

# Sampling networks of ecological interactions

Pedro Jordano<sup>\*a</sup>

<sup>a</sup>Integrative Ecology Group, Estación Biológica de Doñana, Consejo  
Superior de Investigaciones Científicas (EBD-CSIC), Avenida  
Americo Vesputio s/n, E-41092 Sevilla, Spain

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## Summary

1. Sampling ecological interactions presents similar challenges, problems, potential biases, and constraints as sampling individuals and species in biodiversity inventories. Interactions are just pairwise relationships among individuals of two different species, such as those among plants and their seed dispersers in frugivory interactions or those among plants and their pollinators. Sampling interactions is a fundamental step to build robustly estimated interaction networks, yet few analyses have attempted a formal approach to their sampling protocols.

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<sup>\*</sup>jordano@ebd.csic.es

2. Robust estimates of the actual number of interactions (links) within diversified ecological networks require adequate sampling effort that needs to be explicitly gauged. Yet we still lack a sampling theory explicitly focusing on ecological interactions.
3. While the complete inventory of interactions is likely impossible, a robust characterization of its main patterns and metrics is probably realistic. We must acknowledge that a sizable fraction of the maximum number of interactions  $I_{max}$  among, say,  $A$  animal species and  $P$  plant species (i.e.,  $I_{max} = AP$ ) is impossible to record due to forbidden links, i.e., life-history restrictions. Thus, the number of observed interactions  $I$  in robustly sampled networks is typically  $I \ll I_{max}$ , resulting in extremely sparse interaction matrices with low connectance.
4. Reasons for forbidden links are multiple but mainly stem from spatial and temporal uncoupling of partner species encounters and from intrinsically low probabilities of interspecific encounter for many of the potential pairwise interactions. Adequately assessing the completeness of a network of ecological interactions thus needs a deep knowledge of the natural history details embedded, so that forbidden links can be “discounted” when addressing sampling effort.
5. Here I provide a review and outline a conceptual framework for interaction sampling by building an explicit analogue to individuals and species sampling, thus extending diversity-monitoring approaches to the characterization of complex networks of ecological interactions. This is crucial to assess the fast-paced and devastating effects of defaunation-driven loss of key ecological

interactions and the services they provide.

## Keywords

complex networks, food webs, frugivory, mutualism, plant-animal interactions, pollination, seed dispersal

## Introduction

Biodiversity sampling is a labour-intensive activity, and sampling is often not sufficient to detect all or even most of the species present in an assemblage. Gotelli & Colwell (2011).

1 Biodiversity species assessment aims at sampling individuals in collections and  
2 determining the number of species represented. Given that, by definition, samples  
3 are incomplete, these collections enumerate a lower number of the species actually  
4 present. The ecological literature dealing with robust estimators of species rich-  
5 ness and diversity in collections of individuals is immense, and a number of useful  
6 approaches have been used to obtain such estimates (Magurran, 1988; Gotelli &  
7 Colwell, 2001; Hortal, Borges & Gaspar, 2006; Colwell, 2009; Gotelli & Colwell,  
8 2011). Recent effort has been also focused at defining essential biodiversity vari-  
9 ables (EBV) (Pereira *et al.*, 2013) that can be sampled and measured repeatedly  
10 to complement biodiversity estimates. Yet sampling species or taxa-specific EBVs  
11 is just probing a single component of biodiversity; interactions among species are  
12 another fundamental component, the one that supports the existence, but in some

13 cases also the extinction, of species. For example, the extinction of interactions  
14 represents a dramatic loss of biodiversity because it entails the loss of fundamental  
15 ecological functions (Valiente-Banuet *et al.*, 2014). This missed component of bio-  
16 diversity loss, the extinction of ecological interactions, very often accompanies, or  
17 even precedes, species disappearance. Interactions among species are a key com-  
18 ponent of biodiversity and here I aim to show that most problems associated to  
19 sampling interactions in natural communities have to do with problems associated  
20 to sampling species diversity, even worse. I consider pairwise interactions among  
21 species at the habitat level, in the context of alpha diversity and the estimation  
22 of local interaction richness from sampling data (Mao & Colwell, 2005). In the  
23 first part I provide a succinct overview of previous work addressing sampling issues  
24 for ecological interaction networks. In the second part, after a short overview of  
25 asymptotic diversity estimates (Gotelli & Colwell, 2001), I discuss specific ratio-  
26 nales for sampling the biodiversity of ecological interactions. Most of my examples  
27 come from the analysis of plant-animal interaction networks, yet are applicable to  
28 other types of species-species interactions.

29 Interactions can be a much better indicator of the richness and diversity of  
30 ecosystem functions than a simple list of taxa and their abundances and/or re-  
31 lated biodiversity indicator variables (EBVs). Thus, sampling interactions should  
32 be a central issue when identifying and diagnosing ecosystem services (e.g., polli-  
33 nation, natural seeding by frugivores, etc.). Fortunately, all the whole battery of  
34 biodiversity-related tools used by ecologists to sample biodiversity (species, *sensu*  
35 *stricto*) can be extended and applied to the sampling of interactions. Analogs  
36 are evident between these approaches (Colwell, Dunn & Harris, 2012). Monitor-  
37 ing interactions is analogous to any biodiversity sampling [i.e., a species inventory

Jordano (1987); Jordano, Vázquez & Bascompte (2009)] and is subject to similar methodological shortcomings, especially under-sampling (Coddington *et al.*, 2009; Vazquez, Chacoff & Cagnolo, 2009; Dorado *et al.*, 2011; Rivera-Hutinel *et al.*, 2012). For example, when we study mutualistic networks, our goal is to make an inventory of the distinct pairwise interactions that made up the network. We are interested in having a complete list of all the pairwise interactions among species (e.g., all the distinct, species-species interactions, or links, among the pollinators and flowering plants) that can exist in a given community. Sampling these interactions thus entails exactly the same problems, limitations, constraints, and potential biases as sampling individual organisms and species diversity. As Mao & Colwell (2005) put it, these are the workings of Preston’s demon, the moving “veil line” between the detected and the undetected interactions as sample size increases (Preston, 1948).

Early efforts to recognize and solve sampling problems in analyses of interactions stem from research on food webs and to determine how undersampling biases food web metrics (Martinez, 1991; Cohen *et al.*, 1993; Martinez, 1993; Bersier, Banasek-Richter & Cattin, 2002; Brose, Martinez & Williams, 2003; Banasek-Richter, Cattin & Bersier, 2004; Wells & O’Hara, 2012). In addition, the myriad of classic natural history studies documenting animal diets, host-pathogen infection records, plant herbivory records, etc., represent efforts to document interactions occurring in nature. All of them share the problem of sampling incompleteness influencing the patterns and metrics reported. Yet, despite the early recognition that incomplete sampling may seriously bias the analysis of ecological networks (Jordano, 1987), only recent studies have explicitly acknowledged it and attempted to determine its influence (Ollerton & Cranmer, 2002; Nielsen & Bascompte, 2007;

63 Vazquez, Chacoff & Cagnolo, 2009; Gibson *et al.*, 2011; Olesen *et al.*, 2011; Cha-  
 64 coff *et al.*, 2012; Rivera-Hutinel *et al.*, 2012; Olito & Fox, 2014; Bascompte &  
 65 Jordano, 2014; Vizentin-Bugoni, Maruyama & Sazima, 2014; Frund, McCann &  
 66 Williams, 2015). The sampling approaches have been extended to predict patterns  
 67 of coextinctions in interaction assemblages (e.g., hosts-parasites) (Colwell, Dunn &  
 68 Harris, 2012). Most empirical studies provide no estimate of sampling effort, im-  
 69 plicitly assuming that the reported network patterns and metrics are robust. Yet  
 70 recent evidences point out that number of partner species detected, number of  
 71 actual links, and some aggregate statistics describing network patterns, are prone  
 72 to sampling bias (Nielsen & Bascompte, 2007; Dorado *et al.*, 2011; Olesen *et al.*,  
 73 2011; Chacoff *et al.*, 2012; Rivera-Hutinel *et al.*, 2012; Olito & Fox, 2014; Frund,  
 74 McCann & Williams, 2015). Most of these evidences, however, come either from  
 75 simulation studies (Frund, McCann & Williams, 2015) or from relatively species-  
 76 poor assemblages. Even for species-rich, tropical assemblages it might be erroneous  
 77 to conclude that network data routinely come from insufficiently sampled datasets  
 78 (Ollerton & Cranmer, 2002; Chacoff *et al.*, 2012), given the extremely sparse nature  
 79 of these interaction matrices because of the prevalence of forbidden links (which,  
 80 by definition, cannot be documented despite extensive sampling effort). However,  
 81 most certainly, sampling limitations pervade biodiversity inventories in tropical  
 82 areas (Coddington *et al.*, 2009) and we might rightly expect that frequent inter-  
 83 actions may be over-represented and rare interactions may be missed entirely in  
 84 studies of mega-diverse assemblages (Bascompte & Jordano, 2014); but, to what  
 85 extent?

## 86 Sampling interactions: methods

87 When we sample interactions in the field we record the presence of two species that  
 88 interact in some way. For example, Snow and Snow(1988) recorded an interaction  
 89 whenever they saw a bird “touching” a fruit on a plant. We observe and record  
 90 feeding observations, visitation, occupancy, presence in pollen loads or in fecal  
 91 samples, etc., of *individual* animals or plants and accumulate pairwise interactions,  
 92 i.e., lists of species partners and the frequencies with which we observe them.  
 93 Therefore, estimating the sampling completeness of pairwise interactions for a  
 94 whole network, requires some gauging of the sampling completeness (i.e., how the  
 95 number (richness) of distinct pairwise interactions accumulates as sampling effort  
 96 is increased) and/or estimating the uncertainty around the missed links (Wells &  
 97 O’Hara, 2012).

98 Most types of ecological interactions can be illustrated with bipartite graphs,  
 99 with two or more distinct groups of interacting partners (Bascompte & Jordano,  
 100 2014); for illustration purposes I’ll focus more specifically on plant-animal interac-  
 101 tions. Sampling interactions requires filling the cells of an interaction matrix with  
 102 data. The matrix,  $\Delta = AP$ , is a 2D representation of the interactions among,  
 103 say,  $A$  animal species (rows) and  $P$  plant species (columns) (Jordano, 1987; Bas-  
 104 compte & Jordano, 2014). The matrix entries illustrate the values of the pairwise  
 105 interactions visualized in the  $\Delta$  matrix, and can be 0 or 1, for presence-absence  
 106 of a given pairwise interaction, or take a quantitative weight  $w_{ji}$  to represent the  
 107 interaction intensity or unidirectional effect of species  $j$  on species  $i$  (Bascompte  
 108 & Jordano, 2014; Vazquez *et al.*, 2015). The outcomes of most ecological inter-  
 109 actions are dependent on frequency of encounters (e.g., visit rate of pollinators,

number of records of ant defenders, frequency of seeds in fecal samples). Thus, a frequently used proxy for interaction intensities  $w_{ji}$  is just how frequent new interspecific encounters are, whether or not appropriately weighted to estimate interaction effectiveness (Vazquez, Morris & Jordano, 2005).

We need to define two basic steps in the sampling of interactions: 1) which type of interactions we sample; and 2) which type of record we get to document the existence of an interaction. In step #1 we need to take into account whether we are sampling the whole community of interactor species (all the animals, all the plants) or just a subset of them, i.e., a sub matrix  $\Delta_{m,n}$  of  $m < A$  animal species and  $n < P$  plant species of the adjacency matrix  $\Delta_{AP}$ . Subsets can be: a) all the potential plants interacting with a subset of the animals (Fig. 1a); b) all the potential animal species interacting with a subset of the plant species (Fig. 1b); c) a subset of all the potential animal species interacting with a subset of all the plant species (Fig. 1c). While some discussion has considered how to establish the limits of what represents a network (Strogatz, 2001) (in analogy to discussion on food-web limits; Cohen, 1978), it must be noted that situations a-c in Fig. 1 do not represent complete interaction networks. As vividly stated by Cohen *et al.* (1993): “*As more comprehensive, more detailed, more explicit webs become available, smaller, highly aggregated, incompletely described webs may progressively be dropped from analyses of web structure (though such webs may remain useful for other purposes, such as pedagogy)*”. Subnet sampling is generalized in studies of biological networks (e.g., protein interactions, gene regulation), yet it is important to recognize that most properties of subnetworks (even random subsamples) do not represent properties of whole networks (Stumpf, Wiuf & May, 2005).

In step #2 above we face the problem of the type of record we take to sample



135 interactions. This is important because it defines whether we approach the problem  
 136 of filling up the interaction matrix in a “zoo-centric” way or in a “phyto-centric”  
 137 way. Zoo-centric studies directly sample animal activity and document the plants  
 138 ‘touched’ by the animal. For example, analysis of pollen samples recovered from the  
 139 body of pollinators, analysis of fecal samples of frugivores, radio-tracking data, etc.  
 140 Phyto-centric studies take samples of focal individual plant species and document  
 141 which animals ‘arrive’ or ‘touch’ the plants. Examples include focal watches of  
 142 fruiting or flowering plants to record visitation by animals, raising insect herbivores  
 143 from seed samples, identifying herbivory marks in samples of leaves, etc.

144 Most recent analyses of plant-animal interaction networks are phyto-centric;  
 145 just 3.5% of available plant-pollinator ( $N=58$ ) or 36.6% plant-frugivore ( $N=22$ )  
 146 interaction datasets are zoo-centric (see Schleuning *et al.*, 2012). Moreover, most  
 147 available datasets on host-parasite (parasitoid) or plant-herbivore interactions are  
 148 “host-centric” or phyto-centric (e.g., Thébault & Fontaine, 2010; Morris *et al.*,  
 149 2013; Eklöf *et al.*, 2013). This may be related to a variety of causes, like preferred  
 150 methodologies by researchers working with a particular group or system, logistic  
 151 limitations, or inherent taxonomic focus of the research questions. A likely result  
 152 of phyto-centric sampling would be adjacency matrices with large  $A : P$  ratios.  
 153 In any case we don’t have a clear view of the potential biases that taxa-focused  
 154 sampling may generate in observed network patterns, for example by generating  
 155 consistently asymmetric interaction matrices (Dormann *et al.*, 2009). System sym-  
 156 metry has been suggested to influence estimations of generalization levels in plants  
 157 and animals when measured as  $I_A$  and  $I_P$  (Elberling & Olesen, 1999); thus, differ-  
 158 ences in  $I_A$  and  $I_P$  between networks may arise from different  $A : P$  ratios rather  
 159 than other ecological factors (Olesen & Jordano, 2002).

Interestingly enough, quite complete analyses of interaction networks can be obtained when combining both phyto-centric and zoo-centric sampling. For example, Bosch *et al.* (2009) showed that the addition of pollen load data on top of focal-plant sampling of pollinators unveiled a significant number of interactions, resulting in important network structural changes. Connectance increased 1.43-fold, mean plant connectivity went from 18.5 to 26.4, and mean pollinator connectivity from 2.9 to 4.1; moreover, extreme specialist pollinator species (singletons in the adjacency matrix) decreased 0.6-fold. (Olesen *et al.* 2011) identified pollen loads on sampled insects and added the new links to an observation-based visitation matrix, with an extra 5% of links representing the estimated number of missing links in the pollination network. The overlap between observational and pollen-load recorded links was only 33%, underscoring the value of combining methodological approaches. Zoo-centric sampling has recently been extended with the use of DNA-barcoding, for example with plant-herbivore (Jurado-Rivera *et al.*, 2009), host-parasitoid (Wirta *et al.*, 2014), and plant-frugivore interactions (González-Varo, Arroyo & Jordano, 2014). For mutualistic networks we would expect that zoo-centric sampling could help unveiling interactions for rare species or for relatively common species which are difficult to sample by direct observation. Future methodological work may provide significant advances showing how mixing different sampling strategies strengthens the completeness of network data. These mixed strategies may combine, for instance, timed watches at focal plants, spot censuses along walked transects, pollen load or seed contents analyses, monitoring with camera traps, and DNA barcoding records. We might expect increased power of these mixed sampling approaches when combining different methods from both phyto- and zoo-centric perspectives (Bosch *et al.*, 2009; Bluthgen, 2010). Note also

185 that the different methods could be applied in different combinations to the two  
 186 distinct sets of species. However, there are no tested protocols and/or sampling  
 187 designs for ecological interaction studies to suggest an optimum combination of  
 188 approaches. Ideally, pilot studies would provide adequate information for each  
 189 specific study setting.

## 190 Sampling interactions: rationale

191 The number of distinct pairwise interactions that we can record in a landscape  
 192 (an area of relatively homogeneous vegetation, analogous to the one we would  
 193 use to monitor species diversity) is equivalent to the number of distinct classes in  
 194 which we can classify the recorded encounters among individuals of two different  
 195 species. Yet, individual-based interaction networks have been only recently studied  
 196 (Dupont, Trøjelsgaard & Olesen, 2011; Wells & O'Hara, 2012). The most usual  
 197 approach has been to pool individual-based interaction data into species-based  
 198 summaries, an approach that ignores the fact that only a fraction of individuals  
 199 may actually interact given a per capita interaction effect (Wells & O'Hara, 2012).  
 200 Wells & O'Hara (2012) illustrate the pros and cons of the approach. We walk in  
 201 the forest and see a blackbird  $Tm$  picking an ivy  $Hh$  fruit and ingesting it: we  
 202 have a record for  $Tm - Hh$  interaction. We keep advancing and record again a  
 203 blackbird feeding on hawthorn  $Cm$  fruits so we record a  $Tm - Cm$  interaction;  
 204 as we advance we encounter another ivy plant and record a blackcap swallowing a  
 205 fruit so we now have a new  $Sa - Hh$  interaction, and so on. At the end we have  
 206 a series of classes (e.g.,  $Sa - Hh$ ,  $Tm - Hh$ ,  $Tm - Cm$ , etc.), along with their  
 207 observed frequencies. Bunge & Fitzpatrick (1993) review the main aspects and ap-

proaches to estimate the number of distinct classes  $C$  in a sample of observations. Our main problem then turns to estimate the number of true missed links, i.e., those that can't be accounted for by biological constraints and that might suggest undersampling. Thus, the sampling of interactions in nature, as the sampling of species, is a cumulative process. In our analysis, we are not re-sampling individuals, but interactions, so we made interaction-based accumulation curves. If an interaction-based curve points towards a robust sampling, it does mean that no new interactions are likely to be recorded, irrespectively of the species, as it is a whole-network sampling approach (N. Gotelli, pers. com.). We add new, distinct, interactions recorded as we increase sampling effort (Fig. 2). We can obtain an Interaction Accumulation Curve (*IAC*) analogous to a Species cumulating Curve (*SAC*) (see Supplementary Online Material): the observed number of distinct pairwise interactions in a survey or collection as a function of the accumulated number of observations or samples (Colwell, 2009).

Our sampling above would have resulted in a vector  $n = [n_1 \dots n_C]'$  where  $n_i$  is the number of records in the  $i^{th}$  class. As stressed by Bunge & Fitzpatrick (1993), however, the  $i^{th}$  class would appear in the sample if and only if  $n_i > 0$ , and we don't know *a priori* which  $n_i$  are zero. So,  $n$  is not observable. Rather, what we get is a vector  $c = [c_1 \dots c_n]'$  where  $c_j$  is the number of classes represented  $j$  times in our sampling:  $c_1$  is the number of singletons (interactions recorded once),  $c_2$  is the number of twin pairs (interactions wth just two records),  $c_3$  the number of triplets, etc. The problem thus turns to be estimating the number of distinct classes  $C$  from the vector of  $c_j$  values and the frequency of unobserved interactions (see "The real missing links" below).

Estimating the number of interactions with resulting robust estimates of net-

work parameters is a central issue in the study of ecological interaction networks (Jordano, 1987; Bascompte & Jordano, 2014). In contrast with traditional species diversity estimates, sampling networks has the paradox that despite the potentially interacting species being present in the sampled assemblage (i.e., included in the  $A$  and  $P$  species lists), some of their pairwise interactions are impossible to be recorded. The reason is forbidden links. Independently of whether we sample full communities or subset communities we face a problem: some of the interactions that we can visualize in the empty adjacency matrix  $\Delta$  will simply not occur. Thus, independently of the sampling effort we put, we'll never document these pairwise interactions. With a total of  $AP$  "potential" interactions, a fraction of them are impossible to record, because they are forbidden (Jordano, Bascompte & Olesen, 2003; Olesen *et al.*, 2011). Forbidden links are non-occurrences of pairwise interactions that can be accounted for by biological constraints, such as spatio-temporal uncoupling (Jordano, 1987), size or reward mismatching, foraging constraints (e.g., accessibility) (Moré *et al.*, 2012), and physiological-biochemical constraints (Jordano, 1987). We still have extremely reduced information about the frequency of forbidden links in natural communities (Jordano, Bascompte & Olesen, 2003; Stang *et al.*, 2009; Vazquez, Chacoff & Cagnolo, 2009; Olesen *et al.*, 2011; Ibanez, 2012; Maruyama *et al.*, 2014; Vizentin-Bugoni, Maruyama & Sazima, 2014) (Table 1). Forbidden links are thus represented as structural zeroes in the interaction matrix, i.e., matrix cells that cannot get a non-zero value. We might expect different types of  $FL$  to occupy different parts of the  $\Delta$  matrix, with missing cells due to phenological uncoupling,  $FL_P$ , largely distributed in the lower-right half  $\Delta$  matrix and actually missed links  $ML$  distributed in its central part (Olesen *et al.*, 2010). Yet, most of these aspects remain understudied. Therefore, we need to account for

the frequency of these structural zeros in our matrix before proceeding. For example, most measurements of connectance  $C = I/(AP)$  implicitly ignore the fact that by taking the full product  $AP$  in the denominator they are underestimating the actual connectance value, i.e., the fraction of actual interactions  $I$  relative to the *biologically possible* ones, not to the total maximum  $I_{max} = AP$ .

Adjacency matrices are frequently sparse, i.e., they are densely populated with zeroes, with a fraction of them being structural (unobservable interactions) (Bascompte & Jordano, 2014). Thus, it would be a serious interpretation error to attribute the sparseness of adjacency matrices for bipartite networks to undersampling. The actual typology of link types in ecological interaction networks is thus more complex than just the two categories of observed and unobserved interactions (Table 1). Unobserved interactions are represented by zeroes and belong to two categories. Missing interactions may actually exist but require additional sampling or a variety of methods to be observed. Forbidden links, on the other hand, arise due to biological constraints limiting interactions and remain unobservable in nature, irrespectively of sampling effort (Table 1). Forbidden links  $FL$  may actually account for a relatively large fraction of unobserved interactions  $UL$  when sampling taxonomically-restricted subnetworks (e.g., plant-hummingbird pollination networks) (Table 1). Phenological unmatching is also prevalent in most networks, and may add up to explain ca. 25–40% of the forbidden links, especially in highly seasonal habitats, and up to 20% when estimated relative to the total number of unobserved interactions (Table 2). In any case, we might expect that a fraction of the missing links  $ML$  would be eventually explained by further biological reasons, depending on the knowledge of natural details of the particular systems. Our goal as naturalists would be to reduce the fraction of  $UL$  which remain as

283 missing links; to this end we might search for additional biological constraints or  
 284 increase sampling effort. For instance, habitat use patterns by hummingbirds in  
 285 the Arima Valley network (Table 2; Snow & Snow, 1972) impose a marked pattern  
 286 of microhabitat mismatches causing up to 44.5% of the forbidden links. A myr-  
 287 iad of biological causes beyond those included as *FL* in Table 2 may contribute  
 288 explanations for *UL*: limits of color perception and or partial preferences, pres-  
 289 ence of secondary metabolites in fruit pulp and leaves, toxins and combinations  
 290 of monosaccharides in nectar, etc. However, it is surprising that just the limited  
 291 set of forbidden link types considered in Table 1 explain between 24.6–77.2% of  
 292 the unobserved links. Notably, the Arima Valley, Santa Virgínia, and Hato Ratón  
 293 networks have  $> 60\%$  of the unobserved links explained, which might be related  
 294 to the fact that they are subnetworks (Arima Valley, Santa Virgínia) or relatively  
 295 small networks (Hato Ratón). All this means that empirical networks may have  
 296 sizable fractions of structural zeroes. Ignoring this biological fact may contribute  
 297 to wrongly inferring undersampling of interactions in real-world assemblages.

298 To sum up, two elements of inference are required in the analysis of unobserved  
 299 interactions in ecological interaction networks: first, detailed natural history infor-  
 300 mation on the participant species that allows the inference of biological constraints  
 301 imposing forbidden links, so that structural zeroes can be identified in the adja-  
 302 cency matrix. Second, a critical analysis of sampling robustness and a robust  
 303 estimate of the actual fraction of missing links,  $M$ , resulting in a robust estimate  
 304 of  $I$ . In the next sections I explore these elements of inference. The basic proposal  
 305 is to use *IACs* to assess the robustness of interaction sampling, then scale the  
 306 asymptotic estimate of interactions richness to account for the unrealizable *FL*.

## 307 Asymptotic diversity estimates

Let's assume a sampling of the diversity in a specific locality, over relatively homogeneous landscape where we aim at determining the number of species present for a particular group of organisms. To do that we carry out transects or plot samplings across the landscape, adequately replicated so we obtain a number of samples. Briefly,  $S_{obs}$  is the total number of species observed in a sample, or in a set of samples.  $S_{est}$  is the estimated number of species in the community represented by the sample, or by the set of samples, where *est* indicates an estimator. With abundance data, let  $S_k$  be the number of species each represented by exactly  $k$  individuals in a single sample. Thus,  $S_0$  is the number of undetected species (species present in the community but not included in the sample),  $S_1$  is the number of singleton species (represented by just one individual),  $S_2$  is the number of doubleton species (species with two individuals), etc. The total number of individuals in the sample would be:

$$n = \sum_{k=1}^{S_{obs}} S_k$$

308

309 A frequently used asymptotic, bias corrected, non-parametric estimator is  $S_{Chao}$   
 310 (Hortal, Borges & Gaspar, 2006; Chao, 2005; Colwell, 2013):

$$S_{Chao} = S_{obs} + \frac{S_1(S_1 - 1)}{2(S_2 + 1)}$$

311 Another frequently used alternative is the Chao2 estimator,  $S_{Chao2}$  (Gotelli &  
 312 Colwell, 2001), which has been reported to have a limited bias for small sample



313 sizes (Colwell & Coddington, 1994; Chao, 2005):

$$S_{Chao2} = S_{obs} + \frac{S_1^2}{2S_2}$$

314 A plot of the cumulative number of species recorded,  $S_n$ , as a function of some  
 315 measure of sampling effort (say,  $n$  samples taken) yields the species accumulation  
 316 curve (SAC) or collector's curve (Colwell & Coddington, 1994). Such a curve  
 317 eventually reaches an asymptote converging with  $S_{est}$ . Similarly, interaction accu-  
 318 mulation curves (IAC), analogous to SACs, can be used to assess the robustness  
 319 of interactions sampling for plant-animal community datasets (Jordano, 1987; Jor-  
 320 dano, Vázquez & Bascompte, 2009; Olesen *et al.*, 2011). For instance, a random  
 321 accumulator function (e.g., library `vegan` in the R Package, R Development Core  
 322 Team, 2010) which finds the mean IAC and its standard deviation from random  
 323 permutations of the data, or subsampling without replacement (Gotelli & Colwell,  
 324 2001) can be used to estimate the expected number of distinct pairwise interactions  
 325 included in a given sampling of records (Jordano, Vázquez & Bascompte, 2009;  
 326 Olesen *et al.*, 2011). This is analogous to a biodiversity sampling matrix with  
 327 species as rows and sampling units (e.g., quadrats) as columns (Jordano, Vázquez  
 328 & Bascompte, 2009). In this way we effectively extend sampling theory developed  
 329 for species diversity to the sampling of ecological interactions. Yet future theoret-  
 330 ical work will be needed to formally assess the similarities and differences in the  
 331 two approaches and developing biologically meaningful null models of expected  
 332 interaction richness with added sampling effort.

## Assessing sampling effort when recording interactions

The basic method we can propose to estimate sampling effort and explicitly show the analogues with rarefaction analysis in biodiversity research is to vectorize the interaction matrix  $AP$  so that we get a vector of all the potential pairwise interactions ( $I_{max}$ , Table 1) that can occur in a community of  $A$  animal species and  $P$  plant species. The new “species” we aim to sample are the pairwise interactions (Table 3). So, if we have in our community *Turdus merula* ( $Tm$ ) and *Rosa canina* ( $Rc$ ) and *Prunus mahaleb* ( $Pm$ ), our problem will be to sample 2 new “species”:  $Tm - Rc$  and  $Tm - Pm$ . In general, if we have  $A = 1...i$ , animal species and  $P = 1...j$  plant species, we’ll have a vector of “new” species to sample:  $A_1P_1, A_1P_2, ...A_2P_1, A_2P_2, ...A_iP_j$ . We can represent the successive samples where we can potentially get records of these interactions in a matrix with the vectorized interaction matrix and columns representing the successive samples we take (Table 3). This is simply a vectorized version of the interaction matrix.

Rarefaction analysis and diversity-accumulation analysis (Magurran, 1988; Hortal, Borges & Gaspar, 2006) come up immediately with this type of dataset. This procedure plots the accumulation curve for the expected number of distinct pairwise interactions recorded with increasing sampling effort (Jordano, Vázquez & Bascompte, 2009; Olesen *et al.*, 2011). Asymptotic estimates of interaction richness and its associated standard errors and confidence intervals can thus be obtained (Hortal, Borges & Gaspar, 2006) (see Supplementary Online Material). It should be noted that the asymptotic estimate of interaction richness implicitly ignores the fact that, due to forbidden links, a number of pairwise interactions

among the  $I_{max}$  number specified in the adjacency matrix  $\Delta$  cannot be recorded, irrespective of sampling effort. Therefore, the asymptotic value most likely is an overestimate of the actual maximum number of links that can be present in an assemblage. If forbidden links are taken into account, the asymptotic estimate should be lower. Yet, to the best of my knowledge, there is no theory developed to estimate this “biologically real” asymptotic value. Not unexpectedly, most recent analyses of sampling effort in ecological network studies found evidences of undersampling (Chacoff *et al.*, 2012). This needs not to be true, especially when interaction subwebs are studied (Olesen *et al.*, 2011; Vizentin-Bugoni, Maruyama & Sazima, 2014), and once the issue of structural zeroes in the interaction matrices is effectively incorporated in the estimates.

For example, mixture models incorporating detectabilities have been proposed to effectively account for rare species (Mao & Colwell, 2005). In an analogous line, mixture models could be extended to samples of pairwise interactions, also with specific detectability values. These detection rate/odds could be variable among groups of interactions, depending on their specific detectability. For example, detectability of flower-pollinator interactions involving bumblebees could have a higher detectability than flower-pollinator pairwise interactions involving, say, nitidulid beetles. These more homogeneous groupings of pairwise interactions within a network define modules (Bascompte & Jordano, 2014), so we might expect that interactions of a given module (e.g., plants and their hummingbird pollinators; Fig. 1a) may share similar detectability values, in an analogous way to species groups receiving homogeneous detectability values in mixture models (Mao & Colwell, 2005). In its simplest form, this would result in a sample with multiple pairwise interactions detected, in which the number of interaction events recorded for each

distinct interaction found in the sample is recorded (i.e., a column vector in Table 3, corresponding to, say, a sampling day). The number of interactions recorded for the  $i_{th}$  pairwise interaction (i.e.,  $A_i P_j$  in Table 3),  $Y_i$  could be treated as a Poisson random variable with a mean parameter  $\lambda_i$ , its detection rate. Mixture models (Mao & Colwell, 2005) include estimates for abundance-based data (their analogous in interaction sampling would be weighted data), where  $Y_i$  is a Poisson random variable with detection rate  $\lambda_i$ . This is combined with the incidence-based model, where  $Y_i$  is a binomial random variable (their analogous in interaction sampling would be presence/absence records of interactions) with detection odds  $\lambda_i$ . Let  $T$  be the number of samples in an incidence-based data set. A Poisson/binomial density can be written as (Mao & Colwell, 2005):

$$g(y; \lambda) = \begin{cases} \frac{\lambda^y}{y!e^\lambda} & [1] \\ \binom{T}{y} \frac{\lambda^y}{(1+\lambda)^T} & [2] \end{cases}$$

where [1] corresponds to a weighted network, and [2] to a qualitative network.

The detection rates  $\lambda_i$  depend on the relative abundances  $\phi_i$  of the interactions, the probability of a pairwise interaction being detected when it is present, and the sample size (the number of interactions recorded), which, in turn, is a function of the sampling effort. Unfortunately, no specific sampling model has been developed along these lines for species interactions and their characteristic features. For example, a complication factor might be that interaction abundances,  $\phi_i$ , in real assemblages are a function of the abundances of interacting species, that determine interspecific encounter rates; yet they also depend on biological factors that ultimately determine if the interaction occurs when the partner species are

present. For example,  $\lambda_i$  should be set to zero for all  $FL$ . In its simplest form,  $\phi_i$  could be estimated from just the product of partner species abundances, an approach recently used as a null model to assess the role of biological constraints in generating forbidden links and explaining interaction patterns (Vizentin-Bugoni, Maruyama & Sazima, 2014). Yet more complex models (e.g., Wells & O’hara 2012) should incorporate not only interspecific encounter probabilities, but also interaction detectabilities, phenotypic matching and incidence of forbidden links.

## The *real* missing links

Given that a fraction of unobserved interactions can be accounted for by forbidden links, what about the remaining missing interactions? We have already discussed that some of these could still be related to unaccounted constraints, and still others would be certainly attributable to insufficient sampling. Would this always be the case? Multispecific assemblages of distinct taxonomic relatedness, whose interactions can be represented as bipartite networks (e.g., host-parasite, plant-animal mutualisms, plant-herbivore interactions- with two distinct sets of unrelated higher taxa), are shaped by interspecific encounters among individuals of the partner species (Fig. 2). A crucial ecological aspect limiting these interactions is the probability of interspecific encounter, i.e., the probability that two individuals of the partner species actually encounter each other in nature.

Given log-normally distributed abundances of the two species groups, the expected “neutral” probabilities of interspecific encounter ( $PIE$ ) would be simply the product of the two lognormal distributions. Thus, we might expect that for low  $PIE$  values, pairwise interactions would be either extremely difficult to sample, or

426 just simply non-occurring in nature. Consider the Nava de las Correhuelas inter-  
 427 action web (NCH, Table 2), with  $A = 36$ ,  $P = 25$ ,  $I = 181$ , and almost half of the  
 428 unobserved interactions not accounted for by forbidden links, thus  $M = 53.1\%$ .  
 429 Given the robust sampling of this network (Jordano, Vázquez & Bascompte, 2009),  
 430 a sizable fraction of these possible but missing links would be simply not occurring  
 431 in nature, most likely by extremely low  $PIE$ , in fact asymptotically zero. Given  
 432 the vectorized list of pairwise interactions for NCH, I computed the  $PIE$  values for  
 433 each one by multiplying element wise the two species abundance distributions. The  
 434  $PIE_{max} = 0.0597$ , being a neutral estimate, based on the assumption that interac-  
 435 tions occur in proportion to the species-specific local abundances. With  $PIE_{median}$   
 436  $< 1.4 \cdot 10^{-4}$  we may safely expect (note the quantile estimate  $Q_{75\%} = 3.27 \cdot 10^{-4}$ ) that  
 437 a sizable fraction of these missing interactions may simply not occur according to  
 438 this neutral expectation (Jordano, 1987; Olesen *et al.*, 2011) (neutral forbidden  
 439 links, *sensu* Canard *et al.*, 2012). Which is the expected frequency for pairwise  
 440 interactions? and, which is the expected probability for unobserved interactions?  
 441 More specifically, which is the probability of missing interactions,  $M$  (i.e., the  
 442 unobserved ones that cannot be accounted for as forbidden links)?

443 When we consider the vectorized interaction matrix, enumerating all pairwise  
 444 interactions for the  $AP$  combinations, the expected probabilities of finding a given  
 445 interaction can be estimated with a Good-Turing approximation (Good, 1953).  
 446 The technique, developed by Alan Turing and I.J. Good with applications to lin-  
 447 guistics and word analysis (Gale & Sampson, 1995) has been recently applied in  
 448 ecology (Chao *et al.*, 2015), estimates the probability of recording an interaction  
 449 of a hitherto unseen pair of partners, given a set of past records of interactions  
 450 between other species pairs. Let a sample of  $N$  interactions so that  $n_r$  distinct

pairwise interactions have exactly  $r$  records. All Good-Turing estimators obtain the underlying frequencies of events as:

$$P(X) = \frac{(N_X + 1)}{T} \left(1 - \frac{E(1)}{T}\right) \quad (1)$$

where  $X$  is the pairwise interaction,  $N_X$  is the number of times interaction  $X$  is recorded,  $T$  is the sample size (number of distinct interactions recorded) and  $E(1)$  is an estimate of how many different interactions were recorded exactly once. Strictly speaking Equation (1) gives the probability that the next interaction type recorded will be  $X$ , after sampling a given assemblage of interacting species. In other words, we scale down the maximum-likelihood estimator  $\frac{n}{T}$  by a factor of  $\frac{1-E(1)}{T}$ . This reduces all the probabilities for interactions we have recorded, and makes room for interactions we haven't seen. If we sum over the interactions we have seen, then the sum of  $P(X)$  is  $1 - \frac{1-E(1)}{T}$ . Because probabilities sum to one, we have the left-over probability of  $P_{new} = \frac{E(1)}{T}$  of seeing something new, where new means that we sample a new pairwise interaction.

Note, however, that Good-Turing estimators, as the traditional asymptotic estimators, do not account in our case for the forbidden interactions. To account for these *FL* I re-scaled the asymptotic estimates, so that a more meaningful estimate could be obtained (Table 4). The scaling was calculated as  $[Chao1 * (I + ML)] / AP$ , just correcting for the *FL* frequency, given that  $I + ML$  represent the total *feasible* interactions when discounting the forbidden links (Table 1). After scaling, observed  $I$  values (Table 2) are within the *Chao1* and *ACE* asymptotic estimates but below the *ACE* estimates for Hato Ratón and Zackenberg (Table 4). Thus, even after re-scaling for *FL*, it is likely that adequate characterization of most interaction

networks will require intensive sampling effort.

## Discussion

Recent work has inferred that most data available for interaction networks are incomplete due to undersampling, resulting in a variety of biased parameters and network patterns (Chacoff *et al.*, 2012). It is important to note, however, that in practice, many surveyed networks to date have been subnets of much larger networks. This is true for protein interaction, gene regulation, and metabolic networks, where only a subset of the molecular entities in a cell have been sampled (Stumpf, Wiuf & May, 2005). Despite recent attempts to document whole ecosystem meta-networks (Pocock, Evans & Memmott, 2012), it is likely that most ecological interaction networks will illustrate just major ecosystem compartments. Due to their high generalization, high temporal and spatial turnover, and high complexity of association patterns, adequate sampling of ecological interaction networks requires extremely large sampling effort. Undersampling of ecological networks may originate from the analysis of assemblage subsets (e.g., taxonomically or functionally defined), and/or from logistically-limited sampling effort. It is extremely hard to robustly sample the set of biotic interactions even for relatively simple, species-poor assemblages; yet, concluding that all ecological network datasets are undersampled would be unrealistic. The reason stems from a biological fact: a sizeable fraction of the maximum, potential links that can be recorded among two distinct sets of species is simply unobservable, irrespective of sampling effort (Jordano, 1987).

Missing links are a characteristic feature of all plant-animal interaction net-



works, and likely pervade other ecological interactions. Important natural history details explain a fraction of them, resulting in unrealizable interactions (i.e., forbidden interactions) that define structural zeroes in the interaction matrices and contribute to their extreme sparseness. Sampling interactions is a way to monitor biodiversity beyond the simple enumeration of component species and to develop efficient and robust inventories of functional interactions. Yet no sampling theory for interactions is available. Some key components of this sampling are analogous to species sampling and traditional biodiversity inventories; however, there are important differences. Focusing just on the realized interactions or treating missing interactions as the expected unique result of sampling bias would miss important components to understand how mutualisms coevolve within complex webs of interdependence among species.

Contrary to species inventories, a sizable fraction of non-observed pairwise interactions cannot be sampled, due to biological constraints that forbid their occurrence. A re-scaling of traditional asymptotic estimates for interaction richness can be applied whenever the knowledge of natural history details about the study system is sufficient to estimate at least the main causes of forbidden links. Moreover, recent implementations of inference methods for unobserved species (Chao *et al.*, 2015) or for individual-based data (Wells & O'Hara, 2012) can be combined with the forbidden link approach, yet they do not account either for the existence of these ecological constraints.

Ecological interactions provide the wireframe supporting the lives of species, and they also embed crucial ecosystem functions which are fundamental for supporting the Earth system. Yet we still have a limited knowledge of the biodiversity of ecological interactions, but they are being lost (extinct) at a very fast pace, fre-

quently preceding species extinctions (Valiente-Banuet *et al.*, 2014). We urgently need robust techniques to assess the completeness of ecological interactions networks because this knowledge will allow the identification of the minimal components of their ecological complexity that need to be restored to rebuild functional ecosystems after perturbations.

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## Data accessibility

This review does not use new raw data, but includes some re-analyses of previously published material. All the original data supporting the paper, R code, supplementary figures, and summaries of analytical protocols is available at the author's

542 GitHub repository ([https://github.com/pedroj/MS\\_Network-Sampling](https://github.com/pedroj/MS_Network-Sampling)), with  
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## Figure captions

**Figure 1.** Sampling ecological interaction networks (e.g., plant-animal interactions) usually focus on different types of subsampling the full network, yielding submatrices  $\Delta[m, n]$  of the full interaction matrix  $\Delta$  with  $A$  and  $P$  animal and plant species. a) all the potential plants interacting with a subset of the animals (e.g., studying just the hummingbird-pollinated flower species in a community); b) all the potential animal species interacting with a subset of the plant species (e.g., studying the frugivore species feeding on figs *Ficus* in a community); and c) sampling a subset of all the potential animal species interacting with a subset of all the plant species (e.g., studying the plant-frugivore interactions of the rainforest understory).

**Figure 2.** Sampling species interactions in natural communities. Suppose an assemblage with  $A = 3$  animal species (red, species 1–3 with three, two, and 1 individuals, respectively) and  $P = 3$  plant species (green, species a-c with three individuals each) (colored balls), sampled with increasing effort in steps 1 to 6 (panels). In Step 1 we record animal species 1 and plant species 1 and 2 with a total of three interactions (black lines) represented as two distinct interactions:  $1 - a$  and  $1 - b$ . As we advance our sampling (panels 1 to 6, illustrating e.g., additional sampling days) we record new distinct interactions. Note that we actually sample and record interactions among individuals, yet we pool the data across species to get a species by species interaction matrix. Few network analyses have been carried out on individual data (Dupont *et al.*, 2014).

764 **Figures**

Figure 1:

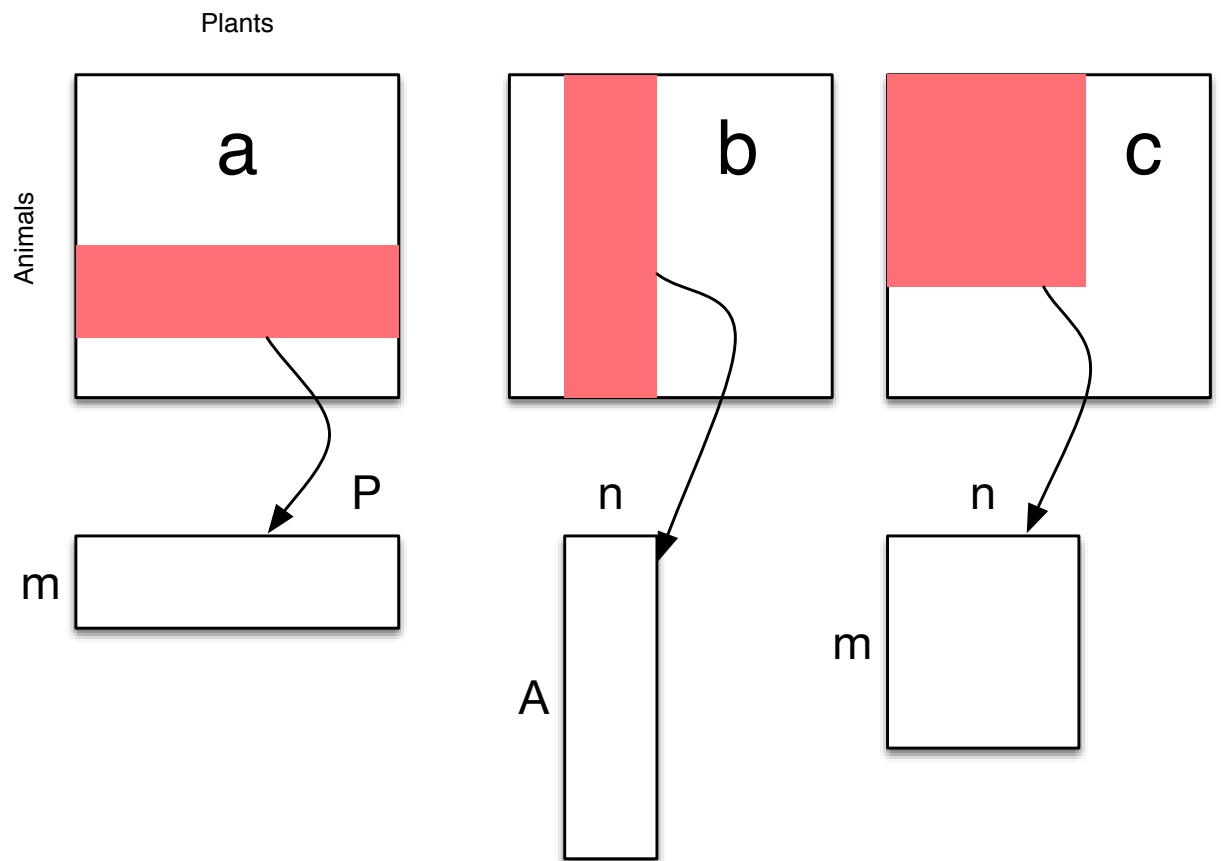
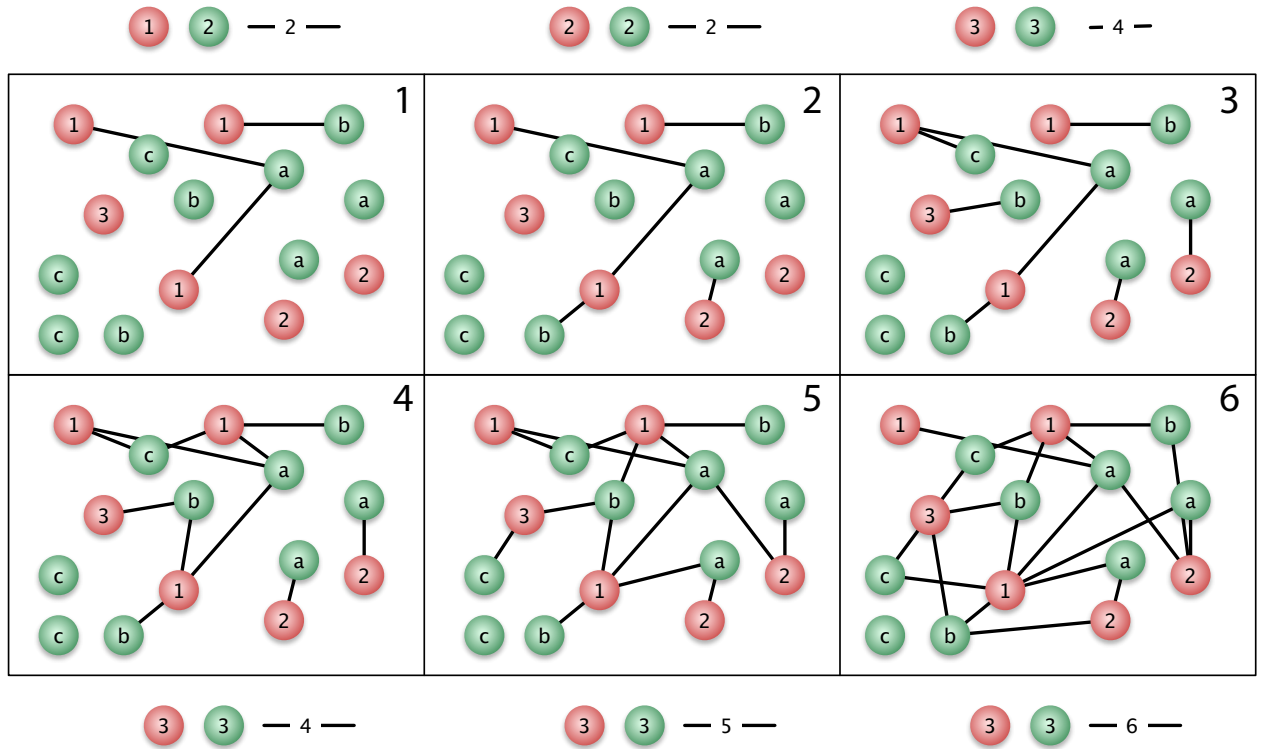


Figure 2:



Jordano - Figure 1

## Table captions

**Table 1.** A taxonomy of link types for ecological interactions (Olesen *et al.* 2011).

$A$ , number of animal species;  $P$ , number of plant species;  $I$ , number of observed links;  $C = 100I/(AP)$ , connectance;  $FL$ , number of forbidden links; and  $ML$ , number of missing links. As natural scientists, our ultimate goal is to eliminate  $ML$  from the equation  $FL = AP - I - ML$ , which probably is not feasible given logistic sampling limitations. When we, during our study, estimate  $ML$  to be negligible, we cease observing and estimate  $I$  and  $FL$ .

**Table 2.** Frequencies of different type of forbidden links in natural plant-animal interaction assemblages.  $AP$ , maximum potential links,  $I_{max}$ ;  $I$ , number of observed links;  $UL$ , number of unobserved links;  $FL$ , number of forbidden links;  $FL_P$ , phenology;  $FL_S$ , size restrictions;  $FL_A$ , accessibility;  $FL_O$ , other types of restrictions;  $ML$ , unknown causes (missing links). Relative frequencies (in parentheses) calculated over  $I_{max} = AP$  for  $I$ ,  $ML$ , and  $FL$ ; for all forbidden links types, calculated over  $FL$ . References, from left to right: Olesen *et al.* 2008; Olesen & Myrthue unpubl.; Snow & Snow 1972 and Jordano *et al.* 2006; Vizentin-Bugoni *et al.* 2014; Jordano *et al.* 2009; Olesen *et al.* 2011.

**Table 3.** A vectorized interaction matrix.

**Table 4.** Sampling statistics for three plant-animal interaction networks (Olesen *et al.* 2011). Symbols as in Table 1;  $N$ , number of records;  $Chao1$  and  $ACE$  are asymptotic estimators for the number of distinct pairwise interactions  $I$  (Hortal



789 *et al.* 2006), and their standard errors;  $C$ , sample coverage for rare interactions  
790 (Chao & Jost 2012). Scaled asymptotic estimators and their confidence intervals  
791 ( $CI$ ) were calculated by weighting *Chao1* and *ACE* with the observed frequencies  
792 of forbidden links.

793

794 **Tables**

Table 1:

Link type	Formulation	Definition
Potential links	$I_{max} = AP$	Size of network matrix, i.e. maximum number of potentially observable interactions; $A$ and $P$ , numbers of interacting animal and plant species, respectively.
Observed links	$I$	Total number of observed links in the network given a sufficient sampling effort. Number of ones in the adjacency matrix.
Unobserved links	$UL = I_{max} - I$	Number of zeroes in the adjacency matrix.
Forbidden links	$FL$	Number of links, which remain unobserved because of linkage constraints, irrespectively of sufficient sampling effort.
Missing links	$ML = AP - I - FL$	Number of links, which may exist in nature but need more sampling effort and/or additional sampling methods to be observed.

Table 2:

Link type	Pollination			Seed dispersal		
	Zackenber	Grundvad	Arima Valley	Sta. Virginia	Hato Ratón	Nava Correhuelas
$I_{max}$	1891	646	522	423	272	825
$I$	268 (0.1417)	212 (0.3282)	185 (0.3544)	86 (0.1042)	151 (0.4719)	181 (0.2194)
$UL$	1507 (0.7969)	434 (0.6718)	337 (0.6456)	337 (0.4085)	169 (0.5281)	644 (0.7806)
$FL$	530 (0.3517)	107 (0.2465)	218 (0.6469)	260 (0.7715)	118 (0.6982)	302 (0.4689)
$FL_P$	530 (1.0000)	94 (0.2166)	0 (0.0000)	120 (0.1624)	67 (0.3964)	195 (0.3028)
$FL_S$	$\dots(\dots)$	8 (0.0184)	30 (0.0890)	140 (0.1894)	31 (0.1834)	46 (0.0714)
$FL_A$	$\dots(\dots)$	5 (0.0115)	150 (0.445) <sup>a</sup>	$\dots(\dots)$	20 (0.1183)	61 (0.0947)
$FL_O$	$\dots(\dots)$	$\dots(\dots)$	38 (0.1128) <sup>b</sup>	$\dots(\dots)$	$\dots(\dots)$	363 (0.5637)
$ML$	977 (0.6483)	327 (0.7535)	119 (0.3531)	77 (0.1042)	51 (0.3018)	342 (0.5311)

<sup>a</sup>, Lack of accessibility due to habitat uncoupling, i.e., canopy-foraging species vs. understory species.

<sup>b</sup>, Colour restrictions, and reward per flower too small relative to the size of the bird.

Table 3:

Interaction	Sample 1	Sample 2	Sample 3	...	Sample $i$
A1 - P2	12	2	0	...	6
A1 - P2	0	0	0	...	1
...	...	...	...	...	...
A5 - P3	5	0	1	...	18
A5 - P4	1	0	1	...	3
...	...	...	...	...	...
A <sub>i</sub> - P <sub>i</sub>	1	0	1	...	2

Table 4:

	Hato Ratón	Nava Correhuelas	Zackenberg
$A$	17	33	65
$P$	16	25	31
$I_{max}$	272	825	1891
$N$	3340	8378	1245
$I$	151	181	268
$C$	0.917	0.886	0.707
$Chao1$	$263.1 \pm 70.9$	$231.4 \pm 14.2$	$509.6 \pm 54.7$
$ACE$	$240.3 \pm 8.9$	$241.3 \pm 7.9$	$566.1 \pm 14.8$
$Scaled\ Chao$	195.4	162.7	308.4
$CI$	[124.5–266.3]	[148.5–176.9]	[253.6–363.1]
$Scaled\ ACE$	178.5	169.7	342.6
$CI$	[169.5–187.4]	[161.8–177.6]	[327.8–357.4]
% <i>unobserved</i> <sup>a</sup>	8.33	15.38	47.80

<sup>a</sup>, estimated with library Jade (R Core Development Team 2010, Chao *et al.* 2015)