

Sampling networks of ecological interactions

Pedro Jordano^{*a}

^aIntegrative Ecology Group, Estación Biológica de Doñana, Consejo
Superior de Investigaciones Científicas (EBD-CSIC), Avenida
Americo Vesputio s/n, E-41092 Sevilla, Spain

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Summary

1. Sampling ecological interactions presents similar challenges, problems, potential biases, and constraints as sampling individuals and species in biodiversity inventories. Sampling interactions is a fundamental step to build robustly estimated interaction networks, yet few analyses have attempted a formal approach to their sampling protocols. Robust estimates of the actual number of interactions (links) within diversified ecological networks require adequate sampling effort that needs to be explicitly gauged. Yet we still lack a sampling theory explicitly focusing on ecological interactions.

^{*}jordano@ebd.csic.es

2. While the complete inventory of interactions is likely impossible, a robust characterization of its main patterns and metrics is probably realistic. We must acknowledge that a sizeable fraction of the maximum number of interactions I_{max} among, say, A animal species and P plant species (i.e., $I_{max} = AP$) is impossible to record due to forbidden links, i.e., life-history restrictions. Thus, the number of observed interactions I in robustly sampled networks is typically $I \ll I_{max}$, resulting in sparse interaction matrices with low connectance.
3. Reasons for forbidden links are multiple but mainly stem from spatial and temporal uncoupling, size mismatches, and intrinsically low probabilities of interspecific encounter for most potential interactions of partner species. Adequately assessing the completeness of a network of ecological interactions thus needs knowledge of the natural history details embedded, so that forbidden links can be accounted for when addressing sampling effort.
4. Here I provide a review and outline a conceptual framework for interaction sampling by building an explicit analogue to individuals and species sampling, thus extending diversity-monitoring approaches to the characterization of complex networks of ecological interactions. Contrary to species inventories, a sizable fraction of non-observed pairwise interactions cannot be sampled, due to biological constraints that forbid their occurrence.
5. Recent implementations of inference methods for unobserved species or for individual-based data can be combined with the assessment of forbidden links. This can help in estimating their relative importance, simply by the difference between the asymptotic estimate of interaction richness *in a*

robustly-sampled assemblage and the maximum richness I_{max} of interactions.

This is crucial to assess the rapid and devastating effects of defaunation-driven loss of key ecological interactions and the services they provide and the analogous losses related to interaction gains due to invasive species and biotic homogenization.

Keywords

complex networks, food webs, frugivory, mutualism, plant-animal interactions, pollination, seed dispersal

Introduction

Biodiversity sampling is a labour-intensive activity, and sampling is often not sufficient to detect all or even most of the species present in an assemblage. Gotelli & Colwell (2011).

1 Biodiversity species assessment aims at sampling individuals in collections and
2 determining the number of species represented. Given that, by definition, samples
3 are incomplete, these collections do not enumerate the species actually present.
4 The ecological literature dealing with robust estimators of species richness and di-
5 versity in collections of individuals is immense, and a number of useful approaches
6 have been used to obtain such estimates (Magurran, 1988; Gotelli & Colwell, 2001;
7 Colwell *et al.*, 2004; Hortal *et al.*, 2006; Colwell, 2009; Gotelli & Colwell, 2011; Chao
8 *et al.*, 2014). Recent effort has been also focused at defining essential biodiversity

9 variables (EBV) (Pereira *et al.*, 2013) that can be sampled and measured repeat-
 10 edly to complement biodiversity estimates. Yet sampling species or taxa-specific
 11 EBVs is just probing a single component of biodiversity; interactions among species
 12 are another fundamental component, one that supports the existence, but in some
 13 cases also the extinction, of species. For example, the extinction of interactions
 14 represents a dramatic loss of biodiversity because it entails the loss of fundamental
 15 ecological functions (Valiente-Banuet *et al.*, 2014). This missed component of bio-
 16 diversity loss, the extinction of ecological interactions, very often accompanies, or
 17 even precedes, species disappearance. Interactions among species are a key com-
 18 ponent of biodiversity and here I aim to show that most problems associated with
 19 sampling interactions in natural communities relate to, and are even worse than,
 20 problems associated with sampling species diversity. I consider pairwise interac-
 21 tions among species at the habitat level, in the context of alpha diversity and the
 22 estimation of local interaction richness from sampling data (Chao *et al.*, 2014). In
 23 the first part I provide a succinct overview of previous work addressing sampling
 24 issues for ecological interaction networks. In the second part, I discuss specific ra-
 25 tionales for sampling the biodiversity of ecological interactions. Finally, I provide
 26 a short overview of asymptotic diversity estimates (Gotelli & Colwell, 2001), and
 27 a discussion of its application to interaction sampling. Most of the examples come
 28 from the analysis of plant-animal interaction networks, yet are applicable to other
 29 types of interspecific interactions.

30 Interactions can be a much better indicator of the richness and diversity of
 31 ecosystem functions than a simple list of taxa and their abundances and/or re-
 32 lated biodiversity indicator variables (EBVs) (Memmott *et al.*, 2006; Valiente-
 33 Banuet *et al.*, 2014). Thus, sampling interactions should be a central issue when

identifying and diagnosing ecosystem services (e.g., pollination, seeding by frugivores, etc.). Fortunately, the whole battery of biodiversity-related tools used by ecologists to sample biodiversity (species, *sensu stricto*) can be extended and applied to the sampling of interactions (see Table 2 in Colwell *et al.*, 2004). Monitoring interactions is a type of biodiversity sampling and is subject to similar methodological shortcomings, especially under-sampling (Jordano, 1987; Jordano *et al.*, 2009; Coddington *et al.*, 2009; Vázquez *et al.*, 2009; Dorado *et al.*, 2011; Rivera-Hutinel *et al.*, 2012). For example, when we study mutualistic networks, our goal is to make an inventory of the distinct pairwise interactions that made up the network. We are interested in having a complete list of all the pairwise interactions among species (e.g., all the distinct, species-species interactions, or links, among the pollinators and flowering plants) that do actually exist in a given community. Sampling these interactions thus entails exactly the same problems, limitations, constraints, and potential biases as sampling individual organisms and species diversity. As Mao & Colwell (2005) put it, these are the workings of Preston’s demon, the moving “veil line” (Preston, 1948) between the detected and the undetected interactions as sample size increases.

Early efforts to recognize and solve sampling problems in analyses of interactions stem from research on food webs and to determine how undersampling biases food web metrics (Martinez, 1991; ?, 1993; Bersier *et al.*, 2002; Brose *et al.*, 2003; Banasek-Richter *et al.*, 2004; Wells & O’Hara, 2012). In addition, the myriad of classic natural history studies documenting animal diets, host-pathogen infection records, plant herbivory records, etc., represent efforts to document interactions occurring in nature. All of them share the problem of sampling incompleteness influencing the patterns and metrics reported. Yet, despite the early recogni-

tion that incomplete sampling may seriously bias the analysis of ecological networks (Jordano, 1987), only recent studies have explicitly acknowledged it and attempted to determine its influence (Ollerton & Cranmer, 2002; Nielsen & Bascompte, 2007; Vázquez *et al.*, 2009; Gibson *et al.*, 2011; Olesen *et al.*, 2011; Chacoff *et al.*, 2012; Rivera-Hutinel *et al.*, 2012; Olito & Fox, 2014; Bascompte & Jordano, 2014; Vizentin-Bugoni *et al.*, 2014, 2016; Frund *et al.*, 2015). The sampling approaches have been extended to predict patterns of coextinctions in interaction assemblages (e.g., hosts-parasites) (Colwell *et al.*, 2012). Most empirical studies provide no indication of sampling effort, implicitly assuming that the reported network patterns and metrics are robust. Yet recent evidences point out that number of partner species detected, number of actual links, and some aggregate statistics describing network patterns, are prone to sampling bias (Nielsen & Bascompte, 2007; Dorado *et al.*, 2011; Olesen *et al.*, 2011; Chacoff *et al.*, 2012; Rivera-Hutinel *et al.*, 2012; Olito & Fox, 2014; Frund *et al.*, 2015). Most of these evidences, however, come either from simulation studies (Frund *et al.*, 2015) or from relatively species-poor assemblages. Most certainly, sampling limitations pervade biodiversity inventories in tropical areas (Coddington *et al.*, 2009) and we might rightly expect that frequent interactions may be over-represented and rare interactions may be missed entirely in studies of mega-diverse assemblages (Bascompte & Jordano, 2014); but, to what extent?

Sampling interactions: methods

When we sample interactions in the field we record the presence of two species that interact in some way. For example, Snow and Snow(1988) recorded an inter-

82 action whenever they saw a bird “touching” a fruit on a plant. We observe and
 83 record feeding observations, visitation, occupancy, presence in pollen loads or in
 84 fecal samples, etc., of *individual* animals or plants and accumulate pairwise inter-
 85 actions, i.e., lists of species partners and the frequencies with which we observe
 86 them. We assume that the matrix (species numbers) is predefined (i.e., all species
 87 interacting are well documented). Therefore, estimating the sampling complete-
 88 ness of pairwise interactions for a whole network, requires some gauging of how the
 89 number (richness) of distinct pairwise interactions accumulates as sampling effort
 90 is increased) and/or estimating the uncertainty around the missed links (Wells &
 91 O’Hara, 2012).

92 Most types of ecological interactions can be illustrated with bipartite graphs,
 93 with two or more distinct groups of interacting partners (Bascompte & Jordano,
 94 2014); for illustration purposes I’ll focus more specifically on plant-animal inter-
 95 actions. Sampling interactions requires filling the cells of an interaction matrix
 96 with data. The matrix, $\Delta = AP$ (the adjacency matrix for the graph representa-
 97 tion of the network), is a 2D inventory of the interactions among, say, A animal
 98 species (rows) and P plant species (columns) (Jordano, 1987; Bascompte & Jor-
 99 dano, 2014). The matrix entries illustrate the values of the pairwise interactions
 100 visualized in the Δ matrix, and can be 0 or 1, for presence-absence of a given
 101 pairwise interaction, or take a quantitative weight w_{ji} to represent the interaction
 102 intensity or unidirectional effect of species j on species i (Bascompte & Jordano,
 103 2014; Vazquez *et al.*, 2015). The outcomes of most ecological interactions are
 104 dependent on frequency of encounters (e.g., visit rate of pollinators, number of
 105 records of ant defenders, frequency of seeds in fecal samples). Thus, a frequently
 106 used proxy for interaction intensities w_{ji} is just how frequent new interspecific

encounters are, whether or not appropriately weighted to estimate interaction effectiveness (Vazquez *et al.*, 2005).

We need to define two basic steps in the sampling of interactions: 1) which type of interactions we sample; and 2) which type of record we get to document the existence of an interaction. In step #1 we need to take into account whether we are sampling the whole community of interactor species (all the animals, all the plants) or just a subset of them, i.e., a sub matrix $\Delta_{m,n}$ of $m < A$ animal species and $n < P$ plant species of the adjacency matrix Δ_{AP} (i.e., the matrix representation of interactions among the partner species). Subsets can be: a) all the potential plants interacting with a subset of the animals (Fig. 1a); b) all the potential animal species interacting with a subset of the plant species (Fig. 1b); c) a subset of all the potential animal species interacting with a subset of all the plant species (Fig. 1c). While some discussion has considered how to establish the limits of what represents a network (Strogatz, 2001) (in analogy to discussion on food-web limits; Cohen, 1978), it must be noted that situations a-c in Fig. 1 do not represent complete interaction networks. Subnet sampling is generalized in studies of biological networks (e.g., protein interactions, gene regulation), yet it is important to recognize that most properties of subnetworks (even random subsamples) do not represent properties of whole networks (Stumpf *et al.*, 2005).

In step #2 above we face the problem of the type of record we take to sample interactions. This is important because it defines whether we approach the problem of filling up the interaction matrix in a “zoo-centric” way or in a “phyto-centric” way. Zoo-centric studies directly sample animal activity and document the plants ‘touched’ by the animal. For example, analysis of pollen samples recovered from the body of pollinators, analysis of fecal samples of frugivores, radio-tracking data, etc.

Phyto-centric studies take samples of focal individual plant species and document which animals ‘arrive’ or ‘touch’ the plants. Examples include focal watches of fruiting or flowering plants to record visitation by animals, raising insect herbivores from seed samples, identifying herbivory marks in samples of leaves, etc.

Most recent analyses of plant-animal interaction networks are phyto-centric; just 3.5% of available plant-pollinator ($N=58$) or 36.6% plant-frugivore ($N=22$) interaction datasets are zoo-centric (see Schleuning *et al.*, 2012). Moreover, most available datasets on host-parasite (parasitoid) or plant-herbivore interactions are “host-centric” or phyto-centric (e.g., Thébault & Fontaine, 2010; Morris *et al.*, 2013; Eklöf *et al.*, 2013). This may be related to a variety of causes, like preferred methodologies by researchers working with a particular group or system, logistic limitations, or inherent taxonomic focus of the research questions. A likely result of phyto-centric sampling would be adjacency matrices with large $A : P$ ratios. In contrast, zoo-centric samplings might be prone to detect plants from outside the habitat, complicating the definition of network boundaries. In any case we don’t have a clear view of the potential biases that taxa-focused sampling may generate in observed network patterns, for example by generating consistently asymmetric interaction matrices (Dormann *et al.*, 2009).

Reasonably complete analyses of interaction networks can be obtained when combining both phyto-centric and zoo-centric sampling. For example, Bosch *et al.* (2009) showed that the addition of pollen load data on top of focal-plant sampling of pollinators unveiled a significant number of interactions, resulting in important network structural changes. Olesen *et al.* (2011) identified pollen loads on sampled insects and added the new links to an observation-based visitation matrix, with an extra 5% of links representing the estimated number of missing links

in the pollination network. The overlap between observational and pollen-load recorded links was only 33%, underscoring the value of combining methodological approaches. Zoo-centric sampling has recently been extended with the use of DNA-barcoding, for example with plant-herbivore (Jurado-Rivera *et al.*, 2009), host-parasitoid (Wirta *et al.*, 2014), and plant-frugivore interactions (González-Varo *et al.*, 2014). For mutualistic networks we would expect that zoo-centric sampling could help unveiling interactions of the animals with rare plant species or for relatively common plants species which are difficult to sample by direct observation. Future methodological work may provide significant advances showing how mixing different sampling strategies strengthens the completeness of network data. These mixed strategies may combine, for instance, timed watches at focal plants, spot censuses along walked transects, pollen load or seed contents analyses, monitoring with camera traps, and DNA barcoding records. However, there are no tested protocols and/or sampling designs for ecological interaction studies to suggest an optimum combination of approaches. Ideally, pilot studies would provide adequate information for each specific study setting.

Sampling interactions: rationale

The number of distinct pairwise interactions that we can record in a landscape (an area of relatively homogeneous vegetation) is equivalent to the number of distinct classes in which we can classify the recorded encounters among *individuals* of two different species. Yet, individual-based interaction networks have been only recently studied (Dupont *et al.*, 2011; Wells & O'Hara, 2012). The most usual approach has been to pool individual-based interaction data into species-based

180 summaries, an approach that ignores the fact that only a fraction of individuals
 181 may actually interact given a per capita interaction effect (Wells & O'Hara, 2012).
 182 Wells & O'Hara (2012) illustrate the pros and cons of the approach. We walk in
 183 the forest and see a blackbird Tm picking an ivy Hh fruit and ingesting it: we
 184 have a record for $Tm - Hh$ interaction. We keep advancing and record again a
 185 blackbird feeding on hawthorn Cm fruits so we record a $Tm - Cm$ interaction;
 186 as we advance we encounter another ivy plant and record a blackcap swallowing a
 187 fruit so we now have a new $Sa - Hh$ interaction, and so on. At the end we have
 188 a series of classes (e.g., $Sa - Hh$, $Tm - Hh$, $Tm - Cm$, etc.), along with their
 189 observed frequencies.

190 We get a vector $c = [c_1 \dots c_n]'$ where c_j is the number of classes represented j
 191 times in our sampling: c_1 is the number of singletons (interactions recorded once),
 192 c_2 is the number of twin pairs (interactions with just two records), c_3 the number
 193 of triplets, etc. The problem thus turns to be estimating the number of distinct
 194 classes C from the vector of c_j values and the frequency of unobserved interactions
 195 (see "The real missing links" below).

196 More specifically, we usually obtain a type of reference sample (Chao *et al.*,
 197 2014) for interactions: a series of repeated samples (e.g., observation days, 1h
 198 watches, etc.) with quantitative information, i.e., recording the number of in-
 199 stances of each interaction type on each day. This replicated abundance data,
 200 can be treated in three ways: 1) Abundance data within replicates: the counts
 201 of interactions, separately for each day; 2) Pooled abundance data: the counts of
 202 interactions, summed over all days (the most usual approach); and 3) Replicated
 203 incidence data: the number of days on which we recorded each interaction. As-
 204 suming a reasonable number of replicates, replicated incidence data is considered

to be the most robust statistically, as it takes account of heterogeneity among days (Colwell *et al.*, 2004, 2012; Chao *et al.*, 2014). Thus, both presence-absence and weighted information on interactions can be accommodated for this purpose.

The species assemblage

When we consider an observed and recorded sample of interactions on a particular assemblage of A_{obs} and P_{obs} species (or a set of replicated samples) as a reference sample (Chao *et al.*, 2014) we may have three sources of undersampling error. These sources are ignored if we treat the reference sample as a true representation of the interactions in a well-defined assemblage: 1) some animal species are actually present but not observed (zero abundance or incidence in the interactions in the reference sample), A_0 ; 2) some plant species are actually present but not observed (zero abundance or incidence in the interactions in the reference sample), P_0 ; 3) some unobserved links (the zeroes in the adjacency matrix, UL) may actually occur but not recorded. Thus a first problem is determining if A_{obs} and P_{obs} truly represent the actual species richness interacting in the assemblage. To this end we might use the replicated reference samples to estimate the true number of interacting animal A_{est} and plant P_{est} species as in traditional diversity estimation analysis (Chao *et al.*, 2014). If there are no uniques (species seen on only one day), then A_0 and P_0 will be zero (based on the Chao2 formula), and we have A_{obs} and P_{obs} as robust estimates of the actual species richness of the assemblage. If A_0 and P_0 are not zero they estimate the minimum number of undetected animal and plant species that can be expected with a sufficiently large number of replicates, taken from the same assemblage/locality by the same methods in the same time

228 period. We can use extrapolation methods (Colwell *et al.*, 2012) to estimate how
 229 many additional replicate surveys it would take to reach a specified proportion g
 230 of A_{est} and P_{est} .

231 The interactions

232 We are then faced with assessing the sampling of interactions I . Table 1 summa-
 233 rizes the main components and targets for estimation of interaction richness. In
 234 contrast with traditional species diversity estimates, sampling networks has the
 235 paradox that despite the potentially interacting species being present in the sam-
 236 pled assemblage (i.e., included in the A_{obs} and P_{obs} species lists), some of their
 237 pairwise interactions are impossible to record. The reason is forbidden links. In-
 238 dependently of whether we sample full communities or subset communities we face
 239 a problem: some of the interactions that we can visualize in the empty adjacency
 240 matrix Δ will simply not occur. With a total of $A_{obs}P_{obs}$ “potential” interactions
 241 (eventually augmented to $A_{est}P_{est}$ in case we have undetected species), a fraction
 242 of them are impossible to record, because they are forbidden (Jordano *et al.*, 2003;
 243 Olesen *et al.*, 2011).

244 Our goal is to estimate the true number of non-null AP interactions, including
 245 interactions that actually occur but have not been observed (I_0) from the repli-
 246 cated incidence frequencies of interaction types: $I_{est} = I_{obs} + I_0$. Note that I_0
 247 estimates the minimum number of undetected plant-animal interactions that can
 248 be expected with a sufficiently large number of replicates, taken from the same
 249 assemblage/locality by the same methods in the same time period. Therefore we
 250 have two types of non-observed links: UL^* and UL , corresponding to the real

251 assemblage species richness and to the observed assemblage species richness, re-
 252 spectively (Table 1).

253 Forbidden links are non-occurrences of pairwise interactions that can be ac-
 254 counted for by biological constraints, such as spatio-temporal uncoupling (Jor-
 255 dano, 1987), size or reward mismatching, foraging constraints (e.g., accessibility)
 256 (Moré *et al.*, 2012), and physiological-biochemical constraints (Jordano, 1987). We
 257 still have very little information about the frequency of forbidden links in natural
 258 communities (Jordano *et al.*, 2003; Stang *et al.*, 2009; Vázquez *et al.*, 2009; Olesen
 259 *et al.*, 2011; Ibanez, 2012; Maruyama *et al.*, 2014; Vizentin-Bugoni *et al.*, 2014)
 260 (Table 1). Forbidden links are thus represented as structural zeroes in the inter-
 261 action matrix, i.e., matrix cells that cannot get a non-zero value. Therefore, we
 262 need to account for the frequency of these structural zeros in our matrix before
 263 proceeding.

264 Our main problem then turns to estimate the number of true missed links,
 265 i.e., those that can't be accounted for by biological constraints and that might
 266 suggest undersampling. Thus, the sampling of interactions in nature, as the sam-
 267 pling of species, is a cumulative process. In our analysis, we are not re-sampling
 268 individuals, but interactions, so we made interaction-based accumulation curves.
 269 If an interaction-based curve suggests a robust sampling, it does mean that no
 270 new interactions are likely to be recorded, irrespectively of the species, as it is a
 271 whole-network sampling approach (N. Gotelli, pers. com.). We add new, distinct,
 272 interactions recorded as we increase sampling effort (Fig. 2). We can obtain an
 273 Interaction Accumulation Curve (*IAC*) analogous to a Species Curve (*SAC*) (see
 274 Supporting Information in the online data availability repository): the observed
 275 number of distinct pairwise interactions in a survey or collection as a function of

the accumulated number of observations or samples (Colwell, 2009).

Empirical data on Forbidden Links

Adjacency matrices are frequently sparse, i.e., they are densely populated with zeroes, with a fraction of them being structural (unobservable interactions) (Bascompte & Jordano, 2014). Thus, it would be a serious interpretation error to attribute the sparseness of adjacency matrices for bipartite networks to just the result of undersampling. The actual typology of link types in ecological interaction networks is thus more complex than just the two categories of observed and unobserved interactions (Table 1). Unobserved interactions are represented by zeroes and belong to two categories. Missing interactions may actually exist but require additional sampling or a variety of methods to be observed. Forbidden links, on the other hand, arise due to biological constraints limiting interactions and remain unobservable in nature, irrespectively of sampling effort (Table 1). Forbidden links FL may actually account for a relatively large fraction of unobserved interactions UL when sampling taxonomically-restricted subnetworks (e.g., plant-hummingbird pollination networks) (Table 1). Phenological uncoupling is also prevalent in most networks, and may add up to explain ca. 25-40% of the forbidden links, especially in highly seasonal habitats, and up to 20% when estimated relative to the total number of unobserved interactions (Table 2). In any case, we might expect that a fraction of the missing links ML would be eventually explained by further biological reasons, depending on the knowledge of natural details of the particular systems. Our goal as naturalists would be to reduce the fraction of UL which remain as missing links; to this end we might search for additional biological

299 constraints or increase sampling effort. For instance, habitat use patterns by hum-
 300 mingbirds in the Arima Valley network (Table 2; Snow & Snow, 1972) impose a
 301 marked pattern of microhabitat mismatches causing up to 44.5% of the forbidden
 302 links. A myriad of biological causes beyond those included as *FL* in Table 1 may
 303 contribute explanations for *UL*: limits of color perception, presence of secondary
 304 metabolites in fruit pulp and leaves, toxins and combinations of monosaccharides
 305 in nectar, etc. For example, aside from *FL*, some pairwise interactions may sim-
 306 ply have an asymptotically-zero probability of interspecific encounter between the
 307 partner species, if they are very rare. However, it is surprising that just the limited
 308 set of forbidden link types considered in Table 1 explain between 24.6-77.2% of
 309 the unobserved links. Notably, the Arima Valley, Santa Virgínia, and Hato Ratón
 310 networks have $> 60\%$ of the unobserved links explained, which might be related
 311 to the fact that they are subnetworks (Arima Valley, Santa Virgínia) or relatively
 312 small networks (Hato Ratón). All this means that empirical networks may have
 313 sizable fractions of structural zeroes. Ignoring this biological fact may contribute
 314 to wrongly inferring undersampling of interactions in real-world assemblages.

315 To sum up, two elements of inference are required in the analysis of unobserved
 316 interactions in ecological interaction networks: first, detailed natural history infor-
 317 mation on the participant species that allows the inference of biological constraints
 318 imposing forbidden links, so that structural zeroes can be identified in the adja-
 319 cency matrix. Second, a critical analysis of sampling robustness and a robust
 320 estimate of the actual fraction of missing links, M , resulting in a robust estimate
 321 of I . In the next sections we explore these elements of inference, using *IACs* as
 322 analogs to *SACs* to assess the robustness of interaction sampling.

Asymptotic diversity estimates

Let's assume a sampling of the diversity in a specific locality, over relatively homogeneous landscape where we aim at determining the number of species present for a particular group of organisms. To do that we carry out transects or plot samplings across the landscape or use any other type of direct or indirect recording method, adequately replicated so we obtain a number of samples. Briefly, S_{obs} is the total number of species observed in a sample, or in a set of samples. S_{est} is the estimated number of species in the community represented by the sample, or by the set of samples, where *est* indicates an estimator. With abundance data, let S_k be the number of species each represented by exactly k individuals in a single sample. Thus, S_0 is the number of undetected species (species present in the community but not included in the sample), S_1 is the number of singleton species (represented by just one individual), S_2 is the number of doubleton species (species with two individuals), etc. The total number of individuals in the sample would be:

$$n = \sum_{k=1}^{S_{obs}} S_k$$

A frequently used asymptotic, bias corrected, non-parametric estimator is S_{Chao1} (Hortal *et al.*, 2006; Chao, 2005; Colwell, 2013):

$$S_{Chao1} = S_{obs} + \frac{S_1(S_1 - 1)}{2(S_2 + 1)}$$

Another frequently used alternative is the Chao2 estimator, S_{Chao2} (Gotelli &

Colwell, 2001), which has been reported to have a limited bias for small sample sizes (Colwell & Coddington, 1994; Chao, 2005). Instead of using counts it uses incidence frequencies (Q_k) among samples (number of species present in just one sample, in two samples, etc.):

$$S_{Chao2} = S_{obs} + \frac{Q_1(Q_1 - 1)}{2(Q_2 + 1)}$$

A plot of the cumulative number of species recorded, S_n , as a function of some measure of sampling effort (say, n samples taken) yields the species accumulation curve (SAC) or collector's curve (Colwell & Coddington, 1994). Similarly, interaction accumulation curves (IAC), analogous to SACs, can be used to assess the robustness of interactions sampling for plant-animal community datasets (Jordano, 1987; Jordano *et al.*, 2009; Olesen *et al.*, 2011), as discussed in the next section.

Assessing sampling effort when recording interactions

The basic method we can propose to estimate sampling effort and explicitly show the analogues with rarefaction analysis in biodiversity research is to vectorize the interaction matrix AP so that we get a vector of all the potential pairwise interactions (I_{max} , Table 1) that can occur in the observed assemblage with A_{obs} animal species and P_{obs} plant species. The new “species” we aim to sample are the pairwise interactions (Table 3). So, if we have in our community *Turdus merula* (Tm) and *Rosa canina* (Rc) and *Prunus mahaleb* (Pm), our problem will be to sample 2 new “species”: $Tm - Rc$ and $Tm - Pm$. In general, if we have $A = 1...i$, animal species

362 and $P = 1 \dots j$ plant species (assuming a complete list of species in the assemblage),
 363 we'll have a vector of "new" species to sample: $A_1P_1, A_1P_2, \dots A_2P_1, A_2P_2, \dots A_iP_j$.
 364 We can represent the successive samples where we can potentially get records of
 365 these interactions in a matrix with the vectorized interaction matrix and columns
 366 representing the successive samples we take (Table 3). This is simply a vectorized
 367 version of the interaction matrix. This is analogous to a biodiversity sampling
 368 matrix with species as rows and sampling units (e.g., quadrats) as columns (Jor-
 369 dano *et al.*, 2009). The package *EstimateS* (Colwell, 2013) includes a complete set
 370 of functions for estimating the mean IAC and its unconditional standard devia-
 371 tion from random permutations of the data, or subsampling without replacement
 372 (Gotelli & Colwell, 2001); it further reports asymptotic estimators for the expected
 373 number of distinct pairwise interactions included in a given reference sample of
 374 interaction records (see also the `specaccum` function in library `vegan` of the R
 375 Package)(R Development Core Team, 2010; Jordano *et al.*, 2009; Olesen *et al.*,
 376 2011). In particular, we may take advantage of replicated incidence data, as it
 377 takes account of heterogeneity among samples (days, censuses, etc.; R.K Colwell,
 378 pers. comm.) (see also Colwell *et al.*, 2004, 2012; Chao *et al.*, 2014).

379 In this way we effectively extend sampling theory developed for species diversity
 380 to the sampling of ecological interactions. Yet future theoretical work will be
 381 needed to formally assess the similarities and differences in the two approaches
 382 and developing biologically meaningful null models of expected interaction richness
 383 with added sampling effort.

384 Diversity-accumulation analysis (Magurran, 1988; Hortal *et al.*, 2006) comes
 385 up immediately with this type of dataset. This procedure plots the accumulation
 386 curve for the expected number of distinct pairwise interactions recorded with in-

creasing sampling effort (Jordano *et al.*, 2009; Olesen *et al.*, 2011). Asymptotic estimates of interaction richness and its associated standard errors and confidence intervals can thus be obtained (Hortal *et al.*, 2006) (see Table 4 and Supplementary Online Material). The characteristic feature of interaction datasets is that, due to forbidden links, a number of pairwise interactions among the I_{max} number specified in the Δ adjacency matrix cannot be recorded, irrespective of sampling effort.

We may expect undersampling specially in moderate to large sized networks with multiple modules (i.e., species subsets requiring different sampling strategies) (Jordano, 1987; Olesen *et al.*, 2011; Chacoff *et al.*, 2012); adequate sampling may be feasible when interaction subwebs are studied (Olesen *et al.*, 2011; Vizentin-Bugoni *et al.*, 2014), typically with more homogeneous subsets of species (e.g., bumblebee-pollinated flowers). In any case the sparseness of the Δ matrix is by no means an indication of undersampling whenever the issue of structural zeroes in the interaction matrices is effectively incorporated in the estimates.

For example, mixture models incorporating detectabilities have been proposed to effectively account for rare species (Mao & Colwell, 2005). In an analogous line, mixture models could be extended to samples of pairwise interactions, also with specific detectability values. These detection rate/odds could be variable among groups of interactions, depending on their specific detectability. For example, detectability of flower-pollinator interactions involving bumblebees could have a higher detectability than flower-pollinator pairwise interactions involving, say, nitidulid beetles. These more homogeneous groupings of pairwise interactions within a network define modules (Bascompte & Jordano, 2014), so we might expect that interactions of a given module (e.g., plants and their hummingbird pollinators; Fig.

1a) may share similar detectability values, in an analogous way to species groups
 receiving homogeneous detectability values in mixture models (Mao & Colwell,
 2005). In its simplest form, this would result in a sample with multiple pairwise
 interactions detected, in which the number of interaction events recorded for each
 distinct interaction found in the sample is recorded (i.e., a column vector in Table
 3, corresponding to, say, a sampling day). The number of interactions recorded for
 the i_{th} pairwise interaction (i.e., $A_i P_j$ in Table 3), Y_i could be treated as a Poisson
 random variable with a mean parameter λ_i , its detection rate. Mixture models
 (Mao & Colwell, 2005) include estimates for abundance-based data (their analogs
 in interaction sampling would be weighted data), where Y_i is a Poisson random
 variable with detection rate λ_i . This is combined with the incidence-based model,
 where Y_i is a binomial random variable (their analogous in interaction sampling
 would be presence/absence records of interactions) with detection odds λ_i . Let
 T be the number of samples in an incidence-based data set. A Poisson/binomial
 density can be written as (Mao & Colwell, 2005):

$$g(y; \lambda) = \begin{cases} \frac{\lambda^y}{y!e^\lambda} & [1] \\ \binom{T}{y} \frac{\lambda^y}{(1+\lambda)^T} & [2] \end{cases}$$

where [1] corresponds to a weighted network, and [2] to a qualitative network.

The detection rates λ_i depend on the relative abundances ϕ_i of the interactions,
 the probability of a pairwise interaction being detected when it is present, and the
 sample size (the number of interactions recorded), which, in turn, is a function
 of the sampling effort. Unfortunately, no specific sampling model has been devel-
 oped along these lines for species interactions and their characteristic features. For

example, a complication factor might be that interaction abundances, ϕ_i , in real assemblages are a function of the abundances of interacting species that determine interspecific encounter rates; yet they also depend on biological factors that ultimately determine if the interaction occurs when the partner species are present. For example, λ_i should be set to zero for all FL . In its simplest form, ϕ_i could be estimated from just the product of partner species abundances, an approach recently used as a null model to assess the role of biological constraints in generating forbidden links and explaining interaction patterns (Vizentin-Bugoni *et al.*, 2014). Yet more complex models (e.g., Wells & O'hara 2012) should incorporate not only interspecific encounter probabilities, but also interaction detectabilities, phenotypic matching and incidence of forbidden links. Mixture models are certainly complex and for most situations of evaluating sampling effort better alternatives include the simpler incidence-based rarefaction and extrapolation (Colwell *et al.*, 2012; Chao *et al.*, 2014).

The *real* missing links

Given that a fraction of unobserved interactions can be accounted for by forbidden links, what about the remaining missing interactions? We have already discussed that some of these could still be related to unaccounted constraints, and still others would be certainly attributable to insufficient sampling. Would this always be the case? A crucial ecological aspect limiting interactions within multispecific assemblages of distinct taxonomic relatedness (Fig. 2) is the probability of interspecific encounter, i.e., the probability that two individuals of the partner species actually encounter each other in nature.

456 Given log-normally distributed abundances of the two species groups, the ex-
 457 pected probabilities of interspecific encounter (PIE) would be simply the product
 458 of the two lognormal distributions. Thus, we might expect that for very low PIE
 459 values, pairwise interactions would be either extremely difficult to sample, or sim-
 460 ply do not occur in nature. Consider the Nava de las Correhuelas interaction
 461 web (NCH, Table 2, 4), with $A = 36$, $P = 25$, $I = 181$, and almost half of the
 462 unobserved interactions not accounted for by forbidden links, thus $M = 53.1\%$
 463 (Jordano *et al.*, 2009). A sizable fraction of these possible but missing links would
 464 be simply not occurring in nature, most likely due to extremely low PIE , in fact
 465 asymptotically zero. Given the vectorized list of pairwise interactions for NCH, I
 466 computed the PIE values for each one by multiplying element-wise the two species
 467 abundance distributions. The $PIE_{max} = 0.0597$, being a neutral estimate, based
 468 on the assumption that interactions occur in proportion to the species-specific local
 469 abundances. With $PIE_{median} < 1.4 \cdot 10^{-4}$ we may safely expect (note the quantile
 470 estimate $Q_{75\%} = 3.27 \cdot 10^{-4}$) that a sizable fraction of these missing interactions
 471 may not occur according to this neutral expectation (Jordano, 1987; Olesen *et al.*,
 472 2011) (neutral forbidden links, *sensu* Canard *et al.*, 2012).

473 When we consider the vectorized interaction matrix, enumerating all pairwise
 474 interactions for the AP combinations, the expected probabilities of finding a given
 475 interaction can be estimated with a Good-Turing approximation (Good, 1953).
 476 The technique, developed by Alan Turing and I.J. Good with applications to lin-
 477 guistics and word analysis (Gale & Sampson, 1995) has been recently extended in
 478 novel ways for ecological analyses (Chao *et al.*, 2015). It estimates the probability
 479 of recording an interaction of a hitherto unseen pair of partners, given a set of past
 480 records of interactions between other species pairs. Let a sample of N interactions

so that n_r distinct pairwise interactions have exactly r records. All Good-Turing estimators obtain the underlying frequencies of events as:

$$P(X) = \frac{(N_X + 1)}{T} \left(1 - \frac{E(1)}{T}\right) \quad (1)$$

where X is the pairwise interaction, N_X is the number of times interaction X is recorded, T is the sample size (number of distinct interactions recorded) and $E(1)$ is an estimate of how many different interactions were recorded exactly once. Strictly speaking Equation (1) gives the probability that the next interaction type recorded will be X , after sampling a given assemblage of interacting species. In other words, we scale down the maximum-likelihood estimator $\frac{n}{T}$ by a factor of $\frac{1-E(1)}{T}$. This reduces all the probabilities for interactions we have recorded, and makes room for interactions we haven't seen. If we sum over the interactions we have seen, then the sum of $P(X)$ is $1 - \frac{1-E(1)}{T}$. Because probabilities sum to one, we have the left-over probability of $P_{new} = \frac{E(1)}{T}$ of seeing something new, where new means that we sample a new pairwise interaction.

Discussion

Recent work has inferred that most data available for interaction networks are incomplete due to undersampling, resulting in a variety of biased parameters and network patterns (Chacoff *et al.*, 2012). It is important to note, however, that in practice, most surveyed networks to date have been subnets of much larger networks. This is also true for protein interaction, gene regulation, and metabolic networks, where only a subset of the molecular entities in a cell have been sam-

501 pled (Stumpf *et al.*, 2005). Despite recent attempts to document whole ecosystem
 502 meta-networks (Pocock *et al.*, 2012), it is likely that most ecological interaction
 503 networks will illustrate just major ecosystem compartments. Due to their high
 504 generalization, high temporal and spatial turnover, and high complexity of associ-
 505 ation patterns, adequate sampling of ecological interaction networks is challenging
 506 and requires extremely large sampling effort. Undersampling of ecological net-
 507 works may originate from the analysis of assemblage subsets (e.g., taxonomically
 508 or functionally defined), and/or from logistically-limited sampling effort. It is ex-
 509 tremely hard to robustly sample the set of biotic interactions even for relatively
 510 simple, species-poor assemblages; thus, we need to assess how robust is the char-
 511 acterization of the adjacency matrix Δ . Concluding that an ecological network
 512 dataset is undersampled just by its sparseness would be unrealistic. The reason
 513 stems from a biological fact: a sizeable fraction of the maximum, potential links
 514 that can be recorded among two distinct sets of species is simply unobservable,
 515 irrespective of sampling effort (Jordano, 1987). In addition, sampling effort needs
 516 to be explicitly gauged because of its potential influence on parameter estimates
 517 for the network.

518 Missing links are a characteristic feature of all plant-animal interaction net-
 519 works, and likely pervade other ecological interactions. Important natural history
 520 details explain a fraction of them, resulting in unrealizable interactions (i.e., for-
 521 bidden interactions) that define structural zeroes in the interaction matrices and
 522 contribute to their extreme sparseness. Sampling interactions is a way to monitor
 523 biodiversity beyond the simple enumeration of component species and to develop
 524 efficient and robust inventories of functional interactions. Yet no sampling theory
 525 for interactions is available. Focusing just on the realized interactions or treating

missing interactions as the expected unique result of sampling bias would miss important components to understand how all sorts of interactions coevolve within complex webs of interdependence among species.

Contrary to species inventories, a sizable fraction of non-observed pairwise interactions cannot be sampled, due to biological constraints that forbid their occurrence. Moreover, recent implementations of inference methods for unobserved species (Chao *et al.*, 2015) or for individual-based data (Wells & O’Hara, 2012) can be combined with the forbidden link approach. They do not account either for the existence of these ecological constraints, but can help in estimating their relative importance, simply by the difference between the asymptotic estimate of interaction richness *in a robustly-sampled* assemblage and the maximum richness I_{max} of interactions.

Ecological interactions provide the wireframe supporting the lives of species, and they also embed crucial ecosystem functions which are fundamental for supporting the Earth system. We still have a limited knowledge of the biodiversity of ecological interactions, and they are being lost (extinct) at a very fast pace, frequently preceding species extinctions (Valiente-Banuet *et al.*, 2014). We urgently need robust techniques to assess the completeness of ecological interactions networks because this knowledge will allow the identification of the minimal components of their ecological complexity that need to be restored to rebuild functional ecosystems after perturbations.

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Data accessibility

This review does not use new raw data, but includes some re-analyses of previously published material. All the original data supporting the paper, R code, supplementary figures, and summaries of analytical protocols is available at the author's GitHub repository (https://github.com/pedroj/MS_Network-Sampling), with DOI: 10.5281/zenodo.29437.

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Figure captions

Figure 1. Sampling ecological interaction networks (e.g., plant-animal interactions) usually focus on different types of subsampling the full network, yielding submatrices $\Delta[m, n]$ of the full interaction matrix Δ with A and P animal and plant species. a) all the potential plants interacting with a subset of the animals (e.g., studying just the hummingbird-pollinated flower species in a community); b) all the potential animal species interacting with a subset of the plant species (e.g., studying the frugivore species feeding on figs *Ficus* in a community); and c) sampling a subset of all the potential animal species interacting with a subset of all the plant species (e.g., studying the plant-frugivore interactions of the rainforest understory).

Figure 2. Sampling species interactions in natural communities. Suppose an assemblage with $A = 3$ animal species (red, species 1–3 with three, two, and 1 individuals, respectively) and $P = 3$ plant species (green, species a-c with three individuals each) (colored balls), sampled with increasing effort in steps 1 to 6 (panels). In Step 1 we record animal species 1 and plant species 1 and 2 with a total of three interactions (black lines) represented as two distinct interactions: 1 – a and 1 – b. As we advance our sampling (panels 1 to 6, illustrating e.g., additional sampling days) we record new distinct interactions. Note that we actually sample and record interactions among individuals, yet we pool the data across species to get a species by species interaction matrix. Few network analyses have been carried out on individual data (Dupont *et al.*, 2014). Above and below each panel are the cumulative number of distinct species and interactions sampled, so

785 that panel 6 illustrates the final network.

786

787 **Figures**

Figure 1:

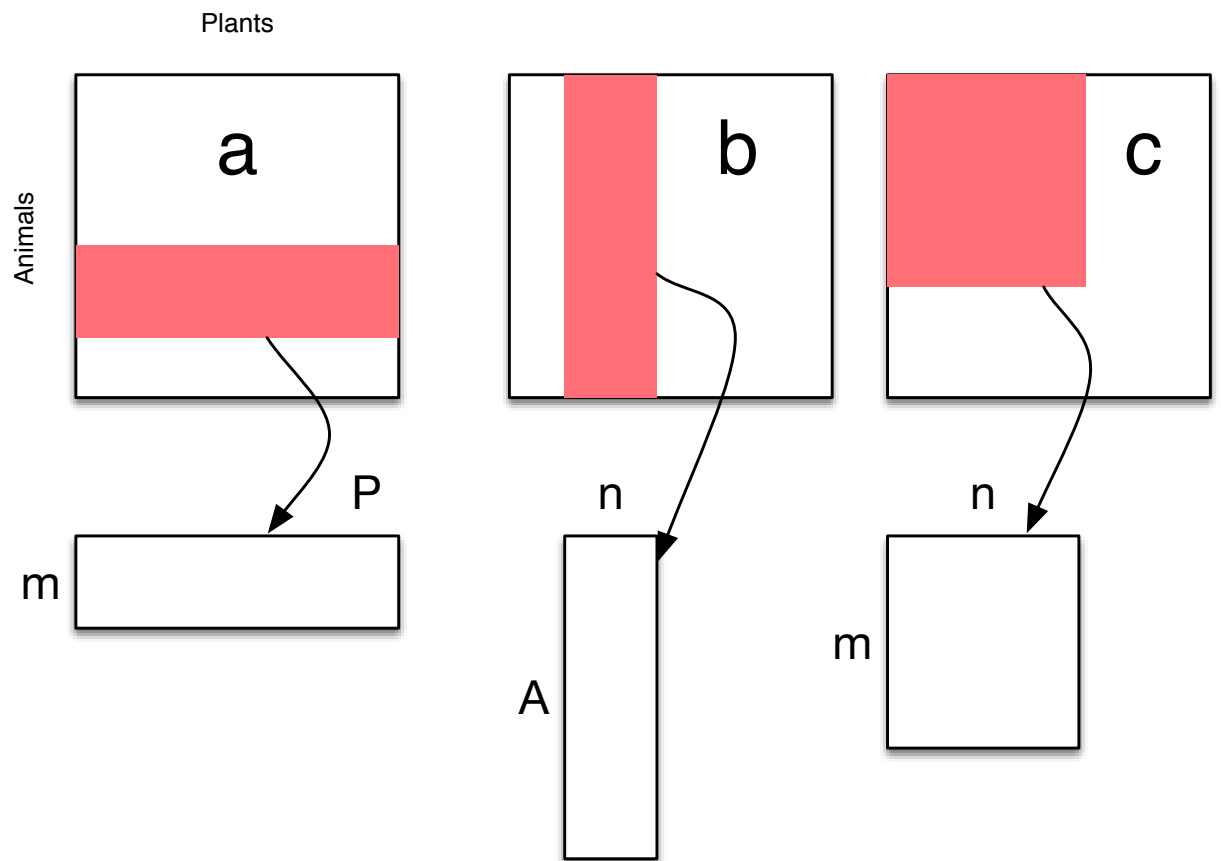
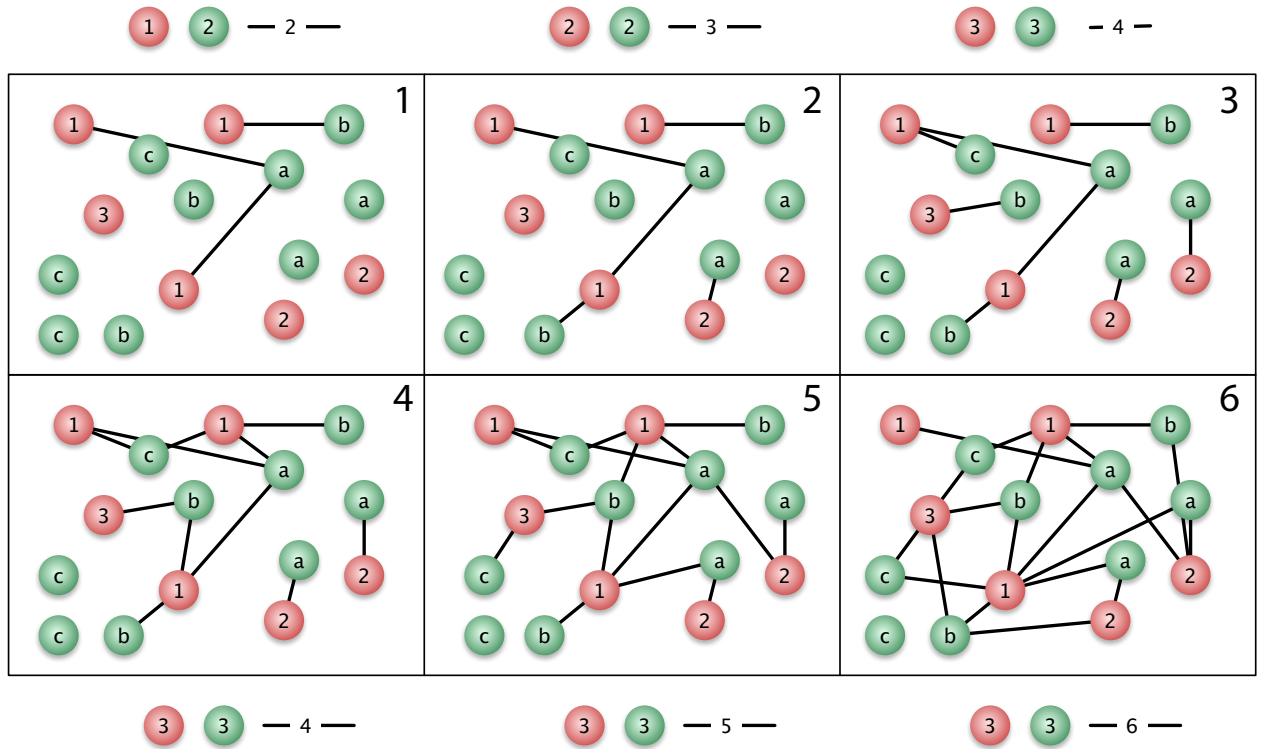


Figure 2:



Jordano - Figure 1

Table captions

Table 1. A taxonomy of link types for ecological interactions (Olesen *et al.* 2011). A , number of animal species; P , number of plant species; I , number of observed links; $C = 100I/(AP)$, connectance; FL , number of forbidden links; and ML , number of missing links. As natural scientists, our ultimate goal is to eliminate ML from the equation $FL = AP - I - ML$, which probably is not feasible given logistic sampling limitations. When we, during our study, estimate ML to be negligible, we cease observing and estimate I and FL .

Table 2. Frequencies of different type of forbidden links in natural plant-animal interaction assemblages. AP , maximum potential links, I_{max} ; I , number of observed links; UL , number of unobserved links; FL , number of forbidden links; FL_P , phenology; FL_S , size restrictions; FL_A , accessibility; FL_O , other types of restrictions; ML , unknown causes (missing links). Relative frequencies (in parentheses) calculated over $I_{max} = AP$ for I , ML , and FL ; for all forbidden links types, calculated over FL . References, from left to right: Olesen *et al.* 2008; Olesen & Myrthue unpubl.; Snow & Snow 1972 and Jordano *et al.* 2006; Vizentin-Bugoni *et al.* 2014; Jordano *et al.* 2009; Olesen *et al.* 2011.

Table 3. A vectorized interaction matrix.

Table 4. Sampling statistics for three plant-animal interaction networks (Olesen *et al.* 2011). Symbols as in Table 1; N , number of records; $Chao1$ and ACE are asymptotic estimators for the number of distinct pairwise interactions I (Hortal

812 *et al.* 2006), and their standard errors; C , sample coverage for rare interactions
813 (Chao & Jost 2012). Scaled asymptotic estimators and their confidence intervals
814 (CI) were calculated by weighting *Chao1* and *ACE* with the observed frequencies
815 of forbidden links.

816

817 Tables

Table 1:

Link type	Formulation	Definition
Potential links	$I_{max} = A_{obs}P_{obs}$	Size of observed network matrix, i.e. maximum number of potentially observable interactions; A_{obs} and P_{obs} , numbers of interacting animal and plant species, respectively. These might be below the real numbers of animal and plant species, A_{est} and P_{est} .
Observed links	I_{obs}	Total number of observed links in the network given a sufficient sampling effort. Number of ones in the adjacency matrix.
True links	I_{est}	Total number of links in the network given a sufficient sampling effort; expected for the augmented $A_{est}P_{est}$ matrix.
Unobserved links	$UL = I_{max} - I_{obs}$	Number of zeroes in the adjacency matrix.
True unobserved links	$UL* = I_{max} - I_{obs}$	Number of zeroes in the augmented adjacency matrix that, eventually, includes unobserved species.
Forbidden links	FL	Number of links, which remain unobserved because of linkage constraints, irrespectively of sufficient sampling effort.
Observed Missing links	$ML = A_{obs}P_{obs} - I_{obs} - FL$	Number of links, which may exist in nature but need more sampling effort and/or additional sampling methods to be observed.
True Missing links	$ML* = A_{est}P_{est} - I_{est} - FL$	Number of links, which may exist in nature but need more sampling effort and/or additional sampling methods to be observed. Augments ML for the $A_{est}P_{est}$ matrix.

Table 2:

Link type	Pollination			Seed dispersal		
	Zackenberg	Grundvad	Arima Valley	Sta. Virginia	Hato Ratón	Nava Correhuelas
I_{max}	1891	646	522	423	272	825
I	268 (0.1417)	212 (0.3282)	185 (0.3544)	86 (0.1042)	151 (0.4719)	181 (0.2194)
UL	1507 (0.7969)	434 (0.6718)	337 (0.6456)	337 (0.4085)	169 (0.5281)	644 (0.7806)
FL	530 (0.3517)	107 (0.2465)	218 (0.6469)	260 (0.7715)	118 (0.6982)	302 (0.4689)
FL_P	530 (1.0000)	94 (0.2166)	0 (0.0000)	120 (0.1624)	67 (0.3964)	195 (0.3028)
FL_S	... (...)	8 (0.0184)	30 (0.0890)	140 (0.1894)	31 (0.1834)	46 (0.0714)
FL_A	... (...)	5 (0.0115)	150 (0.445) ^a	... (...)	20 (0.1183)	61 (0.0947)
FL_O	... (...)	... (...)	38 (0.1128) ^b	... (...)	... (...)	363 (0.5637)
ML	977 (0.6483)	327 (0.7535)	119 (0.3531)	77 (0.1042)	51 (0.3018)	342 (0.5311)

^a, Lack of accessibility due to habitat uncoupling, i.e., canopy-foraging species vs. understory species.

^b, Colour restrictions, and reward per flower too small relative to the size of the bird.

Dots indicate no data available for the FL type.

Table 3:

Interaction	Sample 1	Sample 2	Sample 3	...	Sample i
A1 - P1	12	2	0	...	6
A1 - P2	0	0	0	...	1
...
A5 - P3	5	0	1	...	18
A5 - P4	1	0	1	...	3
...
A _i - P _i	1	0	1	...	2

Table 4:

	Hato Ratón	Nava Correhuelas	Zackenberg
A	17	33	65
P	16	25	31
I_{max}	272	825	1891
N	3340	8378	1245
I	151	181	268
C	0.917	0.886	0.707
$Chao1$	263.1 ± 70.9	231.4 ± 14.2	509.6 ± 54.7
ACE	240.3 ± 8.9	241.3 ± 7.9	566.1 ± 14.8
% <i>unobserved</i> ^a	8.33	15.38	47.80

^a, estimated with library Jade (R Core Development Team 2010, Chao *et al.* 2015)