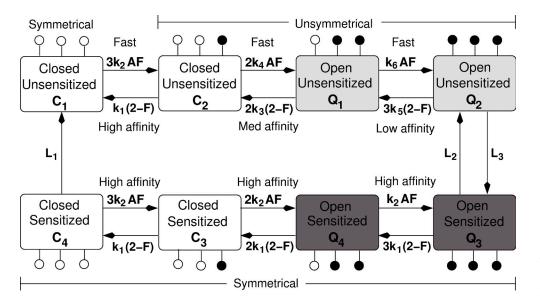


Figure S1. Effects of A438079, a P2X7R-specific antagonist, on agonist-induced receptor activation and deactivation. (A and B) Concentration-dependent effects of A438079 on ATP- (A) and BzATP (B)-induced current response. Note that the current scales are variable. At concentrations indicated above traces, mixture solutions of A438079 and agonist were applied. (C) Dose-dependent effect A438079 on sustained BzATP-induced current.



**S2.** Markov state Figure model adopted from the scheme in Yan et al. (2010) describing the binding and unbinding of agonist (ATP or BzATP) to the P2X7R. C<sub>1</sub>-C<sub>4</sub> are closed states, whereas Q<sub>1</sub>-Q<sub>4</sub> are open states, where Q1 and Q2 possess the same conductance  $g_{12}$ , whereas  $Q_3$ and Q4 possess the conductance  $g_{34}$  ( $g_{12} < g_{34}$ ). The open circles on each state represent unoccupied binding sites, whereas closed circles represent occupied binding sites. C1, C2, Q1, and Q2 are the unsensitized states. whereas Q<sub>3</sub>, Q<sub>4</sub>, C<sub>3</sub>, and C<sub>4</sub> are the sensitized states. Negative cooperativity for agonist binding was assumed to oc-

cur only in the top row (unsensitized states). In other words, binding affinity decreases at each step in the top row (making agonist binding asymmetrical); i.e.,  $3k_2/k_1 > k_4/k_3 > k_6/3k_5$ . Receptor sensitization was assumed to restore both symmetry and backward/forward rates to those belonging to the naive state  $C_1$  (i.e., to  $k_1$  and  $k_2$ ). In other words, negative cooperativity is lost in the bottom row ( $L_4$ , i = 1-3, are the transition rates between unsensitized and sensitized states). The new feature added to this scheme is the inclusion of the allosteric binding of extracellular  $Ca^{2+}$  to the receptor via the two fractions (Hill functions) 2-F, affecting the set of backward rates ( $k_1$ ,  $k_3$ , and  $k_5$ ), and F, affecting the set of forward rates ( $k_2$ ,  $k_4$ , and  $k_6$ ), both of which depend on the concentration of extracellular divalent cations [DC]<sub>e</sub>, including  $Ca^{2+}$ .

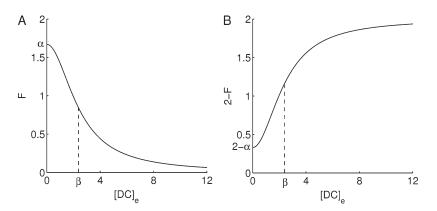


Figure S3. Fitted plots describing the effects of divalent cations. Graphs of the fractions  $F = \alpha \beta^2/(\beta^2 + [\mathrm{DC}]_e^2)$  (A) and  $2 - F(\mathrm{B})$ , the Hill functions describing the dependency of P2X7R allosteric regulation by the concentration of extracellular divalent cations  $[\mathrm{DC}]_e$  (specifically fitted to  $\mathrm{Ca}^{2+}$ ).  $\beta$  represents both the  $\mathrm{IC}_{50}$  for F and  $\mathrm{EC}_{50}$  for 2 - F, whereas  $\alpha$  satisfies  $F(0) = \alpha$ .

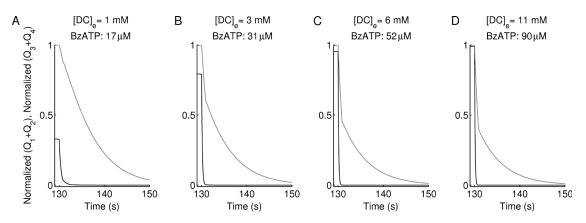


Figure S4. Deactivation phases of the open states  $Q_1 + Q_2$  and  $Q_3 + Q_4$ , normalized by their maximum values, when a single naive model cell is activated by the same free BzATP concentration. This was achieved by simultaneously varying the concentrations of extracellular  $Ca^{2+}$  and BzATP, specified by the values on top of each panel. B–D show that at higher [DC]<sub>e</sub>, the slow component of the deactivation phase of the open state  $Q_3 + Q_4$  gradually recedes in favor of the open state  $Q_1 + Q_2$  when compared with A.

Table S1

Parameter values and distributions used in modeling of P2X7R gating according to the scheme in Fig. S1 and Eqs. 1–9

| Symbol -   | Parameter values and distributions |  |
|--|------------------------------------|--|
|  | Values                             | Distribution                           |
| $\overline{k_1}$                                       | $0.3 \ s^{-1}$                     | Normal, $\sigma = 0.003$               |
| $k_2$  | $40,000~{\rm M.s^{-1}}$            | Normal, $\sigma = 400$                 |
| $k_3$  | $2.4 \ {\rm s^{-1}}$               | Normal, $\sigma = 0.024$               |
| $k_4$  | $50,000 \; \mathrm{M.s^{-1}}$      | Normal, $\sigma = 500$                 |
| $k_5$  | $1.58 \ s^{-1}$                    | Normal, $\sigma = 0.0158$              |
| $k_6$  | $7,000~{\rm M.s^{-1}}$             | Normal, $\sigma = 70$                  |
| $L_1$  | $0.0001~{\rm s}^{-1}$              | NA                                     |
| $L_2$  | $0.004~{\rm s}^{-1}$               | Normal, $\sigma = 4 \times 10^{-5}$    |
| $L_3$  | $0.5 \ s^{-1},  0.1 \ s^{-1a}$     | Normal, $\sigma = 0.005$               |
| α  | 1.67 (unitless)                    | Normal, $\sigma = 0.0167$              |
| β  | $2.4\times10^{-3}~M$               | Uniform, [2-3] $\times 10^{-3}$        |
| $g_{12}$ (Q <sub>1</sub> + Q <sub>2</sub> conductance) | $1.5\times10^{-8}~\mathrm{S}$      | Normal, $\sigma = 1.5 \times 10^{-10}$ |
| $g_{34}$ (Q <sub>3</sub> + Q <sub>4</sub> conductance) | $4.5\times10^{-8}~\mathrm{S}$      | Normal, $\sigma = 4.5 \times 10^{-10}$ |
| V (holding potential)                                  | $60\times 10^{-3}V$                | NA                                     |
| E (reversal potential)                                 | 0 V                                | NA                                     |

NA, not applicable.

## REFERENCES

Yan, Z., A. Khadra, S. Li, M. Tomic, A. Sherman, and S.S. Stojilkovic. 2010. Experimental characterization and mathematical modeling of P2X7 receptor channel gating. *J. Neurosci.* 30:14213–14224. http://dx.doi.org/10.1523/JNEUROSCI.2390-10.2010

<sup>&</sup>lt;sup>a</sup>This value was used only once to generate the second panels (counting from left) in Fig. 6 (A and B; see Yan et al. [2010] for more details).