# **Exercises in Marine Ecological Genetics**

## 04. Population structure and gene flow

- Model the effect of a bottleneck on N<sub>e</sub>
- Calculate F-statistics in R
- Test for population differentiation
- Visualize population-specific  $F_{ST}$  using ggplot

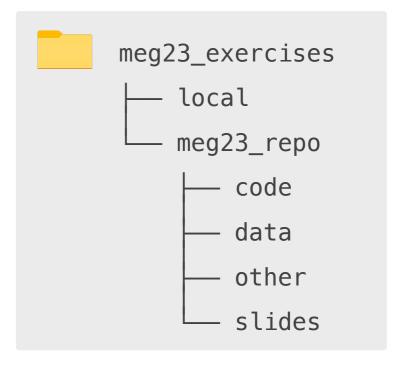
Martin Helmkampf



## Download course materials using git

Go to project directory and check contents

```
cd dir # e.g. Documents/meg23_exercises
ls -l # -l: long format
```



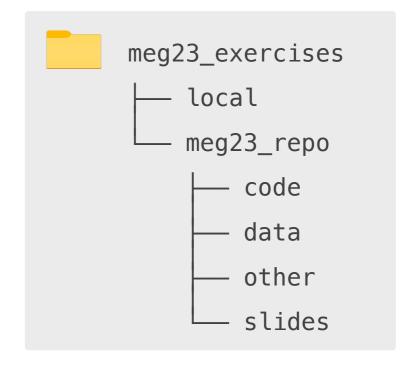
## Download course materials using git

Go to project directory and check contents

```
cd dir # e.g. Documents/meg23_exercises
ls -l # -l: long format
```

Update course repository

```
cd meg23_repo
git pull
```



Please do not save over files in the course repository. Instead, save your own scripts to the local subdirectory (including copies of course scripts you would like to edit), e.g with

```
cp code/04_structure.R ../local/04_structure_lc.R # cp [source] [destination]
```

## In case of an error message ...

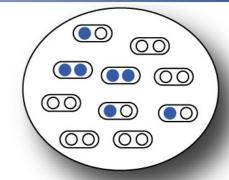
Delete old repository and re-download from GitHub

```
rm -rf meg23_repo
git clone https://github.com/mhelmkampf/meg23_repo.git
```



#### HETEROZYGOSITY

### In one population



 $H_o$  = proportion of heterozygote individuals, observed heterozygosity

$$H_e = 2pq = 1 - p^2 - q^2$$
, expected heterozygosity (assuming HW equilibrium)

$$F = \frac{H_e - H_o}{H_e}$$

Fixation index: proportion by which heterozygosity is reduced or  $F = \frac{H_e - H_o}{H_e}$  increased relative to the heterozygosity of a population at HW equilibrium with the same allele frequencies. equilibrium with the same allele frequencies.

Divided by  $H_e \rightarrow proportion$  (of expected heterozygosity)

Varies between -1 and 1

F < 0: heterozygote excess

*F* > 0 heterozygote deficit (homozygote excess)

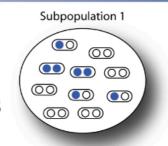
May be averaged over several loci -> reduces bias

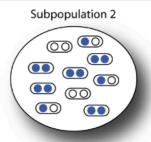
May be extended to *k* alleles



### In n subpopulations

 $H_I = \frac{1}{n} \sum_{i=1}^{n} \hat{H}_i$  Mean observed heterozygosity within subpopulations





$$H_S = \frac{1}{n} \sum_{i=1}^{n} 2p_i q_i$$
 Mean expected heterozygosity within subpopulations (assuming HW within subpops)

$$H_T = 2\overline{p}\overline{q}$$
 Expected heterozygosity of the total population (assuming HW within the total pop)

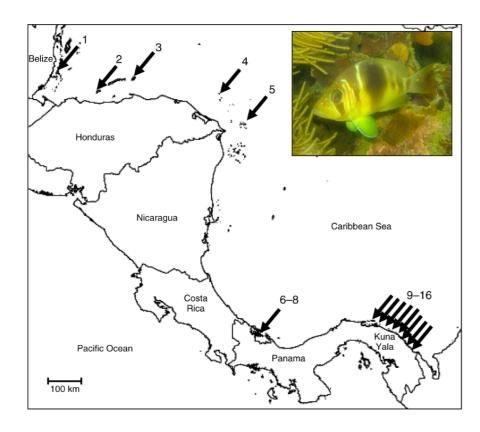
 $\hat{H}_i$  observed heterozygosity in population i  $p_i(q_i)$  frequency of allele A(a) in population i  $\overline{p}(\overline{q})$  mean frequency of allele A(a) n: number of subpopulations

$$F_{IS} = \frac{H_S - H_I}{H_S} \qquad F_{ST} = \frac{H_T - H_S}{H_T} \qquad F_{IT} = \frac{H_T - H_I}{H_T}$$

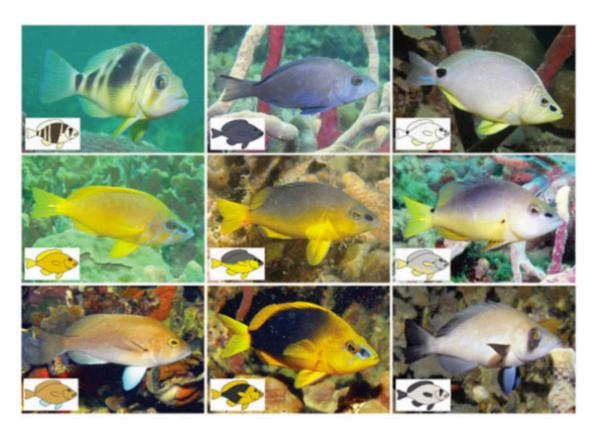
- $F_{IS}$  Average difference between observed and Hardy–Weinberg expected heterozygosity within each subpopulation (due to non-random mating)
- $F_{ST}$  Reduction in heterozygosity due to subpopulation divergence of allele frequencies
- F<sub>IT</sub> Combined departure from HW expected genotype frequencies due to non-random mating within subpopulations and divergence of allele frequencies among subpopulations

### Compare the amount of genetic structure in

- Caribbean populations of H. puella (puella\_caribbean.gen/.txt)
- a time series of H. puella samples from Panama (puella\_temporal.gen/.txt)
- Several species of hamlets from various Caribbean locations (hamlets\_caribbean.gen/.txt)



- Puebla et al. 2009, Ecology



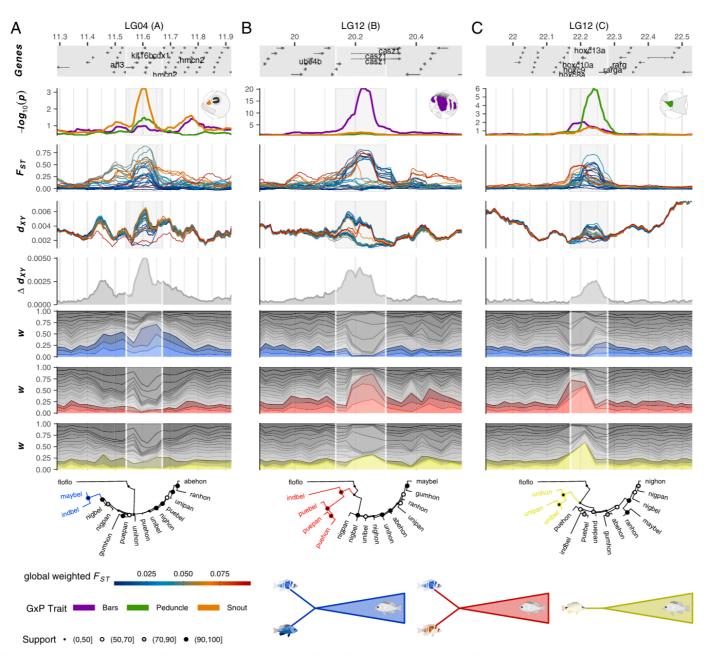
- Puebla et al. 2012, Proc R Soc B

## Calculate global $F_{ST}$ and test for differentiation

Species	Ê <sub>ST</sub>	N <sub>e</sub> m	Reference
Amphibians			
Alytes muletansis (Mallorcan midwife toad)	0.12-0.53	1.8-0.2	Kraaijeveld-Smit et al. 2005
Birds			
Gallus gallus (broiler chicken breed)	0.19	1.0	Emara et al. 2002
Mammals			
Capreolus capreolus (roe deer)	0.097-0.146	2.2-1.4	Wang and Schreiber 2001
Homo sapiens (human)	0.03-0.05	7.8–4.6	Rosenberg et al. 2002
Microtus arvalis (common vole)	0.17	1.2	Heckel et al. 2005
Plants			
Arabidopsis thaliana (mouse-ear cress)	0.643	0.1	Bergelson et al. 1998
Oryza officinalis (wild rice)	0.44	0.3	Gao 2005
Phlox drummondii (annual phlox)	0.17 0.32	1.2 0.5	Levin 1977 Romero et al. 2003
Prunus armeniaca (apricot)	0.32	0.5	Korriero et al. 2003
Fish  Morone saxatilis (striped bass)	0.002	11.8	Brown et al. 2005
Sparisoma viride (stoplight parrotfish)	0.002	12.4	Geertjes et al. 2004
Insects	0.012	12.4	Geer ges et al. 2004
Drosophila melanogaster (fruit fly)	0.112	2.0	Singh and Rhomberg 1987
Glossina pallidipes (tsetse fly)	0.112	1.1	Ouma et al. 2005
Heliconius charithonia (butterfly)	0.003	79.8	Kronforst and Flemming 2001
Corals			3
Seriatopora hystrix	0.089-0.136	2.6-1.6	Maier et al. 200

Species	$\hat{F}_{ST}$	$\widehat{N_e m}$	Reference  Kraaijeveld-Smit et al. 2005  Emara et al. 2002		
Amphibians Alytes muletansis (Mallorcan midwife toad) Birds Gallus gallus (broiler chicken breed)	0.12–0.53 0.19	1.8–0.2 1.0			
Mammals  Capreolus capreolus (roe deer)  Homo sapiens (human)	0.097–0.146 0.03–0.05	2.2–1.4 7.8–4.6	<b>F</b> <sub>ST</sub>	Subpo	pulations are
Microtus arvalis (common vole) Plants	0.17	1.2	< 0.15		very simila
Arabidopsis thaliana (mouse-ear cress) Oryza officinalis (wild rice)	0.643 0.44	0.1 0.3	0.15 - 0.25		simila
Phlox drummondii (annual phlox) Prunus armeniaca (apricot)	0.17 0.32	1.2 0.5	> 0.25		distinc
Fish  Morone saxatilis (striped bass)  Sparisoma viride (stoplight parrotfish)	0.002 0.019	11.8 12.4	Brown et al. 2005 Geertjes et al. 2004	4	
Insects Drosophila melanogaster (fruit fly) Glossina pallidipes (tsetse fly) Heliconius charithonia (butterfly)	0.112 0.18 0.003	2.0 1.1 79.8	Singh and Rhomberg 1987 Ouma et al. 2005 Kronforst and Flemming 2001		
Corals Seriatopora hystrix	0.089-0.136	2.6-1.6	Maier et al. 200	J	

## New R concepts: tidyverse and ggplot



- Hench et al. 2022, PNAS



## New R concepts: tidyverse and ggplot

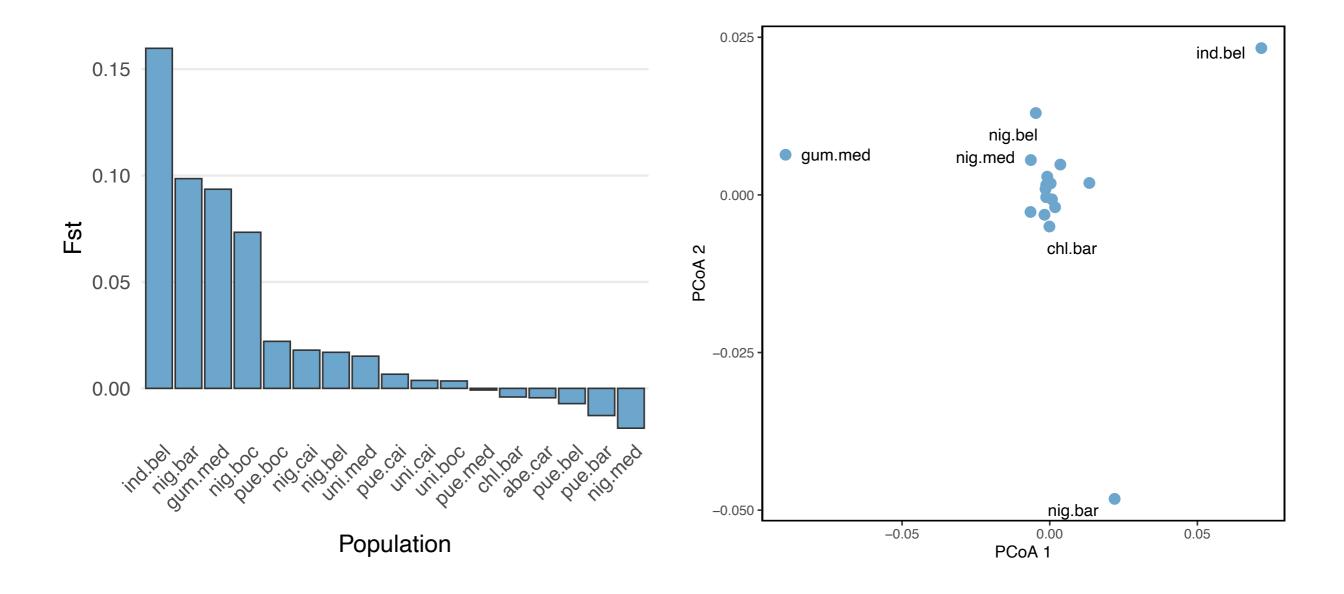
### Tidyverse pipes

```
x <- df %>%  # assign to new object (copy)
mutate() %>%  # 1st operation
arrange() %>%  # 2nd operation
select()  # 3rd operation
```

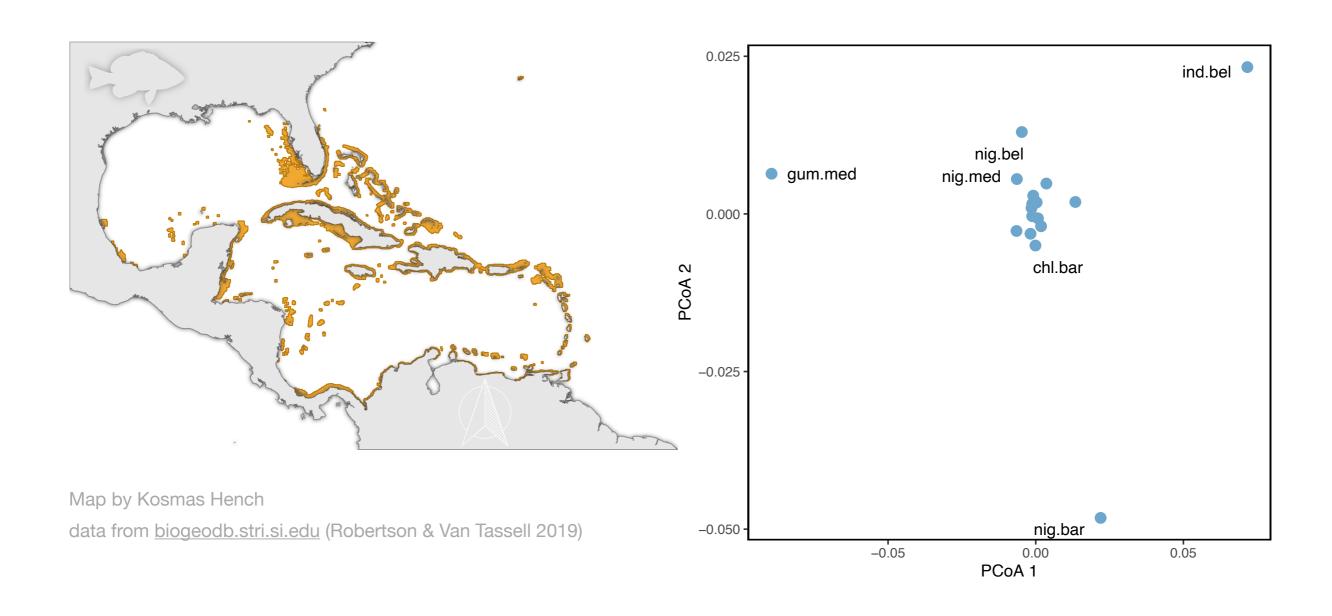
### Basic ggplot syntax

```
ggplot(data = df, aes(x = var1, y = var2)) + # mapping variables
geom_bar() + # geometric shapes representing the data
labs() + # set plot and axis labels
guides() + # customize plot legend
theme() # customize non-data, e.g. fonts, gridlines
```

## Calculate population-specific Fst



## Calculate population-specific Fst



### **Course outline**

Class	Date	Topics	Script
01	Apr 14	Introduction, software installation	01_intro.R
02	Apr 21	Hardy-Weinberg equilibrium	02_hwe.R
03	Apr 28	Genetic drift and effective population size	03_drift.R
04	May 05	Population structure and gene flow	04_structure.R
05	May 12	Isolation by distance (lecture online, exercises in person)	05_ibd.R
_	May 19	Himmelfahrt break	-
06	May 26	Whole-genome sequencing and genome assembly	06_genseq.sh
07	Jun 02	Genotyping, SNPs and population genomics	07_snps.R
08	Jun 09	Recombination and linkage disequilibrium	08_linkage.R
_	Jun 16	Student presentations	_
09	Jun 23	Selection and mutation	09_selection.R
10	Jun 30	DNA barcoding	10_barcode.sh
11	Jul 07	Metabarcoding	11_meta.sh
_	Jul 14	To be determined	_

