Learning Bayesian Networks from gene expression data

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Here we use R package deal1 to analyse yeast cell-cycle regulator genes and learn optimal Bayesian Network.

Install the packages and load and preprocess the data.

YeastCC data: This data package contains an ExpressionSet instance for the yeast cell cycle microarray experiment. The dataset contains gene expression measures (log-ratios, with Cy3-labeled common reference) for 6,178 yeast genes in 77 conditions.

```
# Install deal and load yeastCC data
# install.packages("deal")
library(deal)
#source("https://bioconductor.org/biocLite.R")
#biocLite("yeastCC")
library(yeastCC)

# Load the Yeast cell cycle regulator dataset
yeastCC = yeastCC
# First 3 genes are: [1] "YALOO1C" "YALOO2W" "YALOO3W"
# Last genes are: [1] "YPR202W" "YPR203W" "YPR204W"

# 77 conditions for each gene are measured in log-ratios:
# cln3.1 cln3.2 clb2.2 clb2.1 alpha0 alpha7 alpha14 alpha21 alpha28 alpha35
# YALOO1C 0.15 NA -0.22 0.07 -0.15 -0.21 0.17 -0.42 -0.44
# length(exprs(yeastCC[1]))
```

Turn data object yeastCC into an expression matrix (function exprs) with genes as columns and experiments as rows.

```
expr_df = as.data.frame(exprs(yeastCC))
expr_mt = t(as.matrix(expr_df))
expr_mt[1:5, 1:5]
         YALOO1C YALOO2W YALOO3W YALOO4W YALOO5C
## cln3.1
            0.15
                  -0.07
                           -1.22
                                    -0.09
                                            -0.60
## cln3.2
             NA
                   -0.76
                           -0.27
                                     1.20
                                            1.01
## clb2.2
          -0.22
                   -0.12
                           -0.10
                                            0.24
                                     0.16
## clb2.1
           0.07
                   -0.25
                            0.23
                                    -0.14
                                            0.65
## alpha0
                                    -0.02
                                            -0.05
           -0.15
                   -0.11
                           -0.14
```

Restrict the genes to cell cycle regulators.

The ORF names for the 800 cell cycle regulated genes are stored in orf800.

```
# Create a mask with yeast gene indeces that correspond to 800 cell regulator genes
mask = which(colnames(expr_mt) %in% orf800)
# Subset cell regulators
expr_cc_mt = expr_mt[ , mask]
expr_cc_mt[1:5, 1:5]
          YALO22C YALO40C YALO53W YALO67C YAROO3W
##
## cln3.1
            -0.22
                     3.43
                             0.34
                                    -0.15
                                             0.41
## cln3.2
            -0.86
                     2.75
                                    -1.25
                                              0.25
                             0.90
## clb2.2
           -0.25
                     0.36
                            -0.58
                                             -0.62
                                       NA
                     0.72
## clb2.1
            0.89
                           -0.92
                                      0.07
                                             -0.29
## alpha0
           -0.36
                     1.04
                             0.21
                                      0.01
                                             -0.30
```

Replace missing data (=NA) with each gene's median.

```
replace_na_with_col_medians = function(x) {
    x = replace(x, is.na(x), median(x, na.rm = T))
    return(x)
}

cat("Number of NA's before cleanup is:", length(which(is.na(expr_cc_mt) == TRUE)))

## Number of NA's before cleanup is: 2643

expr_cc_mt = apply(expr_cc_mt, 2, replace_na_with_col_medians)
    cat("Number of NA's after cleanup is:", length(which(is.na(expr_cc_mt) == TRUE)))

## Number of NA's after cleanup is: 0

## Check if correct w/ 1st and 2nd genes
# which(is.na(expr_cc_mt[, 2]))  # cdc28.140 idx61; cdc28.110 idx58
# median(expr_cc_mt[, 2], na.rm = T)  # -0.5 -0.045
```

Calculate the range between the 25th and 75th quantile for each gene and keep genes with a value > 1.6.

The interquartile range of an observation variable is the difference of its upper and lower quartiles. It is a measure of how far apart the middle portion of data spreads in value.

```
# Calculate IQRs for every gene/column, select IQRs with >1.6 and subset the data with them
iqrs = apply(expr_cc_mt, 2, IQR)
mask = which(iqrs > 1.6)
expr16 = as.data.frame(expr_cc_mt[, mask])
expr16[1:5,1:5]
##
          YBR054W YBR088C YER124C YGL028C YGR108W
## cln3.1
           -2.56
                            0.07
                                    -0.31
                                             0.21
                     2.05
## cln3.2
           -1.74
                    1.67
                           -1.29
                                     0.23
                                             0.87
## clb2.2
            2.76
                  -2.06
                           -0.32
                                             2.28
                                    -0.49
## clb2.1
           3.38
                   -1.43
                           -0.10
                                     0.16
                                            1.66
## alpha0
          -1.35
                   -1.15
                            0.92
                                     2.45
                                           -1.88
```

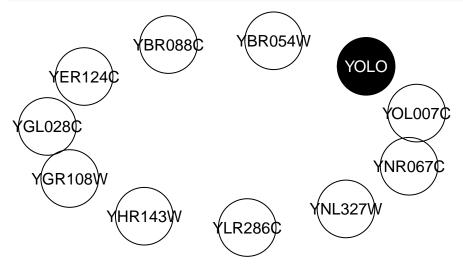
Add a dummy gene with expression 1 over all experiments (deal needs at least one discrete variable).

```
YOLO = as.factor(rep(1, length(expr16[, 1]))) # must be a factor because discrete variable expected in expr16 = cbind(expr16, YOLO) cat("Class of dummy discrete variable should be a factor:", class(expr16[1, 11]))
## Class of dummy discrete variable should be a factor: factor
```

Learn the optimal network

Create an empty network prior.

```
bn_expr16 = network(expr16) # to visually inspect local probs: inspectprob=TRUE
# nodes(bn_expr16) # YOLO node should be discrete!
plot(bn_expr16)
```

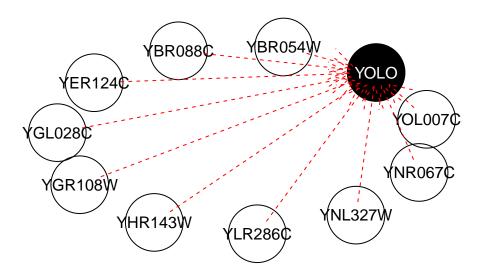


What are the local probability distributions of the genes?

```
localprob(bn_expr16)
## $YBR054W
##
                  s2 Intercept:YBR054W
          1.42234485
                             0.02649351
##
##
## $YBR088C
                  s2 Intercept:YBR088C
##
                          -0.0005194805
        0.9768984314
##
##
## $YER124C
##
                  s2 Intercept:YER124C
          1.41456404
##
                            -0.02246753
##
## $YGL028C
```

```
s2 Intercept:YGL028C
##
##
        1.455174903 -0.009350649
##
## $YGR108W
                s2 Intercept:YGR108W
##
         0.92446915 0.06779221
##
## $YHR143W
##
                s2 Intercept:YHR143W
##
         1.04158249 -0.01097403
##
## $YLR286C
##
               s2 Intercept:YLR286C
       1.9945786136 -0.0007792208
##
##
## $YNL327W
##
                s2 Intercept:YNL327W
         1.41701617 -0.02616883
##
##
## $YNR067C
##
                s2 Intercept:YNR067C
         1.60312299 -0.02415584
##
## $YOLOO7C
                s2 Intercept:YOL007C
##
        1.09477190 -0.01090909
##
## $YOLO
## 1
## 1
```

Ban all outgoing edges for the dummy node (use function banlist).



Create a joint prior with imaginary sample size 5, N=5

```
bn_expr16.prior = jointprior(bn_expr16, N=5)
## Imaginary sample size: 5
head(bn_expr16.prior)
## $jointalpha
## 1
## 5
##
## $jointnu
## 1
## 5
##
## $jointrho
## 1
## 5
##
## $jointmu
                                   YGL028C
##
      YBR054W
                YBR088C
                         YER124C
                                           YGR108W
  :1 0.02649351 -0.0005194805 -0.02246753 -0.009350649 0.06779221
##
##
       YHR143W
                 YLR286C
                          YNL327W
                                   YNR067C
                                            YOLOO7C
##
  :1 -0.01097403 -0.0007792208 -0.02616883 -0.02415584 -0.01090909
##
## $jointsigma
## $jointsigma$`:1`
##
        YBR054W
                YBR088C
                      YER124C YGL028C
                                    YGR108W YHR143W
                                                 YLR286C
## YER124C 0.000000 0.0000000 1.414564 0.000000 0.0000000 0.000000 0.000000
## YGL028C 0.000000 0.0000000 0.000000 1.455175 0.0000000 0.000000 0.000000
## YGR108W 0.000000 0.0000000 0.000000 0.000000 0.9244692 0.000000 0.000000
## YHR143W 0.000000 0.0000000 0.000000 0.000000 1.041582 0.000000
```

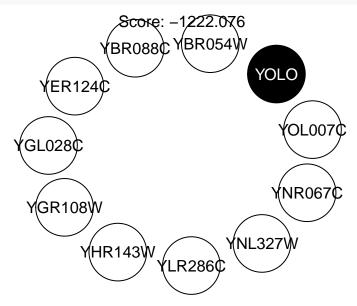
```
YNL327W YNR067C YOL007C
## YBR054W 0.000000 0.000000 0.000000
## YBR088C 0.000000 0.000000 0.000000
## YER124C 0.000000 0.000000 0.000000
## YGL028C 0.000000 0.000000 0.000000
## YGR108W 0.000000 0.000000 0.000000
## YHR143W 0.000000 0.000000 0.000000
## YLR286C 0.000000 0.000000 0.000000
## YNL327W 1.417016 0.000000 0.000000
## YNR067C 0.000000 1.603123 0.000000
## YOL007C 0.000000 0.000000 1.094772
##
##
## $jointphi
## $jointphi$\:1\
        YBR054W YBR088C YER124C YGL028C YGR108W YHR143W YLR286C
## YBR054W 5.689379 0.000000 0.000000 0.00000 0.000000 0.00000
## YER124C 0.000000 0.000000 5.658256 0.0000 0.000000 0.000000 0.000000
## YGL028C 0.000000 0.000000 0.000000 5.8207 0.000000 0.00000 0.000000
## YGR108W 0.000000 0.000000 0.000000 0.0000 3.697877 0.00000 0.000000
## YHR143W 0.000000 0.000000 0.000000 0.00000 4.16633 0.000000
YNL327W YNR067C YOL007C
## YBR054W 0.000000 0.000000 0.000000
## YBR088C 0.000000 0.000000 0.000000
## YER124C 0.000000 0.000000 0.000000
## YGL028C 0.000000 0.000000 0.000000
## YGR108W 0.000000 0.000000 0.000000
## YHR143W 0.000000 0.000000 0.000000
## YLR286C 0.000000 0.000000 0.000000
## YNL327W 5.668065 0.000000 0.000000
## YNR067C 0.000000 6.412492 0.000000
## YOL007C 0.000000 0.000000 4.379088
```

Learn the initial network (use getnetwork to turn the result into an actual network).

```
# master prior
bn_expr16.m_prior = learn(bn_expr16, expr16, bn_expr16.prior)
# initial network
bn_expr16.initial = getnetwork(bn_expr16.m_prior)
cat("Initial network score is:", bn_expr16.initial$score)
```

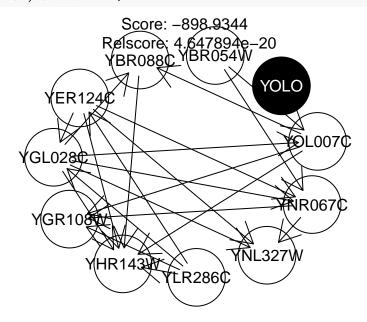
Initial network score is: -1222.076

plot(bn_expr16.initial, showban = FALSE)



Search for the optimal network and plot it

plot(bn_expr16.optimal, showban = F)



Optimal network score is:

bn_expr16.optimal\$score

[1] -898.9344