signalHsmm: prediction of malarial signal peptides

Michał Burdukiewicz^{1*}, Piotr Sobczyk², Paweł Błażej¹, Paweł Mackiewicz¹

¹University of Wrocław, Department of Genomics,

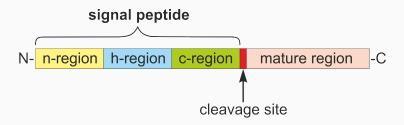
²Wrocław University of Science and Technology, Faculty of Pure and Applied Mathematics,

Signal peptides

Secretory signal peptides:

- are short (20-30 residues) N-terminal amino acid sequences forming α -helices,
- direct a protein to the endomembrane system and next to the extracellular localization,
- are universal enough to direct properly proteins in different secretory systems; artifically introduced bacterial signal peptides can guide proteins in mammals (Nagano and Masuda, 2014) and plants (Moeller et al., 2009),
- tag hormons, immune system proteins, structural proteins, and metabolic enzymes.

Architecture



Signal peptides possess three distinct domains with variable length and characteristic amino acid composition (Hegde and Bernstein, 2006):

- n-region: mostly basic residues (Nielsen and Krogh, 1998),
- h-region: strongly hydrophobic residues (Nielsen and Krogh, 1998),
- c-region: a few polar, uncharged residues (Jain et al., 1994).

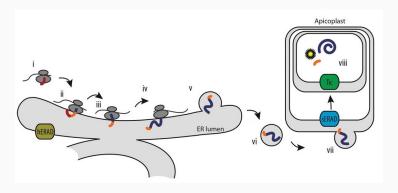
Malarial signal peptides

Apicoplast

Four membrane-bounded plastid responsible for several biochemical pathways including the biosynthesis of fatty acids, isoprenoids and haem.

Apicoplast

The signal peptide is required for targeting the apicoplast protein to the endomembrane system, whereas the transit peptide is required to traffic the protein to the apicoplast.

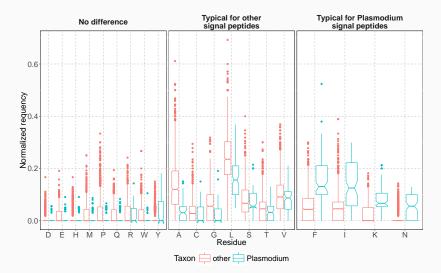


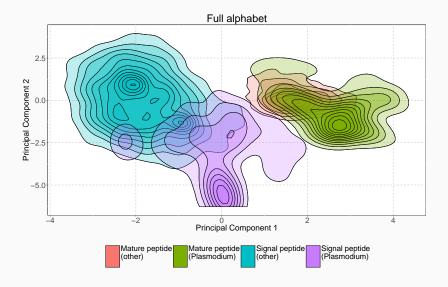
Kalanon and McFadden (2010)

Apicoplast

The absence of a metabolic counterpart in human host make the apicoplast proteins promising targets for anti-malarial drug development.

Heavy adenine-thymine bias of parasitic genomes alters amino acid composition of malarial signal peptides making them hard to predict using software trained on other Eukaryotes.





Methods

Do signal peptides require the exact sequence of amino acids?

Even nonbiological sequences can be effective signal peptides provided they fulfill general requirements (Tonkin et al., 2008).

NH₂-SKINNYSLINKYKINKYTHING-COOH - targets apicoplast.

 $\mathrm{NH}_2\text{-}\mathrm{ITWILLNEVERTARGETPLASTID}\text{-}\mathrm{COOH}$ - does not target apicoplast.

Reduced amino acid alphabets

To date, several reduced amino acid alphabets have been proposed, which have been applied to (among others) protein folding and protein structure prediction.

Novel reduced amino acid alphabets

13 physicochemical properties handpicked from AAIndex database relevant to

Novel reduced amino acid alphabets

Property name	Amino acid scale	
Size	Size	
Size	Molecular weight	
Size	Residue volume	
Size	Bulkiness	
Hydrophobicity	Normalized hydrophobicity scales for $lpha$ -proteins	
Hydrophobicity	Consensus normalized hydrophobicity scale	
Hydrophobicity	Hydropathy index	
Hydrophobicity	Surrounding hydrophobicity in α -helix	
Polarity	Polarity	
Polarity	Mean polarity	
Occurrence in α -helices	Signal sequence helical potential	
Occurrence in α -helices	Normalized frequency of N-terminal helix	
Occurrence in α -helices	Relative frequency in α -helix	

13

Cross-validation

We built 96 reduced amino acid alphabets (each based on one scale per a given property category) and evaluated them in cross-validation experiment using eukaryotic sequences.

Results

Amino acids
D, E, H, K, N, Q, R
G, P, S, T, Y
F, I, L, M, V, W
A, C

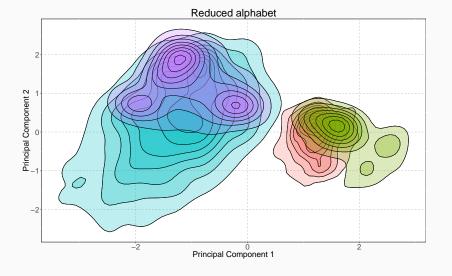
I. Amino acids absent in h-region.

Group	Amino acids
1	D, E, H, K, N, Q, R
Ш	G, P, S, T, Y
Ш	F, I, L, M, V, W
IV	A, C

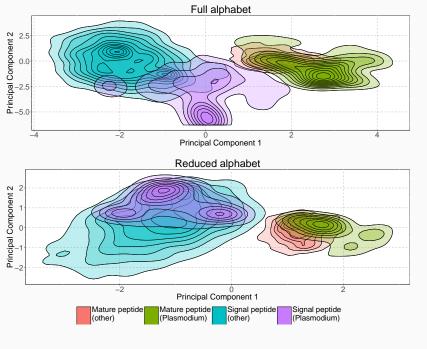
II. Amino acids common in c-region.

Amino acids
D, E, H, K, N, Q, R
G, P, S, T, Y
F, I, L, M, V, W
A, C

III. Amino acids common in h-region.



Signal peptides, after the reduction of the amino acid alphabet, group together despite their different origins.



Benchmark

Benchmark data set: 51 proteins with signal peptide and 211 proteins without signal peptide from members of *Plasmodiidae*.

Benchmark

	MCC	AUC
signalP 4.1 (no tm) (Petersen et al., 2011)	0.6872	0.8667
signalP 4.1 (tm) (Petersen et al., 2011)	0.6196	0.7951
signalP 3.0 (NN) (Bendtsen et al., 2004)	0.7220	0.8938
signalP 3.0 (HMM) (Bendtsen et al., 2004)	0.5553	0.7734
Phobius (Käll et al., 2004)	0.5895	0.7880
signalHsmm-2010	0.7409	0.9262
signalHsmm-2010 (hom. 50%)	0.7621	0.9384
signalHsmm-2010 (raw aa)	0.6853	0.8718
signalHsmm-1987	0.7271	0.9063
signalHsmm-1987 (hom. 50%)	0.7194	0.9090
signalHsmm-1987 (raw aa)	0.6350	0.8350

signalHsmm1987: trained on data set of 496 eukaryotic proteins with signal peptides added before year 1987.

Conclusions

Eukaryotic signal peptides have very similar amino acid composition in their regions considering only the physicochemical properties of residues.

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- Paweł Mackiewicz.
- biogram package
 (https://cran.r-project.org/package=biogram):
 - Piotr Sobczyk,
 - Chris Lauber.

References I

References

- Bendtsen, J. D., Nielsen, H., von Heijne, G., and Brunak, S. (2004). Improved prediction of signal peptides: Signalp 3.0. Journal of Molecular Biology, 340(4):783 – 795.
- Hegde, R. S. and Bernstein, H. D. (2006). The surprising complexity of signal sequences. *Trends in Biochemical Sciences*, 31(10):563–571.

References II

- Jain, R. G., Rusch, S. L., and Kendall, D. A. (1994). Signal peptide cleavage regions. functional limits on length and topological implications. *The Journal of Biological Chemistry*, 269(23):16305–16310.
- Kalanon, M. and McFadden, G. I. (2010). Malaria, Plasmodium falciparum and its apicoplast. *Biochemical Society Transactions*, 38(3):775–782.
- Käll, L., Krogh, A., and Sonnhammer, E. L. L. (2004). A combined transmembrane topology and signal peptide prediction method. *Journal of Molecular Biology*, 338(5):1027–1036.

References III

Moeller, L., Gan, Q., and Wang, K. (2009). A bacterial signal peptide is functional in plants and directs proteins to the secretory pathway. *Journal of Experimental Botany*, 60(12):3337–3352.

Nagano, R. and Masuda, K. (2014). Establishment of a signal peptide with cross-species compatibility for functional antibody expression in both escherichia coli and chinese hamster ovary cells. *Biochemical and Biophysical Research Communications*, 447(4):655 – 659.

References IV

- Nielsen, H. and Krogh, A. (1998). Prediction of signal peptides and signal anchors by a hidden markov model. *Proceedings / ... International Conference on Intelligent Systems for Molecular Biology ; ISMB. International Conference on Intelligent Systems for Molecular Biology*, 6:122–130.
- Petersen, T. N., Brunak, S., von Heijne, G., and Nielsen, H. (2011). SignalP 4.0: discriminating signal peptides from transmembrane regions. *Nature Methods*, 8(10):785–786.
- Tonkin, C. J., Foth, B. J., Ralph, S. A., Struck, N., Cowman, A. F., and McFadden, G. I. (2008). Evolution of malaria parasite plastid targeting sequences. *Proceedings of the National Academy of Sciences of the United States of America*, 105(12):4781–4785.