

Review article

The role of mitochondria in amyotrophic lateral sclerosis



Emma F. Smith, Pamela J. Shaw, Kurt J. De Vos*

Sheffield Institute for Translational Neuroscience (SITraN), Department of Neuroscience, University of Sheffield, Sheffield, S10 2HQ, UK

HIGHLIGHTS

- Mitochondrial dysfunction is one of the earliest pathophysiological events in amyotrophic lateral sclerosis (ALS).
- ALS-associated mutant proteins accumulate in mitochondria and cause mitochondrial damage.
- ALS-associated mitochondrial dysfunction occurs at multiple levels.
- ALS affects mitochondrial respiration and ATP production, calcium handling, dynamics, and apoptotic signalling.

ARTICLE INFO

Article history:

Received 10 March 2017

Accepted 27 June 2017

Available online 30 June 2017

Keywords:

Amyotrophic lateral sclerosis
 Motor neuron disease
 Mitochondria
 Neurodegeneration
 Oxidative stress
 Mitophagy
 Axonal transport
 Oxidative phosphorylation
 Apoptosis

ABSTRACT

Mitochondria are unique organelles that are essential for a variety of cellular processes including energy metabolism, calcium homeostasis, lipid biosynthesis, and apoptosis. Mitochondrial dysfunction is a prevalent feature of many neurodegenerative diseases including motor neuron disorders such as amyotrophic lateral sclerosis (ALS). Disruption of mitochondrial structure, dynamics, bioenergetics and calcium buffering has been extensively reported in ALS patients and model systems and has been suggested to be directly involved in disease pathogenesis. Here we review the alterations in mitochondrial parameters in ALS and examine the common pathways to dysfunction.

© 2017 The Authors. Published by Elsevier Ireland Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

Contents

1. Amyotrophic lateral sclerosis	2
2. The role of mitochondria in ALS pathogenesis	2
2.1. Structural evidence of damaged mitochondria	2
2.2. ALS-associated proteins interact with mitochondria	3
2.3. Defective mitochondrial respiration and ATP production	4
2.4. Oxidative stress	5
2.5. Calcium mishandling	6
2.6. Pro-apoptotic signalling	7
2.7. Impaired mitochondrial dynamics	8
2.7.1. Aberrant mitochondrial fission and fusion	8
2.7.2. Disrupted mitochondrial quality control	8

* Corresponding author.

E-mail address: k.de.vos@sheffield.ac.uk (K.J. De Vos).

2.7.3. Impaired axonal transport of mitochondria	10
3. Mitochondria as therapeutic targets in ALS	11
4. Conclusions	12
Acknowledgements	12
References	12

1. Amyotrophic lateral sclerosis

Motor neuron disease (MND) describes a group of neurological disorders characterised by the selective loss of motor neurons. Amyotrophic lateral sclerosis (ALS), the most common subtype of MND, is characterised by a progressive degeneration of both upper and lower motor neurons, resulting in muscle atrophy, gradual paralysis, and death, usually resulting from respiratory failure. ALS has a worldwide prevalence of 4–6 in 100,000, with differences noted between populations [1,2]. The average age of onset is between 55 and 65 years of age, however familial cases are frequently associated with an earlier age of onset [3]. The average survival is 2–3 years from diagnosis, with only 25% of patients surviving 5 years and 5–10% surviving 10 years post-diagnosis [4].

ALS clinically overlaps with frontotemporal dementia (FTD). FTD is a common cause of dementia in adults under the age of 65 that is characterised by neurodegeneration of the frontal and temporal cortex [5]. Around 50% of ALS patients display evidence of frontal and temporal lobe dysfunction on detailed neuropsychological testing and up to 5–10% of ALS cases present with clinically diagnosed FTD. Similarly, while approximately 15% of FTD patients go on to develop ALS, about half display some degree of motor involvement [6].

Approximately 10% of ALS cases follow a familial, mostly autosomal dominant inheritance pattern (familial ALS). The remaining 90% of cases have no clear genetic basis (sporadic ALS). To date mutations in over 20 genes have been associated with ALS, including superoxide dismutase 1 (SOD1), TAR DNA binding protein (TARDBP; TDP-43), Fused in sarcoma (FUS), and C9orf72 (Table 1; [7]). Together mutations in these genes account for approximately 60% of familial ALS and 11% of sporadic ALS. In agreement with the clinical overlap between ALS and FTD, the GGGGCC (G4C2) repeat expansion mutation in C9orf72 was found to be the most common genetic cause of both ALS and FTD, accounting for approximately 40 and 25% of familial cases, respectively [8–10; reviewed in Ref. [11]]. The genes affected in ALS have been implicated in a wide range of cellular pathways, suggesting that ALS is a multi-factorial disease. Possible pathogenic mechanisms underlying motor neuron degeneration include RNA toxicity, excitotoxicity, disruption of proteostasis, defective axonal transport, oxidative stress, and mitochondrial dysfunction [reviewed in Ref. [12]].

2. The role of mitochondria in ALS pathogenesis

Mitochondria play a central role in cell survival and metabolism. In addition to their well-known role as producers of ATP via oxidative phosphorylation, mitochondria play important roles in phospholipid biogenesis, calcium homeostasis, and apoptosis. Mitochondria are of particular importance in neurons. Neurons have high metabolic requirements – the brain consumes 20% of the body's resting ATP production despite being only 2% of its mass [reviewed in Refs. [13,14]]. Moreover, mitochondria are essential calcium buffering organelles in neurons that modulate local calcium dynamics to, for example, modulate neurotransmitter release [reviewed in Ref. [15]]. Neurons are long-lived cells that persist throughout the lifespan of the individual and as such are more susceptible to the accumulating damage arising from mitochondrial dysfunction [reviewed in Ref. [16]]. Accordingly, the maintenance

of a healthy pool of correctly localised mitochondria is essential for neuronal survival and function. It is not surprising therefore that mitochondrial dysfunction has been linked to a large number neurodegenerative disorders including ALS. Many of the identified ALS genes have a role in mitochondrial-associated functions (Table 1) and evidence gathered from *in vitro* and *in vivo* disease models and from patient studies strongly implicates the dysfunction of mitochondria as a core ALS disease component.

ALS associated mitochondrial dysfunction comes in many guises, including defective oxidative phosphorylation, production of reactive oxygen species (ROS), impaired calcium buffering capacity and defective mitochondrial dynamics. Furthermore, with the possible exception RNA toxicity, mitochondrial dysfunction appears to be directly or indirectly linked to all of the postulated “non-mitochondrial” mechanisms of toxicity associated with ALS, including excitotoxicity, loss of protein homeostasis and defective axonal transport.

2.1. Structural evidence of damaged mitochondria

Structurally altered and aggregated mitochondria, with a swollen and vacuolated appearance, were one of the first changes observed in ALS patient motor neurons [17,18] and in Bunina bodies [19]. Sporadic ALS cases occasionally additionally display axonal swellings consisting of neurofilament accumulations, swollen mitochondria and secondary lysosomes [20]. Morphologically abnormal mitochondria are also consistently reported in animal and cell models of ALS, with a tendency towards a fragmented mitochondrial population being observed. Expression of wild type or ALS mutant TDP-43 (M337V, Q331K, A315T) resulted in aggregated, fragmented and vacuolated mitochondria [21–23]. Overexpression of SOD1 G93A had similar effects and mitochondria were significantly less elongated and more spherical in motor neurons isolated from SOD1 G93A transgenic mice [24–26] and *in vivo* in motor neurons of early symptomatic SOD1 G37R and SOD1 G85R transgenic mice [27]. Furthermore, in SOD1 G93A transgenic mice mitochondria were found in abnormal clusters along the axon [22].

Overexpression of ALS mutant FUS R521G or R521H in cultured motor neurons resulted in mitochondrial shortening which was exacerbated by the presence of FUS in the cytosol [28]. Similar results were seen in both HT22 cells and primary cortical neurons expressing mutant FUS P525L which causes a juvenile form of ALS, and in a FUS P525L transgenic mouse model [29,30]. Subtle fragmentation of the mitochondrial network has also been identified in fibroblasts of ALS patients with C9orf72 repeat expansions [31], and swollen mitochondria were reported in an iPSC model of C9orf72-associated ALS [32]. Aggregation of structurally altered mitochondria was reported in cortical neurons from Als1n knock-out mice [33]. Similarly, mitochondria co-aggregated with TDP-43 in heterozygous knock-in mice bearing an ALS-associated R155H mutation in valosin-containing protein (VCP; also called p97, cdc48 in yeast – further referred to as VCP/p97) [34,35].

Direct evidence that disruption of mitochondrial structure (and as a consequence function; see below) may contribute to the aetiology of ALS comes from the discovery of causative mutations in the mitochondrial protein CHCHD10 which is localised to contact sites between the inner and outer mitochondrial membrane [36]. ALS-associated mutations in CHCHD10 disrupt mitochondrial cristae

Table 1

Impact of ALS-associated genes on mitochondrial function. Pathogenic variants of proteins implicated in ALS have been linked to altered mitochondrial function (Ref. <http://alsod.iop.kcl.ac.uk/home.aspx>; [7]).

ALS locus	Gene	Protein	Potential consequence of mutation on mitochondrial function
ALS 1	SOD1	Cu, Zn superoxide dismutase 1	Mutant protein aggregates in IMS; decreased ATP generation; increased cellular ROS and ROS induced cellular damage; imbalance in calcium homeostasis; induction of apoptosis via VDAC inhibition and Bcl-2 binding; disrupted mitochondrial architecture; impaired mitochondrial network dynamics and axonal transport; impaired mitochondrial clearance by mitophagy; disrupted ER-mitochondria contacts
ALS 2	ALS2	Alsin	Reduced autophagosome formation and decreased mitophagy
ALS 6	FUS	RNA-binding protein FUS	Decreased ATP generation; increased ROS levels; loss of calcium homeostasis and disruption of ER-mitochondria contacts; reduced mitophagy-related gene expression; disrupted mitochondrial architecture; impaired mitochondrial transport via disrupted kinesin gene expression
ALS 8	VAPB	Vesicle-associated membrane protein-associated protein B	Impaired calcium homeostasis; decreased anterograde axonal transport; disrupted ER-mitochondria contacts
ALS 10	TARDBP	TAR DNA-binding protein 43	TDP-43 aggregates in mitochondria and disrupts mtDNA transcription; decreased ATP generation; impaired calcium homeostasis and disrupted ER-mitochondria contacts; disrupted mitochondrial architecture; altered mitochondrial network dynamics and impaired mitochondrial axonal transport; reduced mitophagy related gene expression; impaired mitochondrial clearance by mitophagy
ALS 12	OPTN	Optineurin	Reduced mitochondrial clearance by mitophagy
ALS 14	VCP	Valosin-containing protein	Decreased ATP levels; mitochondrial uncoupling; reduced mitochondrial clearance by mitophagy
ALS 16	SIGMAR1	Sigma non-opioid intracellular receptor 1	Reduced ATP generation; disrupted ER-mitochondria contacts; dysregulated calcium homeostasis; reduced axonal transport
ALS-FTD1	C9orf72	Chromosome 9 open reading frame 72	DPR proteins interact with mitochondrial ribosomal proteins; altered MMP; increased cellular ROS levels; poly(GR) DPR induced oxidative stress; impaired autophagy; disrupted mitochondrial architecture and altered mitochondrial network dynamics
ALS-FTD2	CHCHD10	Coiled-coil-helix-coiled-coil-helix domain containing 10	Disrupted mitochondrial architecture; decreased electron transport chain activity
ALS	SQSTM1	p62/Sequestosome 1	Reduced mitochondrial clearance by mitophagy; reduced MMP

Abbreviations: ATP – adenosine triphosphate; DPR – dipeptide repeat protein; IMS – intermembrane space; MAM – mitochondria associated ER membrane; MMP – mitochondrial membrane potential; ROS – reactive oxygen species.

and have a profound effect on mitochondrial structure [37]. Deformation and loss of mitochondrial cristae have also been reported in C9orf72-related ALS and FTD (C9ALS/FTD) patient fibroblasts, *in vitro* and *in vivo* in SOD1 G93A, TDP-43 A513T and FUS P525L ALS models and in Alsin knockout mice [29,31–33,38,39]. In SOD1 G93A and FUS P525L transgenic mice dilated cristae are already present at disease onset [30,40].

Thus, mitochondrial structure and the mitochondrial network appears to be disrupted in most if not all models of familial ALS and in ALS patients. Moreover, the structural damage to mitochondria and fragmentation of the mitochondrial network was reported to occur in early disease stages in *in vivo* models of ALS indicating that mitochondrial morphological alterations occur potentially as an upstream source of degeneration rather than a consequence [22,23,27].

2.2. ALS-associated proteins interact with mitochondria

A number of proteins that have been linked to familial and sporadic ALS, including SOD1, TDP-43, FUS, C9orf72, and the C9orf72 GGGGCC repeat expansion-associated glycine/arginine (GR) dipeptide repeat protein (DPR), have been shown to interact with mitochondria [29,41–45]. The interaction of these ALS-associated proteins with the mitochondria appears to be instrumental to the induction of mitochondrial damage associated with ALS.

Mutant SOD1 localises to the intermembrane space (IMS), where it has been shown to aggregate, and reduce the activity of the electron transport chain (ETC) complexes (see below) [46,47]. Furthermore, SOD1 aggregates have been proposed to interfere with the activity of voltage-dependent anion channel 1 (VDAC1) which is responsible for the exchange of ATP, ADP and other respira-

tory substrates across the outer mitochondrial membrane (OMM) [reviewed in Ref. [48]]. Direct interaction of ALS mutant SOD1 with VDAC1 inhibits channel conductance and reduces its permeability to ADP at both presymptomatic and symptomatic disease stages in the spinal cord of SOD1 G93A transgenic rats. Furthermore, an approximately 25% reduction in VDAC1 activity accelerated disease in SOD1 G37R transgenic mice [49]. Mutant misfolded SOD1 has also been shown to interact with Bcl-2 family proteins on the OMM, leading to pro-apoptotic changes (see below) [50].

TDP-43 and to a greater extent ALS mutant TDP-43 accumulates in mitochondria where it preferentially binds the mRNAs of the mtDNA-encoded complex I subunits ND3 and ND6 and causes complex I disassembly by impairing their transcription. Accumulation of ALS mutant TDP-43 in mitochondria appears to be mediated by internal mitochondrial targeting sequences in TDP-43 [42]. Since cytoplasmic TDP-43 accumulation is a hallmark pathology in most ALS (but not SOD1 or FUS-related ALS), this may also explain the mitochondrial defects observed in sporadic ALS and other familial ALS cases.

Both wild type FUS and ALS mutant FUS P525L co-purify with mitochondria and this is at least in part due to their interaction with the mitochondrial chaperone heat shock protein (HSP) of 60 kDa (HSP60) [29]. Mitochondrial localisation of FUS correlated with augmented ROS levels [29], and overexpression of FUS has been shown to reduce mitochondrial ATP production [51]. Reducing the association of FUS with mitochondria by knockdown of HSP60 expression ameliorated neurodegeneration in FUS transgenic *Drosophila* [29].

Several mitochondrial proteins have been identified as possible C9orf72-interacting proteins in a BioLP proteomics screen, including members of the IMM solute carrier family, VDAC3 and

translocase of the inner mitochondrial membrane 50 (TIMM50). Furthermore, C9orf72 was detected in mitochondria-enriched fractions [44]. The relevance of this association is not yet clear. Another link between C9ALS/FTD and mitochondria comes from the finding that poly(GR) DPRs preferentially bind to mitochondrial ribosomal proteins [43].

2.3. Defective mitochondrial respiration and ATP production

Reductions in cellular respiration and ATP production are well documented in ALS (Fig. 1). In post-mortem spinal cord of sporadic ALS patients the activity of all ETC complexes, complex I, II, III, and IV was found to be reduced [52,53]. In addition, the activity of complexes I and IV were reported to be impaired in skeletal muscle [54–56] while complex I activity and ATP levels were reduced in lymphocytes of sporadic ALS patients [57]. Counterintuitively, in fibroblasts obtained from skin biopsies of sporadic ALS patients the mitochondrial membrane potential (MMP) was increased compared to healthy controls. Possibly the increased MMP observed in fibroblasts reflects an attempt to rescue inefficient ATP synthesis or the metabolic differences between skin fibroblasts and central nervous system (CNS) cells [58]. Indeed, similar to fibroblasts obtained from sporadic patients, MMP was increased in C9orf72 patient fibroblasts [31] but was significantly decreased in iPSC-derived motor neurons reprogrammed from C9orf72 patient fibroblasts [32].

In SOD1 G93A transgenic mice impaired ATP synthesis and depressed mitochondrial respiration rates in the brain and spinal cord were found well before disease onset and they persist throughout the course of the disease [39,45]. Associated with these changes, decreased complex I+III, II+III and IV activity was observed in the spinal cord of 17-week old symptomatic SOD1 G93A transgenic mice [45] and complex IV activity was reduced in forebrain mitochondria isolated from presymptomatic, symptomatic and end-stage SOD1 G93A transgenic mice [39,45]. Analysis of regional changes in ETC complex activities further showed a selective decrease of complex I and IV activity in the ventral horn of the spinal cord of these mice prior to the onset of disease which further declined as disease progressed [59]. In these cases, the reduced activity of complex IV may relate to impaired association of cytochrome c with the IMM [39,45]. A decrease in complex I activity was also confirmed in *ex vivo* motor cortex slices from SOD1 G93A mice [60] (Fig. 1).

Analysis of mutant SOD1 ALS cell models recaptured the defects found in SOD1 G93A mice. SOD1 I113T patient fibroblast lines showed reduced oxidative phosphorylation compared to non-disease controls [61]. Stable low level expression of SOD1 G93A or G37R in NSC-34 motor neuron-like cell lines reduced the activity of complex II and IV [62,63] accompanied by reduced MMP [64] while doxycycline-induced expression of SOD1 G93A, A4V, H46R, H80R, D90A, or D123H in the same cell line caused a decrease in complex I, II+III and IV activity which was accompanied by reduced cellular ATP levels [46]. In N2A neuroblastoma cells overexpression of SOD1 G37R resulted in reduced complex I activity which correlated with a drop in MMP and reduced levels of cytosolic ATP, but complex II, III and IV activities were unaffected [63].

In contrast to these studies, Bowling et al. found increased complex I activity in post-mortem frontal cortex of familial ALS patients with an SOD1 A4V mutation [65] and the same group reported an increase in complex I activity in the forebrain of SOD1 G93A transgenic mice [66]. Furthermore, when they assessed mitochondrial respiration solely via complex II, the mitochondrial respiration rate was not different in SOD1 G93A transgenic mice compared to wild type SOD1 transgenic mice [67]. Similarly, others found no evidence for reduced mitochondrial respiration via complex I or II in presymptomatic SOD1 G93A transgenic mice [68]. It is not readily

apparent what accounts for the discrepancy between these studies, but these measurements are particularly sensitive to variations in experimental conditions and differences between the SOD1 G93A mouse strains cannot be excluded.

Nevertheless, despite differences between studies and cell models, the overarching trend is that ALS mutant SOD1 decreases ETC activity and causes impaired ATP production. Moreover, the presymptomatic occurrence of oxidative phosphorylation disruption in SOD1 G93A transgenic mice suggests it may have a causative role in this ALS model.

Defective mitochondrial respiration and ATP production have also been observed in models of non-SOD1 related familial ALS. Depolarisation of mitochondria was described in NSC-34 cells expressing wild type or mutant TDP-43 (Q331K and M337V) and primary motor neurons expressing TDP-43 M337V [21,23,69,70]. In NSC-34 cells TDP-43-associated MMP reduction was accompanied by decreased complex I activity [70]. Similarly, in TDP-43 G298S and A382T patient fibroblasts reduced MMP was accompanied by decreased complex I activity, reduced oxygen consumption and decreased ATP levels [42]. However, this may not always be the case as others reported reduced MMP, but no changes in mitochondrial respiration or ATP content in 3 independent TDP-43 A382T patient fibroblast lines [31].

As was the case for TDP-43, overexpression of wild type or mutant FUS P525L caused a reduction in MMP in HEK293 cells [29] and ATP production was impaired upon expression of wild type or ALS mutant FUS R521C or R518K in NSC-34 cells [51]. Similarly overexpression of the sigma non-opioid intracellular receptor 1 (Sig1R) E102Q mutant in N2A cells reduced ATP production [71]. Decreased MMP was further observed in *SQSTM1* knockout MEFs and in SH-SY5Y, mouse cortical neurons and astrocytes in which VCP/p97 expression was ablated using siRNA to model ALS-associated loss of function mutants in p62/Sequestosome1 and VCP/p97, respectively [72,73]. In case of VCP/p97 the same results were obtained in VCP/p97 R155C, R155H and R191Q ALS patient fibroblast lines [73]. In these cell models of VCP/p97-associated ALS the decrease in MMP was accompanied by a decrease in ATP levels but an increase in the rate of mitochondrial respiration and oxygen consumption, suggesting that depletion of MMP is due to uncoupling rather than ETC activity deficits [73]. Finally, analysis of fibroblasts from patients with mutations in the mitochondrial protein CHCHD10 revealed impaired ETC activity at complexes I, II, III and IV which was accompanied by a severe bioenergetics deficit [36,37].

Neurons do not operate as isolated units, but depend on the surrounding cellular environment and supporting cells such as microglia and astrocytes for survival and function. Accordingly, it is now clear that ALS is a non-cell-autonomous disease with astrocytes in particular associated with disease progression [74, reviewed in Refs. [12,75]]. One way in which glia support neurons is by shuttling of lactate, produced by conversion of the glycolysis product pyruvate, to the neighbouring neuron where it is converted back to pyruvate which is subsequently converted to acetyl-CoA and enters the Krebs cycle [76, reviewed in Ref. [77]]. In fact, evidence suggests that glial lactate is the main energy source of neurons (i.e. the Astrocyte-Neuron Lactate Shuttle Hypothesis (ANLSH)). Several lines of evidence suggest that impairment of the ANLSH may be involved in the bioenergetics deficits observed in ALS. The expression of phosphoglycerate kinase 1 (PGK1) which transfers phosphate from 1,3-bisphosphoglycerate (1,3-BPG) to ADP to produce 3-phosphoglycerate (3-PG) and ATP during glycolysis, as well as of the monocarboxylate transporter 4 (MCT4; *Slc16a4*), which is involved in the transport of lactate are downregulated in SOD1 G93A astrocytes [78]. Similarly, MCT1 (*Slc16a1*) expression is downregulated in oligodendrocytes in the spinal cord of SOD1 G93A transgenic mice. Furthermore, consistent with a pos-

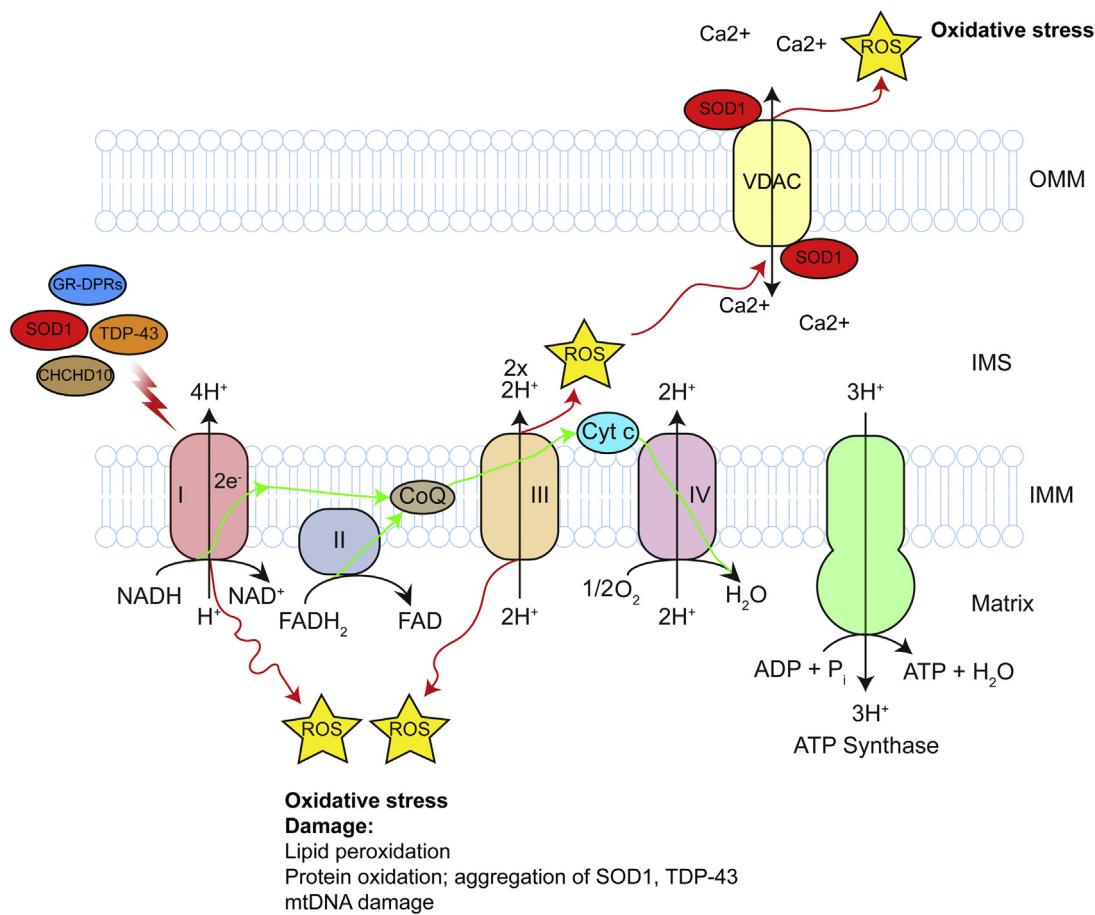


Fig. 1. Defective mitochondrial respiration, ATP production and oxidative stress.

The activity of the electron transport chain is required for the generation of cellular ATP. The activity of the complexes involved in the electron transport chain have been shown to be decreased in SOD1, TDP-43 and CHCHD10-related ALS. This results in decreased MMP and ATP generation, and increased generation of ROS. See text for details.

sible role in neurodegeneration knock down of MTC1 caused motor neuron death *in vivo* [79]. Hence reductions in neuronal respiration and ATP production may be caused by failure of glia to provide respiratory substrates.

Despite differences in mechanisms and specific observations depending on the models studied, decreased ETC activity and ATP levels emerge as a common feature in ALS. It is conceivable that, in line with the high energy demands of neurons, gradual depletion of ATP due to reduced respiration may trigger degeneration.

2.4. Oxidative stress

ROS are a natural by-product of oxidative phosphorylation. The ETC is responsible for the majority of ROS produced in a cell, with free radicals, mainly superoxide anion (O_2^-), produced at complexes I, II and III as a result of incomplete, premature reduction of oxygen [reviewed in Refs. [80,81]]. (Fig. 1). Dismutation of O_2^- , a reaction catalysed by SOD1 and SOD2, gives rise to H_2O_2 which can further generate hydroxyl anions (OH^-) by partial reduction. O_2^- can also react with nitric oxide (NO^-) to form peroxynitrite ($ONOO^-$). ROS generated in cells can act as signalling molecules, but when excessively produced can lead to cellular damage to DNA, proteins and lipids and result in reduced efficiency of cellular processes, induction of inflammatory pathways, excitotoxicity, protein aggregation and endoplasmic reticulum (ER) stress or cell death [reviewed in Refs. [82,83]]. By proximity, mitochondria are particularly susceptible to ROS induced damage to mtDNA, proteins and lipids. Especially mtDNA appears vulnerable and this is

exacerbated by limited DNA repair mechanisms in mitochondria. Resultant mitochondrial damage has been shown to affect mitochondrial function and is implicated in ageing [reviewed in Refs. [84,85]].

Increased levels of ROS and ROS-associated damage have been widely reported in ALS [reviewed in Ref. [86]]. Increased markers of ROS damage have been found in biofluids of patients with sporadic ALS [87–90] as well as in post-mortem tissue [91–94]. Similarly, increased ROS levels were reported in lymphoblasts of familial ALS cases with SOD1 mutations [95] and fibroblasts of patients with C9orf72 G4C2 repeat expansions [31]. In contrast, analysis of lymphoblasts and fibroblast cell lines from sporadic ALS patients or familial ALS patients with a TDP-43 A382T mutation did not show evidence of increased ROS production or oxidative damage [31,95–97], but some were more sensitive to NO^- generating chemicals [98]. Possibly specific culture conditions and respiratory substrates (glucose v galactose) used in these studies can explain the variation in observations. Alternatively, peripheral tissues may not fully model CNS disease.

Oxidative damage to DNA, RNA, proteins and lipids has been widely reported in SOD1 G93A rodent models and cell models [reviewed in Ref. [86]]. DNA and RNA appear to be especially vulnerable to oxidation, and mRNA oxidation has been shown to precede motor neuron degeneration and cause reduced expression of the encoded proteins in SOD1 G93A transgenic mice [94]. Interestingly mRNAs coding for ETC complexes and ATP synthase were selectively susceptible to oxidation [94]. Furthermore, SOD1 itself is a target of oxidative damage [99] and this has been linked

to its misfolding and aggregation [100]. Since misfolded, aggregated SOD1 has been shown to disrupt mitochondrial function and increase superoxide production [49,101,102] a vicious cycle mechanism emerges in which mitochondrial damage and oxidative stress caused by misfolded SOD1 leads to exacerbation of SOD1 misfolding and downstream mitochondrial damage. Interestingly using monoclonal antibodies to misfolded SOD1, misfolded wild type SOD1 species have been shown to be present in sporadic ALS patients [103,104] although this has been disputed by others [105].

Oxidative damage has also been proposed to promote aggregation of TDP-43 via cysteine oxidation and disulphide bond formation and acetylation [106,107]. In agreement, treatment of COS-7 cells with 4-hydroxyneonal (HNE), which is produced in cells by lipid peroxidation, was shown to cause insolubilisation, phosphorylation, and partial cytosolic localisation of TDP-43 [108]. Overexpression of wild type and mutant TDP-43 M337V and Q331K or its C-terminal fragments in NSC-34 have been shown to increase ROS and cause oxidative damage [21]. In addition to these deleterious effects TDP-43 also appears to have a protective function in response to oxidative stress. Indeed, oxidative stress has been shown to induce recruitment of TDP-43 to stress granules [109–111].

Other ALS-associated proteins that have been linked to oxidative stress include FUS P525L which has been shown to augment ROS levels when overexpressed in HEK293 cells [29], and poly(GR) DPRs which have been shown to increase oxidative stress [43]. Interestingly as was the case for TDP-43, an oxidative environment increased FUS inclusions, again pointing toward a detrimental feed-forward loop [112].

2.5. Calcium mishandling

Loss of calcium homeostasis has been observed in *in vitro* and *in vivo* models of mutant SOD1, vesicle-associated membrane protein-associated protein B (VAPB), TDP-43, and FUS-related ALS and in the motor nerve terminals of ALS patients [51,68,113–119]. In SOD1 G93A transgenic mice a significant decrease in mitochondrial calcium loading capacity in the CNS was observed long before disease onset, suggesting an early loss of calcium buffering may be causal in disease [68]. Similarly, analysis of calcium handling in vulnerable hypoglossal motor neurons identified a reduction of uniporter-dependent mitochondrial calcium uptake which was not observed in resistant oculomotor neurons [120]. How these defects in mitochondrial calcium uptake affect motor neurons is not completely clear, but SOD1 G93A motor neurons show a delay in the recovery to basal level of the calcium increase following α-amino-5-methyl-3-hydroxisoxazolone-4-propionate (AMPA) receptor activation [121].

Loss of ER-mitochondria communication has emerged as a major cause of loss of calcium homeostasis in ALS. An estimated 5–20% of mitochondria are closely associated with the ER at contact sites called mitochondria-associated ER membranes (MAM). Several protein complexes that tether ER to mitochondria have been proposed, including homo and heterotypic interactions between mitofusin (Mfn) 1 and 2, interaction of inositol 1,4,5-trisphosphate receptor (IP3R) with VDAC via glucose-regulated protein 75 (Grp75), and the integral ER protein VAPB that binds to protein tyrosine phosphatase-interacting protein 51 (PTPIP51) on the OMM (Fig. 2). Among other functions, ER-mitochondria contact sites allow calcium exchange between the two organelles [reviewed in Refs. [122,123]]. Disruption of ER-mitochondria interactions have been reported in mutant SOD1, Sig1R, TDP-43, and FUS-related ALS [51,115,124]. In the case of mutant TDP-43 and FUS-related ALS, reduced ER-mitochondria communication was caused by a GSK3β-dependent reduction in VAPB-PTPIP51 interaction [51,115]. In contrast, ALS mutant VAPB P56S was shown to

have greater affinity for PTPIP51 and to increase ER-mitochondria association [118]. However, since in ALS patient-derived iPSC neurons VAPB expression is down-regulated because of reduced expression of the VAPB P56S mutant allele [125], it is likely that in VAPB P56S-related ALS ER-mitochondria contacts are actually decreased as well. Whether VAPB-PTPIP51 interaction is also disrupted in mutant SOD1 and Sig1R-related ALS is not known, but in SIGMAR1 knockout mice which model loss of Sig1R function, the interaction between IP3R and VDAC was decreased in motor neurons [124]. Interestingly VAPB expression levels in the spinal cord of sporadic ALS cases are significantly lower compared to healthy controls [126]. Thus, disrupted ER-mitochondria communication may be a general feature in ALS.

In ALS mutant FUS and TDP-43 models, impaired ER-mitochondria communication leads to reduced calcium uptake in mitochondria and an associated rise in cytosolic calcium upon triggering of calcium release from the ER [51,115]. Similarly, in SIGMAR1 knockout mice cytosolic calcium levels were elevated and the time to return to basal levels was significantly longer [124]. As mentioned above, similar loss of calcium homeostasis has been reported in SOD1 G93A transgenic mice. Whether these are due to reduced ER-mitochondria communication has not been formally shown, but neuronal overexpression of wild type human VAPB has been shown to slow disease and increase survival in SOD1 G93A transgenic mice [127]. It is tempting to speculate, that restoring ER-mitochondria contacts is at the basis of this protective effect.

Dysregulation of calcium, possibly by miscommunication between ER and mitochondria, may be a primary cause of motor neuron death in ALS. Indeed, motor neurons possess several properties that make them more vulnerable to calcium dysregulation compared to other neuronal populations. They express a high number of calcium permeable AMPA receptors at the postsynaptic terminal, which results in a greater vulnerability to excitotoxicity thorough excessive calcium influx during excitatory neurotransmission events [117,128,129]. In addition, motor neurons have reduced cytosolic buffering capacity because they express low levels of calcium buffering proteins such as parvalbumin and calbindin D-28k and this makes them more dependent on mitochondria for calcium buffering [130–132, reviewed in Refs. [133,134]]. Furthermore, loss of calcium homeostasis may contribute to a number of other ALS-associated toxic mechanisms, such as axonal transport defects and oxidative stress, and loss of protein homeostasis [135,136, reviewed in Ref. [86]]. Mitochondrial calcium regulates ATP production by activating the rate-limiting enzymes of the Krebs cycle and regulates oxidative phosphorylation and ATP synthesis to match local energy demand. As discussed above, reduced ATP production is a common feature in ALS. Diminished ATP levels may directly impact on axonal transport of mitochondria, vesicles and other cargoes by starving molecular motors of ATP. At the same time, elevated levels of cytosolic calcium may disrupt axonal transport of mitochondria by interacting with the mitochondrial kinesin-1 receptor Miro1 and in turn exacerbate axonal transport deficits and local ATP levels. Indeed, elevated cytosolic calcium levels were shown to disrupt axonal transport of mitochondria in ALS mutant VAPB P56S expressing neurons [114, reviewed in Ref. [135]].

In ALS motor neurons, mitochondrial calcium overload may result from the physiological activity of AMPA receptors with pathologically increased calcium permeability. The resulting chronic calcium overload in mitochondria and concomitant abnormal oxidative phosphorylation has been shown to increase ROS production and oxidative stress [45,137]. Mitochondrial calcium overload is predicted to result in depletion of calcium in the ER which causes protein misfolding and induces ER stress [reviewed in Refs. [133,136]]. Elevated cytosolic calcium levels activate calpain, which has been shown to cleave TDP-43 to generate aggregation-

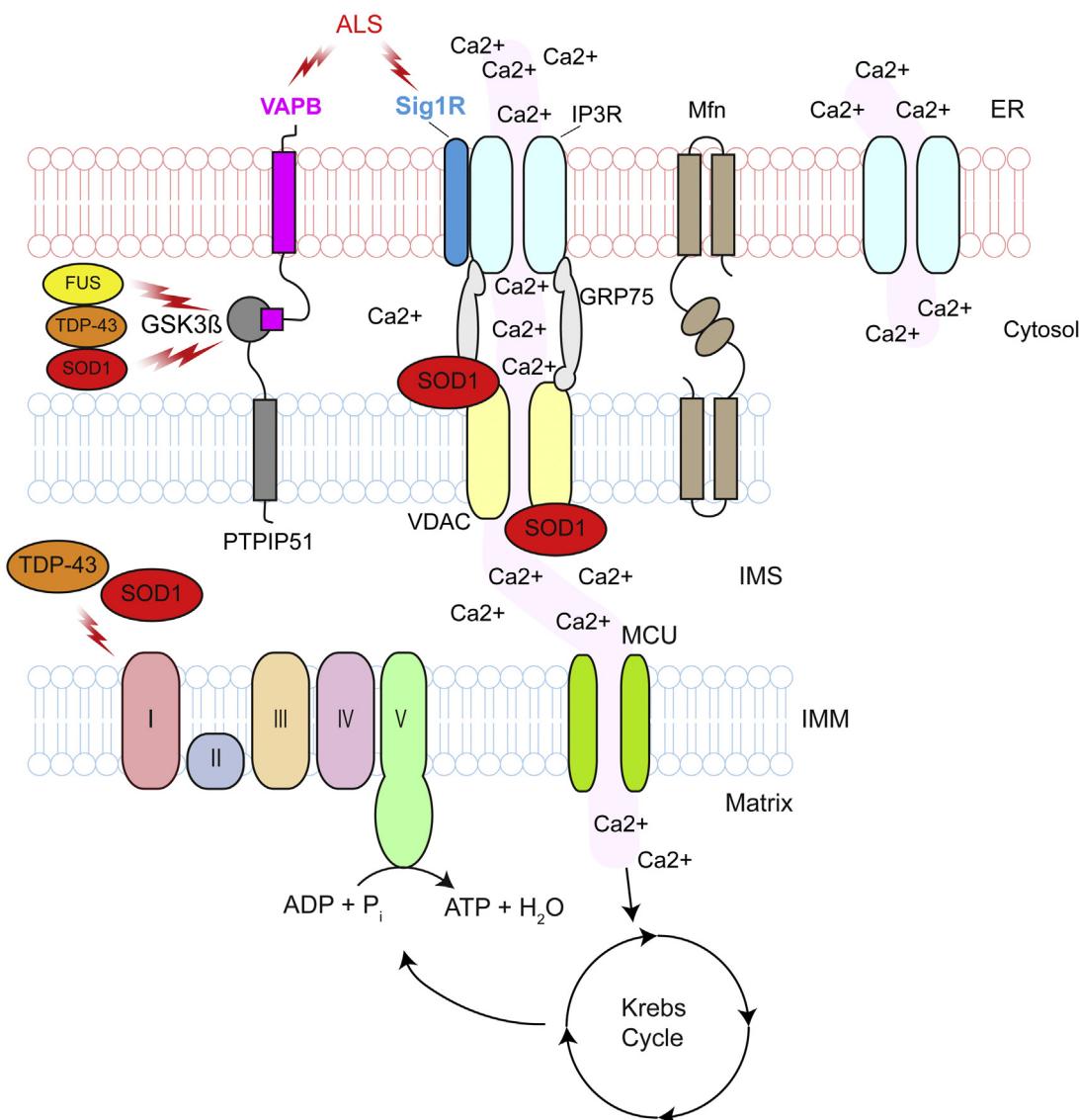


Fig. 2. Loss of calcium homeostasis.

Calcium homeostasis is achieved through the exchange of calcium between the ER and mitochondria at ER-mitochondria contacts. ER-mitochondria contacts are disrupted in SOD1, TDP-43, VAPB, Sig1R and FUS ALS. Disruption of the contacts may lead to increases in cytosolic calcium levels and disruption of calcium dependent cellular processes, including axonal transport, ATP generation and protein homeostasis. MCU, mitochondrial calcium uniporter. See text for details.

prone fragments and could account for the TDP-43 pathology observed in ALS [138,139]. Finally, chronic mitochondrial calcium overload and protein misfolding both induce apoptosis.

2.6. Pro-apoptotic signalling

Apoptosis is a last resort mechanism employed by cells upon suffering irreparable damage [reviewed in Ref. [140]]. Apoptosis serves to remove a damaged cell from the environment in a controlled manner so as to not induce large-scale degeneration. Mitochondria are integral to apoptosis. Upstream apoptotic signalling cascades converge on mitochondria to bring about cytochrome c release, which activates downstream executioner caspases [reviewed in Ref. [140]] (Fig. 3). Apoptosis is regulated by pro- and anti-apoptotic proteins of the Bcl-2 family, which control release of caspase-activating factors from mitochondria [reviewed in Refs. [140,141]]. Activation of apoptotic signalling cascades has been observed in most ALS models, but this is mostly an indirect consequence of other toxic events. However, ALS mutant SOD1 has

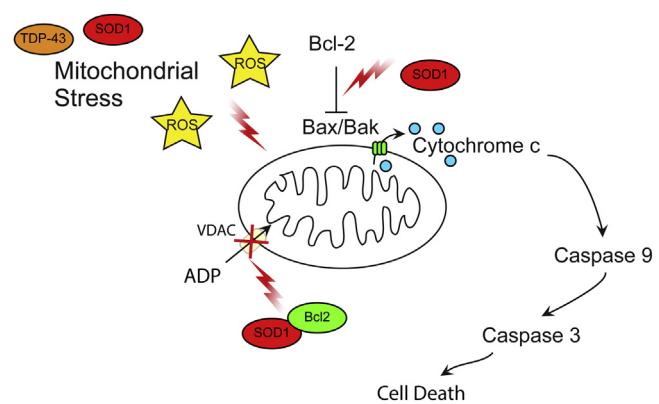


Fig. 3. Pro-apoptotic signalling.

Mitochondrial stress is increased due to dysfunctional cellular pathways resulting in increased ROS generation, and mitochondrial damage. Mutant SOD1 contributes to apoptotic signalling in ALS by binding the anti-apoptotic factor Bcl-2 and promoting a pro-apoptotic conformation of the protein. The Bcl2-SOD1 complex inhibits VDAC ADP permeability and induces mitochondrial hyperpolarisation. See text for details.

been shown to directly influence apoptotic signalling by interaction with Bcl-2. Wild-type and ALS mutant SOD1 (A4V, G37R, G41D and G85R) have been shown to bind the anti-apoptotic factor Bcl-2 in spinal cord samples [50]. When bound to mutant SOD1, the BH3 domain of Bcl-2 is exposed and this causes a pro-apoptotic gain of function of the Bcl-2 protein in both cell and animal models of mutant SOD1 G93A ALS and in mutant SOD1 A4V patient spinal cord [142] (Fig. 3). The toxic mutant SOD1–Bcl-2 complex inhibits mitochondrial permeability to ADP and induces mitochondrial hyperpolarization due to reducing the interaction of Bcl-2 and VDAC1 [143] (Fig. 3).

2.7. Impaired mitochondrial dynamics

Mitochondria are dynamic organelles that undergo fusion and fission events and directed transport. Several lines of research indicate that impaired mitochondrial dynamics may contribute to the aetiology of ALS.

2.7.1. Aberrant mitochondrial fission and fusion

Mitochondria form a dynamic network that evolves according to the energetic requirements of the cell. An individual mitochondrion can fuse to or fission from the mitochondrial network in response to various cues [144]. Fusion allows mitochondria to share mitochondrial metabolites, DNA and proteins, and allows for dissipation of small changes in MMP. Fission facilitates mitochondrial motility and allows the isolation of damaged parts of the network prior to disposal by mitophagy. In healthy cells, mitochondrial network morphology is governed by the interplay between Dynamin-related protein 1 (Drp1) and Fission 1 (Fis1) that promote fission, and Mfn1, Mfn2 and Optic atrophy 1 (Opa1) that promote fusion [reviewed in Ref. [145]] (Fig. 4). Evidence points towards imbalances in these two opposing pathways in ALS.

In SOD1 G93A expressing SH-SY5Y and NSC-34 cell models, a decrease in Opa1 and increase in Drp1 levels resulting in a fragmented mitochondrial network were reported [146]. *In vivo*, the levels of Mfn1 and Opa1 progressively decreased while the levels of activated Drp1 phosphorylated at Ser616 and Fis1 remained stable in the spinal cord of SOD1 G93A transgenic mice [147] (Fig. 4). In contrast, in SOD1 G93A mouse muscle fibres there was no reported change in Mfn1, Mfn2 or Drp1 levels [148].

Overexpression of wild type TDP-43 and to a greater extend ALS mutant TDP-43 Q331K and M337V resulted in reduced mitochondrial length in primary motor neurons which was sensitive to Mfn2 levels [23]. Transgenic mice expressing wild type TDP-43 exhibited increased levels of Fis1 and activated phospho-Ser616 Drp1 and a reduction in Mfn1, which correlated with aberrant mitochondrial morphology and clustering [149]. In contrast, the same group reported no changes in the levels of phospho-Ser616 Drp1, Fis1 or Mfn1 in mutant TDP-43 M337V transgenic mice, despite a similar mitochondrial phenotype [150]. Possibly different pathways lead to fragmentation of the mitochondrial network in wild type and mutant TDP-43 transgenic mice, or, alternatively, external factors linked to disease such as changes in apoptotic factors result in mitochondrial fragmentation. Nevertheless, analysis of TDP-43 A382T patient fibroblasts showed that Fis1 levels were significantly increased compared to control cells [31] (Fig. 4). Counterintuitively, mitochondrial network fragmentation in C9ALS/FTD patient fibroblasts was associated with an increase in Mfn1 levels [31]. Possibly Mfn1 expression is increased to compensate for dysregulation of other fusion and fission factors.

ER-mitochondria contact sites have been shown to regulate mitochondrial morphology. Mitochondrial fission occurs at ER-mitochondria contact sites where ER tubules wrap around and constrict mitochondria to aid division [151]. As discussed above reduced ER-mitochondria contacts are a common obser-

vation in various ALS models. Consistent with reduced fission, decreased ER-mitochondria contacts in motor neurons of *SIGMAR1* knockout correlated with elongated mitochondria compared to controls [124] whereas, in VAPB P56S overexpressing cells increased ER-mitochondria contacts correlated with fragmented mitochondria [118] (Fig. 4). However, in SOD1, TDP-43 and FUS ALS models reduced ER-mitochondria contacts appear to correlate with fragmented mitochondria [23,26,51,115,152,153]. Therefore, there is no straightforward correlation between ALS-associated disruption of ER-mitochondria contacts and changes in mitochondrial network morphology.

The relevance of exacerbated mitochondrial fission in the aetiology of ALS is not clear. Perhaps, smaller mitochondria, which tend to be less energetically favourable and more prone to accumulating ROS induced damage if they cannot fuse back to the network [154], may exacerbate mitochondrial damage. Thus, the fragmented mitochondrial network observed may correlate directly with the reduced MMP, ATP levels and increased ROS reported in ALS. Alternatively, mitochondrial fragmentation may merely reflect mitophagy, a physiological quality control response to mitochondrial damage (see below). Indeed, it is well established that one of the first steps of mitophagy is to isolate damaged mitochondria from the mitochondrial network [reviewed in Ref. [155]]. However, as disease progresses and mitochondrial damage accumulates, mitophagy may become unable to cope and as a consequence mitochondrial fragmentation could become self-propagating. In any case, evidence suggests that mitochondrial fragmentation participates in ALS. Inhibition of mitochondrial fission by overexpression of dominant negative Drp1 K38A rescued SOD1 G93A induced mitochondrial fragmentation and trafficking defects and increased motor neuron viability [156]. Similarly, promotion of mitochondrial fusion by co-expressing Mfn2 in wild type or TDP-43 M333V expressing primary motor neurons rescued mitochondrial fragmentation [23]. Whether rescuing mitochondrial network morphology is sufficient to improve neuron survival *in vivo* remains unclear.

2.7.2. Disrupted mitochondrial quality control

As discussed above, accumulation of fragmented, rounded mitochondria is a common feature in ALS, and may indicate failure of mitophagy to clear damaged mitochondria. The molecular mechanisms of mitophagy have been reviewed in detail elsewhere [157], but briefly, loss of MMP results in accumulation of full length PINK1 on the OMM where it phosphorylates ubiquitin, which in turn promotes the recruitment of the E3 ligase Parkin. Parkin ubiquitinates and promotes the VCP/p97-dependent degradation of various OMM proteins such as Mfn1/2 and Miro1 to isolate and immobilise the damaged mitochondrion. Ubiquitinated mitochondria are then recognised by the canonical autophagy machinery and are delivered to the lysosome for degradation (Fig. 5).

TDP-43 and FUS directly regulate the expression of Parkin [158,159]. Loss of TDP-43 or FUS was shown to decrease Parkin levels [158], and consistent with loss of TDP-43 function in the nucleus decreased Parkin protein levels were observed in sporadic ALS spinal cord motor neurons containing cytoplasmic TDP-43 aggregates [158]. Parkin protein levels were found to be decreased in TDP-43 A513T transgenic mice [38] but this was not observed by others [160]. Thus, since cytosolic TDP-43 aggregation is a near universal phenomenon in ALS, loss of Parkin may impair mitophagy in most ALS cases. Furthermore, since, as discussed above, TDP-43 accumulates in mitochondria and causes mitochondrial damage, a two-hit scenario emerges in which loss of mitophagy exacerbates the consequences of mitochondrial damage. Of note, Parkin has been shown to ubiquitinate TDP-43 and to promote its translocation from the nucleus to the cytosol in an HDAC6-dependent process [160]. Even though the latter is likely to be a reflection of the

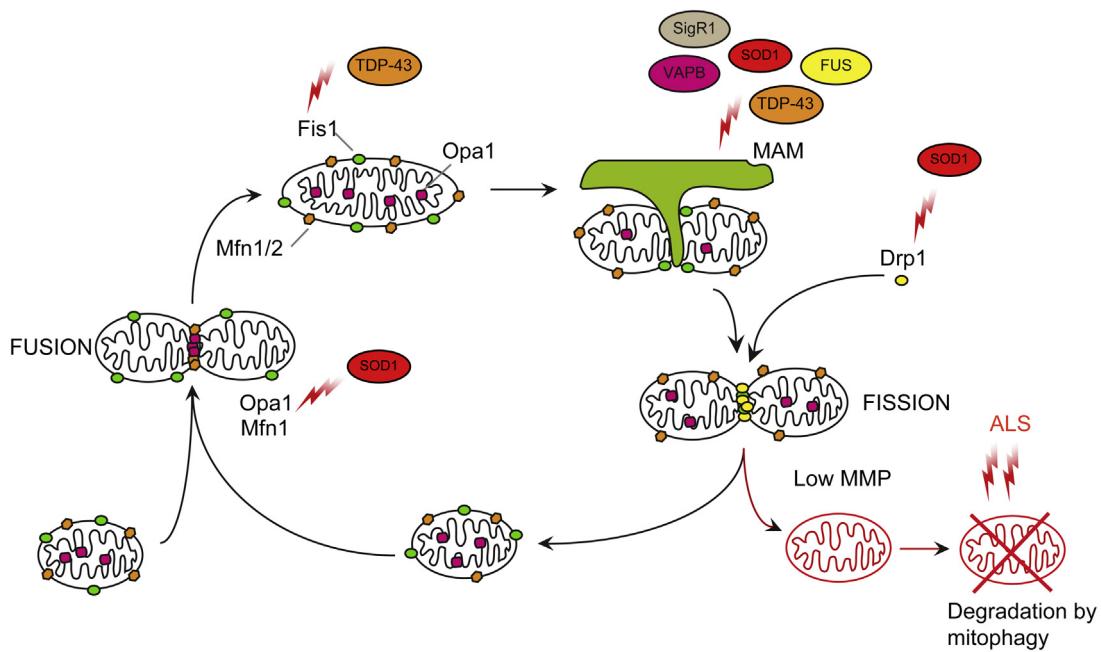


Fig. 4. Aberrant mitochondrial fission and fusion.

Mitochondrial network dynamics are required for correct mitochondrial function. Fission is mediated by Drp1, which is recruited to mitochondria by Fis1. Fusion of the OMM is mediated by Mfn1/2 and the fusion of the IMM by Opa1. Mitochondrial fission is promoted in SOD1 and TDP-43 ALS by increases in the fission factors Fis1 and Drp1 and a decrease in the fusion factors Mfn1 and Opa1. Decreased ER-mitochondria contact sites may also lead to decreased mitochondrial fission events. Finally, dysfunctional removal of damaged organelles through the mitophagy pathway may also contribute to accumulation of fragmented mitochondria. See text for details.

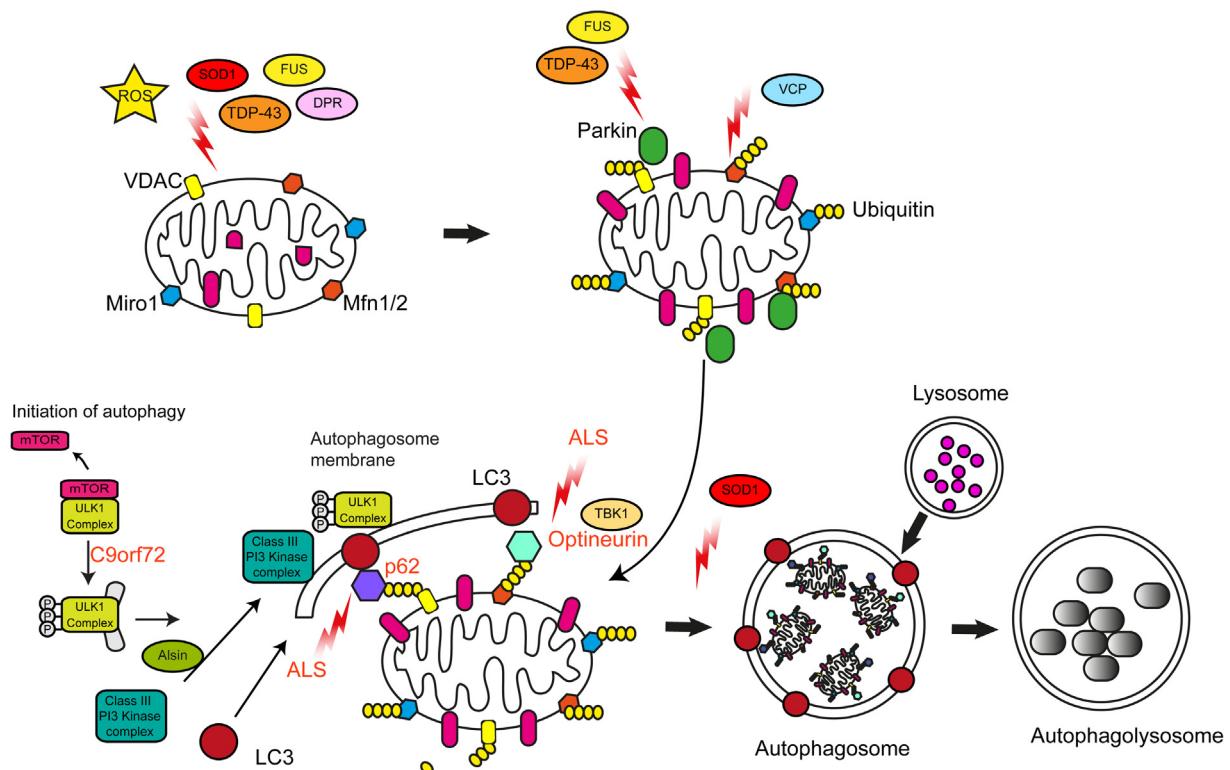


Fig. 5. Disrupted autophagy/mitophagy.

Mitophagy removes dysfunctional mitochondria from the cell. Mitochondria are exposed to high levels of stress due to elevations in ROS levels and aggregates of mutant ALS protein associated with mitochondria. Mitophagy is impaired at multiple stages in ALS. Mutant TDP-43 and FUS mislocalisation may affect Parkin levels. The removal of ubiquinated Mfn1/2 may be impaired by mutant VCP/p97. Recognition of ubiquinated mitochondrial cargos by nascent autophagosomes may be impaired by mutations in p62/Sequestosome-1, Optineurin and TBK1. Other ALS genes such as SOD1, Alsin, and C9orf72 have also been linked to autophagy. See text for details.

role of Parkin and HDAC6 in aggrephagy rather than mitophagy, it does emphasise a close link between Parkin and TDP-43 that merits further examination.

The Type II AAA+ ATPase VCP/p97 is involved in a multitude of processes, but its main function is to dislodge damaged ubiquitinated proteins from large protein complexes and membranes for degradation by the proteasome [reviewed in Refs. [161,162]]. VCP/p97 is recruited to the OMM of damaged mitochondria following Parkin recruitment and extracts ubiquitinylated Mfn1 and 2 from the OMM to allow their degradation by the proteasome [163]. As such VCP/p97 is essential for mitophagy [164]. Missense mutations in VCP/p97 cause of 1–2% of familial ALS cases as well as inclusion body myopathy (IBM) with Paget's disease (PDB) and FTD [165,166]. Although it is not known exactly how disease-associated mutations in VCP/p97 alter its cellular functions, most mutations appear to convey varying degrees of loss of function [reviewed in Ref. [167]]. Consistent with loss of function, MEFs expressing mutant VCP/p97 A232E do not clear mitochondria following damage, despite labelling with Parkin [164]. As discussed above, pathogenic mutations in VCP/p97 induce mitochondrial uncoupling and impair ATP production [73]. Possibly, this phenotype is caused by accumulation of damaged, uncoupled mitochondria that should have been degraded.

Interestingly, not unlike Parkin, VCP/p97 has been linked to TDP-43 metabolism. In SHSY-5Y cells expression of mutant VCP/p97 (R95G, R155H/C, R191Q, A232E) caused TDP-43 redistribution from the nucleus to the cytoplasm [168]. Similarly, in VCP/p97 R152H or A229E transgenic *Drosophila* and spinal cord motor neurons of VCP/p97 R155H or A232E transgenic mice expression of mutant VCP/p97 induced TDP-43 pathology [169]. Furthermore, in *Drosophila*, TBPH, the fly homolog of TDP-43 was identified as a genetic modifier that suppressed mutant VCP/p97 toxicity, indicating that VCP/p97 toxicity is at least in part mediated by a toxic gain of function of TDP-43 [169]. We now know that TDP-43 accumulates in mitochondria and causes damage to the ETC (see above; [42]). Hence a possible explanation for this genetic interaction is that accumulation of cytoplasmic TDP-43 exacerbates loss of mitochondrial quality control by mutations in VCP/p97.

Mitophagy requires autophagy receptors such as optineurin and p62/Sequestosome-1 that accumulate on ubiquitinylated mitochondria and deliver them to the autophagosome by interaction with LC3-II on autophagic membranes. Optineurin and p62/Sequestosome-1 are substrates of Tank-binding kinase 1 (TBK1); phosphorylation by TBK1 promotes ubiquitin and LC3 binding and enhances mitophagy [reviewed in Refs. [170–172]].

Mutations in the *OPTN*, *SQSTM1* and *TBK1* genes which encode Optineurin, p62/Sequestosome-1, and TBK1, respectively, have been associated with ALS [173–175, reviewed in Ref. [11]] (Table 1). ALS-associated mutations in these genes appear to cause loss of function phenotypes. The ALS-associated Optineurin E478G mutation maps to the ubiquitin binding domain of Optineurin and the mutant protein is no longer recruited to damaged mitochondria [176,177]. Along the same lines, the ALS-associated L341V mutation in p62/Sequestosome-1 maps to the LC3-interacting region (LIR) domain and disrupts interaction with LC3 [178]. The ALS mutant forms of TBK1, TBK1 del690–713 and E696K, prevent the interaction between TBK1 and Optineurin and impair clearance of LC3 labelled autophagic cargos [179,180]. If ALS-associated TBK1 mutants also affect mitophagy remains to be determined, but the role of TBK1 in mitophagy is well established. Furthermore, the activation of TBK1 is dependent on the activity of the upstream activity of PINK1/Parkin which as discussed above is affected in ALS [179,180]. Consistent with failure to deliver damaged mitochondria to the lysosome, LC3-II positive mitochondrial aggregates have been observed in wild type and ALS mutant TDP-43 Q331K and M337V expressing NSC-34 cells as well as in SOD1 G93A transgenic

mouse spinal cord [21,181]. In case of SOD1 G93A the observed failure to clear damaged mitochondria appears to be caused by lysosomal deficits caused by impairment of retrograde trafficking of late endosomes rather than specific mitophagy deficits [182].

Interestingly TBK1 has also been found to interact with C9orf72 in complex with SMCR8 and WDR41, and C9orf72 controls autophagy via regulation of the small Rab GTPases Rab1a, Rab8a and Rab39b [183–186, reviewed in Ref. [187]]. Another ALS and autophagy-associated regulator of small Rab GTPases is Alsin, a guanine nucleotide exchange factor (GEF) for Rab5. Loss-of-function mutations in Alsin cause a recessive juvenile form of ALS, ALS2 [188,189]. Loss of Alsin in mice impairs autophagy and corticospinal motor neurons exhibit fused mitochondria engulfed by vacuole structures consistent with failure to remove damaged mitochondria by mitophagy [33,190]. Interestingly loss of Alsin in corticospinal motor neurons correlated with a progressive decline in Rab1a levels suggesting a direct link to C9orf72 [33]. Thus, a number of ALS-associated insults appear to converge on autophagy and mitophagy.

2.7.3. Impaired axonal transport of mitochondria

Motor neurons are highly polarised cells and require membrane-bound vesicles, organelles, proteins, lipids and RNA to be transported from the soma to the axon terminal and vice versa. Impaired axonal transport of mitochondria is a well-documented phenomenon in ALS. In fact, axonal transport defects are one of the earliest pathophysiological events in ALS motor neurons, indicating that they may be a primary cause of motor neuron loss [reviewed in Refs. [135,191]].

Long distance transport of mitochondria is mediated by the molecular motors kinesin-1 (previously referred to as conventional kinesin or KIF5) and cytoplasmic dynein that move along microtubules. Kinesin-1 moves toward the faster growing plus end of microtubules whereas cytoplasmic dynein moves toward the opposite minus end. Because axonal microtubules are uniformly orientated with their plus ends pointing away from the cell body, kinesin-1 transports mitochondria toward the axon terminal (anterograde transport) and cytoplasmic dynein ferries mitochondria to the cell body (retrograde transport) (Fig. 6).

Defects in both anterograde and retrograde transport of mitochondria have been reported in a number of *in vitro* and *in vivo* experimental models. ALS mutant SOD1 G93A, A4V, G85R or G37R caused reduced anterograde but not retrograde axonal transport of mitochondria in cultured cortical neurons and an identical phenotype was observed in embryonic motor neurons isolated from SOD1 G93A transgenic mice [26]. Similarly, overexpression of the ALS mutant VAPB P56S caused a selective block in anterograde transport of mitochondria [114]. Reduced overall mitochondrial transport was observed in primary motor neurons expressing wild-type TDP-43 and to a greater extent ALS mutant TDP-43 Q331K or M337V [23] and in Sig1R deficient motor neurons [124]. Interestingly, normal levels of axonal transport of mitochondria were reported in cortical neurons expressing wild type or mutant TDP-43 M337V or A315T at 5–7 days in culture, suggesting a possible cell type specificity [192]. Time-lapse recordings in single axons in the intact sciatic nerve of presymptomatic SOD1 G93A and TDP-43 A315T transgenic mice and rats confirmed deficits in axonal transport of mitochondria *in vivo* [22,193–195].

Mitochondrial transport defects result in redistribution of mitochondrial in axons. *In vitro*, the number of axonal mitochondria was significantly reduced and the remaining mitochondria were spaced further apart in primary neurons expressing ALS mutant SOD1 [26]. *In vivo* the number of axonal mitochondria was reduced in motor neurons of early symptomatic SOD1 G37R and SOD1 G85R transgenic mice and the motor axon terminals of SOD1 G93A transgenic rats and TDP-43 transgenic mice [27,194,196]. Furthermore, the

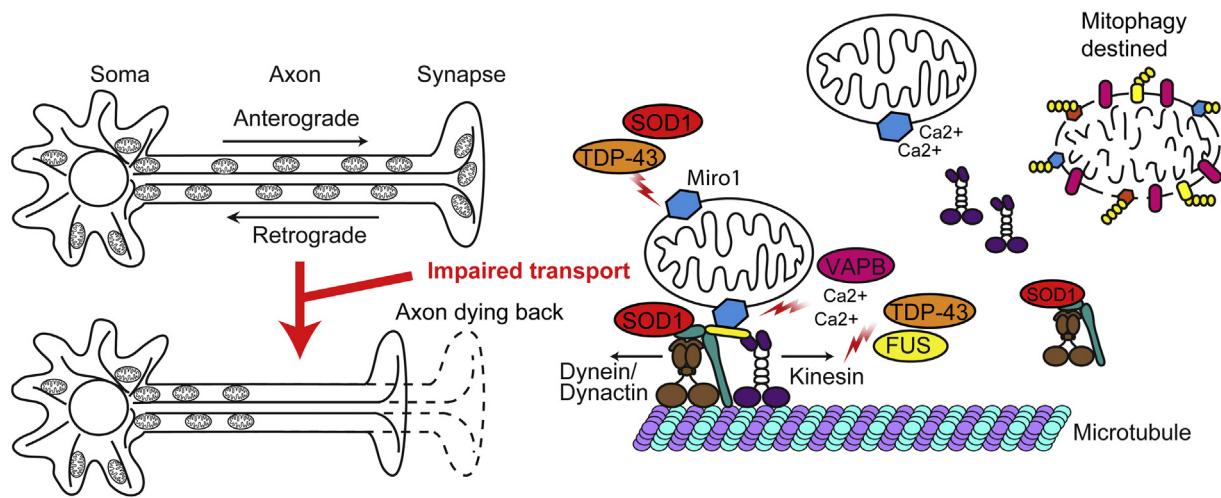


Fig. 6. Disrupted axonal transport.

Neurons depend on axonal transport of mitochondria for their ATP requirement and calcium buffering. Axonal transport of mitochondria is impaired in ALS. Miro1, the OMM protein involved in mitochondrial trafficking, is reduced in SOD1 and TDP-43-associated ALS. Mutant SOD1 binds the dynein/dynactin complex required for retrograde transport sequestering it in the cytosol. FUS and TDP-43 are proposed to regulate kinesin expression levels. Increases in cytosolic calcium levels due to decreased ER-mitochondria contacts in SOD1, TDP-43, FUS, Sig1R and VAPB-related ALS may contribute to detachment of the mitochondria from kinesin, through aberrant Miro1 regulation. Loss of mitochondria from the axon terminal may lead to the dying back of the axon. See text for details.

distribution of the remaining mitochondria was no longer homogeneous throughout the axons, with abnormal mitochondrial clusters observed along the axon [22,27,196,197]. Similar mislocalisation and aggregation of mitochondria has been observed in the soma, dendrites and proximal axons of motor neurons in spinal cord sections from ALS patients, suggesting that disruption to mitochondrial transport is a general phenomenon in ALS [18].

A number of possible mechanisms underlying mitochondrial axonal transport defects have been proposed. In rare cases, pathogenic mutations in the axonal transport machinery, such as cytoplasmic dynein, kinesin-1 and α -tubulin directly disrupt axonal transport. In most cases however the disruption of axonal transport of mitochondria appears to be indirect; proposed mechanisms include microtubule destabilisation, pathogenic kinase signalling, protein aggregation, and mitochondrial damage [reviewed in Ref. [135]]. Axonal transport of mitochondria is controlled by the integral OMM protein Miro1 [reviewed in Ref. [198]]. Miro1 is an atypical Rho GTPase comprised of two GTPase domains separated by two calcium-binding E-helix-loop-F-helix (EF)-hand motifs [199]. Miro1 is anchored in the OMM and connects mitochondria to kinesin-1 and cytoplasmic dynein via the adapter proteins TRAK1 and 2 [200–205].

Miro1 regulates mitochondrial trafficking in response to physiological calcium stimuli and mitochondrial damage. Binding of calcium to the Miro1 EF-hand motifs modulates the interaction of kinesin-1 with Miro1 and impedes anterograde transport of mitochondria [206,207]. During mitophagy, PINK1 phosphorylates Miro1 in response to mitochondrial damage [208–210]. Phosphorylation marks Miro1 for Parkin-dependent proteasomal degradation and results in detachment of kinesin-1 from mitochondria and arrested mitochondrial movement [209].

As discussed above, mitochondrial calcium handling is dysregulated in ALS. Hence ALS-associated mitochondrial axonal transport defects may be the direct result of aberrant calcium-mediated regulation of Miro1. In agreement with such a mechanism, cytosolic calcium levels are elevated in cellular ALS models and in motor neurons from transgenic ALS models [114,119] and, at least in case of VAPB P56S there is direct evidence that elevated calcium disrupts transport of mitochondria via Miro1 [114] (Fig. 6). Similarly, ALS mutant SOD1, TDP-43, and Sig1R-associated reductions in ER-mitochondria contacts result in transiently elevated cytosolic

calcium levels due to reduced calcium-uptake in mitochondria that coexist with axonal transport deficits, but the possible involvement of Miro1 remains to be determined [115,124,152].

Decreased levels of Miro1 have been reported in SOD1 G93A and TDP-43 M337V transgenic mice, as well as in the spinal cord of ALS patients [211]. Since both SOD1 G93A and TDP-43 M337V damage mitochondria (see above), these results suggest that ALS-associated mitochondrial damage leads to halting of mitochondrial transport via PINK1/Parkin-dependent degradation of Miro1.

In some cases, ALS-associated proteins may directly affect the axonal transport machinery. Both TDP-43 and FUS have been shown to regulate the expression of several kinesins, including kinesin-1, and TDP-43 binds TRAK1 mRNA [159,212,213]. ALS-mutant SOD1 directly binds to cytoplasmic dynein in SOD1 G93A and G37R cell and mouse models and the interaction becomes more prevalent throughout disease progression [214–216]. Thus, in case of mutant SOD1, axonal transport defects may in part be due to sequestration of dynein.

There are several ways in which defective axonal transport of mitochondria may contribute to disease. Defective axonal transport of mitochondria may lead to imbalances in ATP generation and calcium buffering at synaptic terminals. In conjunction with mitochondrial damage this may cause the dying back of the axon, a feature implicated in ALS [217] (Fig. 6). Retrograde transport defects are associated with defects in the removal of damaged organelles by mitophagy, which could explain the mitochondrial aggregates found in the axons of ALS patients [20,218]. Furthermore, defective axonal transport of mitochondria may also affect the transport of other axonal cargoes such as signalling endosomes which appear closely linked to ALS pathology [reviewed in Ref. [135]].

3. Mitochondria as therapeutic targets in ALS

Finding a cure for ALS has thus far been unsuccessful. Targeting mitochondrial dysfunction presents an attractive treatment option due to the widespread prevalence of mitochondrial dysfunction in disease. Potential therapeutic strategies have aimed to decrease ROS generation, increase mitochondrial biogenesis, inhibit apoptotic pathways or dampen excitotoxicity.

The only FDA drug approved for use in ALS is Riluzole. However, Riluzole only extends survival by approximately 3months. The mechanism by which Riluzole acts to improve survival is unclear. Some evidence suggests that Riluzole decreases ROS through induction of glutathione synthesis [219]. However little effect was observed on basal levels of ROS, and therefore Riluzole may only act in this way in the context of elevated ROS levels. Riluzole has been found to display a range of effects on glutamatergic signalling at the synapse, leading ultimately to attenuation in calcium handling requirements. Indeed, treatment *in vitro* with Riluzole prevents motor and cortical neuron loss induced by sustained glutamate increase and elevated ROS levels [220]. Furthermore, Riluzole has been demonstrated to reduce inward calcium currents and rescue the axonal transport of neurofilaments [221,222]. Attenuation of inward calcium currents is predicted to alleviate stresses on cytosolic and mitochondrial calcium buffering mechanisms and potentially decrease cytosolic calcium levels with knock-on effects on ROS generation and mitochondrial function. However, with the limited effect on prognosis offered by Riluzole, alternative therapies are required for the treatment of ALS.

Despite many drug-based and gene therapy approaches targeting mitochondrial dysfunction in ALS, neither treatments aimed at increasing mitochondrial function and survival nor those aimed at reducing oxidative stress have yielded significant results in clinical trials, despite promising trials in animal models [223]. For example, several drugs targeting mitochondrial function and/or ROS such as Coenzyme Q10, Dexpramipexole, Olesoxime and Creatine all showed initial success in animal models but were unsuccessful in human clinical trials [224–231]. Reducing ROS using Edaravone, a free radical scavenger was successful in mouse models but had only a small effect on human disease progression [232,233]. Similarly, minocycline, an anti-apoptotic and anti-inflammatory drug which extended survival in mouse models, failed in a human phase III randomised trial [234]. The discrepancy between the results from mouse models and human trials highlights the disparity between models and the human disease, and the need for better disease models [235]. In addition, the wide range of mitochondrial dysfunction in ALS means that a single therapy is unlikely to attenuate all aspects of dysfunction. Therefore, identifying which of the mitochondrial dysfunctions are relevant to disease causation and progression will continue to be important for the development of neuroprotective therapies in ALS. Targeting specific proteins dysregulated in ALS may provide alternative therapy avenues. Indeed a blocking peptide that prevents the abnormal interaction between mutant SOD1 and Bcl-2 has been shown to prevent apoptosis and improve mitochondrial function in SOD1 G93A transgenic mice [143].

4. Conclusions

Despite the surge in the number genes found to be associated with ALS since the turn of the millennium, the aetiology ALS remains largely unknown. Mitochondrial dysfunction has emerged as a common, early phenomenon in ALS. The appearance of deficits in oxidative phosphorylation, calcium buffering and mitochondrial transport prior to the onset of disease symptoms *in vivo* in disease models suggests an important role for loss of mitochondrial integrity in the aetiology of ALS.

Nevertheless, clinical trials targeting mitochondria have been disappointing, indicating that mitochondrial dysfunction alone may not be a primary cause of disease. Possibly the widespread presence of mitochondrial dysfunction in ALS models and patients, may be secondary to other disease-related insults such as protein aggregation and excitotoxicity. In such a scenario, mitochondrial function would gradually decay until a point-of-no-return thresh-

old is reached and the motor neuron dies. Such a mechanism would be predicted to conspire with the age-associated decline in mitochondrial function and may explain the late onset of the majority of ALS cases. In any case, since, with the possible exception of Riluzole, treatments targeting other proposed causes of ALS have been equally unsuccessful in clinical trials, it transpires that any successful therapeutic strategy may have to address multiple targets in a combination therapy.

Importantly, mitochondrial dysfunction has also been widely reported in other neurodegenerative diseases such as Parkinson's, Huntington's and Alzheimer's disease [236–238]. Similarities in defects in mitochondrial clearance and bioenergetic functions exist between these diseases and ALS despite different clinical presentation. Therefore, neurodegenerative conditions appear to be group of diseases with common dysfunction but with varying endpoints. Possibly discovery and study of common mechanisms underlying these diseases, including mitochondrial dysfunction, will increase our understanding of the essential requirements for neuronal survival that can inform future neuroprotective therapies.

Acknowledgements

This work was funded by grants from the UK Medical Research Council (MRC; MR/K005146/1 and MR/M013251/1 to KJDV). EFS is supported by a Motor Neurone Disease Association Prize Studentship (DeVos/Oct13/870-892 to KJDV). PJS is supported as an NIHR Senior Investigator and by funding awards from the UK Medical Research Council, the National Institute for Health Research, the Motor Neurone Disease Association and the EU Horizon 2020 programme.

References

- [1] A. Chiò, G. Logroscino, B.J. Traynor, J. Collins, J.C. Simeone, L.A. Goldstein, L.A. White, Global epidemiology of amyotrophic lateral sclerosis: a systematic review of the published literature, *Neuroepidemiology* 41 (2013) 118–130.
- [2] P. Mehta, W. Kaye, L. Bryan, T. Larson, T. Copeland, J. Wu, O. Muravov, K. Horton, Prevalence of amyotrophic lateral sclerosis – United States, 2012–2013, *MMWR Surveill. Summ.* 65 (2016) 1–12.
- [3] M. Orsini, A.B. Oliveira, O.J. Nascimento, C.H. Reis, M.A. Leite, J.A. de Souza, C. Pupe, O.G. de Souza, V.H. Bastos, M.R. de Freitas, S. Teixeira, C. Bruno, E. Davidovich, B. Smidt, Amyotrophic lateral sclerosis: new perspectives and update, *Neurol. Int.* 7 (2015) 5885.
- [4] M.C. Kiernan, S. Vucic, B.C. Cheah, M.R. Turner, A. Eisen, O. Hardiman, J.R. Burrell, M.C. Zoinig, Amyotrophic lateral sclerosis, *Lancet* 377 (2011) 942–955.
- [5] A.L. Boxer, M. Gold, E. Huey, F.-B. Gao, E.A. Burton, T. Chow, A. Kao, B. Leavitt, B. Lamb, M. Grether, D. Knopman, N.J. Cairns, I.R. Mackenzie, L. Mitic, E.D. Roberson, D. Van Kammen, M. Cantillon, K. Zahs, S. Salloway, J. Morris, G. Tong, H. Feldman, H. Fillit, S. Dickinson, Z. Khachaturian, M. Sutherland, R. Farsc, B.L. Miller, J. Cummings, Frontotemporal degeneration, the next therapeutic frontier: molecules and animal models for FTD drug development, *Alzheimer's Dement.* 9 (2013) 176–188.
- [6] B. Swinnen, W. Robberecht, The phenotypic variability of amyotrophic lateral sclerosis, *Nat. Rev. Neurol.* 10 (2014) 661–670.
- [7] O. Abel, J.F. Powell, P.M. Andersen, A. Al-Chalabi, ALSoD. A user-friendly online bioinformatics tool for amyotrophic lateral sclerosis genetics, *Hum. Mutat.* 33 (2012) 1345–1351.
- [8] M. DeJesus-Hernandez, I. Mackenzie, B. Boeve, A. Boxer, M. Baker, N. Rutherford, A. Nicholson, N. Finch, H. Flynn, J. Adamson, N. Kouri, A. Wojtas, P. Sengdy, G.Y. Hsiung, A. Karydas, W. Seeley, K. Josephs, G. Coppola, D. Geschwind, Z. Wszolek, H. Feldman, D. Knopman, R. Petersen, B. Miller, D. Dickson, K. Boylan, N. Graff-Radford, R. Rademakers, Expanded GGGGCC hexanucleotide repeat in noncoding region of C9ORF72 causes chromosome 9p-linked FTD and ALS, *Neuron* 72 (2011) 245–256.
- [9] A. Renton, E. Majounie, A. Waite, J. Simón-Sánchez, S. Rollinson, J. Gibbs, J. Schymick, H. Laaksovirta, J. van Swieten, L. Myllykangas, H. Kalimo, A. Paetau, Y. Abramzon, A. Remes, A. Kaganovich, S. Scholz, J. Duckworth, J. Ding, D. Harmer, D. Hernandez, J. Johnson, K. Mok, M. Ryten, D. Trabzuni, R. Guerreiro, R. Orrell, J. Neal, A. Murray, J. Pearson, I. Jansen, D. Sondervan, H. Seelaar, D. Blake, K. Young, N. Halliwell, J. Callister, G. Toulson, A. Richardson, A. Gerhardt, J. Snowden, D. Mann, D. Neary, M. Nalls, T. Peuralinna, L. Jansson, V.M. Isoviita, A.L. Kaivorinne, M. Hölttä-Vuori, E. Ikonomi, R. Sulkava, M. Benatar, J. Wuu, A. Chiò, G. Restagno, G. Borghero, M. Sabatelli, D. Heckerman, E. Rogava, L. Zinman, J. Rothstein, M. Sendtner, C. Drepper, E. Eichler, C. Alkan, Z. Abdullaev, S. Pack, A. Dutra, E. Pak, J. Hardy,

- A. Singleton, N. Williams, P. Heutink, S. Pickering-Brown, H. Morris, P. Tienari, B. Traynor, A hexanucleotide repeat expansion in C9ORF72 is the cause of chromosome 9p21-linked ALS-FTD, *Neuron* 72 (2011) 257–268.
- [10] M. Van Blitterswijk, M. DeJesus-Hernandez, R. Rademakers, How do C9ORF72 repeat expansions cause ALS and FTD: can we learn from other non-coding repeat expansion disorders? *Curr. Opin. Neurol.* 25 (2014) 689–700.
- [11] A.E. Renton, A. Chiò, B.J. Traynor, State of play in amyotrophic lateral sclerosis genetics, *Nat. Neurosci.* 17 (2014) 17–23.
- [12] L. Ferraiuolo, J. Kirby, A.J. Grierson, M. Sendtner, P.J. Shaw, Molecular pathways of motor neuron injury in amyotrophic lateral sclerosis, *Nat. Rev. Neurol.* 7 (2011) 616–630.
- [13] D.G. Nicholls, S.L. Budd, Mitochondria and neuronal survival, *Physiol. Rev.* 80 (2000) 315–360.
- [14] E. Engl, D. Attwell, Non-signalling energy use in the brain, *J. Physiol. (Lond.)* 593 (2015) 3417–3429.
- [15] R. Rizzuto, D. De Stefani, A. Raffaello, C. Mammucari, Mitochondria as sensors and regulators of calcium signalling, *Nat. Rev. Mol. Cell Biol.* 13 (2012) 566–578.
- [16] B.A. Payne, P.F. Chinnery, Mitochondrial dysfunction in aging: much progress but many unresolved questions, *Biochim. Biophys. Acta* 1847 (2015) 1347–1353.
- [17] T. Atsumi, The ultrastructure of intramuscular nerves in amyotrophic lateral sclerosis, *Acta Neuropathol. (Berl.)* 55 (1981) 193–198.
- [18] S. Sasaki, M. Iwata, Mitochondrial alterations in the spinal cord of patients with sporadic amyotrophic lateral sclerosis, *J. Neuropathol. Exp. Neurol.* 66 (2007) 10–16.
- [19] M.N. Hart, P.A. Cancilla, S. Frommes, A. Hirano, Anterior horn cell degeneration and Bunina-type inclusions associated with dementia, *Acta Neuropathol. (Berl.)* 38 (1977) 225–228.
- [20] K. Okamoto, S. Hirai, M. Shoji, Y. Senoh, T. Yamazaki, Axonal swellings in the corticospinal tracts in amyotrophic lateral sclerosis, *Acta Neuropathol. (Berl.)* 80 (1990) 222–226.
- [21] K. Hong, Y. Li, W. Duan, Y. Guo, H. Jiang, W. Li, C. Li, Full-length TDP-43 and its C-terminal fragments activate mitophagy in NSC34 cell line, *Neurosci. Lett.* 530 (2012) 144–149.
- [22] J. Magrané, C. Cortez, W. Gan, G. Manfredi, Abnormal mitochondrial transport and morphology are common pathological denominators in SOD1 and TDP43 ALS mouse models, *Hum. Mol. Genet.* 23 (2014) 1413–1424.
- [23] W. Wang, L. Li, W.L. Lin, D.W. Dickson, L. Petruccielli, T. Zhang, X. Wang, The ALS disease-associated mutant TDP-43 impairs mitochondrial dynamics and function in motor neurons, *Hum. Mol. Genet.* 22 (2013) 4706–4719.
- [24] M.C. Dal Canto, M.E. Gurney, Development of central nervous system pathology in a murine transgenic model of human amyotrophic lateral sclerosis, *Am. J. Pathol.* 145 (1994) 1271–1279.
- [25] C.M.J. Higgins, C. Jung, Z. Xu, ALS-associated mutant SOD1G93A causes mitochondrial vacuolation by expansion of the intermembrane space by involvement of SOD1 aggregation and peroxisomes, *BMC Neurosci.* 4 (2003).
- [26] K.J. De Vos, A.L. Chapman, M.E. Tennant, C. Manser, E.L. Tudor, K.F. Lau, J. Brownlees, S. Ackerley, P.J. Shaw, D.M. McLoughlin, C.E. Shaw, P.N. Leigh, C.C.J. Miller, A.J. Grierson, Familial amyotrophic lateral sclerosis-linked SOD1 mutants perturb fast axonal transport to reduce axonal mitochondria content, *Hum. Mol. Genet.* 16 (2007) 2720–2728.
- [27] C. Vande Velde, K.K. McDonald, Y. Boukhdemi, M. McAlonis-Downes, C.S. Lobsiger, S. Bel Hadj, A. Zandona, J.-P. Julien, S.B. Shah, D.W. Cleveland, Misfolded SOD1 associated with motor neuron mitochondria alters mitochondrial shape and distribution prior to clinical onset, *PLoS One* 6 (2011) e22031.
- [28] M.L. Tradewell, Z. Yu, M. Tibshirani, M. Boulanger, H.D. Durham, S. Richard, Arginine methylation by Prmt1 regulates nuclear-cytoplasmic localization and toxicity of FUS/TLS harbouring ALS-linked mutations, *Hum. Mol. Genet.* 21 (2012) 136–149.
- [29] J. Deng, M. Yang, Y. Chen, X. Chen, J. Liu, S. Sun, H. Cheng, Y. Li, E.H. Bigio, M. Mesulam, Q. Xu, S. Du, K. Fushimi, L. Zhu, J.Y. Wu, FUS interacts with HSP60 to promote mitochondrial damage, *PLoS Genet.* 11 (2015) e1005357.
- [30] A. Sharma, A.K. Lyashchenko, L. Lu, S.E. Nasrabad, M. Elmaleh, M. Mendelsohn, A. Nemes, J.C. Tapia, G.Z. Mentis, N.A. Schneider, ALS-associated mutant FUS induces selective motor neuron degeneration through toxic gain of function, *Nat. Commun.* 7 (2016) 10465.
- [31] E. Onesto, C. Colombrita, V. Gumina, M.O. Borghi, S. Dusi, A. Doretti, G. Fagiolari, F. Invernizzi, M. Moggio, V. Tiranti, V. Silani, A. Ratti, Gene-specific mitochondrial dysfunctions in human TARDBP and C9ORF72 fibroblasts, *Acta Neuropathol. Commun.* 4 (2016) 47.
- [32] R. Dafinca, J. Scaber, N.A. Ababneh, T. Lalic, G. Weir, H. Christian, J. Vowles, A.G.L. Douglas, A. Fletcher-Jones, C. Browne, M. Nakanishi, M.R. Turner, R. Wade-Martins, S.A. Cowley, K. Talbot, C9orf72 hexanucleotide expansions are associated with altered endoplasmic reticulum calcium homeostasis and stress granule formation in induced pluripotent stem cell-derived neurons from patients with amyotrophic lateral sclerosis and frontotemporal dementia, *Stem Cells* 34 (2016) 2063–2078.
- [33] M. Gautam, J.H. Jara, G. Sekerkova, M.V. Yasvoina, M. Martina, P.H. Ozdinler, Absence of alsin function leads to corticospinal motor neuron vulnerability via novel disease mechanisms, *Hum. Mol. Genet.* 25 (2016) 1074–1087.
- [34] H.Z. Yin, A. Nalbandian, C.I. Hsu, S. Li, K.J. Llewellyn, T. Mozaffar, V.E. Kimonis, J.H. Weiss, Slow development of ALS-like spinal cord pathology in mutant valosin-containing protein gene knock-in mice, *Cell Death Dis.* 3 (2012) e374.
- [35] A. Nalbandian, K.J. Llewellyn, M. Badadani, H.Z. Yin, C. Nguyen, V. Katherine, G. Watts, J. Mukherjee, J. Vesa, V. Caiozzo, T. Mozaffar, J.H. Weiss, V.E. Kimonis, A progressive translational mouse model of human valosin-containing protein disease: the VCPR155H/+ mouse, *Muscle Nerve* 47 (2013) 260–270.
- [36] S. Bannwarth, S. Ait-El-Mkadem, A. Chaussenet, E.C. Genin, S. Lucas-Gervais, K. Fragaki, L. Berg-Alonso, Y. Kageyama, V. Serre, D.G. Moore, A. Verschueren, C. Rouzier, I. Le Ber, G. Augé, C. Cochaud, F. Lespinasse, K. N'Guyen, A. de Septeuville, A. Brice, P. Yu-Wai-Man, H. Sesaki, J. Pouget, V. Paquis-Flucklinger, A mitochondrial origin for frontotemporal dementia and amyotrophic lateral sclerosis through CHCHD10 involvement, *Brain* 137 (2014) 2329–2345.
- [37] E.C. Genin, M. Plutino, S. Bannwarth, E. Villa, E. Cisneros-Barroso, M. Roy, B. Ortega-Vila, K. Fragaki, F. Lespinasse, E. Pinero-Martos, G. Augé, D. Moore, F. Burté, S. Lucas-Gervais, Y. Kageyama, K. Itoh, P. Yu-Wai-Man, H. Sesaki, J.E. Ricci, C. Vives-Bauza, V. Paquis-Flucklinger, CHCHD10 mutations promote loss of mitochondrial cristae junctions with impaired mitochondrial genome maintenance and inhibition of apoptosis, *EMBO Mol. Med.* 8 (2016) 58–72.
- [38] C. Stribi, A. Samara, D. Trümbach, R. Peis, M. Neumann, H. Fuchs, V. Gailus-Durner, M. Hrabé de Angelis, B. Rathkolb, E. Wolf, J. Beckers, M. Horsch, F. Neff, E. Kremmer, S. Koob, A.S. Reichert, W. Hans, J. Rozman, M. Klingenspor, M. Aichler, A.K. Walch, L. Becker, T. Klopstock, L. Glasl, S.M. Höller, W. Wurst, T. Floss, Mitochondrial dysfunction and decrease in body weight of a transgenic knock-in mouse model for TDP-43, *J. Biol. Chem.* 289 (2014) 10769–10784.
- [39] I.G. Kirkinezos, S.R. Bacman, D. Hernandez, J. Oca-Cossio, L.J. Arias, M.A. Perez-Pinzon, W.G. Bradley, C.T. Moraes, Cytochrome c association with the inner mitochondrial membrane is impaired in the CNS of G93A-SOD1 mice, *J. Neurosci.* 25 (2005) 164–172.
- [40] J. Kong, Z. Xu, Massive mitochondrial degeneration in motor neurons triggers the onset of amyotrophic lateral sclerosis in mice expressing a mutant SOD1, *J. Neurosci.* 18 (1998) 3241–3250.
- [41] C.M. Higgins, C. Jung, H. Ding, Z. Xu, Mutant Cu, Zn superoxide dismutase that causes motoneuron degeneration is present in mitochondria in the CNS, *J. Neurosci.* 22 (2002).
- [42] W. Wang, L. Wang, J. Lu, S.L. Siedlak, H. Fujioka, J. Liang, S. Jiang, X. Ma, Z. Jiang, E.L. da Rocha, M. Sheng, H. Choi, P.H. Lerou, H. Li, X. Wang, The inhibition of TDP-43 mitochondrial localization blocks its neuronal toxicity, *Nat. Med.* 22 (2016) 869–878.
- [43] R. Lopez-Gonzalez, Y. Lu, T.F. Gendron, A. Karydas, H. Tran, D. Yang, L. Petruccielli, B.L. Miller, S. Almeida, F.B. Gao, Poly(GR) in C9ORF72-related ALS/FTD compromises mitochondrial function and increases oxidative stress and DNA damage in iPSC-derived motor neurons, *Neuron* 92 (2016) 383–391.
- [44] A.M. Blokhuis, M. Koppers, E.J. Groen, D.M. van den Heuvel, S. Dini Modigliani, J.J. Anink, K. Fumoto, F. van Diggelen, A. Snelting, P. Sodaar, B.M. Verheijen, J.A. Demmers, J.H. Veldink, E. Aronica, I. Bozzoni, J. den Hertog, L.H. van den Berg, R.J. Pasterkamp, Comparative interactomics analysis of different ALS-associated proteins identifies converging molecular pathways, *Acta Neuropathol. (Berl.)* 132 (2016) 175–196.
- [45] M. Mattiazzi, M. D'Aurelio, C.D. Gajewski, K. Martushova, M. Kiaei, M. Flint Beal, G. Manfredi, Mutated human SOD1 causes dysfunction of oxidative phosphorylation in mitochondria of transgenic mice, *J. Biol. Chem.* 277 (2002) 29626–29633.
- [46] A. Ferri, M. Cozzolino, C. Crosio, M. Nencini, A. Casciati, E.B. Gralla, G. Rotilio, J.S. Valentine, M.T. Carri, Familial ALS-superoxide dismutases associate with mitochondria and shift their redox potentials, *Proc. Natl. Acad. Sci. U. S. A.* 103 (2006) 13860–13865.
- [47] C. Vijayvergiya, M.F. Beal, J. Buck, G. Manfredi, Mutant superoxide dismutase 1 forms aggregates in the brain mitochondrial matrix of amyotrophic lateral sclerosis mice, *J. Neurosci.* 25 (2005) 2463–2470.
- [48] V. Shoshan-Barmatz, D. Ben-Hail, VDAC, a multi-functional mitochondrial protein as a pharmacological target, *Mitochondrion* 12 (2012) 24–34.
- [49] A. Israelson, N. Arbel, S. Da Cruz, H. Ilieva, K. Yamanaka, V. Shoshan-Barmatz, D.W. Cleveland, Misfolded mutant SOD1 directly inhibits VDAC1 conductance in a mouse model of inherited ALS, *Neuron* 67 (2010) 575–587.
- [50] P. Pasinelli, M.E. Belford, N. Lennon, B.J. Bacska, B.T. Hyman, D. Trott, R.H. Brown Jr., Amyotrophic lateral sclerosis-associated SOD1 mutant proteins bind and aggregate with Bcl-2 in spinal cord mitochondria, *Neuron* 43 (2004) 19–30.
- [51] R. Stoica, S. Paillusson, P. Gomez-Suaga, J.C. Mitchell, D.H. Lau, E.H. Gray, R.M. Sancho, G. Vizcay-Barrena, K.J. De Vos, C.E. Shaw, D.P. Hanger, W. Noble, C.C. Miller, ALS/FTD-associated FUS activates GSK-3beta to disrupt the VAPB-PTPIP51 interaction and ER-mitochondria associations, *EMBO Rep.* 17 (2016) 1326–1342.
- [52] F.R. Wiedemann, G. Manfredi, C. Mawrin, M. Flint Beal, E.A. Schon, Mitochondrial DNA and respiratory chain function in spinal cords of ALS patients, *J. Neurochem.* 80 (2002) 616–625.
- [53] G.M. Borthwick, M.A. Johnson, P.G. Ince, P.J. Shaw, D.M. Turnbull, Mitochondrial enzyme activity in amyotrophic lateral sclerosis: implications for the role of mitochondria in neuronal cell death, *Ann. Neurol.* 46 (1999) 787–790.
- [54] F.R. Wiedemann, K. Winkler, A.V. Kuznetsov, C. Bartels, S. Vielhaber, H. Feistner, W.S. Kunz, Impairment of mitochondrial function in skeletal

- muscle of patients with amyotrophic lateral sclerosis.** *J. Neurol. Sci.* 156 (1998) 65–72.
- [55] S. Vielhaber, D. Kunz, K. Winkler, F.R. Wiedemann, E. Kirches, H. Feistner, H.J. Heinze, C.E. Elger, W. Schubert, W.S. Kunz, Mitochondrial DNA abnormalities in skeletal muscle of patients with sporadic amyotrophic lateral sclerosis, *Brain* 123 (2000) 1339–1348.
- [56] V. Crugnola, C. Lamperti, V. Lucchini, D. Ronchi, L. Peverelli, A. Prelle, M. Sciacca, A. Bordonali, E. Fassone, F. Fortunato, S. Corti, V. Silani, N. Bresolin, S. Di Mauro, G.P. Comi, M. Moggi, Mitochondrial respiratory chain dysfunction in muscle from patients with amyotrophic lateral sclerosis, *Arch. Neurol.* 67 (2010) 849–854.
- [57] P. Ghiasi, S. Hosseinkhani, A. Noori, S. Nafissi, K. Khajeh, Mitochondrial complex I deficiency and ATP/ADP ratio in lymphocytes of amyotrophic lateral sclerosis patients, *Neurol. Res.* 34 (2012) 297–303.
- [58] K. Kirk, C. Gennings, J.C. Hupf, S. Tadesse, M. D'Aurelio, H. Kawamata, F. Valsecchi, H. Mitsumoto, G. Manfredi, Bioenergetic markers in skin fibroblasts of sporadic amyotrophic lateral sclerosis and progressive lateral sclerosis patients, *Ann. Neurol.* 76 (2014) 620–624.
- [59] C. Jung, C.M.J. Higgins, Z. Xu, Mitochondrial electron transport chain complex dysfunction in a transgenic mouse model for amyotrophic lateral sclerosis, *J. Neurochem.* 83 (2002) 535–545.
- [60] S. Loizzo, M. Pieri, A. Ferri, M.T. Carrà, C. Zona, A. Fortuna, S. Vella, Dynamic NAD(P)H post-synaptic autofluorescence signals for the assessment of mitochondrial function in a neurodegenerative disease: monitoring the primary motor cortex of G93A mice an amyotrophic lateral sclerosis model, *Mitochondrion* 10 (2010) 108–114.
- [61] S.P. Allen, S. Rajan, L. Duffy, H. Mortiboys, A. Higginbottom, A.J. Grierson, P.J. Shaw, Superoxide dismutase 1 mutation in a cellular model of amyotrophic lateral sclerosis shifts energy generation from oxidative phosphorylation to glycolysis, *Neurobiol. Aging* 35 (2014) 1499–1509.
- [62] F.M. Menzies, M.R. Cookson, R.W. Taylor, D.M. Turnbull, Z.M. Chrzanowska-Lightowler, L. Dong, D.A. Figlewicz, P.J. Shaw, Mitochondrial dysfunction in a cell culture model of familial amyotrophic lateral sclerosis, *Brain* 125 (2002) 1522–1533.
- [63] E. Coussee, P. De Smet, E. Bogaert, I. Elens, P. Van Damme, P. Willems, W. Koopman, L. Van Den Bosch, G. Callewaert, G37R SOD1 mutant alters mitochondrial complex I activity, Ca²⁺ uptake and ATP production, *Cell Calcium* 49 (2011) 217–225.
- [64] K. Richardson, S.P. Allen, H. Mortiboys, A.J. Grierson, S.B. Wharton, P.G. Ince, P.J. Shaw, P.R. Heath, The effect of SOD1 mutation on cellular bioenergetic profile and viability in response to oxidative stress and influence of mutation-type, *PLoS One* 8 (2013) e68256.
- [65] A.C. Bowling, J.B. Schulz, R.H. Brown Jr., M.F. Beal, Superoxide dismutase activity, oxidative damage, and mitochondrial energy metabolism in familial and sporadic amyotrophic lateral sclerosis, *J. Neurochem.* 61 (1993) 2322–2325.
- [66] S.E. Browne, A.C. Bowling, M.J. Baik, M. Gurney, R.H. Brown Jr., M.F. Beal, Metabolic dysfunction in familial but not sporadic, amyotrophic lateral sclerosis, *J. Neurochem.* 71 (1998) 281–287.
- [67] S. Wendt, A. Dedeoglu, O. Speer, T. Wallimann, M.F. Beal, O.A. Andreassen, Reduced creatine kinase activity in transgenic amyotrophic lateral sclerosis mice, *Free Rad. Biol. Med.* 32 (2002) 920–926.
- [68] M. Damiano, A.A. Starkov, S. Petri, K. Kipiani, M. Kiae, M. Mattiazzi, M. Flint Beal, G. Manfredi, Neural mitochondrial Ca²⁺ capacity impairment precedes the onset of motor symptoms in G93A Cu/Zn-superoxide dismutase mutant mice, *J. Neurochem.* 96 (2006) 1349–1361.
- [69] W. Duan, X. Li, J. Shi, Y. Guo, Z. Li, C. Li, Mutant TAR DNA-binding protein-43 induces oxidative injury in motor neuron-like cell, *Neuroscience* 169 (2010) 1621–1629.
- [70] J. Lu, W. Duan, Y. Guo, H. Jiang, Z. Li, J. Huang, K. Hong, C. Li, Mitochondrial dysfunction in human TDP-43 transfected NSC34 cell lines and the protective effect of dimethoxy curcumin, *Brain Res. Bull.* 89 (2012) 185–190.
- [71] H. Tagashira, Y. Shinoda, N. Shioda, K. Fukunaga, Methyl pyruvate rescues mitochondrial damage caused by S100A1 mutation related to amyotrophic lateral sclerosis, *Biochim. Biophys. Acta* 1840 (2014) 3320–3334.
- [72] M.L. Seibenhener, Y. Du, M.T. Diaz-Meco, J. Moscat, M.C. Wooten, M.W. Wooten, A role for sequestosome 1/p62 in mitochondrial dynamics, Import and genome integrity, *Biochim. Biophys. Acta – Mol. Cell Res.* 1833 (2013) 452–459.
- [73] F. Bartolome, H.C. Wu, V.S. Burchell, E. Preza, S. Wray, C.J. Mahoney, N.C. Fox, A. Calvo, A. Canosa, C. Moglia, J. Mandrioli, A. Chiò, R.W. Orrell, H. Houlden, J. Hardy, A.Y. Abramov, H. Plun-Favreau, Pathogenic VCP mutations induce mitochondrial uncoupling and reduced ATP levels, *Neuron* 78 (2013) 57–64.
- [74] A.M. Clement, M.D. Nguyen, E.A. Roberts, M.L. Garcia, S. Boillée, M. Rule, A.P. McMahon, W. Doucette, D. Siwek, R.J. Ferrante, R.H. Brown Jr., J.P. Julien, L.S.B. Goldstein, D.W. Cleveland, Wild-type nonneuronal cells extend survival of SOD1 mutant motor neurons in ALS mice, *Science* 302 (2003) 113–117.
- [75] K.A. Staats, L. Van Den Bosch, Astrocytes in amyotrophic lateral sclerosis: direct effects on motor neuron survival, *J. Biol. Phys.* 35 (2009) 337–346.
- [76] L. Pellerin, P.J. Magistretti, Glutamate uptake into astrocytes stimulates aerobic glycolysis: a mechanism coupling neuronal activity to glucose utilization, *Proc. Natl. Acad. Sci. U. S. A.* 91 (1994) 10625–10629.
- [77] M. Bélanger, I. Allaman, Pierre J. Magistretti, Brain energy metabolism: focus on astrocyte-neuron metabolic cooperation, *Cell Metab.* 14 (2011) 724–738.
- [78] L. Ferraiuolo, A. Higginbottom, P.R. Heath, S. Barber, D. Greenald, J. Kirby, P.J. Shaw, Dysregulation of astrocyte-motoneuron cross-talk in mutant superoxide dismutase 1-related amyotrophic lateral sclerosis, *Brain* 134 (2011) 2627–2641.
- [79] Y. Lee, B.M. Morrison, Y. Li, S. Lengacher, M.H. Farah, P.N. Hoffman, Y. Liu, A. Tsingalia, L. Jin, P.W. Zhang, L. Pellerin, P.J. Magistretti, J.D. Rothstein, Oligodendroglia metabolically support axons and contribute to neurodegeneration, *Nature* 487 (2012) 443–448.
- [80] J.F. Turrens, Mitochondrial formation of reactive oxygen species, *J. Physiol. (Lond.)* 552 (2003) 335–344.
- [81] M.P. Murphy, How mitochondria produce reactive oxygen species, *Biochem. J.* 417 (2009) 1–13.
- [82] G.H. Kim, J.E. Kim, S.J. Rhee, S. Yoon, The role of oxidative stress in neurodegenerative diseases, *Exp. Neurobiol.* 24 (2015) 325–340.
- [83] S. Gandhi, A.Y. Abramov, Mechanism of oxidative stress in neurodegeneration, *Oxid. Med. Cell. Longev.* 2012 (2012) 428010.
- [84] H. Cui, Y. Kong, H. Zhang, Oxidative stress, mitochondrial dysfunction, and aging, *J. Signal Transduc.* 2012 (2012) 646354.
- [85] J.K. Andersen, Oxidative stress in neurodegeneration: cause or consequence? *Nat. Med.* 10 (Suppl) (2004) S18–S25.
- [86] S.C. Barber, P.J. Shaw, Oxidative stress in ALS: Key role in motor neuron injury and therapeutic target, *Free Radic. Biol. Med.* 48 (2010) 629–641.
- [87] R.G. Smith, Y.K. Henry, M.P. Mattson, S.H. Appel, Presence of 4-hydroxyxynonenal in cerebrospinal fluid of patients with sporadic amyotrophic lateral sclerosis, *Ann. Neurol.* 44 (1998) 696–699.
- [88] E.P. Simpson, Y.K. Henry, J.S. Henkel, R.G. Smith, S.H. Appel, Increased lipid peroxidation in sera of ALS patients: a potential biomarker of disease burden, *Neurology* 62 (2004) 1758–1765.
- [89] H. Mitsumoto, R. Santella, X. Liu, M. Bogdanov, J. Zipprich, H.C. Wu, J. Mahata, M. Kilti, K. Bednarz, D. Bell, P.H. Gordon, M. Hornig, M. Mehrazin, A. Naini, M. Flint Beal, P. Factor-Litvak, Oxidative stress biomarkers in sporadic ALS, *Amyotroph. Lateral Scler.* 9 (2008) 177–183.
- [90] M. Bogdanov, R.H. Brown Jr., W. Matson, R. Smart, D. Hayden, H. O'Donnell, M. Flint Beal, M. Cudkowicz, Increased oxidative damage to DNA in ALS patients, *Free Radic. Biol. Med.* 29 (2000) 652–658.
- [91] P.J. Shaw, P.G. Ince, G. Falkous, D. Mantle, Oxidative damage to protein in sporadic motor neuron disease spinal cord, *Ann. Neurol.* 38 (1995) 691–695.
- [92] N. Shibata, R. Nagai, K. Uchida, S. Horiuchi, S. Yamada, A. Hirano, M. Kawaguchi, T. Yamamoto, S. Sasaki, M. Kobayashi, Morphological evidence for lipid peroxidation and protein glycoxidation in spinal cords from sporadic amyotrophic lateral sclerosis patients, *Brain Res.* 917 (2001) 97–104.
- [93] P.S. Fitzmaurice, I.C. Shaw, H.E. Kleiner, R.T. Miller, T.J. Monks, S.S. Lau, J.D. Mitchell, P.G. Lynch, Evidence for DNA damage in amyotrophic lateral sclerosis, *Muscle Nerve* 19 (1996) 797–798.
- [94] Y. Chang, Q. Kong, X. Shan, G. Tian, H. Ilieva, D.W. Cleveland, J.D. Rothstein, D.R. Borchelt, P.C. Wong, C.L.G. Lin, Messenger RNA oxidation occurs early in disease pathogenesis and promotes motor neuron degeneration in ALS, *PLoS One* 3 (2008).
- [95] M. Said Ahmed, W.Y. Hung, J.S. Zu, P. Hockberger, T. Siddique, Increased reactive oxygen species in familial amyotrophic lateral sclerosis with mutations in SOD1, *J. Neurol. Sci.* 176 (2000) 88–94.
- [96] G. Sala, F. Trombin, L. Mattavelli, S. Beretta, L. Tremolizzo, S. Andreoni, E. Calabrese, L. Sanvitto, C. Ferrarese, Lack of evidence for oxidative stress in sporadic amyotrophic lateral sclerosis fibroblasts, *Neurodegener. Dis.* 6 (2009) 9–15.
- [97] G.A. Jansen, R.J. Wanders, G.J. Jobsis, P.A. Bolhuis, J.M. de Jong, Evidence against increased oxidative stress in fibroblasts from patients with non-superoxide-dismutase-1 mutant familial amyotrophic lateral sclerosis, *J. Neurol. Sci.* 139 (Suppl) (1996) 91–94.
- [98] T. Aguirre, L. Van Den Bosch, K. Goetschalckx, P. Tilkin, G. Mathijss, J.J. Cassiman, W. Robberecht, Increased sensitivity of fibroblasts from amyotrophic lateral sclerosis patients to oxidative stress, *Ann. Neurol.* 43 (1998) 452–457.
- [99] P.K. Andrus, T.J. Fleck, M.E. Gurney, E.D. Hall, Protein oxidative damage in a transgenic mouse model of familial amyotrophic lateral sclerosis, *J. Neurochem.* 71 (1998) 2041–2048.
- [100] R. Rakshit, P. Cunningham, A. Furtoš-Matei, S. Dahan, X.F. Qi, J.P. Crow, N.R. Cashman, L.H. Kondejewski, A. Chakrabarty, Oxidation-induced misfolding and aggregation of superoxide dismutase and its implications for amyotrophic lateral sclerosis, *J. Biol. Chem.* 277 (2002) 47551–47556.
- [101] S. Pickles, L. Destroismaisons, S.L. Peyrard, S. Cadot, G.A. Rouleau, R.H. Brown, J.P. Julien, N. Arbour, C.V. Velde, Mitochondrial damage revealed by immunoselection for ALS-linked misfolded SOD1, *Hum. Mol. Genet.* 22 (2013) 3947–3959.
- [102] S. Pickles, S. Semmler, H.R. Broom, L. Destroismaisons, L. Legroux, N. Arbour, E. Meiering, N.R. Cashman, C. Vande Velde, ALS-linked misfolded SOD1 species have divergent impacts on mitochondria, *Acta Neuropathol. Commun.* 4 (2016) 43.
- [103] T.E. Brotherton, Y. Li, D. Cooper, M. Gearing, J.P. Julien, J.D. Rothstein, K. Boylan, J.D. Glass, Localization of a toxic form of superoxide dismutase 1 protein to pathologically affected tissues in familial ALS, *Proc. Natl. Acad. Sci. U. S. A.* 109 (2012) 5505–5510.
- [104] D.A. Bosco, G. Morfini, N.M. Karabacak, Y. Song, F. Gros-Louis, P. Pasinelli, H. Goolsby, B.A. Fontaine, N. Lemay, D. McKenna-Yasek, M.P. Frosch, J.N. Agar, J.P. Julien, S.T. Brady, R.H. Brown, Wild-type and mutant SOD1 share an

- aberrant conformation and a common pathogenic pathway in ALS, *Nat. Neurosci.* 13 (2010) 1396–1403.
- [105] J.I. Ayers, G. Xu, O. Pletnikova, J.C. Troncoso, P.J. Hart, D.R. Borchelt, Conformational specificity of the C4F6 SOD1 antibody; low frequency of reactivity in sporadic ALS cases, *Acta Neuropathol. Commun.* 2 (2014) 55.
- [106] T.J. Cohen, A.W. Hwang, T. Unger, J.Q. Trojanowski, V.M. Lee, Redox signalling directly regulates TDP-43 via cysteine oxidation and disulphide cross-linking, *EMBO J.* 31 (2012) 1241–1252.
- [107] T.J. Cohen, A.W. Hwang, C.R. Restrepo, C.X. Yuan, J.Q. Trojanowski, V.M. Lee, An acetylation switch controls TDP-43 function and aggregation propensity, *Nat. Commun.* 6 (2015) 5845.
- [108] C. Kabuta, K. Kono, K. Wada, T. Kabuta, 4-Hydroxyxynonenal induces persistent insolubilization of TDP-43 and alters its intracellular localization, *Biochem. Biophys. Res. Commun.* 463 (2015) 82–87.
- [109] C. Colombrita, E. Zennaro, C. Fallini, M. Weber, A. Sommacal, E. Buratti, V. Silani, A. Ratti, TDP-43 is recruited to stress granules in conditions of oxidative insult, *J. Neurochem.* 111 (2009) 1051–1061.
- [110] J. Meyerowitz, S.J. Parker, L.J. Vella, D.C. Ng, K.A. Price, J.R. Liddell, A. Caragounis, Q.X. Li, C.L. Masters, T. Nonaka, M. Hasegawa, M.A. Bogoyevitch, K.M. Kanninen, P.J. Crouch, A.R. White, C-Jun N-terminal kinase controls TDP-43 accumulation in stress granules induced by oxidative stress, *Mol. Neurodegener.* 6 (2011).
- [111] K.K. McDonald, A. Aulas, L. Destroismaisons, S. Pickles, E. Beleac, W. Camu, G.A. Rouleau, C. Vande Velde, TAR DNA-binding protein 43 (TDP-43) regulates stress granule dynamics via differential regulation of G3BP and TIA-1, *Hum. Mol. Genet.* 20 (2011) 1400–1410.
- [112] M.J. Finelli, K.X. Liu, Y. Wu, P.L. Oliver, K.E. Davies, Oxr1 improves pathogenic cellular features of ALS-associated FUS and TDP-43 mutations, *Hum. Mol. Genet.* 24 (2015) 3529–3544.
- [113] M.T. Carrà, A. Ferri, A. Battistoni, L. Famhy, R. Gabbianelli, F. Poccia, G. Rotilio, Expression of a Cu, Zn superoxide dismutase typical of familial amyotrophic lateral sclerosis induces mitochondrial alteration and increase of cytosolic Ca²⁺ concentration in transfected neuroblastoma SH-SY5Y cells, *FEBS Lett.* 414 (1997) 365–368.
- [114] G.M. Mórotz, K.J. De Vos, A. Vagnoni, S. Ackerley, C.E. Shaw, C.C.J. Miller, Amyotrophic lateral sclerosis-associated mutant VAPB/P56s perturbs calcium homeostasis to disrupt axonal transport of mitochondria, *Hum. Mol. Genet.* 21 (2012) 1979–1988.
- [115] R. Stoica, K.J. De Vos, S. Paillusson, S. Mueller, R.M. Sancho, K.F. Lau, G. Vizcay-Barrena, W.L. Lin, Y.F. Xu, J. Lewis, D.W. Dickson, L. Petruccielli, J.C. Mitchell, C.E. Shaw, C.C. Miller, ER-mitochondria associations are regulated by the VAPB-PTPIP51 interaction and are disrupted by ALS/FTD-associated TDP-43, *Nat. Commun.* 5 (2014) 3996.
- [116] L. Siklós, J. Engelhardt, Y. Harati, R.G. Smith, F. Joó, S.H. Appel, Ultrastructural evidence for altered calcium in motor nerve terminals in amyotrophic lateral sclerosis, *Ann. Neurol.* 39 (1996) 203–216.
- [117] L. Van Den Bosch, W. Vandenberghe, H. Klaassen, E. Van Houtte, W. Robberecht, Ca²⁺-permeable AMPA receptors and selective vulnerability of motor neurons, *J. Neurol. Sci.* 180 (2000) 29–34.
- [118] K.J. De Vos, G.M. Mórotz, R. Stoica, E.L. Tudor, K. Lau, S. Ackerley, A. Warley, C.E. Shaw, C.C. Miller, VAPB interacts with the mitochondrial protein PTP1B to regulate calcium homeostasis, *Hum. Mol. Genet.* 21 (2012) 1299–1311.
- [119] L. Siklos, J.I. Engelhardt, M.E. Alexianu, M.E. Gurney, T. Siddique, S.H. Appel, Intracellular calcium parallels motoneuron degeneration in SOD-1 mutant mice, *J. Neuropathol. Exp. Neurol.* 57 (1998) 571–587.
- [120] A. Fuchs, S. Kutterer, T. Mühlung, J. Duda, B. Schütz, B. Liss, B.U. Keller, J. Roeper, Selective mitochondrial Ca²⁺ uptake deficit in disease endstage vulnerable motoneurons of the SOD1 G93A mouse model of amyotrophic lateral sclerosis, *J. Physiol. (Lond.)* 591 (2013) 2723–2745.
- [121] E. Guatteo, I. Caruncho, M. Pieri, F. Albo, N. Canu, N.B. Mercuri, C. Zona, Altered calcium homeostasis in motor neurons following AMPA receptor but not voltage-dependent calcium channels' activation in a genetic model of amyotrophic lateral sclerosis, *Neurobiol. Dis.* 28 (2007) 90–100.
- [122] S. Paillusson, R. Stoica, P. Gomez-Suaga, D.H. Lau, S. Mueller, T. Miller, C.C. Miller, There's something wrong with my MAM; the ER-mitochondria axis and neurodegenerative diseases, *Trends Neurosci.* 39 (2016) 146–157.
- [123] G. Manfredi, H. Kawamata, Mitochondria and endoplasmic reticulum crosstalk in amyotrophic lateral sclerosis, *Neurobiol. Dis.* 90 (2016) 35–42.
- [124] N. Bernard-Marissal, J.J. Medard, H. Azzedine, R. Chrast, Dysfunction in endoplasmic reticulum-mitochondria crosstalk underlies SIGMAR1 loss of function mediated motor neuron degeneration, *Brain* 138 (2015) 875–890.
- [125] M. Mitne-Neto, M. Machado-Costa, M.C. Marchetto, M.H. Bengtson, C.A. Joazeiro, H. Tsuda, H.J. Bellen, H.C. Silva, A.S. Oliveira, M. Lazar, A.R. Muotri, M. Zatz, Downregulation of VAPB expression in motor neurons derived from induced pluripotent stem cells of ALS8 patients, *Hum. Mol. Genet.* 20 (2011) 3642–3652.
- [126] G. Agnostonou, M.T. Akbar, P. Paul, C. Angelinetta, T.J. Steiner, J. de Belleroche, Vesicle associated membrane protein B (VAPB) is decreased in ALS spinal cord, *Neurobiol. Aging* 31 (2010) 969–985.
- [127] J.Y. Kim, A. Jang, R. Reddy, W.H. Yoon, J.L. Jankowsky, Neuronal overexpression of human VAPB slows motor impairment and neuromuscular denervation in a mouse model of ALS, *Hum. Mol. Genet.* 25 (2016) 4661–4673.
- [128] T.L. Williams, N.C. Day, P.G. Ince, R.K. Kamboj, P.J. Shaw, Calcium-permeable α -amino-3-hydroxy-5-methyl-4-isoxazole propionic acid receptors: a molecular determinant of selective vulnerability in amyotrophic lateral sclerosis, *Ann. Neurol.* 42 (1997) 200–207.
- [129] P.J. Shaw, C.J. Eggert, Molecular factors underlying selective vulnerability of motor neurons to neurodegeneration in amyotrophic lateral sclerosis, *J. Neurol. Supplement* 247 (2000) 17–27.
- [130] M.E. Alexianu, B.K. Ho, A.H. Mohamed, V. La Bella, R.G. Smith, S.H. Appel, The role of calcium-binding proteins in selective motoneuron vulnerability in amyotrophic lateral sclerosis, *Ann. Neurol.* 36 (1994) 846–858.
- [131] P. Ince, N. Stout, P. Shaw, J. Slade, W. Hunziker, C.W. Heizmann, K.G. Bainbridge, Parvalbumin and calbindin D-28k in the human motor system and in motor neuron disease, *Neuropathol. Appl. Neurobiol.* 19 (1993) 291–299.
- [132] J. Palecek, M.B. Lips, B.U. Keller, Calcium dynamics and lowering in motoneurones of the mouse spinal cord, *J. Physiol. (Lond.)* 520 (1999) 485–502.
- [133] J. Grosskreutz, L. Van Den Bosch, B.U. Keller, Calcium dysregulation in amyotrophic lateral sclerosis, *Cell Calcium* 47 (2010) 165–174.
- [134] V. Tadic, T. Prell, J. Lautenschlaeger, J. Grosskreutz, The ER-mitochondria calcium cycle and ER stress response as therapeutic targets in amyotrophic lateral sclerosis, *Front. Cell. Neurosci.* 8 (2014) 147.
- [135] K.J. De Vos, M. Hafezparast, Neurobiology of axonal transport defects in motor neuron diseases: opportunities for translational research? *Neurobiol. Dis.* (2017), <http://dx.doi.org/10.1016/j.nbd.2017.02.004>.
- [136] C.P. Webster, E.F. Smith, P.J. Shaw, K.J. De Vos, Protein homeostasis in amyotrophic lateral sclerosis: therapeutic opportunities? *Front. Neurosci.* (2017), <http://dx.doi.org/10.3389/fnins.2017.00123>.
- [137] S.G. Carriero, S.L. Sensi, H.Z. Yin, J.H. Weiss, AMPA exposures induce mitochondrial Ca²⁺ overload and ROS generation in spinal motor neurons in vitro, *J. Neurosci.* 20 (2000) 240–250.
- [138] D. Aggad, J. Verrie, A. Tauffenberger, J.A. Parker, TDP-43 toxicity proceeds via calcium dysregulation and necrosis in aging *Caenorhabditis elegans* motor neurons, *J. Neurosci.* 34 (2014) 12093–12103.
- [139] T. Yamashita, T. Hideyama, K. Hachiga, S. Teramoto, J. Takano, N. Iwata, T.C. Saido, S. Kwak, A role for calpain-dependent cleavage of TDP-43 in amyotrophic lateral sclerosis pathology, *Nat. Commun.* 3 (2012) 1307.
- [140] A. Ashkenazi, G. Salvesen, Regulated cell death: signaling and mechanisms, *Annu. Rev. Cell Dev. Biol.* 30 (2014) 337–356.
- [141] L.D. Walensky, BCL-2 in the crosshairs: tipping the balance of life and death, *Cell Death Dif.* 13 (2006) 1339–1350.
- [142] S. Pedrini, D. Sau, S. Guareschi, M. Bogush, R.H. Brown Jr., N. Naniche, A. Kia, D. Trott, P. Pasinelli, ALS-linked mutant SOD1 damages mitochondria by promoting conformational changes in Bcl-2, *Hum. Mol. Genet.* 19 (2010) 2974–2986.
- [143] W. Tan, N. Naniche, A. Bogush, S. Pedrini, D. Trott, P. Pasinelli, Small peptides against the mutant SOD1/Bcl-2 toxic mitochondrial complex restore mitochondrial function and cell viability in mutant SOD1-mediated ALS, *J. Neurosci.* 33 (2013) 11588–11598.
- [144] L.M. Westrate, J.A. Drocco, K.R. Martin, W.S. Hlavacek, J.P. MacKeigan, Mitochondrial morphological features are associated with fission and fusion events, *PLoS One* 9 (2014).
- [145] D.C. Chan, Fusion and fission: interlinked processes critical for mitochondrial health, *Annu. Rev. Genet.* 46 (2012) 265–287.
- [146] A. Ferri, P. Fiorenzo, M. Nencini, M. Cozzolino, M.G. Pesaresi, C. Valle, S. Sepe, S. Moreno, M.T. Carrà, Glutaredoxin 2 prevents aggregation of mutant SOD1 in mitochondria and abolishes its toxicity, *Hum. Mol. Genet.* 19 (2010) 4529–4542.
- [147] W. Liu, F. Tian, N. Morimoto, T. Yamashita, Y. Ikeda, K. Deguchi, K. Abe, Dynamic changes of mitochondrial fusion and fission proteins in a murine model of amyotrophic lateral sclerosis, *Curr. Neurovasc. Res.* 10 (2013) 222–230.
- [148] G. Luo, J. Yi, C. Ma, Y. Xiao, F. Yi, T. Yu, J. Zhou, Defective mitochondrial dynamics is an early event in skeletal muscle of an amyotrophic lateral sclerosis mouse model, *PLoS One* 8 (2013) e82112.
- [149] Y.F. Xu, T.F. Gendron, Y.J. Zhang, W.L. Lin, S. D'Alton, H. Sheng, M.C. Casey, J. Tong, J. Knight, X. Yu, R. Rademakers, K. Boylan, M. Hutton, E. McGowan, D.W. Dickson, J. Lewis, L. Petruccielli, Wild-type human TDP-43 expression causes TDP-43 phosphorylation, mitochondrial aggregation, motor deficits, and early mortality in transgenic mice, *J. Neurosci.* 30 (2010) 10851–10859.
- [150] Y.F. Xu, Y.J. Zhang, W.L. Lin, X. Cao, C. Stelter, D.W. Dickson, J. Lewis, L. Petruccielli, Expression of mutant TDP-43 induces neuronal dysfunction in transgenic mice, *Mol. Neurodegener.* 6 (2011).
- [151] J.R. Friedman, L.L. Lackner, M. West, J.R. DiBenedetto, J. Nunnari, G.K. Voeltz, ER tubules mark sites of mitochondrial division, *Science* 334 (2011) 358–362.
- [152] S. Watanabe, H. Ilieva, H. Tamada, H. Nomura, O. Komine, F. Endo, S. Jin, P. Mancias, H. Kiyama, K. Yamanaka, Mitochondria-associated membrane collapse is a common pathomechanism in SIGMAR1- and SOD1-linked ALS, *EMBO Mol. Med.* 8 (2016) 1421–1437.
- [153] K. Fukunaga, Y. Shinoda, H. Tagashira, The role of SIGMAR1 gene mutation and mitochondrial dysfunction in amyotrophic lateral sclerosis, *J. Pharm. Sci.* 127 (2015) 36–41.
- [154] H. Hoitzing, I.G. Johnston, N.S. Jones, What is the function of mitochondrial networks? A theoretical assessment of hypotheses and proposal for future research, *Bioessays* 37 (2015) 687–700.
- [155] G. Ashrafi, T.L. Schwarz, The pathways of mitophagy for quality control and clearance of mitochondria, *Cell Death Dif.* 20 (2013) 31–42.

- [156] W. Song, Y. Song, B. Kincaid, B. Bossy, E. Bossy-Wetzel, Mutant SOD1 G93A triggers mitochondrial fragmentation in spinal cord motor neurons: neuroprotection by SIRT3 and PGC-1 α , *Neurobiol. Dis.* 51 (2013) 72–81.
- [157] A. Hamacher-Brady, N.R. Brady, Mitophagy programs: mechanisms and physiological implications of mitochondrial targeting by autophagy, *Cell. Mol. Life Sci.* 73 (2016) 775–795.
- [158] C. Lagier-Tourenne, M. Polymenidou, K.R. Hutt, A.Q. Vu, M. Baughn, S.C. Huelga, K.M. Clutario, S.C. Ling, T.Y. Liang, C. Mazur, E. Wancewicz, A.S. Kim, A. Watt, S. Freier, G.G. Hicks, J.P. Donohue, L. Shiu, C.F. Bennett, J. Ravits, D.W. Cleveland, G.W. Yeo, Divergent roles of ALS-linked proteins FUS/TLS and TDP-43 intersect in processing long pre-mRNAs, *Nat. Neurosci.* 15 (2012) 1488–1497.
- [159] M. Polymenidou, C. Lagier-Tourenne, K.R. Hutt, S.C. Huelga, J. Moran, T.Y. Liang, S.C. Ling, E. Sun, E. Wancewicz, C. Mazur, H. Kordasiewicz, Y. Sedaghat, J.P. Donohue, L. Shiu, C.F. Bennett, G.W. Yeo, D.W. Cleveland, Long pre-mRNA depletion and RNA missplicing contribute to neuronal vulnerability from loss of TDP-43, *Nat. Neurosci.* 14 (2011) 459–468.
- [160] M.L. Hebron, I. Lonskaya, K. Sharpe, P.P. Weerasinghe, N.K. Algarzae, A.R. Shekoyan, C.E. Moussa, Parkin ubiquitinates Tar-DNA binding protein-43 (TDP-43) and promotes its cytosolic accumulation via interaction with histone deacetylase 6 (HDAC6), *J. Biol. Chem.* 288 (2013) 4103–4115.
- [161] D. Avci, M.K. Lemberg, Clipping or extracting: two ways to membrane protein degradation, *Trends Cell Biol.* 25 (2015) 611–622.
- [162] H. Meyer, C.C. Weihl, The VCP/p97 system at a glance: connecting cellular function to disease pathogenesis, *J. Cell Sci.* 127 (2014) 3877–3883.
- [163] A. Tanaka, M.M. Cleland, S. Xu, D.P. Narendra, D.F. Suen, M. Karbowksi, R.J. Youle, Proteasome and p97 mediate mitophagy and degradation of mitofusins induced by Parkin, *J. Cell Biol.* 191 (2010) 1367–1380.
- [164] N.C. Kim, E. Tresse, R.M. Kolaitis, A. Molliex, R.E. Thomas, N.H. Alami, B. Wang, A. Joshi, R.B. Smith, G.P. Ritson, B.J. Winborn, J. Moore, J.Y. Lee, T.P. Yao, L. Pallanca, M. Kundu, J.P. Taylor, VCP is essential for mitochondrial quality control by PINK1/Parkin and this function is impaired by VCP mutations, *Neuron* 78 (2013) 65–80.
- [165] J.O. Johnson, J. Mandrioli, M. Benatar, Y. Abramzon, V.M. Van Deerlin, J.Q. Trojanowski, J.R. Gibbs, M. Brunetti, S. Gronka, J. Wuu, J. Ding, L. McCluskey, M. Martinez-Lage, D. Falcone, D.G. Hernandez, S. Arepalli, S. Chong, J.C. Schymick, J. Rothstein, F. Landi, Y.D. Wang, A. Calvo, G. Mora, M. Sabatelli, M.R. Monsurro, S. Battistini, F. Salvi, R. Spataro, P. Sola, G. Borghero, G. Galassi, S.W. Scholz, J.P. Taylor, G. Restagno, A. Chiò, B.J. Traynor, Exome sequencing reveals VCP mutations as a cause of familial ALS, *Neuron* 68 (2010) 857–864.
- [166] M. Koppers, M.M. van Blitterswijk, L. Vlam, P.A. Rowicka, P.W.J. van Vught, E.J.N. Groen, W.G.M. Spliet, J. Engelen-Lee, H.J. Schelhaas, M. de Visser, A.J. van der Kooi, W.L. van der Pol, R.J. Pasterkamp, J.H. Veldink, L.H. van den Berg, VCP mutations in familial and sporadic amyotrophic lateral sclerosis, *Neurobiol. Aging* 33 (837) (2012) e837.
- [167] W.K. Tang, D. Xia, Mutations in the human AAA+ chaperone p97 and related diseases, *Front. Mol. Biosci.* 3 (2016) 79.
- [168] M.A. Gitcho, J. Strider, D. Carter, L. Taylor-Reinwald, M.S. Forman, A.M. Goate, N.J. Cairns, VCP mutations causing frontotemporal lobar degeneration disrupt localization of TDP-43 and induce cell death, *J. Biol. Chem.* 284 (2009) 12384–12398.
- [169] S.K. Custer, M. Neumann, H. Lu, A.C. Wright, J.P. Taylor, Transgenic mice expressing mutant forms VCP/p97 recapitulate the full spectrum of IBMPFD including degeneration in muscle, brain and bone, *Hum. Mol. Genet.* 19 (2010) 1741–1755.
- [170] V. Majcher, A. Goode, V. James, R. Layfield, Autophagy receptor defects and ALS-FTLD, *Mol. Cell. Neurosci.* 66 (2015) 43–52.
- [171] B. Richter, D.A. Sliter, L. Herhaus, A. Stolz, C. Wang, P. Beli, G. Zaffagnini, P. Wild, S. Martens, S.A. Wagner, R.J. Youle, I. Dikic, Phosphorylation of OPTN by TBK1 enhances its binding to Ub chains and promotes selective autophagy of damaged mitochondria, *Proc. Natl. Acad. Sci. U. S. A.* 113 (2016) 4039–4044.
- [172] P. Wild, H. Farhan, D.G. McEwan, S. Wagner, V.V. Rogov, N.R. Brady, B. Richter, J. Korac, O. Waidmann, C. Choudhary, V. Dotsch, D. Bumann, I. Dikic, Phosphorylation of the autophagy receptor optineurin restricts Salmonella growth, *Science* 333 (2011) 228–233.
- [173] A. Freischmidt, T. Wieland, B. Richter, W. Ruf, V. Schaeffer, K. Muller, N. Marroquin, F. Nordin, A. Hubers, P. Weydt, S. Pinto, R. Press, S. Millecamps, N. Molko, E. Bernard, C. Desnuelle, M.H. Soriani, J. Dorst, E. Graf, U. Nordstrom, M.S. Feiler, S. Putz, T.M. Boeckters, T. Meyer, A.S. Winkler, J. Winkelmann, M. de Carvalho, D.R. Thal, M. Otto, T. Brannstrom, P. Volk, P. Kursula, K.M. Danzer, P. Lichtner, I. Dikic, T. Meitinger, A.C. Ludolph, T.M. Strom, P.M. Andersen, J.H. Weishaupt, Haploinsufficiency of TBK1 causes familial ALS and fronto-temporal dementia, *Nat. Neurosci.* 18 (2015) 631–636.
- [174] H. Maruyama, H. Morino, H. Ito, Y. Izumi, H. Kato, Y. Watanabe, Y. Kinoshita, M. Kamada, H. Nodera, H. Suzuki, O. Komure, S. Matsuu, K. Kobatake, N. Morimoto, K. Abe, N. Suzuki, M. Aoki, A. Kawata, T. Hirai, T. Kato, K. Ogasawara, A. Hirano, T. Takumi, H. Kusaka, K. Hagiwara, R. Kaji, H. Kawakami, Mutations of optineurin in amyotrophic lateral sclerosis, *Nature* 465 (2010) 223–226.
- [175] F. Fecto, J. Yan, S.P. Vemula, E. Liu, Y. Yang, W. Chen, J.G. Zheng, Y. Shi, N. Siddique, H. Arrat, S. Donkervoort, S. Ajroud-Driss, R.L. Sufit, S.L. Heller, H.X. Deng, T. Siddique, SQSTM1 mutations in familial and sporadic amyotrophic lateral sclerosis, *Arch. Neurol.* 68 (2011) 1440–1446.
- [176] W.C. Shen, H.Y. Li, G.C. Chen, Y. Chern, P.H. Tu, Mutations in the ubiquitin-binding domain of OPTN/optineurin interfere with autophagy-mediated degradation of misfolded proteins by a dominant-negative mechanism, *Autophagy* 11 (2015) 685–700.
- [177] Y.C. Wong, E.L.F. Holzbaur, Optineurin is an autophagy receptor for damaged mitochondria in parkin-mediated mitophagy that is disrupted by an ALS-linked mutation, *Proc. Natl. Acad. Sci. U. S. A.* 111 (2014) E4439–E4448.
- [178] A. Goode, K. Butler, J. Long, J. Cavey, D. Scott, B. Shaw, J. Sollnerberger, C. Gell, T. Johansen, N.J. Oldham, M.S. Searle, R. Layfield, Defective recognition of LC3B by mutant SQSTM1/p62 implicates impairment of autophagy as a pathogenic mechanism in ALS-FTLD, *Autophagy* 12 (2016) 1094–1104.
- [179] M. Lazarou, D.A. Sliter, L.A. Kane, S.A. Sarraf, C. Wang, J.L. Burman, D.P. Sideris, A.I. Fogel, R.J. Youle, The ubiquitin kinase PINK1 recruits autophagy receptors to induce mitophagy, *Nature* 524 (2015) 309–314.
- [180] J.M. Heo, A. Ordureau, J.A. Paulo, J. Rinehart, J.W. Harper, The PINK1-PARKIN mitochondrial ubiquitylation pathway drives a program of OPTN/NDP52 recruitment and TBK1 activation to promote mitophagy, *Mol. Cell* 60 (2015) 7–20.
- [181] F. Fornai, P. Longone, L. Cafaro, O. Kastsiuchenka, M. Ferrucci, M.L. Manca, G. Lazzeri, A. Spalloni, N. Bellio, P. Lenzi, N. Modugno, G. Siciliano, C. Isidoro, L. Murri, S. Ruggieri, A. Paparelli, Lithium delays progression of amyotrophic lateral sclerosis, *Proc. Natl. Acad. Sci. U. S. A.* 105 (2008) 2052–2057.
- [182] Y. Xie, B. Zhou, M.Y. Lin, S. Wang, K.D. Foust, Z.H. Sheng, Endolysosomal deficits augment mitochondria pathology in spinal motor neurons of asymptomatic FALS mice, *Neuron* 87 (2015) 355–370.
- [183] C. Selleri, M.L. Campanari, C. Julie Corbier, A. Gaucheret, I. Kolb-Cheynel, M. Oulad-Abdelghani, F. Ruffenach, A. Page, S. Ciura, E. Kabashi, N. Charlet-Berguerand, Loss of C9ORF72 impairs autophagy and synergizes with polyQ Ataxin-2 to induce motor neuron dysfunction and cell death, *EMBO J.* 35 (2016) 1276–1297.
- [184] C.P. Webster, E.F. Smith, C.S. Bauer, A. Moller, G.M. Hautbergue, L. Ferraiuolo, M.A. Myszcynska, A. Higginbottom, M.J. Walsh, A.J. Whitworth, B.K. Kaspar, K. Meyer, P.J. Shaw, A.J. Grierson, K.J. De Vos, The C9orf72 protein interacts with Rab1a and the ULK1 complex to regulate initiation of autophagy, *EMBO J.* 35 (2016) 1656–1676.
- [185] P.M. Sullivan, X. Zhou, A.M. Robins, D.H. Paushter, D. Kim, M.B. Smolka, F. Hu, The ALS/FTLD associated protein C9orf72 associates with SMCR8 and WDR41 to regulate the autophagy-lysosome pathway, *Acta Neuropathol. Commun.* 4 (2016) 51.
- [186] M. Yang, C. Liang, K. Swaminathan, S. Herrlinger, F. Lai, R. Shiekhattar, J.F. Chen, A C9orf72/SMCR8-containing complex regulates ULK1 and plays a dual role in autophagy, *Sci. Adv.* 2 (2016) e1601167.
- [187] C.P. Webster, E.F. Smith, A.J. Grierson, K.J. De Vos, C9orf72 plays a central role in Rab GTPase-dependent regulation of autophagy, *Small GTPases* (2016) 1–10.
- [188] Y. Yang, A. Hentati, H.X. Deng, O. Dabbagh, T. Sasaki, M. Hirano, W.Y. Hung, K. Ouahchi, J. Yan, A.C. Azim, N. Cole, G. Gascon, A. Yagmour, M. Ben-Hamida, M. Pericak-Vance, F. Hentati, T. Siddique, The gene encoding alsin a protein with three guanine-nucleotide exchange factor domains, is mutated in a form of recessive amyotrophic lateral sclerosis, *Nat. Genet.* 29 (2001) 160–165.
- [189] S. Hadano, C.K. Hand, H. Osuga, Y. Yanagisawa, A. Otomo, R.S. Devon, N. Miyamoto, J. Showguchi-Miyata, Y. Okada, R. Singaraja, D.A. Figlewicz, T. Kwiatkowski, B.A. Hosler, T. Sagie, J. Skaug, J. Nasir, R.H. Brown, S.W. Scherer, G.A. Rouleau, M.R. Hayden, J.E. Ikeda, A gene encoding a putative GTPase regulator is mutated in familial amyotrophic lateral sclerosis 2, *Nat. Genet.* 29 (2001) 166–173.
- [190] S. Hadano, S.C. Benn, S. Kakuta, A. Otomo, K. Sudo, R. Kunita, K. Suzuki-Utsunomiya, H. Mizumura, J.M. Shefner, G.A. Cox, Y. Iwakura, J.E. Brown Jr., R.H. Ikeda, Mice deficient in the Rab5 guanine nucleotide exchange factor ALS2/alsin exhibit age-dependent neurological deficits and altered endosome trafficking, *Hum. Mol. Genet.* 15 (2006) 233–250.
- [191] K.J. De Vos, A.J. Grierson, S. Ackerley, C.C.J. Miller, Role of axonal transport in neurodegenerative diseases, *Annu. Rev. Neurosci.* 31 (2008) 151–173.
- [192] N.H. Alami, R.B. Smith, M.A. Carrasco, L.A. Williams, C.S. Winborn, S.S. Han, E. Kiskinis, B. Winborn, B.D. Freibaum, A. Kanagaraj, A.J. Clare, N.M. Badders, B. Bilican, E. Chaum, S. Chandran, C.E. Shaw, K.C. Eggan, T. Maniatis, J.P. Taylor, Axonal transport of TDP-43 mRNA granules is impaired by ALS-causing mutations, *Neuron* 81 (2014) 536–543.
- [193] L.G. Bilsland, E. Sahai, G. Kelly, M. Golding, L. Greensmith, G. Schiavo, Deficits in axonal transport precede ALS symptoms in vivo, *Proc. Natl. Acad. Sci. U. S. A.* 107 (2010) 20523–20528.
- [194] J. Magrané, M.A. Sahawneh, S. Przedborski, A.G. Estévez, G. Manfredi, Mitochondrial dynamics and bioenergetic dysfunction is associated with synaptic alterations in mutant sod1 motor neurons, *J. Neurosci.* 32 (2012) 229–242.
- [195] P. Marinković, M.S. Reuter, M.S. Brill, L. Godinho, M. Kerschensteiner, T. Misgeld, Axonal transport deficits and degeneration can evolve independently in mouse models of amyotrophic lateral sclerosis, *Proc. Natl. Acad. Sci. U. S. A.* 109 (2012) 4296–4301.
- [196] X. Shan, P.M. Chiang, D.L. Price, P.C. Wong, Altered distributions of Gemini of coiled bodies and mitochondria in motor neurons of TDP-43 transgenic mice, *Proc. Natl. Acad. Sci. U. S. A.* 107 (2010) 16325–16330.
- [197] J.R. Sotelo-Silveira, P. Lepanto, V. Elizondo, S. Horjales, F. Palacios, L. Martinez-Palma, M. Marin, J.S. Beckman, L. Barbeito, Axonal mitochondrial

- clusters containing mutant SOD1 in transgenic models of ALS, *Antioxidants Redox Signal.* 11 (2009) 1535–1545.
- [198] M.J. Devine, N. Birsu, J.T. Kittler, Miro sculpts mitochondrial dynamics in neuronal health and disease, *Neurobiol. Dis.* 90 (2016) 27–34.
- [199] S. Fransson, A. Ruusala, P. Aspenstrom, The atypical Rho GTPases Miro-1 and Miro-2 have essential roles in mitochondrial trafficking, *Biochem. Biophys. Res. Commun.* 344 (2006) 500–510.
- [200] A.F. MacAskill, K. Brickley, F.A. Stephenson, J.T. Kittler, GTPase dependent recruitment of Grif-1 by Miro1 regulates mitochondrial trafficking in hippocampal neurons, *Mol. Cell. Neurosci.* 40 (2009) 301–312.
- [201] E.E. Glater, L.J. Megeath, R.S. Stowers, T.L. Schwarz, Axonal transport of mitochondria requires milton to recruit kinesin heavy chain and is light chain independent, *J. Cell Biol.* 173 (2006) 545–557.
- [202] K. Brickley, M.J. Smith, M. Beck, F.A. Stephenson, GRIF-1 and OIP106, members of a novel gene family of coiled-coil domain proteins: association *in vivo* and *in vitro* with kinesin, *J. Biol. Chem.* 280 (2005) 14723–14732.
- [203] K. Brickley, F.A. Stephenson, Trafficking kinesin protein (TRAK)-mediated transport of mitochondria in axons of hippocampal neurons, *J. Biol. Chem.* 286 (2011) 18079–18092.
- [204] M. van Spronsen, M. Mikhaylova, J. Lipka, M.A. Schlager, D.J. van den Heuvel, M. Kuijpers, P.S. Wulf, N. Keijzer, J. Demmers, L.C. Kapitein, D. Jaarsma, H.C. Gerritsen, A. Akhmanova, C.C. Hoogenraad, TRAK/Milton motor-adaptor proteins steer mitochondrial trafficking to axons and dendrites, *Neuron* 77 (2013) 485–502.
- [205] G. Morlino, O. Barreiro, F. Baixaui, J. Robles-Valero, J.M. Gonzalez-Granado, R. Villa-Bellotsa, J. Cuena, C.O. Sanchez-Sorzano, E. Veiga, N.B. Martin-Cofreces, F. Sanchez-Madrid, Miro-1 links mitochondria and microtubule Dynein motors to control lymphocyte migration and polarity, *Mol. Cell. Biol.* 34 (2014) 1412–1426.
- [206] A.F. Macaskill, J.E. Rinholm, A.E. Twelvetrees, I.L. Arancibia-Carcamo, J. Muir, A. Fransson, P. Aspenstrom, D. Attwell, J.T. Kittler, Miro1 is a calcium sensor for glutamate receptor-dependent localization of mitochondria at synapses, *Neuron* 61 (2009) 541–555.
- [207] X. Wang, T.L. Schwarz, The mechanism of Ca²⁺ –dependent regulation of kinesin-mediated mitochondrial motility, *Cell* 136 (2009) 163–174.
- [208] A. Weihofen, K.J. Thomas, B.L. Ostaszewski, M.R. Cookson, D.J. Selkoe, Pink1 forms a multiprotein complex with Miro and Milton, linking Pink1 function to mitochondrial trafficking, *Biochemistry* 48 (2009) 2045–2052.
- [209] X. Wang, D. Winter, G. Ashrafi, J. Schlehe, Y.L. Wong, D. Selkoe, S. Rice, J. Steen, M.J. LaVoie, T.L. Schwarz, PINK1 and Parkin target Miro for phosphorylation and degradation to arrest mitochondrial motility, *Cell* 147 (2011) 893–906.
- [210] E. Shlevkov, T. Kramer, J. Schapansky, M.J. LaVoie, T.L. Schwarz, Miro phosphorylation sites regulate Parkin recruitment and mitochondrial motility, *Proc. Natl. Acad. Sci. U. S. A.* 113 (2016) E6097–E6106.
- [211] F. Zhang, W. Wang, S.L. Siedlak, Y. Liu, J. Liu, K. Jiang, G. Perry, X. Zhu, X. Wang, Miro1 deficiency in amyotrophic lateral sclerosis, *Front. Aging Neurosci.* 7 (2015) 100.
- [212] C. Colombrita, E. Onesto, F. Megiorni, A. Pizzuti, F.E. Baralle, E. Buratti, V. Silani, A. Ratti, TDP-43 and FUS RNA-binding proteins bind distinct sets of cytoplasmic messenger RNAs and differently regulate their post-transcriptional fate in motoneuron-like cells, *J. Biol. Chem.* 287 (2012) 15635–15647.
- [213] S. Xiao, T. Sanelli, S. Dib, D. Sheps, J. Findlater, J. Bilbao, J. Keith, L. Zinman, E. Rogaeva, J. Robertson, RNA targets of TDP-43 identified by UV-CLIP are deregulated in ALS, *Mol. Cell. Neurosci.* 47 (2011) 167–180.
- [214] F. Zhang, A.L. Strom, K. Fukada, S. Lee, L.J. Hayward, H. Zhu, Interaction between familial amyotrophic lateral sclerosis (ALS)-linked SOD1 mutants and the dynein complex, *J. Biol. Chem.* 282 (2007) 16691–16699.
- [215] P. Shi, A.-L. Ström, J. Gal, H. Zhu, Effects of ALS-related SOD1 mutants on dynein- and KIF5-mediated retrograde and anterograde axonal transport, *Biochim. Biophys. Acta (BBA) – Mol. Basis Dis.* 1802 (2010) 707–716.
- [216] M. Tateno, S. Kato, T. Sakurai, N. Nukina, R. Takahashi, T. Araki, Mutant SOD1 impairs axonal transport of choline acetyltransferase and acetylcholine release by sequestering KAP3, *Hum. Mol. Genet.* 18 (2009) 942–955.
- [217] E.B. Moloney, F. de Winter, J. Verhaagen, ALS as a distal axonopathy: molecular mechanisms affecting neuromuscular junction stability in the presymptomatic stages of the disease, *Front. Neurosci.* 8 (2014).
- [218] S. Sasaki, M. Iwata, Ultrastructural study of synapses in the anterior horn neurons of patients with amyotrophic lateral sclerosis, *Neurosci. Lett.* 204 (1996) 53–56.
- [219] Y. Deng, Z.F. Xu, W. Liu, B. Xu, H.B. Yang, Y.G. Wei, Riluzole-triggered GSH synthesis via activation of glutamate transporters to antagonize methylmercury-induced oxidative stress in rat cerebral cortex, *Oxid. Med. Cell. Longev.* 2012 (2012) 534705.
- [220] J.Y. Koh, D.K. Kim, J.Y. Hwang, Y.H. Kim, J.H. Seo, Antioxidative and proapoptotic effects of riluzole on cultured cortical neurons, *J. Neurochem.* 72 (1999) 716–723.
- [221] A. Siniscalchi, A. Bonci, N.B. Mercuri, G. Bernardi, Effects of riluzole on rat cortical neurones: an *in vitro* electrophysiological study, *Br. J. Pharmacol.* 120 (1997) 225–230.
- [222] A. Stevenson, D.M. Yates, C. Manser, K.J. De Vos, A. Vagnoni, P.N. Leigh, D.M. McLoughlin, C.C.J. Miller, Riluzole protects against glutamate-induced slowing of neurofilament axonal transport, *Neurosci. Lett.* 454 (2009) 161–164.
- [223] N.A. Goyal, T. Mozaffar, Experimental trials in amyotrophic lateral sclerosis: a review of recently completed, ongoing and planned trials using existing and novel drugs, *Expert Opin. Investig. Drugs* 23 (2014) 1541–1551.
- [224] T. Bordet, B. Buisson, M. Michaud, C. Drouot, P. Galéa, P. Delaage, N.P. Akentieva, A.S. Evers, D.F. Covey, M.A. Ostuni, J.J. Lacapère, C. Massaad, M. Schumacher, E.M. Steidl, D. Maux, M. Delage, C.E. Henderson, R.M. Pruss, Identification and characterization of cholest-4-en-3-one oxime (TRO19622), a novel drug candidate for amyotrophic lateral sclerosis, *J. Pharmacol. Exp. Ther.* 322 (2007) 709–720.
- [225] M.E. Cudkowicz, L.H. van den Berg, J.M. Shefner, H. Mitsumoto, J.S. Mora, A. Ludolph, O. Hardiman, M.E. Bozik, E.W. Ingessoll, D. Archibald, A.L. Meyers, Y. Dong, W.R. Farwell, D.A. Kerr, Dexampramipexole versus placebo for patients with amyotrophic lateral sclerosis (EMPOWER): a randomised, double-blind, phase 3 trial, *Lancet Neurol.* 12 (2013) 1059–1067.
- [226] L.J. Martin, Olesoxime, a cholesterol-like neuroprotectant for the potential treatment of amyotrophic lateral sclerosis, *IDrugs* 13 (2010) 568–580.
- [227] D.M. Pastula, D.H. Moore, R.S. Bedlack, Creatine for amyotrophic lateral sclerosis/motor neuron disease, *Cochrane Database Syst. Rev.* 12 (2012) CD005225.
- [228] J.M. Shefner, M.E. Cudkowicz, D. Schoenfeld, T. Conrad, J. Taft, M. Chilton, L. Urbinelli, M. Qureshi, H. Zhang, A. Pestronk, J. Caress, P. Donofrio, E. Sorenson, W. Bradley, C. Lomen-Hoerth, E. Pioro, K. Rezania, M. Ross, R. Pascuzzi, T. Heiman-Patterson, R. Tandan, H. Mitsumoto, J. Rothstein, T. Smith-Palmer, D. MacDonald, D. Burke, A clinical trial of creatine in ALS, *Neurology* 63 (2004) 1656–1661.
- [229] C. Sunyach, M. Michaud, T. Arnoux, N. Bernard-Marissal, J. Aebsicher, V. Latyszenok, C. Gouarne, C. Raoul, R.M. Pruss, T. Bordet, B. Pettmann, Olesoxime delays muscle denervation astrogliosis, microglial activation and motoneuron death in an ALS mouse model, *Neuropharmacology* 62 (2012) 2346–2352.
- [230] P. Kaufmann, J.L.P. Thompson, G. Levy, R. Buchsbaum, J. Shefner, L.S. Krivickas, J. Katz, Y. Rollins, R.J. Barohn, C.E. Jackson, E. Tiryaki, C. Lomen-Hoerth, C. Armon, R. Tandan, S.A. Rudnicki, K. Rezania, R. Sufit, A. Pestronk, S.P. Novella, T. Heiman-Patterson, E.J. Kasarskis, E.P. Pioro, J. Montes, R. Arbing, D. Vecchio, A. Barsdorf, H. Mitsumoto, B. Levin, Phase II trial of CoQ10 for ALS finds insufficient evidence to justify phase III, *Ann. Neurol.* 66 (2009) 235–244.
- [231] T. Lenglet, L. Lacomblez, J.L. Abitbol, A. Ludolph, J.S. Mora, W. Robberecht, P.J. Shaw, R.M. Pruss, V. Cuvier, V. Meininger, A phase II–III trial of olesoxime in subjects with amyotrophic lateral sclerosis, *Eur. J. Neurol.* 21 (2014) 529–536.
- [232] H. Yoshino, A. Kimura, Investigation of the therapeutic effects of edaravone a free radical scavenger, on amyotrophic lateral sclerosis (Phase II study), *Amyotroph. Lateral Scler.* 7 (2006) 241–245.
- [233] K. Ikeda, Y. Iwasaki, Edaravone, a free radical scavenger, delayed symptomatic and pathological progression of motor neuron disease in the wobbler mouse, *PLoS One* 10 (2015) e0140316.
- [234] P.H. Gordon, D.H. Moore, R.G. Miller, J.M. Florence, J.L. Verheijde, C. Doorish, J.F. Hilton, G.M. Spitalny, R.B. MacArthur, H. Mitsumoto, H.E. Neville, K. Boylan, T. Mozaffar, J.M. Belsh, J. Ravits, R.S. Bedlack, M.C. Graves, L.F. McCluskey, R.J. Barohn, R. Tandan, Efficacy of minocycline in patients with amyotrophic lateral sclerosis: a phase III randomised trial, *Lancet Neurol.* 6 (2007) 1045–1053.
- [235] M. Benatar, Lost in translation: treatment trials in the SOD1 mouse and in human ALS, *Neurobiol. Dis.* 26 (2007) 1–13.
- [236] B.J. Ryan, S. Hoek, E.A. Fon, R. Wade-Martins, Mitochondrial dysfunction and mitophagy in Parkinson's: from familial to sporadic disease, *Trends Biochem. Sci.* 40 (2015) 200–210.
- [237] D.D. Martin, S. Ladha, D.E. Ehrnhofer, M.R. Hayden, Autophagy in Huntington disease and huntingtin in autophagy, *Trends Neurosci.* 38 (2015) 26–35.
- [238] Q. Cai, P. Tammineni, Alterations in mitochondrial quality control in Alzheimer's disease, *Front. Cell. Neurosci.* 10 (2016).