# Population and ODE-based models using Stan and Torsten

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### Outline

- 1. Course information
- 2. Introduction and modeling framework | Charles Margossian
- 3. Models in pharmacometrics | Charles Margossian
- 4. ODEs in Stan and Torsten | Charles Margossian
- 5. Numerical ODE integrators | Yi Zhang
- 6. Population models | Charles Margossian
- 7. ODE group integrators | Yi Zhang
- 8. PMX population solvers | Yi Zhang
- 9. Additional nonlinear ODE example | Charles Margossian

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### Instructors

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  - Columbia University, Department of Statistics
- Yi Zhang
  - ► Metrum Research Group

#### TA

- Steve Bronder
  - ► Capital One

## Outline

## Day 1

- Introduction and modeling framework
- Pharmacometrics models
- Ordinary differential equation(ODE) based models
- Numerical ODE integrators

## Day 2

- Population models
- Group/Population ODE integrators and MPI parallelisation
- Group/Population solvers and MPI parallelisation

# Logistics

METWORX  $^{\text{TM}},$  cloud-based modeling & simulation platform by Metrum Research Group.









## Logistics

### Workshop package

- R scripts and Stan files to do the exercises
- ▶ These slides
- Outline of the course
- pAdditional documentation

### We will be using:

- ▶ Torsten v0.87
- ► RStan v2.19.2
- ggplot, plyr, tidyr, dplyr

## Outline

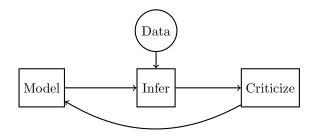
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## Preliminary question

- ▶ Why Bayesian in a field such as pharmacometrics?
- Example Bayesian aggregation of average data: an application in drug development [Weber et al., 2018]

# Modeling framework

## Box's loop



### Inference

- find the set of parameters consistent with our model and our data
- approximate this set with draws from the posterior distribution

# Sampling algorithm

- ▶ Use the NUTS to sample  $\pi(\theta|y)$
- ▶ Requires users the specify  $\log \pi(\theta, y) = \log \pi(y|\theta) + \log \pi(\theta)$

# The "criticism" step

This step can be broken up in two parts:

- 1. did we sample from the correct distribution?
- 2. does our model capture the characteristics of the data we care about?

## Diagnosing the inference algorithm

- look at the trace and the density plots
- ▶ look at  $\hat{R}$  and effective number of samples
- have any warning messages been issued, i.e. divergent transitions?

# Example: fitting a linear model

Likelihood:

$$Y \sim \text{Normal}(x\beta, \sigma^2)$$

Prior:

$$\beta \sim \text{Normal}(2,1)$$

$$\sigma^2 \sim \! \mathrm{Normal}(1,1)$$

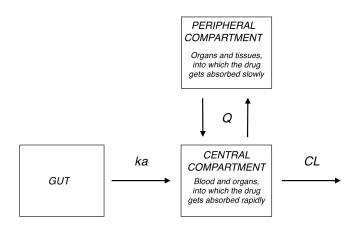
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# What is the effect of a treatment on a patient?

- pharmacokinetics (PK): how is the drug absorbed in the body?
- pharmacodynamics (PD): once it is absorbed, what are its effects?

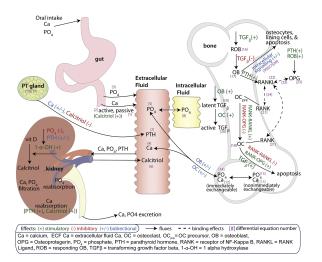
## Example: Two compartment model



# Two compartment model

$$y'_{
m gut} = -k_a y_{
m gut}$$
 $y'_{
m cent} = k_a y_{
m gut} - \left(\frac{CL}{V_{
m cent}} + \frac{Q}{V_{
m cent}}\right) y_{
m cent} + \frac{Q}{V_{
m peri}} y_{
m peri}$ 
 $y'_{
m peri} = \frac{Q}{V_{
m cent}} y_{
m cent} - \frac{Q}{V_{
m peri}} y_{
m peri}$ 

# Example 2: Bone mineral density model from [Peterson and Riggs, 2012]



## Two compartment model

Denote  $\theta = \{CL, Q, VC, VP, K_a\}$ , the ODE coefficients. Then  $y' = f(y, t, \theta)$ 

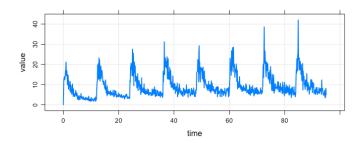
Given an initial condition  $y_0 = y(t_0)$ , solving the above ODE gives us the  $\{natural\ evolution\}$  of the system at any given time point.

### The event schedule

#### An event can be:

- Sate changer: an (exterior) intervention that alters the state of the system; for example a bolus dosing or the beginning of an infusion.
- Observation: a measurement of a quantity of interest at a certain time.

# Drug concentration in a patient's blood



### The event schedule

- ▶ There is no general theory for the event schedule :(
- ► The modeling software NONMEM®proposes a convention for pharmacometrics, which we adopt in Torsten.

### Torsten functions

Torsten functions offers additional built-in functions to simulate data from a compartment model.



Each Torsten function requires users to specify:

- ▶ a system of ODEs and a method to solve it.
- An event schedule.

### Torsten functions

- Analytically solutions for the one/two cpt models.
- Event schedule
- ▶ ODE coefficients, e.g.  $\theta = \{CL, Q, VC, VP, ka\}$  for two-cpt model.
- bio-availibility fraction and lag times.

## Example

### Clinical trial

- ► Single patient
- Bolus doses with 1200 mg, administered every 12 hours, for a total of 15 doses.
- Many observations for the first, second, and last doses.
- Additional observation every 12 hours.

Note: the observation are plasma drug concentration measurement.

See data/twoCpt.data.r.

## Example

### Model

- two compartment model with first-order absorption
- prior information based on clinical trial conducted on a large population
- normal error for the plasma drug concentration measurement.

## Example

### Prior

```
CL ~ lognormal(log(10), 0.25);

Q ~ lognormal(log(15), 0.5);

VC ~ lognormal(log(35), 0.25);

VP ~ lognormal(log(105), 0.5);

ka ~ lognormal(log(2.5), 1);

sigma ~ cauchy(0, 1);
```

### Likelihood

$$\log(cObs) \sim \operatorname{Normal}\left(\log\left(\frac{y_2}{VC}\right), \sigma^2\right)$$

Exercise 1: write and fit this model, using twoCptModel.r and model/twoCptModel.stan. Exercise 2: Write a generated quantities block and do posterior predictive checks.

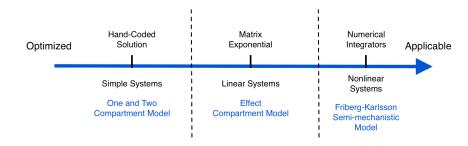
### Resources

- ➤ Torsten repository: https://github.com/metrumresearchgroup/Torsten
- ► Torsten User manual (on GitHub and in the workshop folder).

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## Arsenal of tools



For some examples, see [Margossian and Gillespie, 2017a].

- the "optimized applicable" spectrum is a heuristic; counter-examples can be built.
- coding effort may also be a criterion



## Matrix exponential

Consider a system of linear ODEs:

$$y'(t) = Ky(t)$$

where K is a constant matrix.

Then

$$y(t) = e^{tK}y_0$$

# Matrix Exponential

$$e^{tK} = \sum_{n=0}^{\infty} \frac{(tK)^n}{n!} = I + tK + \frac{(tK)^2}{2} + \frac{(tK)^3}{3!} + \dots$$

## Matrix Exponential

For example, the two compartment model generates the following matrix:

$$K = \begin{bmatrix} -ka & 0 & 0 \\ ka & -(CL+Q)/Vc & Q/Vp \\ 0 & Q/V_c & -Q/V_p \end{bmatrix}$$

### Linear ODE solver in Torsten

### Numerical integrator

```
real[ , ] pmx_integrate_ode_rk45(ODE_RHS, real[] y0, real t0,
    real[] ts, real[] theta, real[] x_r, int[] x_i, real rtol =
    1.e-6, real atol = 1.e-6, int max_step = 1e6);
```

- ▶ ODE\_RHS: ODE right-hand-side f in  $y' = f(y, t, \theta, x_r, x_i)$ .
- ▶ y0: initial condition at time t0.
- ▶ t0: initial time.
- ts: times at which we require a solution.
- ▶ theta: parameters to be passed to f.
- x\_r: real data to be passed to f.
- x\_i: integer data to be passed to f.
- rtol, atol, and max\_step are optional control parameters for relative tolerance, absolute tolerance, and max number of time steps, respectively. Their default values have no theoretical justification.

# System function

#### Torsten function

Exercise 3: Write, fit, and diagnose the two compartment model using the pmx\_solve\_rk45 function.

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# Nonlinear ODEs without analytical solution

kinetics of an autocatalytic reaction [Robertson, 1966]

The structure of the reactions is

$$A \xrightarrow{k_1} B$$
,  $B + B \xrightarrow{k_2} C + B$ ,  $B + C \xrightarrow{k_3} C + A$ ,

where  $k_1$ ,  $k_2$ ,  $k_3$  are the rate constants and A, B and C are the chemical species involved. The corresponding ODEs are

$$x'_{1} = -k_{1}x_{1} + k_{3}x_{2}x_{3}$$

$$x'_{2} = k_{1}x_{1} - k_{2}x_{2}^{2} - k_{3}x_{2}x_{3}$$

$$x'_{3} = k_{2}x_{2}^{2}$$

Given  $k_1 = 0.04$ ,  $k_2 = 3.0e7$ ,  $k_3 = 1.0e4$ , we make inference regarding the initial condition for  $x_1(t = 0)$ .

## Nonlinear ODEs without analytical solution

$$x'_1 = -k_1x_1 + k_3x_2x_3$$

$$x'_2 = k_1x_1 - k_2x_2^2 - k_3x_2x_3$$

$$x'_3 = k_2x_2^2$$

Given  $k_1 = 0.04$ ,  $k_2 = 3.0e7$ ,  $k_3 = 1.0e4$ , we make inference regarding the initial condition for  $x_1(t = 0)$ .

#### Exercise

Write Stan function for the above ODE's RHS.

## Stan function for autocatalytic kinetics

```
x_2' = k_1 x_1 - k_2 x_2^2 - k_3 x_2 x_3
                     x_3' = k_2 x_2^2
functions{
  real[] reaction(real t, real[] x, real[] theta, real[] r,
   \hookrightarrow int[] i){
    real dxdt[3];
    real k1 = theta[1];
    real k2 = theta[2];
    real k3 = theta[3]:
    dxdt[1] = -k1*x[1] + k3*x[2]*x[3];
    dxdt[2] = k1*x[1] - k3*x[2]*x[3] - k2*(x[2])^2;
    dxdt[3] = k2*(x[2])^2;
    return dxdt;
```

 $x_1' = -k_1x_1 + k_3x_2x_3$ 

▶ What's the initial conditions for  $x_2$  and  $x_3$ ?

### Numerical integrators

- Runge-Kutta 4th/5th (rk45)
  - non-stiff equations
  - Most popular, try this if you don't know the nature of the ODE, or what you're doing, or both.
- Backward differentiation formula (bdf)
  - stiff equations
  - More expensive to use
- Adams-Moulton
  - non-stiff equations
  - higher-order of accuracy(do you really need it?)
  - scales better with number of steps

### Numerical integrators

Integrators	Stan	Torsten
rk45	integrate_ode_rk45	pmx_integrate_ode_rk45
BDF	integrate_ode_bdf	pmx_integrate_ode_bdf
Adams	integrate_ode_adams	<pre>pmx_integrate_ode_adams</pre>

- ▶ ODE\_RHS: ODE right-hand-side f in  $y' = f(y, t, \theta, x_r, x_i)$ .
- ▶ y0: initial condition at time t0.
- t0: initial time.
- ts: times at which we require a solution.
- theta: parameters to be passed to f.
- x\_r: real data to be passed to f.
- ▶ x\_i: integer data to be passed to f.



- ▶ In each of 8 experiments performed *x*3 is observed.
- ▶ Hierarchical model for x0[1]

```
model {
  y0_mu ~ lognormal(log(2.0), 0.5);
  for (i in 1:nsub) {
    y0_1[i] ~ lognormal(y0_mu, 0.5);
  }
  sigma ~ cauchy(0, 0.5);
  obs ~ lognormal(log(x3), sigma);
}
```

#### Data available for the inference

#### Given above data and model, write the rest of Stan code.

- ► Hint: use chem.stan as template, also see chem.data.R and chem.init.R.
- ▶ Reaction begins with A(on which is also what we'd like to make inference), the other two spiecies are non-existent at the beginning of the reaction.
- ▶ Which numerical integrator are you using? Why?

#### How to build & run?

#### Edit/Add cmdstan/make/local

#### Build in cmdstan

make path\_to\_workshop/RScript/model/chemical\_reactions/chem

#### Run

```
./chem sample adapt delta=0.95 random seed=1104508041 data \rightarrow file=chem.data.R init=chem.init.R
```

#### Outline

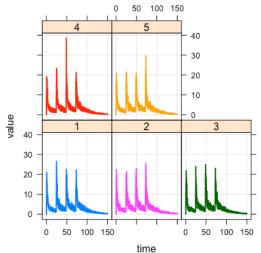
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## Data pooled into groups

- sport measurements are grouped by players
- people's voting intention can be grouped by states, social status, etc.
- medical measurements are grouped by patients

## Data pooled into groups

- medical measurements are grouped by patients
  - ► Simulated with mrgsolve https://mrgsolve.github.io/



#### Hierarchical model

With a hierarchical model, we can

- do partial pooling.
- estimate how similar the groups are to one another.
- estimate individual parameters.

$$\theta = (\theta_1, ..., \theta_L) \sim \textit{p}(\theta | \theta_{\text{pop}})$$

$$y = (y_1, ..., y_N) \sim p(y|\theta, x)$$

#### Hierarchical model

$$heta = ( heta_1,..., heta_L) \sim p( heta| heta_{ ext{pop}})$$
 $y = (y_1,...,y_N) \sim p(y| heta,x)$ 
 $heta_1 \qquad heta_2 \qquad heta_L$ 
 $heta_1 \qquad heta_2 \qquad heta_L$ 
 $heta_1 \qquad heta_2 \qquad heta_2 \qquad heta_L$ 

# Example 3: Hierarchical two compartment model

#### Likelihood function:

$$\log heta \sim ext{Normal}(\log heta_{ ext{pop}}, \Omega)$$

$$\Omega = \left( egin{array}{cccccc} \omega_1 & 0 & 0 & 0 & 0 & 0 \ 0 & \omega_2 & 0 & 0 & 0 & 0 \ 0 & 0 & \omega_3 & 0 & 0 & 0 \ 0 & 0 & 0 & \omega_4 & 0 \ 0 & 0 & 0 & 0 & \omega_5 \end{array} 
ight)$$

$$\log(cObs) \sim \text{Normal}\left(\log\left(\frac{y_2}{VC}\right), \sigma^2\right)$$

# Exercise 6: Write, fit, and diagnose a hierarchical two

compartment model for a population of 10 patients. Use data/twoCptPop.data.r and twoCptPop.r.}

- ▶ Start by running 3 chains with 30 iterations.
- ▶ Do you get any warning messages?

## Divergent transitions

- ► Do you get any warning messages?

  There were 29 divergent transitions after warmup.
- ▶ A divergent transition occurs when we fail to accurately compute a Hamiltonian trajectory.
- ▶ This is because we *approximate* trajectories.
- Our sampler may not be refined enough to explore the entire typical set.

# Divergent transitions

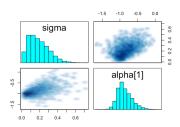
Consider the following hierarchical model:

$$\alpha_i \sim \text{Normal}(\mu, \sigma)$$
 $y_i \sim p(y|\alpha_i)$ 

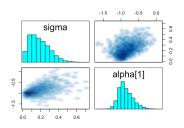
## Divergent transitons

$$\alpha_i \sim \text{Normal}(\mu, \sigma)$$

Fitting this model yields the following pairs plot:



## Divergent transitons



- ► This geometric shape is known as Neil's funnel [Neil, 2003].
- ▶ Its interactions with HMC is described in [Betancourt and Girolmi, 2015].
- ▶ It occurs in hierarchical models when we have sparse data and a centered prior.

#### Proposition

Reparameterize the model to avoid the funnel shape. We will do so by standardizing  $\alpha.$ 

$$\alpha_{\mathrm{std},i} := \frac{\alpha_i - \mu}{\sigma}$$

Then

$$\alpha_{\rm std} \sim {
m Normal}(0,1)$$

Then

$$\alpha_i = \mu + \sigma \alpha_{\mathrm{std},i}$$

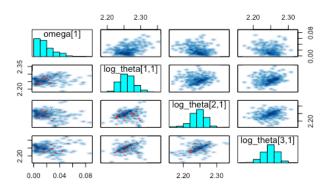
Hence

$$y_i \sim p(\mu + \sigma \alpha_{\mathrm{std},i})$$

Same data generating process; but how does this affect the geometry of the posterior?

Our model is a little more complicated than the above example:

- ▶ a lot of parameters (100 +)!
- multiple population parameters and hierarchical structures.
- ▶ these parameters follow a log normal distribution (so we need a pairs plot with  $\log \theta$ ).



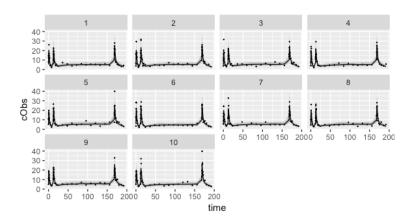
Exercise 6: Reparametrize the two compartment population model and fit it.}

- ▶ First, work out the appropriate parametrization. You should start with  $\log \theta_i \sim \text{Normal}(\theta_{\text{pop},i},\omega)$
- Write, fit, and check the inference (run 100 chains).
- What kind of predictive checks can we do?

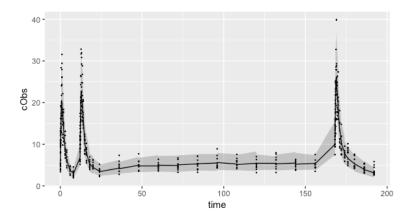
#### Need:

- predictions at an individual level
- predictions at a population level As always, this comes down to properly writing the data generating process in the generated quantities block.

# Individual predictions



# Population predictions



## Further reading

For a very good case study on hierarchical models, see, Bob Carpenter's *Pooling with Hierarchical Models for Repeated Binary Trials* 

https://mc-stan.org/users/documentation/case-studies/pool-binary-trials.html

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### ODE group integrators

#### Single ODE system

```
pmx_integrate_ode_rk45
pmx_integrate_ode_bdf
pmx_integrate_ode_adams
```

#### ODE group

```
pmx_integrate_ode_group_rk45
pmx_integrate_ode_group_bdf
pmx_integrate_ode_group_adams
```

#### Single ODE system

```
real[,]
pmx_integrate_ode_xxx(
          f,
          real[] y0, real t0,
          real[] ts,
          real[] theta,
          real[] x_r, int[] x_i,
          ...);
```

#### ODE group

```
matrix
pmx_integrate_ode_group_xxx(
    f,
    real[ , ] y0, real t0,
    int[] len, real[] ts,
    real[ , ] theta,
    real[ , ] x_r, int[ , ] x_i,
    ...);
```

## ODE group integrators

#### Single ODE system

#### ODE group

```
matrix
pmx_integrate_ode_group_xxx(
    f,
    real[ , ] y0, real t0,
    int[] len, real[] ts,
    real[ , ] theta,
    real[ , ] x_r, int[ , ] x_i,
    ...);
```

- len specifies the length of data for each subject within the above ragged arrays, and the size of len is the size of the population.
- ► The group integrators return a single matrix ragged column-wise. The number of rows equals to the size of ODE system.

## autocatalytic reaction model: ODE group version

- Change the loop with the numerical integrator to use group integrator.
- Remeber the return of the group integrator is a matrix
  - nb. of rows: nb. of states
  - ▶ nb. of cols: nb. of *total* results-extraction time points.

#### Build and run

Edit/Add cmdstan/make/local

Build in cmdstan

make ../example-models/chemical\_reactions/chem\_group

Run

mpiexec -n 2 -l ./chem\_group sample adapt delta=0.95 random  $\hookrightarrow$  seed=1104508041 data file=chem.data.R init=chem.init.R

- What does output say?
- How many cores can you use until performance saturates? Why?
- (optional)Can you do it using Stan's map\_rect? Is there a difference in style, output, and performance?

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# PMX population solvers

Single ODE system	ODE group
pmx_solve_rk45	pmx_solve_group_rk45
pmx_solve_bdf	pmx_solve_group_bdf
pmx_solve_adams	${\tt pmx\_solve\_group\_adams}$

## Individual solvers

## Population solvers

```
matrix
pmx_solve_group_bdf(f, int nCmt,
   int[] len, real[] time,
   real[] amt, real[] rate,
   real[] ii, int[] evid,
   int[] cmt, real[] addl,
   int[] ss, real[,] theta,
   real[,] biovar, real[,] tlag,
   real rel_tol, real abs_tol,
   int max_step);
```

## PMX population solvers

#### matrix

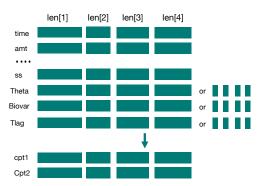
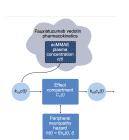


Figure: arguments and output of pmx\_solve\_group\_xxx

We analyze the time to the first grade 2+ peripheral neuropathy (PN) event in patients treated with an antibody-drug conjugate (ADC) delivering monomethyl auristatin E (MMAE). We will simulate and analyze data using a simplified version of the model reported in [Lu et al., 2017].

- ► Fauxlatuzumab vedotin 1.2 mg/kg IV boluses q3w × 6 does.
- ▶ 19 patients with 6 right-censored (simulated data).

#### Model scheme



#### Note

- To keep things simpler, we use the simulated individual CL and V values, and only model PD part of the problem.
- ► PN hazard is substantially delayed relative to PK exposure.
- Hazard increases over time to an extent not completely described by P



Likelihood for time to first PN  $\geq$  2 event in the  $i^{th}$  patient:

$$\begin{split} &L\left(\theta|t_{\text{PN},i}, \text{censor}_{i}, X_{i}\right) \\ &= \begin{cases} &h_{i}\left(t_{\text{PN},i}|\theta, X_{i}\right) e^{-\int_{0}^{t_{\text{PN},i}} h_{i}\left(u|\theta, X_{i}\right)du}, & \text{censor}_{i} = 0 \\ &e^{-\int_{0}^{t_{\text{PN},i}} h_{i}\left(u|\theta, X_{i}\right)du}, & \text{censor}_{i} = 1 \end{cases} \end{split}$$

where

$$t_{\text{PN}} \equiv \text{time to first PN} \geq 2 \text{ or right censoring event}$$
 $\theta \equiv \text{model parameters}$ 
 $X \equiv \text{independent variables} / \text{covariates}$ 
 $\text{censor} \equiv \left\{ \begin{array}{l} 1, & \text{PN} \geq 2 \text{ event is right censored} \\ 0, & \text{PN} \geq 2 \text{ event is observed} \end{array} \right.$ 

One can see the expression

$$e^{-\int_0^{t_{\text{PN},i}} h_i(u|\theta,X_i)du}$$

as the survival function at time t.



Hazard of PN grade 2+ based on the Weibull distribution, with drug effect proportional to effect site concentration of MMAE:

$$egin{aligned} h_j(t) &= eta E_{ ext{drug}j}(t)^eta t^{(eta-1)} \ E_{ ext{drug}j}(t) &= lpha c_{ej}(t) \ c'_{ej}(t) &= k_{e0} \left( c_j(t) - c_{ej}(t) 
ight). \end{aligned}$$

Overall ODE system including integration of the hazard function:

$$x_1' = -\frac{CL}{V}x_1 \tag{1}$$

$$x_2' = k_{e0} \left( \frac{x_1}{V} - x_2 \right) \tag{2}$$

$$x_3' = h(t) \tag{3}$$

where  $x_2(t) = c_e(t)$  and  $x_3(t) = \int_0^t h(u)du$  aka cumulative hazard.



## "just walk in a minute ago, literally" mode

Apply pmx\_solve\_group\_rk45 function

#### Intermediate mode

Code pmx\_solve\_group\_rk45 function and its args. Use input data file ttp2n.data2.R as hint.

#### hard mode

Code ODE, pmx\_solve\_group\_rk45 function and its args, and the likelihood for harzard and censor event. Use input data file ttp2n.data2.R and model block as hint.

"why bother" mode

## Edit/Add cmdstan/make/local

#### Build in cmdstan

```
make ../example-models/ttpn2/ttpn2_group
```

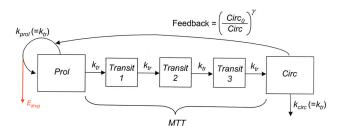
#### Run

- ▶ The parallel performance is not optimal, why?
- ► Can you do it using Stan's map\_rect?

## Outline

- 1. Course information
- 2. Introduction and modeling framework | Charles Margossian
- 3. Models in pharmacometrics | Charles Margossian
- 4. ODEs in Stan and Torsten | Charles Margossian
- 5. Numerical ODE integrators | Yi Zhang
- 6. Population models | Charles Margossian
- 7. ODE group integrators | Yi Zhang
- 8. PMX population solvers | Yi Zhang

# Friberg-Karlsson semi-mechanistic model [Friberg et al., 2002]



# ODE system of F-K model

$$y'_{\text{prol}} = k_{\text{tr}} y_{\text{prol}} (1 - E_{\text{drug}}) \left(\frac{\textit{Circ}_0}{\textit{y}_{\text{circ}}}\right)^{\gamma} - k_{\text{tr}} y_{\text{prol}}$$

$$y'_{\text{tr}1} = k_{\text{tr}} y_{\text{prol}} - k_{\text{tr}} y_{\text{tr}1}$$

$$y'_{\text{tr}2} = k_{\text{tr}} y_{\text{tr}1} - k_{\text{tr}} y_{\text{tr}2}$$

$$y'_{\text{tr}3} = k_{\text{tr}} y_{\text{tr}2} - k_{\text{tr}} y_{\text{tr}3}$$

$$y'_{\text{circ}} = k_{\text{tr}} y_{\text{tr}3} - k_{\text{tr}} y_{\text{circ}}$$

where  $E_{\rm drug} = \alpha \frac{y_{\rm cent}}{V_{\rm cent}}$ , ktr = 4/MTT, and  $\alpha \approx 3e - 4$ .

- $\triangleright$   $y_{\text{cent}}$  is obtained from a two compartment model.
- Our PK/PD model therefore has a total of 8 equations.
- This problem can be solved using pmx\_solve\_\*.

# Coupled PK-PD system

Alternatively, we may elect to solve the PK ODEs analytically and the PD ODEs numerically.

 This can yield some speedup, in particular for problems that require ODE solutions and sensitivities (e.g [Margossian and Gillespie, 2017b]).

# Coupled PK-PD system

- we now pass a "reduced system".
- we specify the number of ODEs to be solved numerically, not the number of compartments.
- theta now contains the parameters for the two cpt model, followed by the parameters that get passed to the numerical solver:

```
theta = {CL, Q, VC, VP, ka, /* ... */ };
```

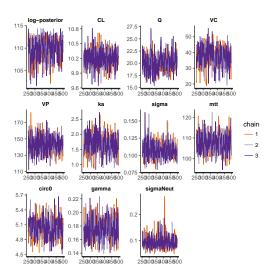
# Reduced system

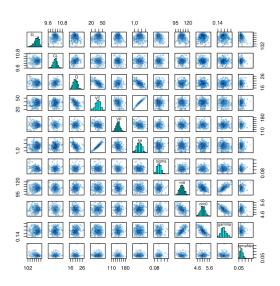
Exercise 4 (optional): Write, fit, and diagnose a Friberg-Karlsson model with a two compartment with first order absorption PK. Use FKModel.r and data/FKModel.data.r.

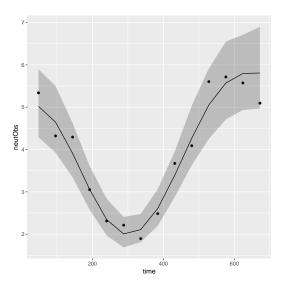
Write, fit, and diagnose a Friberg-Karlsson model with a two compartment with first order absorption PK. Use FKModel.r and data/FKModel.data.r.

- ➤ You may either use pmx\_solve\_\* or pmx\_solve\_twocpt\_\*.
- ▶ Use  $\alpha = 3e 4$  and estimate all other 8 ODE coefficients, i.e.  $\theta = \{\mathit{CL}, \mathit{Q}, \mathit{VC}, \mathit{VP}, \mathit{ka}, \mathit{MTT}, \mathit{circ0}, \gamma\}.$
- ▶ The initial state for the neutrophil count is *Circ*<sub>0</sub>. Either edit the event schedule to reflect this at time 0, or write the solution to your ODEs as a deviation from the baseline.

- ► This exercise entails a few subtleties; in the interest of time we won't go through it in class.
- ► Here are however results I get from 3 chains with 500 iterations you can use as a benchmark.







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