**Sean Maden**

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# Summary

* Delivered illuminating insights about complex biomedical data; executed complex data processing and analyses.
* Led productive research at acclaimed institutions; published 14 peer-reviewed manuscripts (including 7 as first author).
* Developed open-access databases, workflows, software packages, and dashboards for high-dimensional genomics.
* Delivered polished posters and presentations using excellent initiative, project management, and communication.

# Education

**Ph.D., Computational Biology, Oregon Health & Science University, Portland, OR June 2018 to June 2022**

Graduate courses: biostatistics, epidemiology, machine learning, and bioinformatics; conference funding award; 5 peer-reviewed journal publications (3 first-author); 2 software packages (R, Python); 1 database; dissertation: “Applications of public omics data”

**B.A., Biology, Reed College, Portland, OR September 2007 to May 2011**

Rigorous liberal arts and sciences; fellowship completed; earned research funding; thesis topic: microsatellite genetic diversity

**High School, Full-I.B. Diploma, Biology, Edmonds-Woodway HS, Edmonds, WA September 2003 to May 2007**

Completed Full-IB curriculum, final exam, Senior project; completed AP and IB coursework; graduated top 5% of class

**Experience**

**Postdoctoral Research Fellow, Johns Hopkins School of Public Health, Baltimore, MD June 2022 to June 2024**

• Delivered manuscripts and software for critical arm of multifaceted deconvolution research initiative.

* Leveraged multiomics data integration to derive novel insights about cell size divergence biases.
* Completed high-throughput, high-dimensional processing and analysis using HPC SGE environments.

**Data Analyst Assistant, Fred Hutch, Seattle, WA October 2015 to June 2018**

* Determined cancer risk from molecular subtyping, biopsy pathologist reports, and AUC/ROC critical assessment.
* Completed high-throughput data processing and high-dimensional analyses using HPC Slurm environments.

**Bioinformatics Intern, Fred Hutch, Seattle, WA April to October 2015**

* Lead statistical insights from analysis of colorectal cancer epigenetic profiles and EHR data.
* Quantified biomarker associations from colorectal cancer microarray data processed using a custom ETL pipeline.

**Laboratory Assistant, University of Washington Medical Genetics, Seattle, WA January to October 2013**

• Determined Paraoxonase (PON)-1 enzyme activity from population modeling and controlling genetic background.

* Completed novel genome-wide association studies (GWAS), PLINK variant array processing, and Manhattan plots.

**Awards**

* **MOMACS Grad Student Travel Stipend (May 2019)** Modeling the World’s Systems conference, Washington, D.C.
* **SAS-BWF Fellow (May 2016-2017)** Collaborative colorectal cancer biomarker research, Fred Hutch, Seattle, WA
* **Undergraduate Research Grant (January to March 2010)** Genetics research for thesis, Reed College, Portland, OR
* **Fischer Memorial Fellow (May to September 2010)** Amphibian population research, Reed College, Portland, OR

**Programming**

* **lute (2023, coauthor: Stephanie Hicks, language: R/Bioconductor)** Data transformation & experiment framework.
* **cellScaleFactors (2023, language: R/Bioconductor)** Mined/scraped cell size bias-correction factors & metadata.
* **intronomer (2022, coauthor: Julianne David, language: Python)** Long-read retained intron detection and analysis.
* **recountmethylation (2021, language: database, R/Bioconductor)** High-throughput QC and analysis of public data.
* **recountmethylation (2021, language: Python, SQL, HDF5)** Epigenetics microarray data server and database.
* **cgageR (2018, language: R/Bioconductor)** Ensemble age prediction from multiple machine learning models.
* **cgmappeR (2018, language: R/shiny)** Query and visualize genome ideograms, sequences, and sample data.

# Skills

**Statistics:** Machine learning, decomposition, cluster analysis, transformation, bias correction, ROC/AUC

**Data analytics:** Benchmarks, workflows, normalization, automation, high-throughput, feature engineering, dashboards

**Data engineering:** Relational (SQL), Document (MongoDB), Graph (PostgreSQL), Sparse (HDF5), ETL, JSON, XML

**Compute environments:** High-performance compute (Slurm, SGE), remote servers, AWS, Azure, Windows, OSX, Linux, POSIX

**Software development:** Version control (Git/GitHub), data servers (Synapse, NCBI), APIs, Jupyter, Markdown, open-access

**Programming languages:** R/Bioconductor, Python, Bash/Shell, HTML, CSS, JavaScript, NextFlow, Snakemake

# Peer-reviewed preprints and publications (\*first author;)

1. **Maden SK\***, Huuki-Myers LA, Kwon SH, Collado-Torres L, Maynard KR, Hicks SC. “lute: estimating the cell composition of heterogeneous tissue with varying cell sizes using gene expression.” bioRxiv. 2024.
2. Huuki-Myers LA\*, Montgomery KD\*, Kwon SH, Cinquemani S, **Maden SK**, Eagles NJ, Kleinman JE, Hyde TM, Hicks SC, Maynard KR, Collado-Torres L. “Benchmark of cellular deconvolution methods using a multi-assay reference dataset from postmortem human prefrontal cortex.” bioRxiv. 2024.
3. **Maden SK\***, Kwon SH, Huuki-Myers LA, Collado-Torres L, Hicks SC, Maynard KR. “Challenges and opportunities to computationally deconvolve heterogeneous tissue with varying cell sizes using singlecell RNA-sequencing datasets.” Genome Biol 24, 288. 2023.
4. **Maden SK\*** , Walsh B, Ellrott K, Hansen KD, Thompson RF, Nellore A. “Recountmethylation enables flexible analysis of public blood DNA methylation array data.” Bioinformatics Advances 3, 1. 2023.
5. David JK\*, **Maden SK\***, Wood MA, Thompson RF, Nellore A. “Retained introns in long RNA-seq reads are not reliably detected in sample-matched short reads.” Genome Biology 23, 240. 2022.
6. **Maden SK\***, Thompson RF, Hansen KD, Nellore A. “Human methylome variation across Infinium 450K data on the Gene Expression Omnibus.” NAR Genomics and Bioinformatics 3, lqab025. 2021.
7. David JK\*, **Maden SK**, Weeder BR, Thompson RF, Nellore A. “Putatively cancer-specific exon–exon junctions are shared across patients and present in developmental and other non-cancer cells.” NAR Cancer 2, zcaa001. 2020.
8. Nguyen A\*, David JK, **Maden SK**, Wood MA, Weeder BR, Nellore A, Thompson RF. “Human Leukocyte Antigen Susceptibility Map for Severe Acute Respiratory Syndrome Coronavirus 2.” Journal of Virology 94, e00510–20. 2020.
9. Wang T\*, **Maden SK**, Luebeck GE, Christopher LI, Newcomb PA, Ulrich CM, Joo JE, Buchanan DD, Milne RL, Southey MC, Carter KT, Willbanks AR, Luo Y, Yu M, Grady WM. “Dysfunctional epigenetic aging of the normal colon and colorectal cancer risk.” Clinical Epigenetics 12, 5. 2020.
10. Luebeck GE\*, Hazelton WD, Curtius K, **Maden SK**, Yu M, Carter KT, Burke W, Lampe PD, Li CI, Ulrich CM, Newcomb PA, Westerhoff M, Kaz AM, Luo Y, Inadomi JM, Grady WM. “Implications of Epigenetic Drift in Colorectal Neoplasia.” Cancer Research 79, 495–504. 2019.
11. Yu M\*, **Maden SK\***, Stachler MS\*, Ayers AMJ, Guo Y, Carter KT, Willbanks A, Heinzerling TJ, O’Leary RM, Xu S, Bass A, Chandar AK, Chak A, Elliott R, Willis JE, Markowitz SD, Grady WM. “Subtypes of Barrett’s oesophagus and oesophageal adenocarcinoma based on genome-wide methylation analysis.” Gut 68, 389–399. 2019.
12. Smith J\*, **Maden SK\***, Lee D\*, Buie R, Peddu V, Shean R, Busby B. “Consensus Machine Learning for Gene Target Selection in Pediatric AML Risk.” bioRxiv. 2019.
13. Luebeck GE\*, Curtius K, Hazelton WD, **Maden S**, Yu M, Thota PN, Patil DT, Chak A, Willis JE, Grady WM. “Identification of a key role of widespread epigenetic drift in Barrett’s esophagus and esophageal adenocarcinoma.” Clinical Epigenetics 9, 113. 2017.
14. Kim DS\*, **Maden SK**, Burt AA, Ranchalis JE, Furlong CE, Jarvik GP. “Dietary fatty acid intake is associated with paraoxonase 1 activity in a cohort-based analysis of 1,548 subjects.” Lipids in Health and Disease 12, 183. 2013.