FIND A GENE - MEG ROBINSON

[Q1] Tell me the name of a protein you are interested in. Include the species and the accession number. This can be a human protein or a protein from any other species as long as it's function is known. If you do not have a favorite protein, select human RBP4 or KIF11. Do not use beta globin as this is in the worked example report that I provide you with online.

Name: Retinol-binding protein 4

Accession: P02753

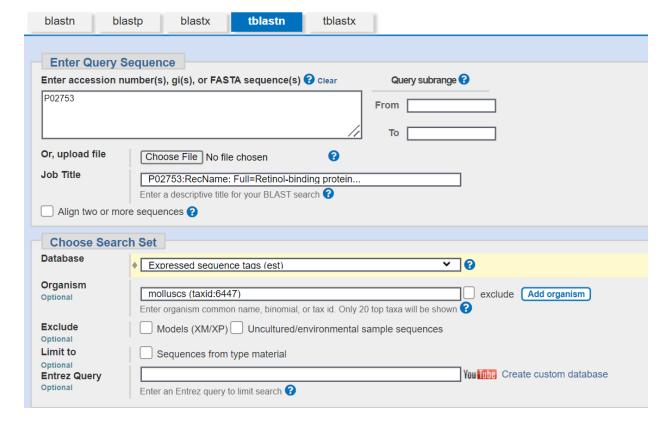
Species: Homo sapiens (Human)

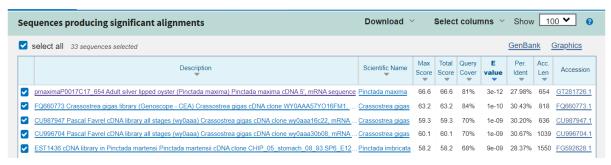
[Q2] Perform a BLAST search against a DNA database, such as a database consisting of genomic DNA or ESTs. The BLAST server can be at NCBI or elsewhere. Include details of the BLAST method used, database searched and any limits applied (e.g. Organism). Also include the output of that BLAST search in your document. On the BLAST results, clearly indicate a match that represents a protein sequence, encoded from some DNA sequence, that is homologous to your query protein. I need to be able to inspect the pairwise alignment you have selected, including the E value and score. It should be labeled a "genomic clone" or "mRNA sequence", etc. - but include no functional annotation.

Method: TBLASTN (2.7.1) search against zebra fish ESTs

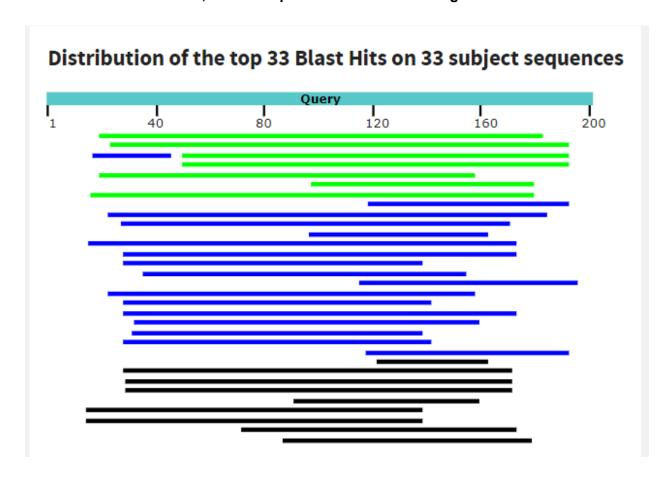
Database: Expressed Sequence Tags (est)

Organism: molluscs (taxid:6447))





Chosen match: Accession GT281726.1 Adult silver lipped oyster (Pinctada maxima) Pinctada maxima cDNA 5', mRNA sequence. See below for alignment details.



pmaximaP0017C17_654 Adult silver lipped oyster (Pinctada maxima) Pinctada maxima cDNA 5', mRNA sequence Sequence ID: GT281726.1 Length: 654 Number of Matches: 1

Range 1: 142 to 618 GenBank Graphics

\forall	N	ovt	Match	Α.	Pre	vioi	IC	NΛ	al	tch.
7	1.3	CVL	IVICILLII		1 10	A IOI	40	IVI		

Score		Expect Method	Identities	Positives	Gaps	Frame
66.6 b	its(16	 3e-12 Compositional matrix adjust 	. 47/168(28%)	79/168(47%)	14/168(8%)	+1
Query	20	RDCRVSSFRVKENFDKARFSGTWYAM			•	
Sbjct	142	+DC +S+F+ + NF+ +F G WY + KDCVISNFQTQSNFEADKFVGKWYEIEWMTH				
Query	75	AKGRVRLLNNWDVCADMVGTFTDTEDPAKFF R N +C+ T + AK+				
Sbjct	316	TAFRSNPNKTICSLQNAVMYRTSN-AKYI				
Query	135	QYSCRLLNLDGTCADSYSFVFSRDPNGLPPI YSC + N+DGTC + FSR L				
Sbjct	478	IYSCHVQNIDGTCKTWVAKTFSR-KRTLDDI				

[Q3] Gather information about this "novel" protein. At a minimum, show me the protein sequence of the "novel" protein as displayed in your BLAST results from [Q2] as FASTA format (you can copy and paste the aligned sequence subject lines from your BLAST result page if necessary) or translate your novel DNA sequence using a tool called EMBOSS Transeq at the EBI. Don't forget to translate all six reading frames; the ORF (open reading frame) is likely to be the longest sequence without a stop codon. It may not start with a methionine if you don't have the complete coding region. Make sure the sequence you provide includes a header/subject line and is in traditional FASTA format.

Translated Sequence:

>GT281726.1 | pmaximaP0017C17_654 Adult silver lipped oyster (Pinctada maxima) Pinctada maxima cDNA 5', mRNA sequence

KDCVISNFQTQSNFEADKFVGKWYEIEWMTHQAENPNDFW--DDYVTNYTLNDDGSFSLF TAFRSN--PNKTICSLQNAVMYRTSN-AKYDV---AVSSCRQIRHSPQWIISTDYIRYAI IYSCHVONIDGTCKTWVAKTFSR-KRTLDDRYISLAHDTYKDLCLNRH

>Human RBP4 | NP_001310447.1 | retinol-binding protein 4 isoform b [Homo sapiens]

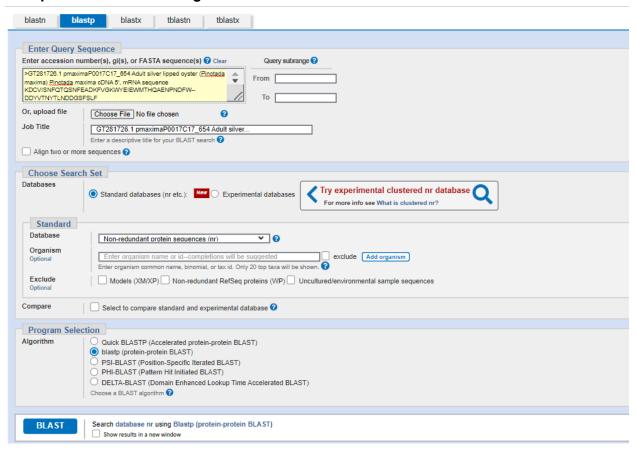
RDCRVSSFRVKENFDKARFSGTWYAM----AKKDPEGLFLQDNIVAEFSVDETGQMSAT AKGRVRLLNNWDVCADMVGTFTDTEDPAKFKMKYWGVASFLQKGNDDHWIVDTDYDTYAV QYSCRLLNLDGTCADSYSFVFSRDPNGLPPEAQKIVRQRQEELCLARQ

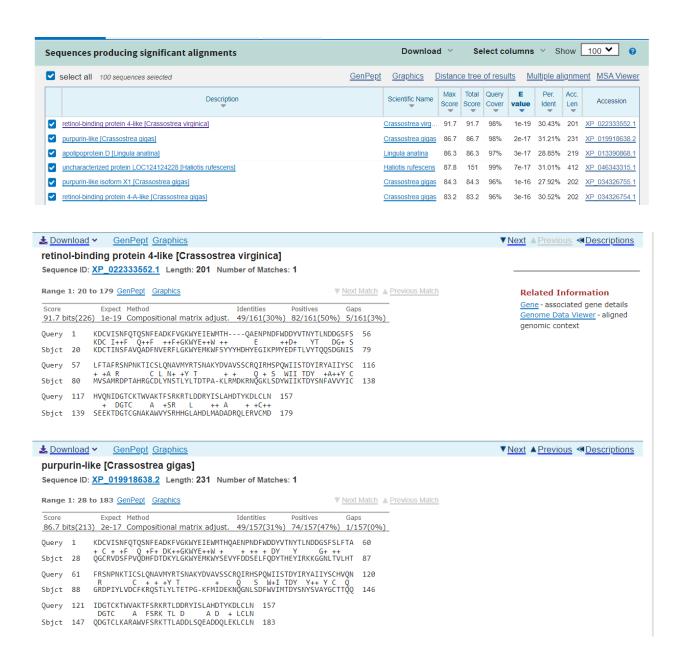
SOURCE Pinctada maxima (silver-lipped pearl oyster)

Eukaryota; Metazoa; Spiralia; Lophotrochozoa; Mollusca; Bivalvia; Autobranchia; Pteriomorphia; Pterioida; Pterioidea; Pteriidae; Pinctada.

[Q4] Prove that this gene, and its corresponding protein, are novel. For the purposes of this project, "novel" is defined as follows. Take the protein sequence (your answer to [Q3]), and use it as a query in a blastp search of the nr database at NCBI. • If there is a match with 100% amino acid identity to a protein in the database, from the same species, then your protein is NOT novel (even if the match is to a protein with a name such as "unknown"). Someone has already found and annotated this sequence, and assigned it an accession number. • If the top match reported has less than 100% identity, then it is likely that your protein is novel, and you have succeeded. • If there is a match with 100% identity, but to a different species than the one you started with, then you have likely succeeded in finding a novel gene. • If there are no database matches to the original query from [Q1], this indicates that you have partially succeeded: yes, you may have found a new gene, but no, it is not actually homologous to the original query. You should probably start over.

A BLASTP search against NR database (see setup in first screen-shot below) yielded a top hit result is to a protein from Crassostrea virginica (eastern oyster) but with not 100% identity (30.43%). See additional screen shots below for top hits and selected alignment details:





[Q5] Generate a multiple sequence alignment with your novel protein, your original query protein, and a group of other members of this family from different species. A typical number of proteins to use in a multiple sequence alignment for this assignment purpose is a minimum of 5 and a maximum of 20 - although the exact number is up to you. Include the multiple sequence alignment in your report. Use Courier font with a size appropriate to fit page width. Side-note: Indicate your sequence in the alignment by choosing an appropriate name for each sequence in the input unaligned sequence file (i.e. edit the sequence file so that the species, or short common, names (rather than accession numbers) display in the output alignment and in the subsequent answers below). The goal in this step is to create an interesting alignment for building a phylogenetic tree that illustrates species divergence.

Re-labeled sequences for alignment:

- >Human_original_seq | NP_001310447.1 | retinol-binding protein 4 isoform b [Homo sapiens]
 RDCRVSSFRVKENFDKARFSGTWYAM----AKKDPEGLFLQDNIVAEFSVDETGQMSAT
 AKGRVRLLNNWDVCADMVGTFTDTEDPAKFKMKYWGVASFLQKGNDDHWIVDTDYDTYAV
 QYSCRLLNLDGTCADSYSFVFSRDPNGLPPEAQKIVRQRQEELCLARQ
- > Silver_lipped_oyster_novel_protein | taken from BLAST | Pinctada maxima cDNA 5', mRNA sequence | GT281726.1 KDCVISNFQTQSNFEADKFVGKWYEIEWMTHQAENPNDFW--DDYVTNYTLNDDGSFSLF TAFRSN--PNKTICSLQNAVMYRTSN-AKYDV---AVSSCRQIRHSPQWIISTDYIRYAI IYSCHVQNIDGTCKTWVAKTFSR-KRTLDDRYISLAHDTYKDLCLNRH
- > Eastern_Oyster | Crassostrea virginica KDCVISNFQTQSNFEADKFVGKWYEIEWMTH----QAENPNDFWDDYVTNYTLNDDGSFS LFTAFRSNPNKTICSLQNAVMYRTSNAKYDVAVSSCRQIRHSPQWIISTDYIRYAIIYSC HVONIDGTCKTWVAKTFSRKRTLDDRYISLAHDTYKDLCLN
- > Scallop | XP_021358553.1 | retinol-binding protein 4-like [Mizuhopecten yessoensis]
 RDCRLSSFQLQPDFNLAQFEGYWYSLTINRYWLAVPRWFPVRQSNVQVNYRLLSDGSLEV
 KTGGEFMFPFCDYIAGKGYIPDKTQPQKLEVQFDTLLTRTSRKNPYWVVSTDYEGFAVIY
 SCWKEREDGTC
- > Snail | XP_041352779.1 | retinol-binding protein 4-like [Gigantopelta aegis]
 KDCNVHNITVQPDFDLQKYAGTWYEMKWLATVYIPPNQLYQDYRHIYTYAGRGENVTVDI
 AGRDPANINKCFYNTARLMETDDDAK---MVFATVKERYS-YWVVKTDYTSYAVVYGCYS
 VTSDGACNGTRSWIWSRTKTLSQDKMAIAETVISETLCVNQ
- > Mandarin_fish | XP_044034678.1 | retinol-binding protein 4 [Siniperca chuatsi]
 QDCQVANIQVMQNFDKTRYAGTWYVIA----KKDPEGLFLLDNIMAQFTVADDGKMTAT
 AKGRVIILNNWEMCADMLATFEETSDPAKFRMKYWGVASYLQTGNDDHWVIDTDYDNYAI
 HYSCRLQDSDGTCLDSYSMIFSRHLDGLRPEDQRTV---HQKKMDLCL
- > Painted_turtle | XP_005301331.1 | retinol-binding protein 4 [Chrysemys picta bellii]
 RDCRVSNFRVQENFDKARYTGTWYAIA----KKDPEGLFLQDNVVAQFTIDENGQMSAT
 AKGRVRLFNNWDVCADMIGSFTDTEDPAKFKMKYWGVASFLQKGNDDHWVVDTDYDTYAL
 HYSCRQLNDDGTCADSYSFVFSRDPKGLSPEVQRIIRQRQVDLCLDR

Alignment:

Obtained using MUSCLE (version 3.8) at EBI:

CLUSTAL multiple sequence alignment by MUSCLE (3.8)

Snail
Mandarin_fish
Human_original_seq
Painted_turtle
Silver_lipped_oyster_novel_prote
Eastern_Oyster
Scallop

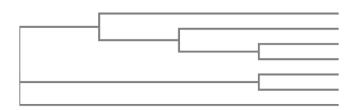
Snail
Mandarin_fish
Human_original_seq
Painted_turtle
Silver_lipped_oyster_novel_prote
Eastern_Oyster
Scallop

Snail
Mandarin_fish
Human_original_seq
Painted_turtle
Silver_lipped_oyster_novel_prote
Eastern_Oyster
Scallop

NVTVDIAGRDPANINKCFYNTARLMETDDDAKMVFA-----TVKERYSYWVVKTDYTS
TATAKGRVIILNNWEMCADMLATFEETSDPAKFRMKYWGVASYLQTGNDDHWVIDTDYDN
SATAKGRVRLLNNWDVCADMVGTFTDTEDPAKFKMKYWGVASFLQKGNDDHWVVDTDYDT
SATAKGRVRLFNNWDVCADMIGSFTDTEDPAKFKMKYWGVASFLQKGNDDHWVVDTDYDT
SLFTAFRSNPNKTICSLQNAVMY---RTSNAKYDVA---VSSCRQIRHSPQWIISTDYIR
SLFTAFRSNPNKTICSLQNAVMY---RTSNAKYDVA---VSSCRQIRHSPQWIISTDYIR
EVKTG--GEFMFPFCDYIAGKGYIPDKTQPQKLEVQFDTLLT-RTSRKNPYWVVSTDYEG

YAVVYGCYSVTSDGACNGTRSWIWSRTKTLSQDKMAIAETVISETLCVNQ-YAIHYSCRLQDSDGTCLDSYSMIFSRHLDGLRPEDQRTVHQKKMDLCL---YAVQYSCRLLNLDGTCADSYSFVFSRDPNGLPPEAQKIVRQRQEELCLARQYALHYSCRQLNDDGTCADSYSFVFSRDPKGLSPEVQRIIRQRQVDLCLDRYAIIYSCHVQNIDGTCKTWVAKTFSRKRT-LDDRYISLAHDTYKDLCLNRHYAIIYSCHVQNIDGTCKTWVAKTFSRKRT-LDDRYISLAHDTYKDLCLN--FAVIYSCWKEREDGTC------

[Q6] Create a phylogenetic tree, using either a parsimony or distance-based approach. Bootstrapping and tree rooting are optional. Use "simple phylogeny" online from the EBI or any respected phylogeny program (such as MEGA, PAUP, or Phylip). Paste an image of your Cladogram or tree output in your report.



Snail 0.35884
Mandarin_fish 0.17833
Human_original_seq 0.08599
Painted_turtle 0.0745
Silver_lipped_oyster_novel_prote -0.00055
Eastern_Oyster 0.00055
Scallop 0.34334

Q7-Q10

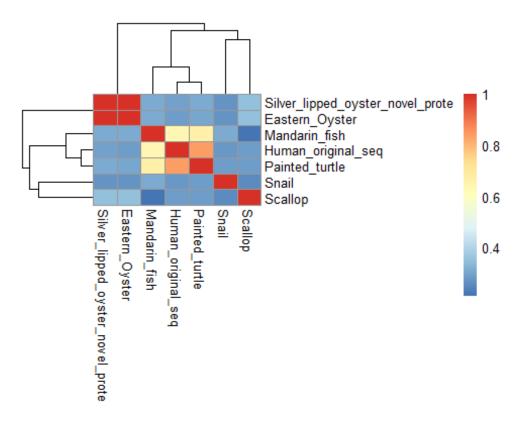
Meg Robinson

3/11/2022

Question 7

[Q7] Generate a sequence identity based heatmap of your aligned sequences using R. If necessary convert your sequence alignment to the ubiquitous FASTA format (Seaview can read in clustal format and "Save as" FASTA format for example). Read this FASTA format alignment into R with the help of functions in the Bio3D package. Calculate a sequence identity matrix (again using a function within the Bio3D package). Then generate a heatmap plot and add to your report. Do make sure your labels are visible and not cut at the figure margins.

```
library(bio3d)
## Warning: package 'bio3d' was built under R version 4.1.2
library(pheatmap)
## Warning: package 'pheatmap' was built under R version 4.1.2
aln <- read.fasta("cluster.fasta")
iden <- seqidentity(aln)
pheatmap(iden, margins = c(12,12))</pre>
```



Question 8

[Q8] Using R/Bio3D (or an online blast server if you prefer), search the main protein structure database for the most similar atomic resolution structures to your aligned sequences. List the top 3 unique hits (i.e. not hits representing different chains from the same structure) along with their Evalue and sequence identity to your query. Please also add annotation details of these structures. For example include the annotation terms PDB identifier (structureId), Method used to solve the structure (experimentalTechnique), resolution (resolution), and source organism (source).

Use the sequence with the highest identity

```
rowmean <- rowMeans(iden)</pre>
rowmean
##
                                Snail
                                                           Mandarin_fish
                            0.3811429
                                                                0.4931429
##
##
                  Human_original_seq
                                                          Painted_turtle
##
                            0.5180000
                                                                0.5265714
## Silver lipped oyster novel prote
                                                          Eastern Oyster
                                                                0.5032857
##
                            0.5040000
##
                              Scallop
##
                            0.3904286
highest.name <- names(which.max(rowmean))</pre>
highest.val <- max(rowMeans(iden))*100
```

```
print(paste("The species with the highest average row identity is",
highest.name))
## [1] "The species with the highest average row identity is Painted_turtle"
print(paste("The average row identity of this species is",
round(highest.val,2), "%"))
## [1] "The average row identity of this species is 52.66 %"
```

Now we will continue with the FASTA of the turtle

```
turtle <- read.fasta("turtle.fasta")</pre>
turtle.blast <- blast.pdb(turtle)</pre>
## Searching ... please wait (updates every 5 seconds) RID = 2RPPBTPA016
##
## Reporting 111 hits
# top 3 hits ID
library(dplyr)
## Warning: package 'dplyr' was built under R version 4.1.2
##
## Attaching package: 'dplyr'
## The following objects are masked from 'package:stats':
##
##
       filter, lag
## The following objects are masked from 'package:base':
##
##
       intersect, setdiff, setequal, union
df.blast <- turtle.blast$hit.tbl</pre>
df.blast.id <- head(df.blast$subjectids,3)</pre>
df.eval.iden <- select(head(df.blast,3), c("evalue","identity"))</pre>
rownames(df.eval.iden) <-</pre>
as.vector(select(head(df.blast,3),"subjectids"))[1:3,]
df.eval.iden
             evalue identity
## 1IIU A 1.52e-113 91.304
## 409S_A 9.07e-106
                       83.951
## 3FMZ A 9.23e-106
                       83.951
df.pdb <- lapply(df.blast.id,pdb.annotate) %>% bind rows()
df.pdb.select <- select(df.pdb,</pre>
c("structureId", "experimentalTechnique", "resolution", "source"))
df.pdb.select
```

```
## structureId experimentalTechnique resolution source
## 1IIU_A 1IIU X-ray 2.5 Gallus gallus
## 409S_A 409S X-ray 2.3 Homo sapiens
## 3FMZ_A 3FMZ X-ray 2.9 Homo sapiens
```

Now we can combine our two dataframes

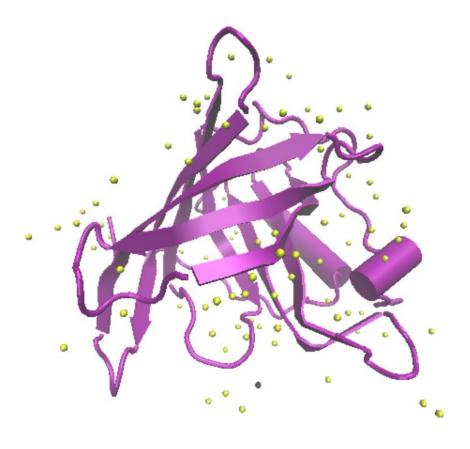
```
df.final <- cbind(df.eval.iden, df.pdb.select)</pre>
colnames(df.final) <- c("Evalue", "Identity", "ID", "Technique",</pre>
"Resolution", "Source")
df.final
##
             Evalue Identity
                               ID Technique Resolution
                                                                Source
## 1IIU A 1.52e-113
                                       X-ray
                                                    2.5 Gallus gallus
                      91.304 1IIU
## 409S A 9.07e-106
                      83.951 4095
                                       X-ray
                                                    2.3 Homo sapiens
## 3FMZ_A 9.23e-106
                      83.951 3FMZ
                                       X-ray
                                                    2.9 Homo sapiens
```

Question 9

[Q9] Generate a molecular figure of one of your identified PDB structures using VMD. You can optionally highlight conserved residues that are likely to be functional. Please use a white or transparent background for your figure (i.e. not the default black). Based on sequence similarity. How likely is this structure to be similar to your "novel" protein?

Based on the sequence similarity, this is very likely to be similar in structure to my novel protein Pinctada maxima due to the high sequence similarity (>91%).

(I used 1IIU for Q9)



Question 10

[Q10] Perform a "Target" search of ChEMBEL (https://www.ebi.ac.uk/chembl/) with your novel sequence. Are there any Target Associated Assays and ligand efficiency data reported that may be useful starting points for exploring potential inhibition of your novel protein?

CHEMBL details 3 Binding Assays (CHEMBL3707846, CHEMBL3707845, CHEMBL3705836 – all the same) and 1 Inhibition Assay (CHEMBL3887722); No ligand efficiency data.

https://www.ebi.ac.uk/chembl/g/#search_results/assays/query=%3E%20Silver_lipped_o yster_novel_protein%20%7C%20taken%20from%20BLAST%20%7C%20Pinctada%20ma xima%20cDNA%205%26%23x27%3B%2C%20mRNA%20sequence%20%7C%20GT2817 26.1%20KDCVISNFQTQSNFEADKFVGKWYEIEWMTHQAENPNDFW--DDYVTNYTLNDDGSFSLF%20TAFRSN--PNKTICSLQNAVMYRTSN-AKYDV---AVSSCRQIRHSPQWIISTDYIRYAI%20IYSCHVQNIDGTCKTWVAKTFSR-KRTLDDRYISLAHDTYKDLCLNRH

In the binding assay they tested the affinity of various compounds at the NE, DA and 5HT transporters of HEK293E cell lines. There are no references provided or useful information.