

Predicting properties of biological sequences using n-gram analysis

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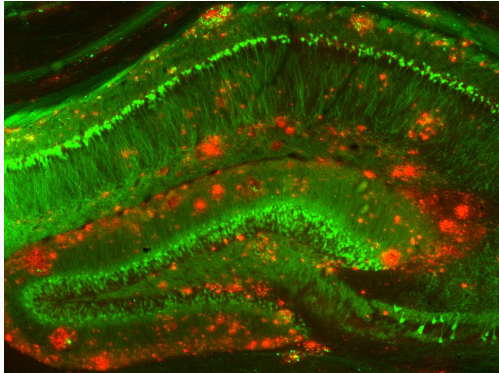
In silico research allows scientists to more efficiently design experimental studies.

Examples:

- prediction of protein properties (presence of signal peptides, amyloidogenicity),
- predicting culture conditions.

Amyloid proteins

Amyloid are proteins associated with various diseases (e.g., Alzheimer's, Creutzfeldt-Jakob's and Huntington's diseases) which are able to form harmful aggregates.

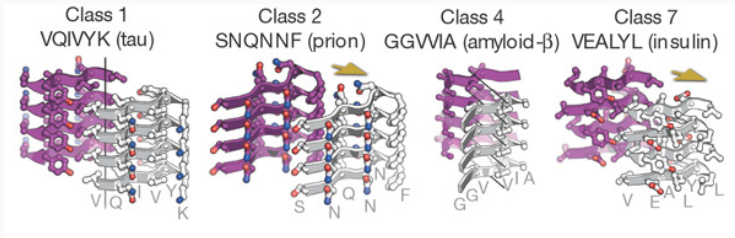


Amyloid aggregates (red) around neurons (green). Strittmatter Laboratory, Yale University.

Amyloid proteins

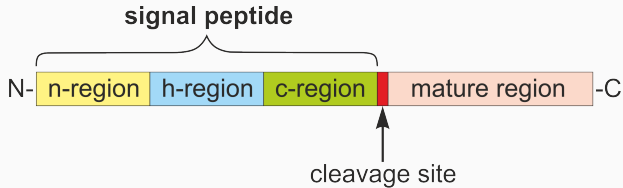
Hot-spots:

- short (6-15 amino acids),
- very high variability of amino acid composition,
- initiate amyloid aggregation,
- create specific "zipper-like" β -structures.



Sawaya et al. (2007)

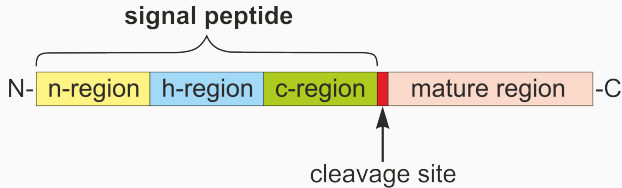
Signal peptides



Signal peptides:

- are short (20-30 residues) N-terminal amino acid sequences forming α -helices,

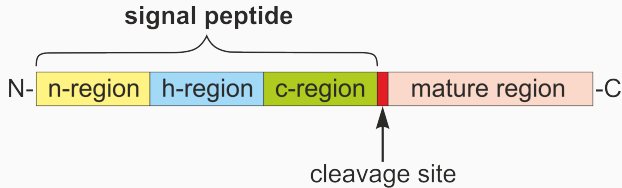
Signal peptides



Signal peptides:

- are short (20-30 residues) N-terminal amino acid sequences forming α -helices,
- direct proteins to the endomembrane system and next to extra- or intracellular localizations,

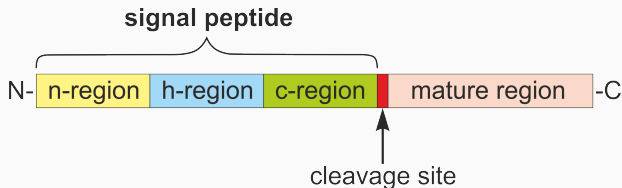
Signal peptides



Signal peptides:

- are short (20-30 residues) N-terminal amino acid sequences forming α -helices,
- direct proteins to the endomembrane system and next to extra- or intracellular localizations,
- are universal enough to direct properly proteins in different secretory systems.

Signal peptides

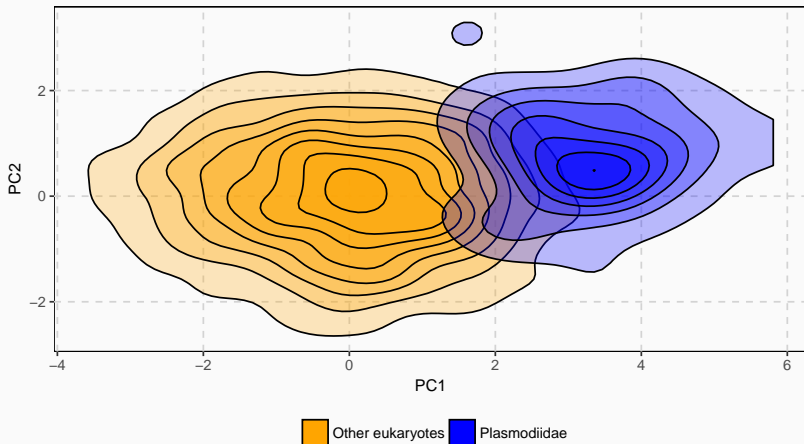


Signal peptides possess three distinct domains with variable length and characteristic amino acid composition (Hegde and Bernstein, 2006):

- n-region: mostly basic residues (Nielsen and Krogh, 1998),
- h-region: strongly hydrophobic residues (Nielsen and Krogh, 1998),
- c-region: a few polar, uncharged residues.

Signal peptides

Amino acid composition of signal peptides differ between *Plasmodium* sp. and other eukaryotes. Therefore, predictors of signal peptides do not detect malarial signal peptides accurately.



n-grams

Computational analysis of biological sequences requires converting them to features understandable by machines.

The optimal conversion of information:

- lossless,
- concise.

n-grams (k-tuples, k-mers):

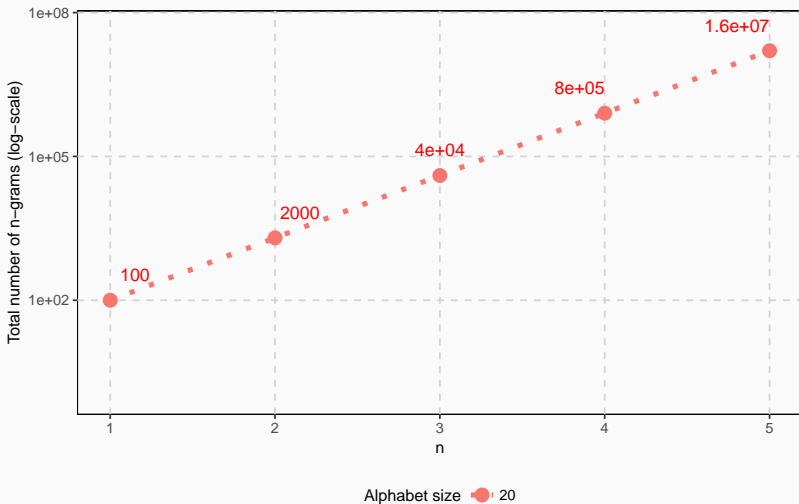
- subsequences (continuous or gapped) of n residues,
- considers the context of a specific residue.

| | P1 | P2 | P3 | P4 | P5 |
|----|----|----|----|----|----|
| S1 | M | R | K | L | Y |

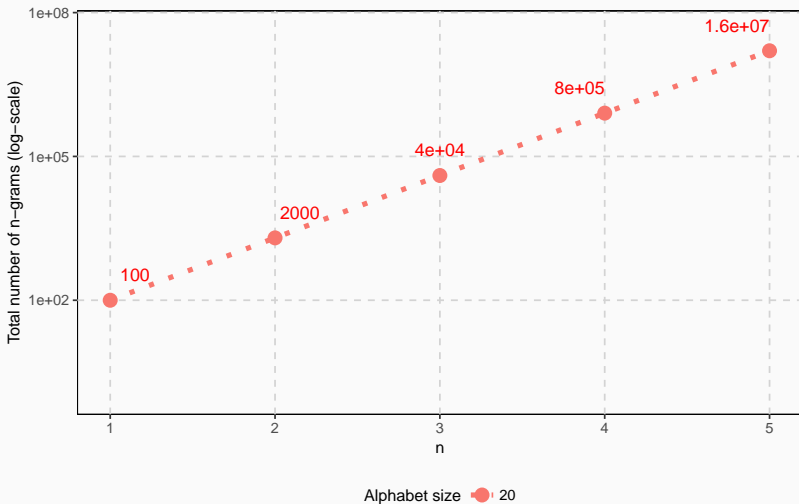
2-grams: MR, RK, KL, LY

2-grams (gap 1): M – K, R – L, K – Y

3-grams: MRK, RKL, KLY



n-grams create very large datasets, which are hard to process and analyze.



Longer n-grams are more informative, but create larger feature spaces.

Permutation Tests

Informative n-grams are usually selected using permutation tests.

During a permutation test we shuffle randomly class labels and compute a defined statistic (e.g. information gain). Values of statistic for permuted data are compared with the value of statistic for original data.

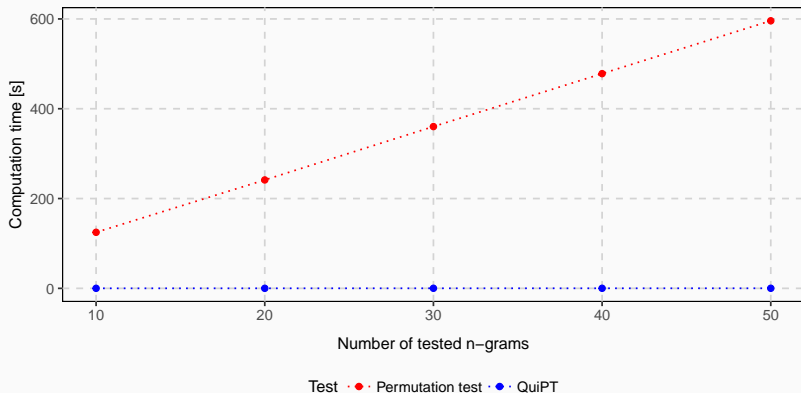
$$\text{p-value} = \frac{N_{T_P > T_R}}{N}$$

$N_{T_P > T_R}$: number of cases, where T_P (permuted test statistic) has more extreme values than T_R (test statistic for original data).

N : number of permutations.

Quick Permutation Test is a fast alternative to permutation tests for n-gram data. It also allows precise estimation of p-value.

QuiPT is available as part of the **biogram** R package.



QuiPT is faster than classical permutation tests and returns exact p-values.

Simplified alphabets

Simplified alphabets:

- are based on grouping amino acids with similar physicochemical properties,
- ease computational analysis of a sequence (Murphy et al., 2000),
- create more interpretable models.

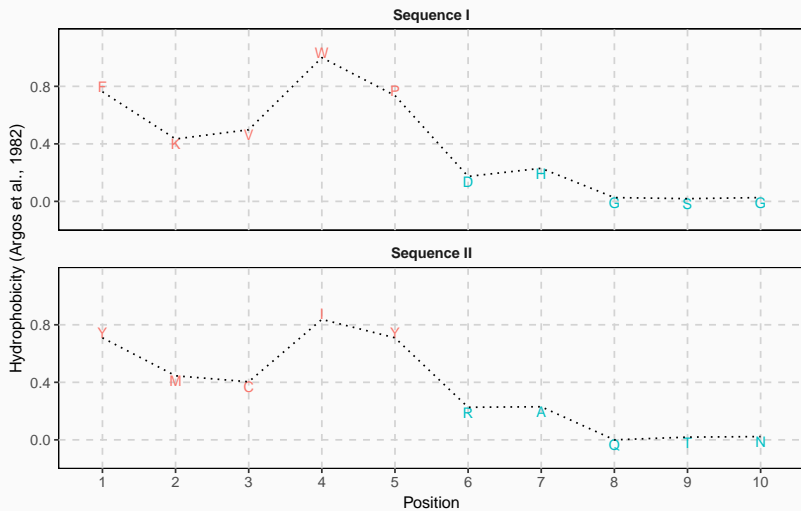
Two sequences that are drastically different considering their amino acids composition can have the same physicochemical properties.

Sequence I:

FKVWPDHGSG

Sequence II:

YMCIIYRAQTN

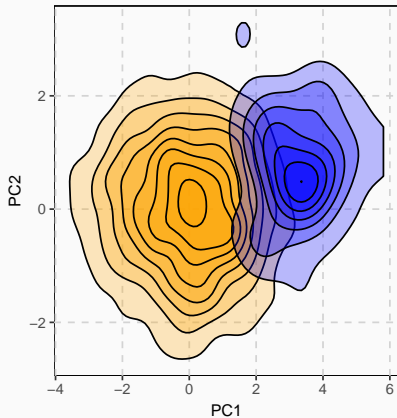


| Subgroup | Amino acid |
|----------|------------------------------|
| 1 | C, I, L, K, M, F, P, W, Y, V |
| 2 | A, D, E, G, H, N, Q, R, S, T |

Sequence I: FKVWPDHGSG 1111122222

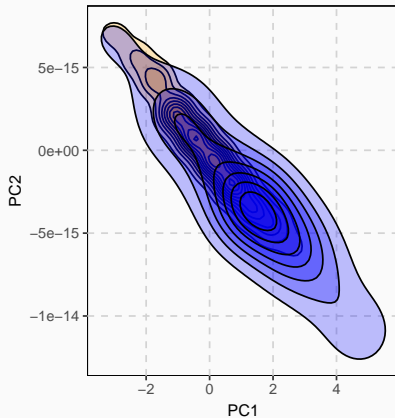
Sequence II: YMCIIYRAQTN 1111122222

Full alphabet



Other eukaryotes Plasmodiidae

Simplified alphabet

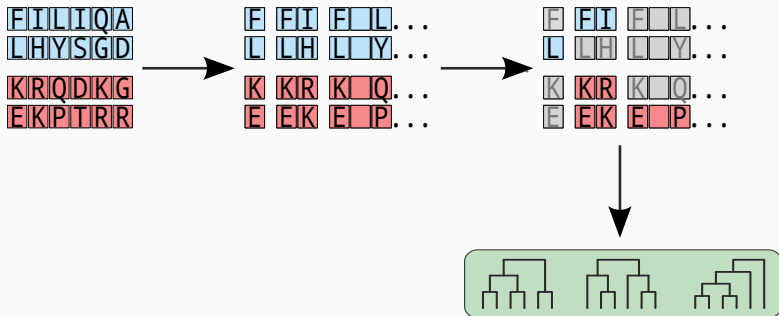


Other eukaryotes Plasmodiidae

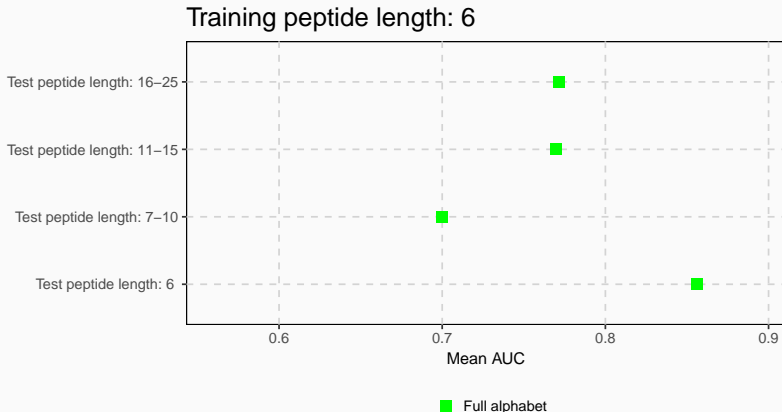
PCA of amino acid frequency in signal peptides.

Prediction of amyloidogenicity

AmyloGram: n-gram based tool for prediction of amyloid proteins (Burdukiewicz et al., 2016).



Cross-validation



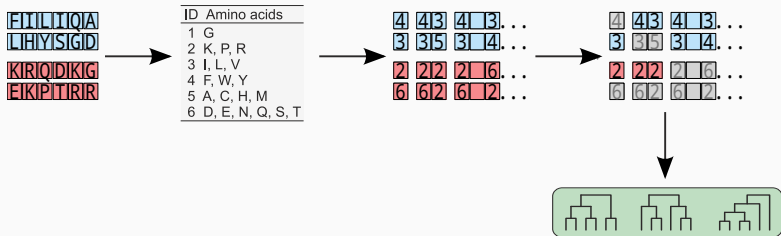
AUC (Area Under the Curve) measures the performance of a classifier (1 - classifier always properly recognizes amyloid proteins, 0 - classifier never properly recognizes amyloid proteins).

Does amyloidogenicity depend on the exact sequence of amino acids?

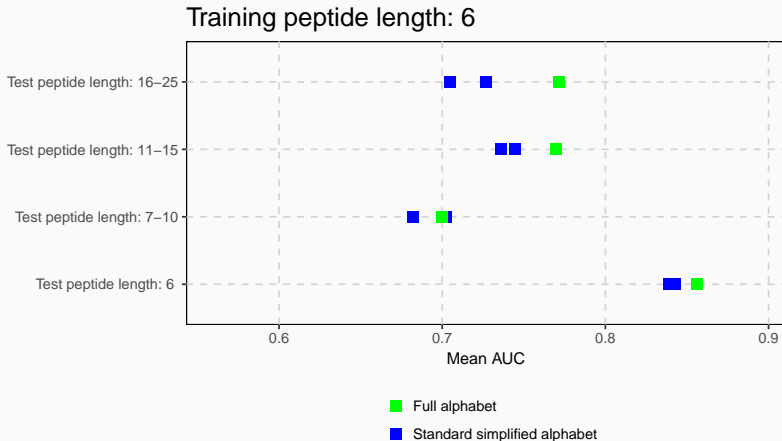
Standard simplified amino acid alphabets

To date, several simplified amino acid alphabets have been proposed, which have been applied to (among others) protein folding and protein structure prediction (Kosiol et al., 2004; Melo and Marti-Renom, 2006).

Standard simplified amino acid alphabets



Cross-validation

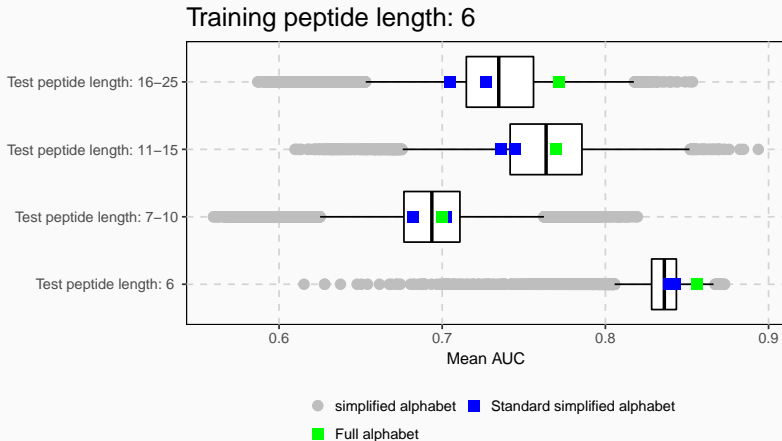


Standard simplified amino acid alphabets do not enhance discrimination between amyloidogenic and non-amyloidogenic proteins.

Novel simplified amino acid alphabets

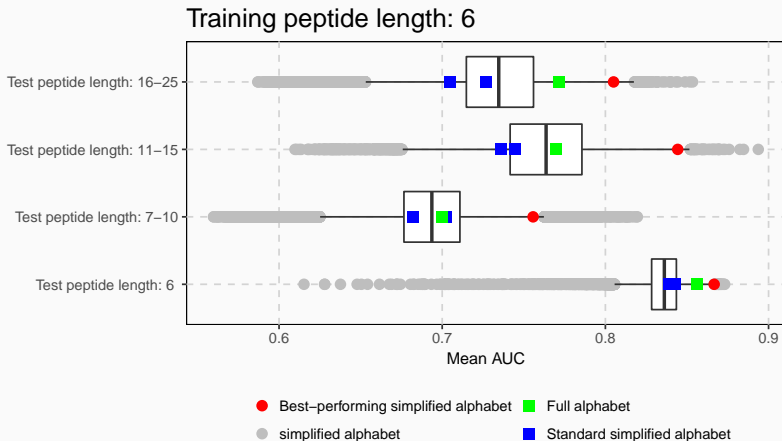
- 17 measures handpicked from AAIndex database:
 - size of residues,
 - hydrophobicity,
 - solvent surface area,
 - frequency in β -sheets,
 - contactivity.
- 524 284 amino acid simplified alphabets with different level of amino acid alphabet reduction (three to six amino acid groups).

Cross-validation



Hinges of boxes correspond to the 0.25 and 0.75 quartiles. The bar inside the box represents the median. The gray circles correspond to the simplified alphabets with the AUC outside the 0.95 confidence interval.

The best-performing simplified alphabet



The best-performing simplified alphabet

| Subgroup ID | Amino acids |
|-------------|------------------|
| 1 | G |
| 2 | K, P, R |
| 3 | I, L, V |
| 4 | F, W, Y |
| 5 | A, C, H, M |
| 6 | D, E, N, Q, S, T |

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Group 3 and 4 - hydrophobic amino acids.

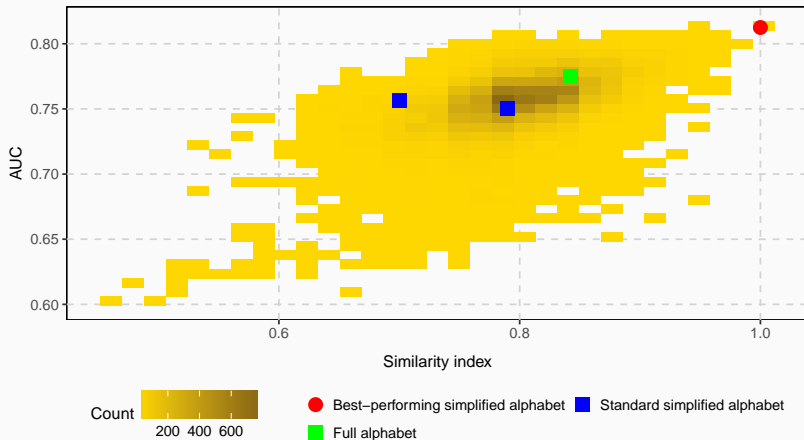
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Group 2 - charged breakers of β -structures.

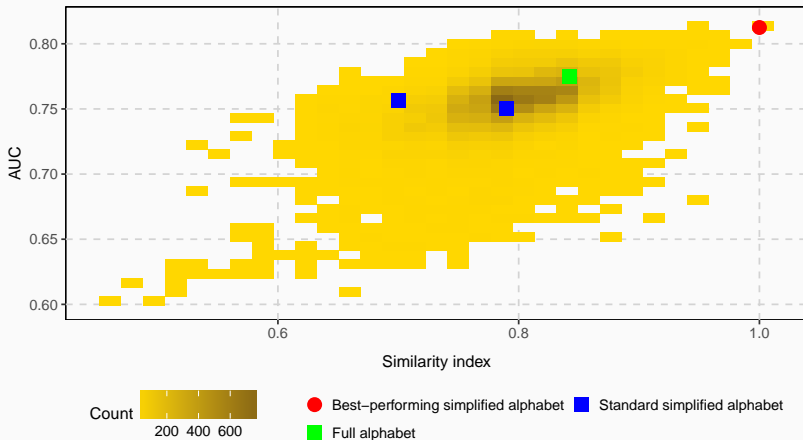
Is the best-performing simplified amino alphabet associated with amyloidogenicity?

Similarity index



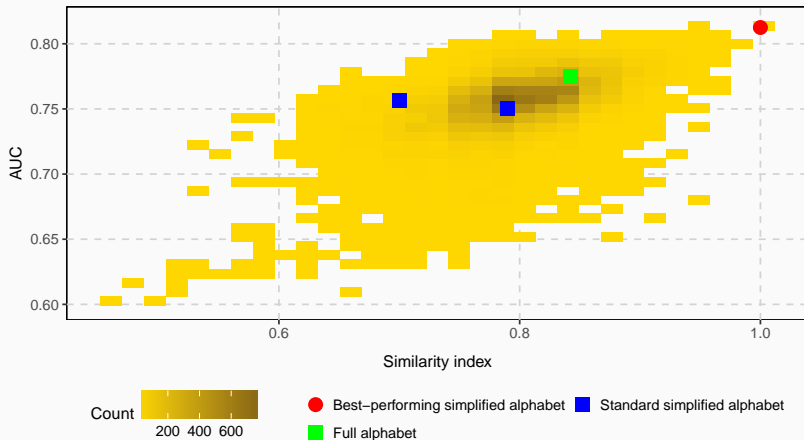
Similarity index (Stephenson and Freeland, 2013) measures the similarity between two simplified alphabets (1 - identical, 0, totally dissimilar).

Similarity index



The color of a square is proportional to the number of simplified alphabets in its area.

Similarity index



The correlation between mean AUC and similarity index is significant ($p\text{-value} \leq 2.2^{-16}$; $\rho = 0.51$).

Benchmark results

| Classifier | AUC | MCC |
|---|---------------|---------------|
| AmyloGram | 0.8972 | 0.6307 |
| PASTA 2.0 (Walsh et al., 2014) | 0.8550 | 0.4291 |
| FoldAmyloid (Garbuzynskiy et al., 2010) | 0.7351 | 0.4526 |
| APPNN (Família et al., 2015) | 0.8343 | 0.5823 |

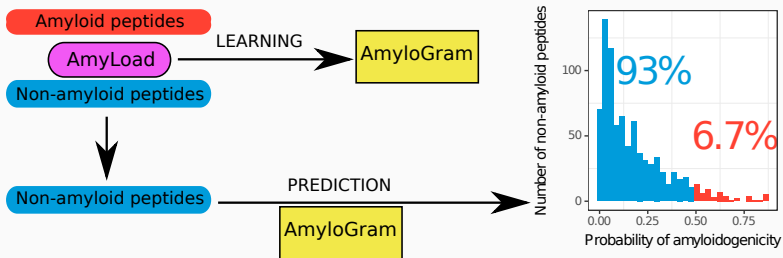
The predictor based on the best-performing alphabet, called AmyloGram, was benchmarked against the most popular tools for the detection of amyloid peptides using an external data set *pep424*.

Benchmark results

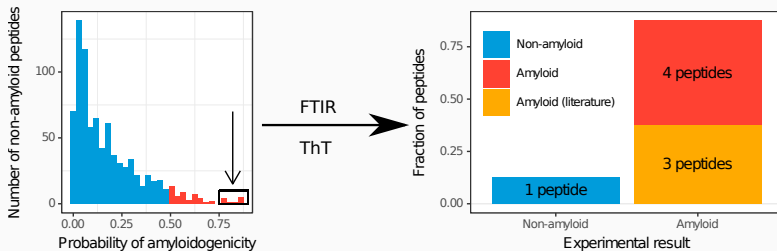
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MCC (Matthew's Correlation Coefficient) measures the performance of a classifier (1 - classifier always properly recognizes amyloid proteins, -1 - classifier never properly recognizes amyloid proteins).

Experimental validation

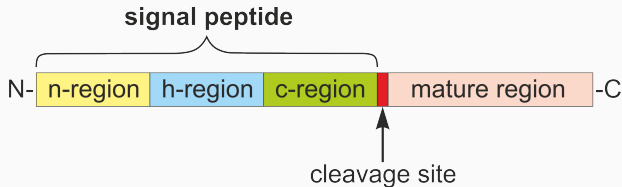


Experimental validation



Other applications

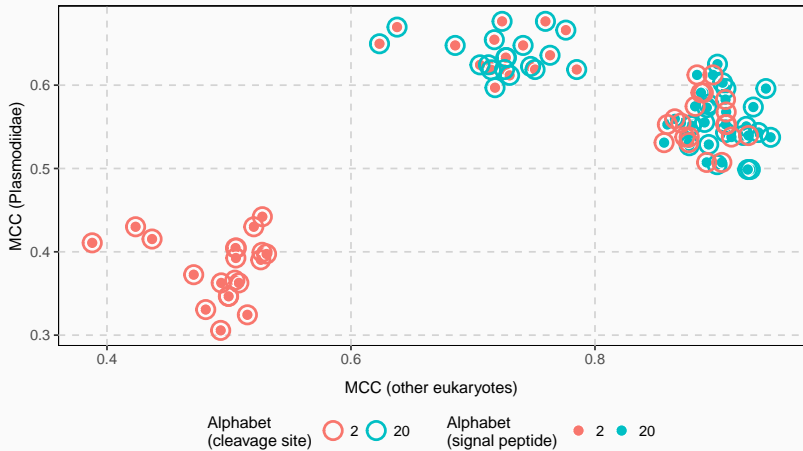
Signal peptide prediction



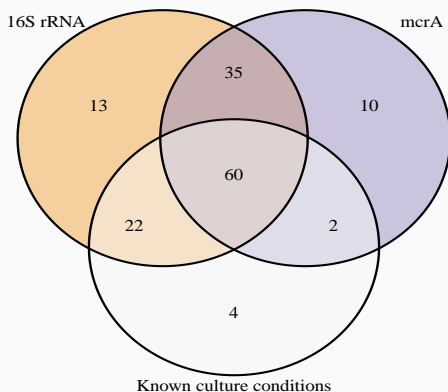
SignalP 4.1 (Petersen et al., 2011) combines output of two separate predictors:

- cleavage site,
- signal peptide.

Signal peptide prediction



Prediction of culturing conditions



`metanogen.biotech.uni.wroc.pl` (Jabłoński et al., 2015)

Prediction of culturing conditions

Results of nested cross-validation of MethanoGram.

| Culturing condition | Mean error |
|--|------------|
| Growth rate [h^{-1}] | 0.35 |
| Growth doubling time [h] | 27.19 |
| Optimal growth temp. [$^{\circ}\text{C}$] | 8.89 |
| Optimal growth pH | 0.47 |
| Optimal growth NaCl [mol/dm^3] | 0.21 |

Summary

1. Created algorithms effectively filtering n-grams.
2. Introduced new methods for search of simplified amino acids.
3. Implemented novel algorithms in the **R** package *AmyloGram*.
4. Applied the n-gram analysis framework to:
 - prediction of amyloids (AmyloGram),
 - prediction of atypical signal peptides,
 - prediction of culture conditions of methanogenes (MethanoGram).

Summary

Web servers:

- **AmyloGram:**
`http://www.smorfland.uni.wroc.pl/shiny/AmyloGram/`
- **MethanoGram:** `http://www.smorfland.uni.wroc.pl/shiny/MethanoGram/`
- **signalHsmm:** `http://www.smorfland.uni.wroc.pl/shiny/signalHsmm/`

Software packages:

- **biogram:**
`https://cran.r-project.org/package=biogram`
- **AmyloGram:**
`https://cran.r-project.org/package=AmyloGram`

Mentors:

- **Paweł Mackiewicz (University of Wrocław).**
- Lars Kaderali (University of Greifswald).
- Małgorzata Kotulska (Wrocław University of Science and Technology).
- Marcin Łukaszewicz (University of Wrocław).
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References

- Burdukiewicz, M., Sobczyk, P., Rödiger, S., Duda-Madej, A., Mackiewicz, P., and Kotulska, M. (2016). Prediction of amyloidogenicity based on the n-gram analysis. Technical Report e2390v1, PeerJ Preprints.
- Família, C., Dennison, S. R., Quintas, A., and Phoenix, D. A. (2015). Prediction of Peptide and Protein Propensity for Amyloid Formation. *PLOS ONE*, 10(8):e0134679.

References II

- Garbuzynskiy, S. O., Lobanov, M. Y., and Galzitskaya, O. V. (2010). FoldAmyloid: a method of prediction of amyloidogenic regions from protein sequence. *Bioinformatics (Oxford, England)*, 26(3):326–332.
- Hegde, R. S. and Bernstein, H. D. (2006). The surprising complexity of signal sequences. *Trends in Biochemical Sciences*, 31(10):563–571.
- Jabłoński, S., Rodowicz, P., and Łukaszewicz, M. (2015). Methanogenic archaea database containing physiological and biochemical characteristics. *Int J Syst Evol Microbiol*, 65(4):1360–1368.

References III

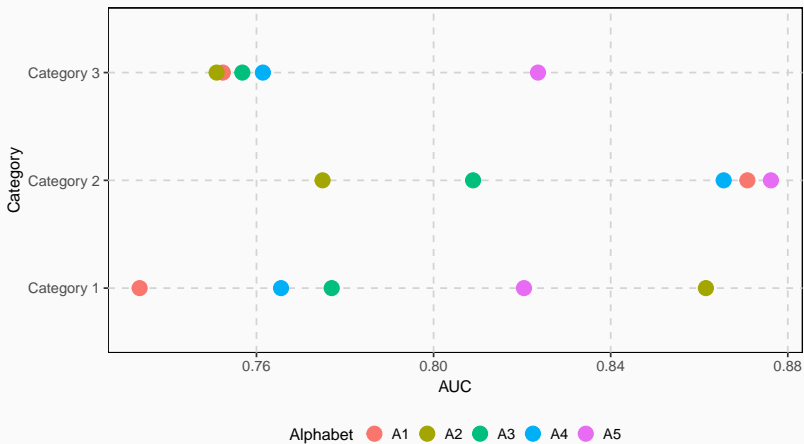
- Kosiol, C., Goldman, N., and Buttimore, N. H. (2004). A new criterion and method for amino acid classification. *Journal of Theoretical Biology*, 228(1):97–106.
- Melo, F. and Marti-Renom, M. A. (2006). Accuracy of sequence alignment and fold assessment using reduced amino acid alphabets. *Proteins*, 63(4):986–995.
- Murphy, L. R., Wallqvist, A., and Levy, R. M. (2000). Simplified amino acid alphabets for protein fold recognition and implications for folding. *Protein Engineering*, 13(3):149–152.

References IV

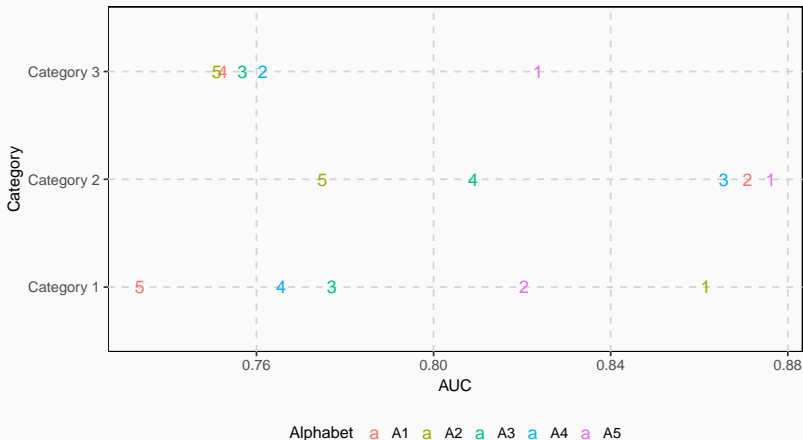
- Nielsen, H. and Krogh, A. (1998). Prediction of signal peptides and signal anchors by a hidden Markov model. *Proceedings / ... International Conference on Intelligent Systems for Molecular Biology ; ISMB. International Conference on Intelligent Systems for Molecular Biology*, 6:122–130.
- Petersen, T. N., Brunak, S., von Heijne, G., and Nielsen, H. (2011). SignalP 4.0: discriminating signal peptides from transmembrane regions. *Nature Methods*, 8(10):785–786.
- Sawaya, M. R., Sambashivan, S., Nelson, R., Ivanova, M. I., Sievers, S. A., Apostol, M. I., Thompson, M. J., Balbirnie, M., Wiltzius, J. J. W., McFarlane, H. T., Madsen, A. , Riek, C., and Eisenberg, D. (2007). Atomic structures of amyloid cross-spines reveal varied steric zippers. *Nature*, 447(7143):453–457.

- Stephenson, J. D. and Freeland, S. J. (2013). Unearthing the root of amino acid similarity. *Journal of Molecular Evolution*, 77(4):159–169.
- Walsh, I., Seno, F., Tosatto, S. C. E., and Trovato, A. (2014). PASTA 2.0: an improved server for protein aggregation prediction. *Nucleic Acids Research*, 42(W1):W301–W307.

Ranking alphabets

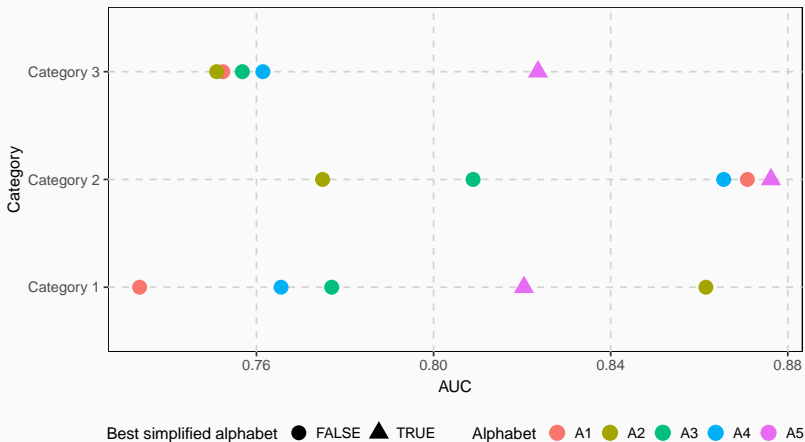


Ranking alphabets



We rank alphabets separately in all length categories assuming the rank 1 for the best AUC, rank 2 for the second best AUC and so on.

Ranking alphabets



The best-performing alphabet has the lowest sum of ranks.