloads phenotype data from GEO for a specific study, discovers control samples and returns all sample phenotype metadata

**Usage**

```
GSEmetadata(x, y)
```

**Arguments**

x                      a GSE series ID  
y                      a GEO platform id (GPL)

**Value**

a data frame of metadata with columns: sample IDs, Class and all other GEO phenotype data found in series matrix



**Examples**

```
metadata<-GSEmetadata("GSE11761","GPL570")
```

---

GSEtoDisease	<i>Annotate a study (GSE) with a disease term from the Disease Ontology by exploiting both PubTator and GEO</i>
--------------	---

---

**Description**

Given a GSE id this function annotates the study with a disease term from the Disease Ontology (D-O): <http://disease-ontology.org/> It provides the most specific disease term, meaning the term with the highest depth in the D-O.

**Usage**

```
GSEtoDisease(GSE)
```

**Arguments**

GSE                      a GSE series ID

**Value**

a character vector of all related diseases, separated by ;

**Examples**

```
diseases<-GSEtoDisease("GSE10245")
```

---

GSEtoDiseaseGEO	<i>Annotate a study (GSE) with a disease term from the Disease Ontology by exploiting only GEO</i>
-----------------	--

---

**Description**

Given a GSE id this function annotates the study with a disease term from the Disease Ontology (D-O): <http://disease-ontology.org/> It provides the most specific disease term, meaning the term with the highest depth in the D-O.

**Usage**

```
GSEtoDiseaseGEO(GSE)
```

**Arguments**

GSE                      a GSE series ID

**Value**

a character vector of all related diseases, separated by ;

**Examples**

```
diseases<-GSEtoDiseaseGEO("GSE10245")
```

---

```
preprocessGEO
```

---

```
Preprocess CEL files with SCAN
```

---

**Description**

This function calls the SCAN method as described in Piccolo SR, Sun Y, Campbell JD, Lenburg ME, Bild AH and Johnson WE (2012). A single-sample microarray normalization method to facilitate personalized-medicine workflows. *Genomics*, 100(6), pp. 337-344.

**Usage**

```
preprocessGEO(x, y)
```

**Arguments**

x                      the path where the CEL files are stored  
y                      the number of cores to run in parallel

**Value**

a matrix of dimensions: probes x samples with the normalized expression values

**Examples**

```
Assuming that CEL files are located in working directory  
preprocessGEO(getwd(),3)
```

---

```
preprocessGEOmethylation
```

---

```
Preprocess IDAT files from Illumina HumanMethylation450 BeadChip
```

---

**Description**

This function utilizes minfi Package to convert data into methylation measurements.

**Usage**

```
preprocessGEOmethylation(x)
```

**Arguments**

x                      a character array with the paths to the idat files

**Value**

a matrix of dimensions: probes x samples with the normalized methylation values

**Examples**

Assuming there is a directory named GSE78279 in the working directory where idat files are stored and "GSM2071074\_8655685078\_R03C02" and "GSM2071074\_8655685078\_R03C02" are the file names for the idat files, then x should be:

```
x<-c("GSM2071074_8655685078_R03C02", "GSM2071075_8655685078_R04C02")
x<-file.path(getwd(), "GSE78279", x)
dataNorm<-preprocessGEOmethylation(x)
```

---

`recountIDtoGSE`*Map a recount dataset ID to GSE ID*

---

**Description**

Map a recount dataset ID to GSE ID

**Usage**

```
recountIDtoGSE(x)
```

**Arguments**

x                      a recount dataset ID

**Value**

a GSE series ID

**Examples**

```
recountIDtoGSE("SRP032775")
```

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