90 Gerald E. Gaull et al.

correlation between the changes that occur and the pathogenesis of the disease can only be speculative at this time.

## **B.** Metabolism of Catecholamines

The possibility of defective catecholamine biosynthesis (Fig. 4) in phenylketonuria was suggested by the observation of decreased concentrations of catecholamines in the plasma (286,287) and urine (287) of these patients. The excretion of normal amounts of catecholamines, but the excretion of decreased amounts of the metabolite vanillylmandelic acid, also has been reported. (288) On a low-phenylalanine diet there were increases in plasma and urine catecholamines, (287) in urinary vanillylmandelic acid, (288) and in the accumulation of the metabolite homovanillic acid in cerebrospinal fluid after the administration of probenecid. (93) Curtius et al. (289) administered deuterated L-tyrosine to patients with phenylketonuria and found a lower excretion of metabolites of dopamine and norepinephrine when the patients had a high plasma phenylalanine concentration than when the plasma phenylalanine concentration was low. Although such findings do not necessarily reflect the situation in the brain, the recent analysis by McKean<sup>(93)</sup> has shown that the norepinephrine content of the caudate nucleus, the brain stem, and the

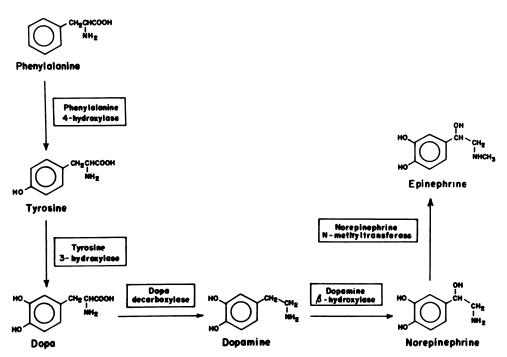


Fig. 4. Steps in the synthesis of the catecholamines.

126 Gerald E. Gaull et al.

93. C. M. McKean, The effects of high phenylalanine concentrations on serotonin and catecholamine metabolism in the human brain, *Brain Res.* 47:469-476 (1972).

- 94. Y. H. Loo and K. Mack, Effect of vitamin B<sub>6</sub> on phenylalanine metabolism in the brain of normal and p-chlorophenylalanine-treated rats, J. Neurochem. 19:2385-2394 (1972).
- 95. D. J. Edwards and K. Blau, Aromatic acids derived from phenylalanine in the tissues of rats with experimentally induced phenylketonuria-like characteristics, *Biochem. J.* 130:495-503 (1972).
- 96. D. J. Edwards and K. Blau, Phenethylamines in brain and liver of rats with experimentally induced phenylketonuria-like characteristics, *Biochem. J.* 132:95-100 (1973).
- 97. A. L. Miller, R. A. Hawkins, and R. L. Veech, Phenylketonuria: Phenylalanine inhibits brain pyruvate kinase in vivo, Science (Wash. D.C.) 179:904-906 (1973).
- 98. A. N. Davison, Nutrition and amino acid imbalance as factors influencing brain development, *Biochem. Soc. Spec. Publ.* 1:27-37 (1974).
- J. M. Saavedra, Enzymatic isotopic assay for and presence of β-phenylethylamine in brain, J. Neurochem. 22:211-216 (1974).
- 100. J. M. Saavedra and J. Axelrod, Demonstration and distribution of phenylethanolamine in brain and other tissues, *Proc. Natl. Acad. Sci.* (USA) 70:769-772 (1973).
- Y. H. Loo, Characterization of a new phenylalanine metabolite in phenylketonuria, J. Neurochem. 14:813-821 (1967).
- 102. T. L. Perry and R. T. Jones, The amino acid content of human cerebrospinal fluid in normal individuals and in mental defectives, J. Clin. Invest. 40:1363-1372 (1961).
- 103. C. M. McKean and D. E. Boggs, Influence of high concentrations of phenylalanine on the amino acids of cerebrospinal fluid and blood, *Proc. Soc. Exp. Biol. Med.* 122:987-991 (1966).
- 104. Y. Mardens, J. Dumon, F. Hayez, S. Vrydagh, A. Cools, and G. Myle, Observation de deux paires de jumeaux monozygotiques atteints de phénylcétonurie, J. Génét, Hum. 16:42-77 (1967).
- M. van Sande, Y. Mardens, K. Adriaenssens, and A. Lowenthal, The free amino acids in human cerebrospinal fluid, J. Neurochem. 17:125-135 (1970).
- 106. C.-D. Quentin, A. W. Behbehani, F. J. Schulte, and V. Neuhoff, Microanalysis with <sup>14</sup>C-dansyl chloride of amino acids and amines in the cerebrospinal fluid of patients with phenyl-ketonuria. I. Analysis in untreated phenylketonuria, *Neuropädiatrie* 5:138-145 (1974).
- 107. C.-D. Quentin, A. W. Behbehani, F. J. Schulte, and V. Neuhoff, Microanalysis with <sup>14</sup>C-dansyl chloride of amino acids and amines in the cerebrospinal fluid of patients with phenyl-ketonuria. III. Analysis of amino acids after loading with L-phenylalanine, Neuropädiatrie 5:271-278 (1974).
- T. L. Perry, S. Hansen, B. Tischler, R. Bunting, and S. Diamond, Glutamine depletion in phenylketonuria. A possible cause of the mental defect, N. Engl. J. Med. 282:761-766 (1970).
- P. W. K. Wong, J. L. Berman, M. W. Partington, S. K. Vickery, M. E. O'Flynn, and D. Y.-Y. Hsia, Glutamine in PKU, N. Engl. J. Med. 285:580 (1971).
- D. R. Lines and H. A. Waisman, Urinary amino acid excretion in phenylketonuric, hyperphenylalaninemic, and normal patients, J. Pediatr. 78:474-480 (1971).
- J. P. Colombo, Plasma glutamine in a phenylketonuric family with normal and mentally defective members, Arch. Dis. Child. 46:720-721 (1971).
- 112. R. Blasberg and A. Lajtha, Heterogeneity of the mediated transport systems of amino acid uptake in brain, *Brain Res.* 1:86-104 (1966).
- 113. A. Neidle, J. Kandera, and M. Chedekel, Amino acid efflux and protein turnover in mouse brain slices, Fed. Proc. 29:911 Abs (1970).
- 114. T. M. Andrews, R. O. McKeran, R. W. E. Watts, K. McPherson, and R. Lax, A relation-ship between the granulocyte phenylalanine content and the degree of disability in phenyl-ketonuria, Q. J. Med. 42:805-817 (1973).
- 115. A. L. Prensky and H. W. Moser, Brain lipids, proteolipids, and free amino acids in maple syrup urine disease, *J. Neurochem.* 13:863-874 (1966).

Wilson's Disease 251

per. (49) In almost every patient with Wilson's disease the capacity of this excretory route is limited, since he cannot synthesize normal amounts of ceruloplasmin. (50)

Second, evidence from several independent studies indicates that the biliary excretion of copper is considerably diminished in Wilson's disease, apparently because of a defect in hepatic lysosomes. (51)

Little attention has been paid to how remarkably small a positive balance of copper is sufficient to result in the accumulations seen in patients with Wilson's disease: no more than 10-20 mg, out of well over a gram of copper eaten each year, is retained, a 1% positive balance.

## A. Pathology

The effects of copper on the structure of the liver and the brain of patients with Wilson's disease have little specificity to distinguish them from the changes induced by other toxins. In the liver the copper is associated with a sequence of stages—fat accumulation; inflammation; the appearance of delicate, and then of coarse, fibrous bands; and hepatocellular necrosis resulting in collapse of parenchyma and regeneration—which eventuate in classical postnecrotic, nodular cirrhosis, (62,53) unique only in two histopathological respects. First, histochemical stains demonstrate that the excess hepatic copper in children with Wilson's disease is diffusely distributed throughout the cytoplasm of liver cells, in contrast to its localization in lysosomal granules in hepatocytes of normal newborn babies, (64) in whom physiologically large hepatic copper concentrations do not cause pathological consequences, probably because copper sequestered by hepatic lysosomes is innocuous. (63)

Second, electron microscopic study of biopsy samples of liver from patients with other types of liver disease, with Wilson's disease, and with no hepatic disorder has shown that specific mitochondrial changes—enlargement, bizarre shapes, changes in matrix density, presence of vacuoles and inclusions, and separation of the normally apposed inner and outer membranes—are seen only in patients with Wilson's disease (Fig. 1).<sup>(55)</sup> They are found only during the early stages of the disease and not when the cirrhosis is established.<sup>(63)</sup>

In the brain the distribution of copper and its pathological effects can be seen only in fatal cases and not in biopsy tissue, so that the natural sequence of pathological events and the effects of therapy, rather well known for the liver, are much less so for the brain.

The ventricular system is usually enlarged because of atrophy of basal ganglia. The lenticular nuclei show symmetrical atrophy and yellow or reddish brown discoloration. Gross cystic changes, noted by Wilson, (56) are seen in the putamen and globus pallidus (Fig. 2) Despite the name given it by Wilson, the cerebral degeneration in this disease is not limited to the lenticular nuclei, but it may invade the cerebral hemispheres and the cerebellum, so that the term hepatocerebral degeneration is more appropriate (Fig. 3),(57) though even that omits reference to several instances in which central pontine myelinolysis has been seen.(58)

- 93. Y. Itokawa and J. R. Cooper, The enzymatic synthesis of triphosphothiamin, *Biochim. Biophys. Acta* 158:180-182 (1968).
- 94. R. L. Barchi and P. E. Braun, A membrane associated thiamine triphosphatase from rat brain, J. Biol. Chem. 247:7668-7673 (1972).
- 95. J. H. Pincus, Y. Itokawa, and J. R. Cooper, Enzyme-inhibiting factor in subacute necrotizing encephalomyelopathy, *Neurology* 19:841-845 (1969).
- 96. M. Hamburgh and L. B. Flexner, Biochemical and physiological differentiation during morphogenesis. XXI. Effect of hypothyroidism and hormone therapy on enzyme activities of the developing cerebral cortex of the rat, *J. Neurochem.* 1:279-288 (1957).
- 97. Y. S. Kim and J. P. Lambooy, Induction of a specific enzyme inadequacy in infant rats by the use of a homologue of riboflavin, J. Nutr. 101:819-830 (1971).
- 98. T. Nagatsu, T. Yamamoto, and M. Harada, Purification and properties of human brain mitochondrial monoamine oxidase, *Enzymologia* 39:15-25 (1970).
- 99. M. Harada and T. Nagatsu, Identification of flavin in the purified beef brain mitochondrial monoamine oxidase, *Experientia* 25:583-584 (1969).
- 100. H. B. Burch, O. H. Lowry, A. M. Padilla, and A. M. Combs, Effects of riboflavin deficiency and realimentation on flavin enzymes of tissues, J. Biol. Chem. 223:29-45 (1956).
- 101. S. Schapiro and C. J. Percin, Thyroid hormone induction of  $\alpha$ -glycerophosphate dehydrogenase in rats of different ages, *Endocrinology* 79:1075-1078 (1966).
- 102. M. J. Blunt and C. P. Wendell-Smith, Glial  $\alpha$ -glycerophosphate dehydrogenase and central myelination, *Nature* (Lond.) **216**:605-606 (1967).
- 103. P. L. Wendell, Distribution of glutathione reductase and detection of glutathione-cystine transhydrogenase in rat tissues, *Biochim. Biophys. Acta* 159:179-181 (1968).
- 104. R. S. Rivlin, Medical Progress: Riboflavin metabolism, N. Engl. J. Med. 283:463-472 (1970).
- 105. K. Kanig, Die vitamine in der neurologie, Bibl. Psychiatr. Neurol. 138:60-89 (1969).
- 106. T. Arakawa, T. Mizuno, F. Chiba, K. Sakai, S. Watanabe, and T. Tamura, Frequency analysis of electroencephalograms and latency of photically induced average evoked responses in children with ariboflavinosis, *Tohoku J. Exp. Med.* 94:327-335 (1968).
- M. K. Horwitt, O. W. Hills, C. C. Harvey, E. Liebert, and D. L. Steinberg, Effects of dietary depletion of riboflavin, J. Nutr. 39:357-373 (1949).
- 108. M. K. Horwitt, C. C. Harvey, O. W. Hills, and E. Liebert, Correlation of urinary excretion of riboflavin with dietary intake and symptoms of ariboflavinosis, J. Nutr. 41:247-264 (1950).
- O. W. Hills, E. Liebert, D. L. Steinberg, and M. K. Horwitt, Clinical aspects of dietary depletion of riboflavin, Arch. Intern. Med. 87:682-693 (1951).
- 110. M. Lane, C. P. Alfrey, C. E. Mengel, M. A. Doherty, and J. Doherty, The rapid induction of human riboflavin deficiency with galactoflavin, *J. Clin. Invest.* 43:357-373 (1964).
- 111. R. W. Engle and P. H. Phillips, Lack of nerve degeneration in uncomplicated vitamin B<sub>1</sub> deficiency in chick and rat, J. Nutr. 16:585-596 (1938).
- 112. R. W. Engle and P. H. Phillips, Effect of certain nutritional deficiencies on various phosphorus-containing fractions of chick brain, Proc. Soc. Exp. Biol. Med. 37:553-556 (1937).
- 113. R. W. Engle and P. H. Phillips, Effect of riboflavin-low diets upon nerves, growth and reproduction in the rat, Proc. Soc. Exp. Biol. Med. 40: 597-598 (1939).
- 114. G. V. Mann, P. L. Watson, A. McNally, and J. Goddard, Primate nutrition. II. Riboflavin deficiency in the Cebus monkey and its diagnosis, J. Nutr. 47:225-241 (1952).
- 115. H. R. Street, G. R. Cowgill, and H. M. Zimmerman, Further observations of riboflavin deficiency in the dog, J. Nutr. 22:7-24 (1941).
- S. W. Lippincott and H. P. Morris, Pathological changes associated with riboflavin deficiency in the mouse, J. Natl. Cancer Inst. 2:601-610 (1942).
- 117. M. M. Wintrobe, W. H. Buschke, R. H. Follis, Jr., and S. Humphreys, Riboflavin deficiency in swine with special reference to occurrence of cataracts, *Bull. Johns Hopkins Hosp.* 75:102-114 (1944).
- 118. C. S. Lai and G. A. Ransome, Burning-feet syndrome: Case due to malabsorption and responding to riboflavin, *Br. Med. J.* 2:151-152 (1970).