MethanoGram

Michał Burdukiewicz, Przemysław Gagat, Michał Gaworski, Sławomir Jabłoński, Paweł Mackiewicz, Marcin Łukaszewicz

Contents

Introduction	1
Tuning and evaluation of MethanoGram	1
Datasets	1
Tuning procedure	2
Results of tuning	3

Introduction

MethanoGram is a predictor of culture conditions of methanogenes. Using random forests trained on n-gram encoded 16 rRNA and mcrA sequences, MethanoGram is able to estimate:

- growth rate,
- growth doubling time [h],
- optimal growth temperature,
- optimal growth pH,
- optimal growth NaCl.

Here we document the process of tuning and evalutation of set of classifiers constituting MethanoGram.

Tuning and evaluation of MethanoGram

Datasets

To train MethanoGram we used n-grams (subsequences of length n) extrated from mcrA and 16 rRNA sequences found in the PhyMet² database. We chose only sequences for which we were able to identify all culture conditions described in the database (both optimal and non-optimal). Thus, we chose only records that have known 16 rRNA sequence, mcrA sequence and all culture conditions (growth rate, growth doubling time, optimal growth temperature, growth temperature, optimal growth pH, growth pH, optimal growth NaCl, growth NaCl).

We considered two different sets of 16 rRNA sequences and three different sets of mcrA sequences. We removed all sequences containing atypical or unknown nucleotides (b, d, k, m, n, r, s, v, w, y). After purification steps described above we ended with 60 methanogenes (Fig. 1).

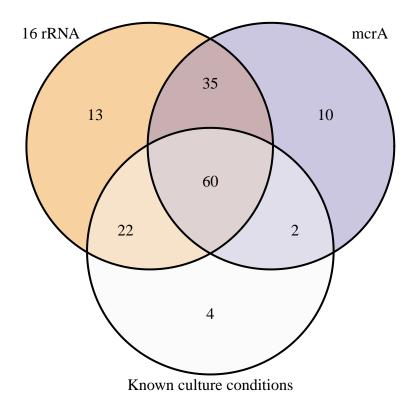


Figure 1: The Venn diagram of methanogens species in the analysis.

Tuning procedure

We chose the random forests implementated in the **ranger** R package for estimation of culture conditions, because of its speed and high accuracy. To find optimal values of hyperparameters, we performed a nested cross-validation of random forest classifiers. The inner loop was a 5-fold cross-validation and the outer loop was more demanding 3-fold cross-validation.

We have optimized three hyperparameters related to the random forest algorithm: a number of variables to possibly split at in each node, a number of trees in the forest and a minimal node size. In the tuning procedure we also incorporated different data sources, levels of feature selection and n-gram lengths.

n-gram length

We considered continous 2-, 3-, 4- and 5-grams. The number of possible n-grams for a nucleotide sequence is equal to 4^n , so the number of feature ranges between 16 (for 2-grams) to 1024 (for 5-grams). Since further increases in the n-gram size were not providing the algorithm with satisfying descrese in the error, we did not considered longer n-grams.

n-gram source

The algorithm was trained on n-grams extracted from:

- 16 rRNA,
- mcrA,
- 16 rRNA and mcrA.

In the third case, n-grams were annotated by their source. For example, in the case of bigrams, GA_RNA and GA_mcrA (bigram GA coming from RNA and mcrA) were treated as two different features.

Feature selection

To select the most informative n-grams, we used Pearson's correlation between the feature and the target as implemented in the Rfast package. We retained a fraction of features instead of an absolute number of features to keep the feature selection consistent between datasets with varying number of features. We considered following fractions of features: 0.25 and 0.5. In all cases, the strictest feature selection (0.25) proved to create the most efficient classifiers.

We also considered a predictor without any feature selection, but it consistently had the worst performance (results not shown).

Number of variables to possibly split at in each node (mtry)

In addition to the standard number of variables to possibly split at in each node for regresion tasks ($\frac{1}{3}$ of all considered features), we have also examined $\frac{1}{2}$ and $\frac{1}{4}$. In most cases, $\frac{1}{4}$ was the most optimal.

Minimal node size

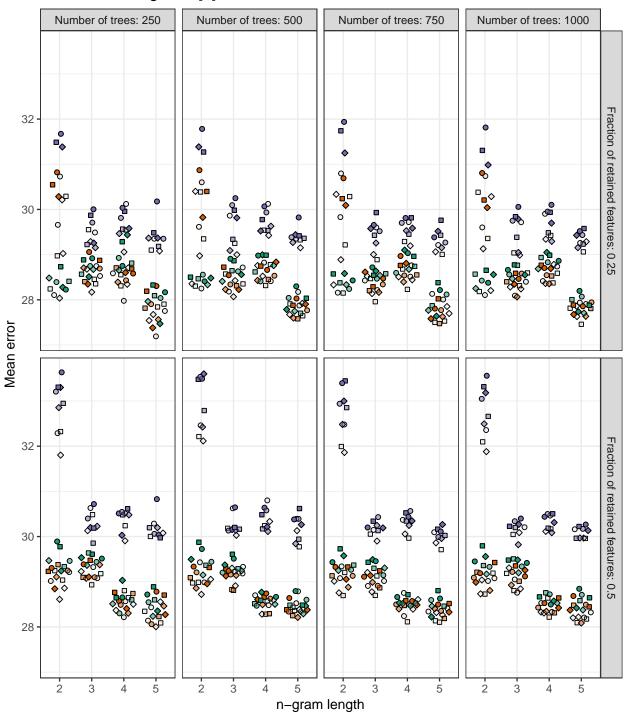
Aside from the optimal number of variables to possibly split advised by literature (5), we have also considered 3 and 7 variables. There were no visible patterns in the optimal value of the minimum node size, aside from the fact that the value advised by the literature was never producing the best-performing predictors.

Results of tuning

Below are included beeswarm plots for all parameters tuned for MethanoGram. The shape of points represents minimal node size and transparency distinguish between different mumber of variables to possibly split at in each node (mtry).

The best combinations of parameters for each condition can be also accessed in the table 1.

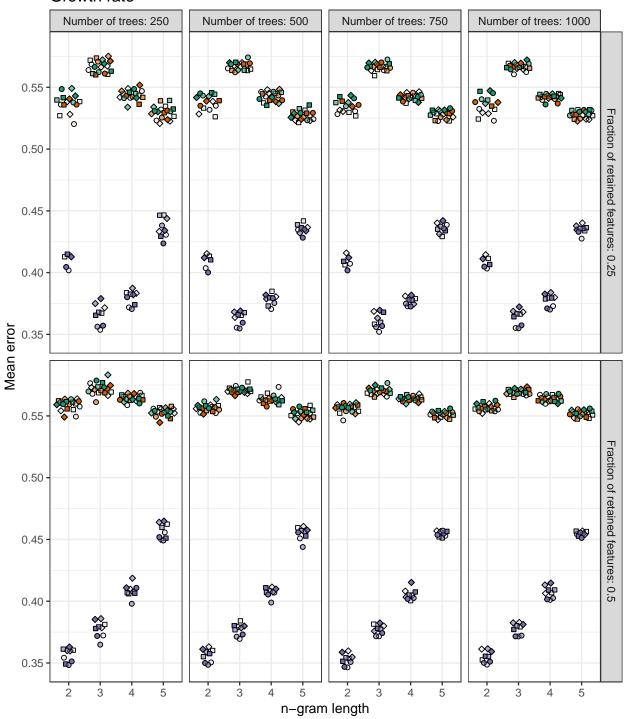
Growth doubling time [h]



Minimum node size \circ 3 \square 5 \diamond 7

Fraction of of variables to possibly split at in each node $\frac{1}{4} \cdot \frac{1}{3} \cdot \frac{1}{2}$

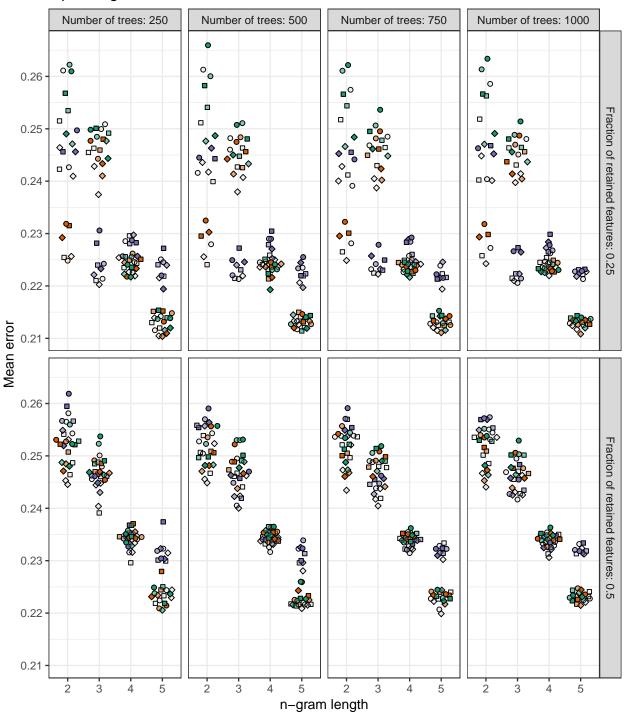
Growth rate



Minimum node size \circ 3 \circ 5 \diamond 7

Fraction of of variables to possibly split at in each node \circ $\frac{1}{4}$ \circ $\frac{1}{3}$ \bullet

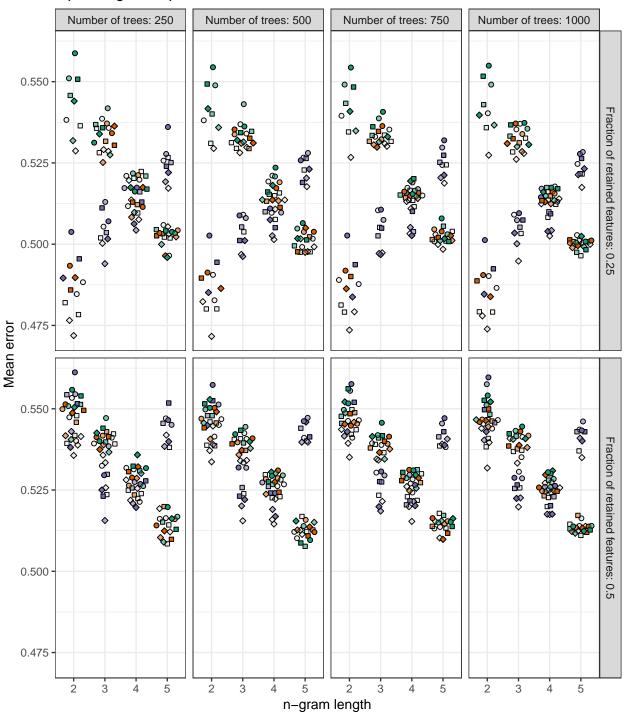
Optimal growth NaCl



Minimum node size \circ 3 \circ 5 \diamond 7

Fraction of of variables to possibly split at in each node \circ $\frac{1}{4}$ \circ $\frac{1}{3}$ \circ $\frac{1}{2}$

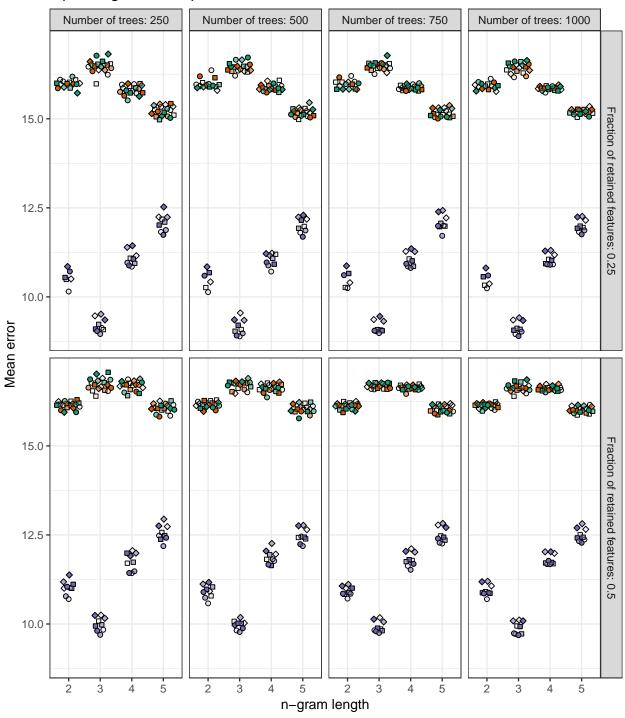
Optimal growth pH



Minimum node size \circ 3 \circ 5 \diamond 7

Fraction of of variables to possibly split at in each node \circ $\frac{1}{4}$ \circ $\frac{1}{3}$ \bullet $\frac{1}{2}$ Data source \circ Both \circ 16 rRNA \circ mcrA

Optimal growth temp.



Minimum node size ○ 3 □ 5 ◇ 7

Fraction of of variables to possibly split at in each node $\frac{1}{4} \cdot \frac{1}{3} \cdot \frac{1}{2}$

Mean error	0.35	27.19	8.89	0.47	0.21
Minimal node size	3	အ	အ	7	7
Number of trees	750	250	200	200	250
\mid mtry \mid \mid	0.25	0.25	0.25	0.25	0.33
Feature selection	0.25	0.25	0.25	0.25	0.25
Source	mcrA	16 rRNA	mcrA	mcrA	16 rRNA
n-gram length	3	ಬ	က	2	ಬ
Condition	Growth rate	\mid Growth doubling time [h]	Optimal growth temp.	Optimal growth pH	Optimal growth NaCl

Table 1: Results of nested cross-validation. mtry denotes fraction of variables to possibly split at in each node.