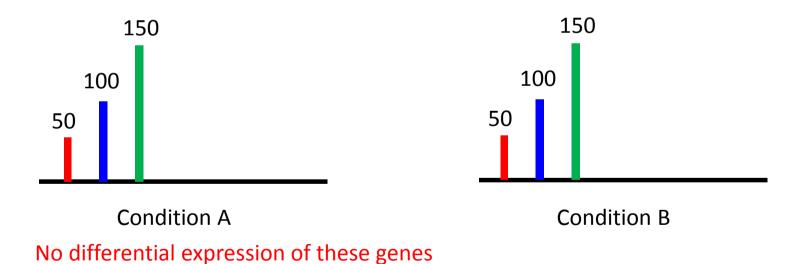
Thought experiment (I)

Suppose

- Two RNA populations (samples): A and B
- The same three genes expressed in both samples
- Numbers indicate number of transcripts / cell

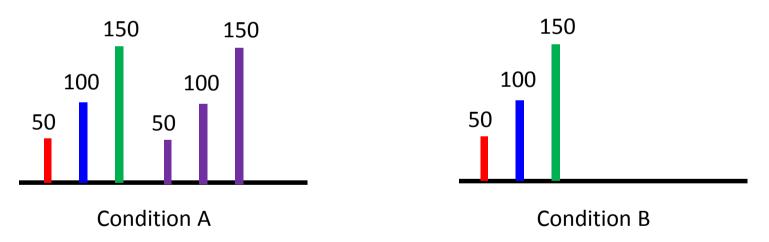




Thought experiment (II)

Suppose

- Two RNA populations (samples): A and B
- The same 3 genes expressed in both samples
- Numbers indicate number of transcripts / cell
- Now condition A has 3 additional genes not in B with equal number and expression

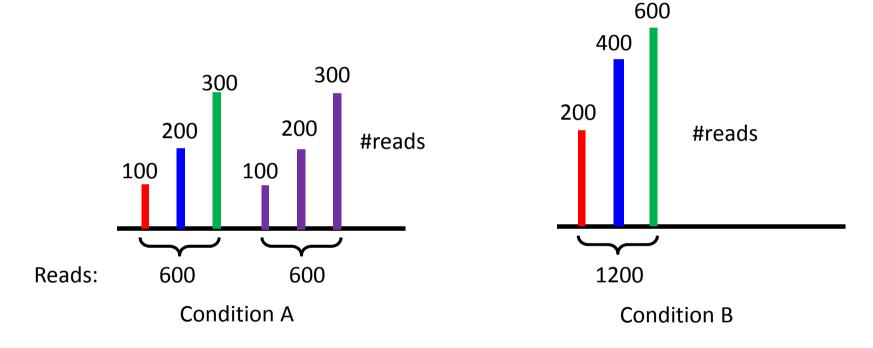


Still no differential expression of first three genes. However, RNA production in A is twice the one in B.



Thought experiment (III)

Suppose we sequence both samples with the same depth (1200 reads)
These reads get 'distributed' over the expressed genes



- (1) Correct normalization factor would adjust condition A by a factor of two
- (2) Proportion of reads attributed to a gene in one library depends on expression properties of whole sample → If a sample has larger RNA output , RNA-seq will undersample many genes



RPKM would fail in this example

$$RPKM = \frac{\text{# mapped reads} \times 10^6}{\text{total number of mapped reads}}$$

(assuming transcript lengths are the same)

In this example:

Condition A, first (red) gene:
$$RPKM = \frac{100 \times 10^6}{1200} = 83333$$

Condition B, first (red) gene:
$$RPKM = \frac{200 \times 10^6}{1200} = 166666$$

RPKM normalization would result in differential expression: we did not take total RNA production into account



When does RKPM fail?

- If samples have largely different RNA production
 - Many unique genes and/or highly expressed genes
 - If many genes in one sample have a very high expression compared to the other samples
- If RNA sample is contaminated
 - Reads that represent the contamination will take away reads from the true sample, thus dropping the number of reads of interest.
- If you can assume that your samples are 'comparable' then RKPM is OK
 - e.g., technical replicates

