# Class 09: Structural Bioinformatics

Max Gruber

5/3/23

### 1: Introduction to the RCSB Protein Data Bank (PDB)

To read the file, we are going to use the command read.csv.

```
pdb_stats <- read.csv('Data Export Summary.csv', row.names = 1)</pre>
```

# Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy?

I need to sum all the elements of the X.ray column.

```
pdb_stats$X.ray
[1] "154,766" "9,083" "8,110" "2,664" "163" "11"
```

We are going to use gsub to remove the commas.

```
xray_without_commas <- gsub(',',"",pdb_stats$X.ray)
as.numeric(xray_without_commas)</pre>
```

[1] 154766 9083 8110 2664 163 11

```
em_without_commas <- gsub(',',"",pdb_stats$EM)
as.numeric(em_without_commas)</pre>
```

[1] 10155 1802 3176 94 9 0

```
total_without_commas <- gsub(',',"",pdb_stats$Total)</pre>
  as.numeric(total_without_commas)
[1] 177403 10925 11575
                            4223 204
                                                22
I use the sum command to get the sum.
  n_xray <- sum(as.numeric(xray_without_commas))</pre>
  n_xray
[1] 174797
  n_em <- sum(as.numeric(em_without_commas))</pre>
  n_em
[1] 15236
  n_total <- sum(as.numeric(total_without_commas))</pre>
  n_total
[1] 204352
Now I can find the percentage.
  p_xray <- (n_xray)/ n_total</pre>
  p_xray
[1] 0.8553721
  p_{em} \leftarrow (n_{em})/n_{total}
  p_em
```

[1] 0.07455763

```
((n_xray + n_em) / n_total)*100
[1] 92.99297
92.99%
```

Q2: What proportion of structures in the PDB are protein?

```
protein_without_commas <- gsub(',',"",pdb_stats[1,7])
as.numeric(protein_without_commas)

[1] 177403

n_protein <- sum(as.numeric(protein_without_commas))
n_protein/n_total

[1] 0.8681246
.8681, or 86.81%, are protein.</pre>
```

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

1,292 was one possible answer. Overall, it was difficult to search/find an exact value for the specific criteria because different values came up depending on how and where search criteria was entered.

## 2. Visualizing the HIV-1 protease structure

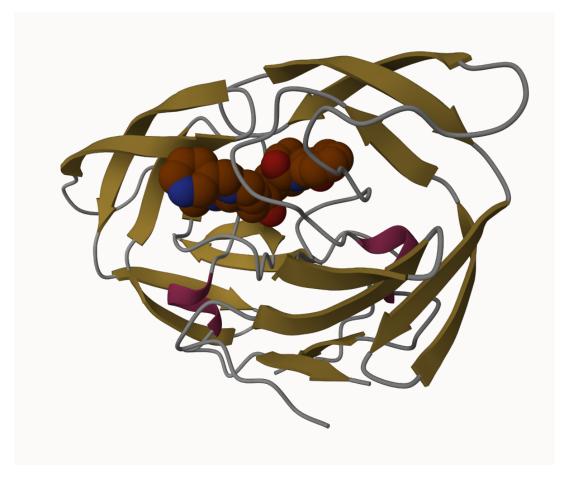
Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

We see one because in x-ray crystallography it's hard to detect hydrogen (as its density is so small), and so in the structures only oxygen is seen (one atom) because it's simply easier to see.

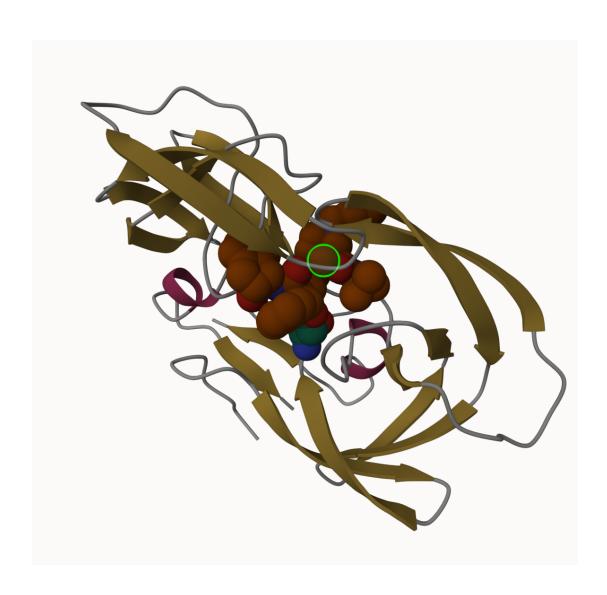
Q5: There is a critical "conserved" water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have?

Yes, you can identify the water molecule, and it has a residue number of 308 (HOH 308).

Q6: Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain and the critical water (we recommend "Ball & Stick" for these side-chains). Add this figure to your Quarto document.



And a second image with the catalytic residues ASP 25 in each chain and the critical water (highlighted in green).



# 3. Introduction to Bio3D in R

```
library(bio3d)
pdb <- read.pdb("1hsg")

Note: Accessing on-line PDB file
pdb</pre>
```

+ attr: atom, xyz, seqres, helix, sheet, calpha, remark, call

#### Q7: How many amino acid residues are there in this pdb object?

There are 198 amino acid residues in this pdb object.

#### Q8: Name one of the two non-protein residues?

One of the two non-protein residues is HOH, or water.

#### Q9: How many protein chains are in this structure?

There are 2 protein chains in this structure.

#### head(pdb\$atom)

```
type eleno elety alt resid chain resno insert
                                                                 z o
                                                    Х
1 ATOM
                N < NA >
                                          <NA> 29.361 39.686 5.862 1 38.10
          1
                         PRO
                                 Α
                                       1
2 ATOM
          2
                         PRO
                                      1 <NA> 30.307 38.663 5.319 1 40.62
               CA <NA>
                                 Α
3 ATOM
                C <NA>
                         PRO
                                       1 <NA> 29.760 38.071 4.022 1 42.64
          3
                                 Α
                                      1 <NA> 28.600 38.302 3.676 1 43.40
4 ATOM
          4
                O <NA>
                         PRO
                                 Α
```

```
5 ATOM
          5
              CB <NA>
                        PRO
                                      1 <NA> 30.508 37.541 6.342 1 37.87
                                Α
6 ATOM
               CG <NA>
                        PRO
                                         <NA> 29.296 37.591 7.162 1 38.40
                                Α
                                      1
 segid elesy charge
1 <NA>
           N
               <NA>
2 <NA>
           C <NA>
3 <NA>
           C <NA>
4 <NA>
           O <NA>
5 <NA>
           C <NA>
6 <NA>
           С
               <NA>
```

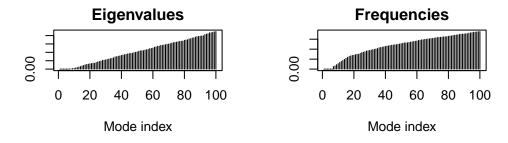
#### Predicting functional motions of a single structure by NMA

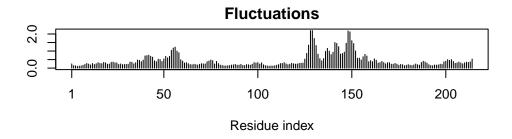
```
adk <- read.pdb('6s36')
 Note: Accessing on-line PDB file
  PDB has ALT records, taking A only, rm.alt=TRUE
  adk
       read.pdb(file = "6s36")
  Total Models#: 1
     Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)
    Protein Atoms#: 1654 (residues/Calpha atoms#: 214)
    Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
     Non-protein/nucleic Atoms#: 244 (residues: 244)
    Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]
  Protein sequence:
     MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAAVKSGSELGKQAKDIMDAGKLVT
     DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVDKI
     VGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG
     YYSKEAEAGNTKYAKVDGTKPVAEVRADLEKILG
+ attr: atom, xyz, segres, helix, sheet,
       calpha, remark, call
```

```
m <- nma(adk)
```

Building Hessian... Done in 0.012 seconds. Diagonalizing Hessian... Done in 0.26 seconds.

plot(m)





mktrj(m, file = "adk\_m7.pdb")

## 4. Comparative structure analysis of Adenylate Kinase

Q10. Which of the packages above is found only on BioConductor and not CRAN?

The msa package.

Q11. Which of the above packages is not found on BioConductor or CRAN?

The bio3d view package.

Q12. True or False? Functions from the devtools package can be used to install packages from GitHub and BitBucket?

True

#### Search and retrieve ADK structures

```
#install.packages("bio3d")
#install.packages("devtools")
#install.packages("BiocManager")

#BiocManager::install("msa")
#devtools::install_bitbucket("Grantlab/bio3d-view")

library(bio3d)
aa <- get.seq("1ake_A")

Warning in get.seq("1ake_A"): Removing existing file: seqs.fasta

Fetching... Please wait. Done.</pre>
```

Q13. How many amino acids are in this sequence, i.e. how long is this sequence?

214