

# Preventing relapse after CD19 CAR T-cell therapy for pediatric ALL: the role of transplant and enhanced CAR T cells

Aimee C. Talleur, Swati Naik, and Stephen Gottschalk

Department of Bone Marrow Transplantation and Cellular Therapy, St Jude Children's Research Hospital, Memphis, TN

CD19-specific chimeric antigen receptor (CAR) T-cell therapy has become an integral part of our treatment armamentarium for pediatric patients with relapsed or refractory B-cell acute lymphoblastic leukemia (B-ALL). However, despite initial remission rates of greater than 80%, durable remission occurs in only 40% to 50% of patients. In this review we summarize our current knowledge of the role of consolidative hematopoietic cell transplantation in the management of pediatric patients who achieved a minimal residual disease-negative complete response post CD19 CAR T-cell therapy. In addition, we review approaches to enhance effector function CD19 CAR T cells, focusing on how to improve persistence and prevent the emergence of CD19<sup>+</sup> B-ALL blasts.

## LEARNING OBJECTIVES

- Understand the role of hematopoietic stem cell transplantation post CD19 CAR T-cell therapy
- Explain common mechanisms of ALL recurrence post CD19 CAR T-cell therapy
- Explain approaches to enhance the anti-ALL activity of CD19 CAR T cells

## Introduction

CD19-redirected chimeric antigen receptor (CAR) T-cell therapy is an effective therapeutic modality for pediatric patients with relapsed and/or refractory (R/R) acute lymphoblastic leukemia (B-ALL), a patient cohort that historically was largely incurable.<sup>1,2</sup> However, despite initial remission rates of greater than 80% across studies, durable remission occurs in only 40% to 50% of patients. This includes the use of the US Food and Drug Administration-approved product tisagenlecleucel, as well as other CD19-directed CAR T-cell products evaluated in clinical trials.<sup>3-8</sup> While data suggesting risk factors for disease nonresponse or relapse have been reported, including high leukemic disease burden and a history of nonresponse to other CD19-directed therapies, it is currently unknown for which patients stand-alone CD19 CAR T-cell therapy is curative.<sup>8-12</sup>

Outcomes after CAR T-cell relapse are dismal,<sup>13</sup> and it is therefore critical to identify patients at high risk of relapse through close monitoring for CAR T-cell persistence in conjunction with frequent disease evaluations in the first year post CAR T-cell infusion. While many factors have to be taken into consideration when making post-CAR T-cell therapy decisions, a universal algorithm is yet to be developed.

Therefore, a key focus in the field includes efforts to better identify patients at higher risk of disease recurrence post CAR T-cell therapy, as well as investigation of novel treatment approaches aimed to enhance CAR T-cell efficacy.<sup>14</sup>

In this educational review, we present 2 cases highlighting the current challenges and then review the role of hematopoietic cell transplantation (HCT) post CD19 CAR T-cell therapy and approaches to enhance the anti-ALL activity of CD19 CAR T cells. Thus, the broad learning objectives are to i) understand the role of HCT post CD19 CAR T-cell therapy and ii) explain common mechanisms of therapeutic failure and approaches to enhance the anti-ALL activity of CD19 CAR T cells.

## CLINICAL CASE 1

A 6-year-old boy with standard-risk B-ALL was treated with standard chemotherapy, achieved remission at the end of induction therapy, and then suffered a relapse during maintenance therapy. He had persistent CD19<sup>+</sup> B-ALL after 2 cycles of intensive reinduction chemotherapy. He received CD19-redirected CAR T-cell therapy and

achieved remission, with no detectable clonal cells by next-generation sequencing (NGS) testing. He subsequently proceeded to a consolidative HCT with a matched related donor and remains in remission 2 years post HCT.

## CLINICAL CASE 2

An 18-year-old man with primary refractory Philadelphia chromosome-like B-ALL received CD19-redirected CAR T-cell therapy and achieved remission. At 8 months post-CAR infusion, a loss of B-cell aplasia (BCA) was noted. A bone marrow biopsy performed at that time revealed detectable disease by NGS testing, with a rising copy number on a short interval repeat marrow. In the setting of loss of BCA and rising NGS, the patient was considered at risk for impending relapse, received treatment with blinatumomab, proceeded to HCT, and remains in remission without excessive HCT-related toxicity.

## HCT post CD19 CAR T-cell therapy: experience to date

Data on the use of consolidative HCT post CD19 CAR T-cell therapy in pediatric patients are limited and come largely from single-center experience using varying CAR T-cell products (Table 1).<sup>3,4,6-8,10,15-17</sup> While at present it is difficult to draw overarching conclusions about the role of consolidative HCT in this patient population, the current experience with HCT post CD19 CAR T-cell therapy can be used to better understand potential predictors of relapse and identify patients who might benefit from HCT.

Tisagenlecleucel (Kymriah) is the only commercially available CAR T-cell product for pediatric patients with R/R CD19+ ALL. It consists of ex vivo activated and expanded autologous T cells genetically modified with a lentiviral vector encoding a CD19 CAR with a 41BB.zeta signaling domain (CD19/41BB). With a follow-up of 38.8 months, the seminal trial and the phase 2 global

registration trial (ELIANA) reported high rates of initial complete response (CR) with relapse-free survival of 76% and 59%, respectively.<sup>3,4,16</sup> Analyses of real-world use of tisagenlecleucel by the Center for International Blood and Marrow Transplant Research and the Pediatric Real-World CAR Consortium demonstrated similar response rates.<sup>13,17</sup> Notably, very few patients in these studies proceeded to a consolidative HCT, and there are limited data on those who did.

Other CD19/41BB-CAR T-cell products were evaluated in pediatric patients with R/R B-ALL in early-phase clinical trials. The PLAT-02 study, a phase 1/2 trial, utilized an institutional product infused with a defined CD4/CD8 ratio.<sup>5</sup> Among 64 treated patients, 50 were considered eligible for potential HCT post CAR T-cell therapy. Of these, 23 proceeded to HCT (second HCT, n=10) at a median of 3 months post infusion. Rates of relapse were lower in the consolidative HCT group (5/23 patients) compared to the non-HCT group (19/27 patients). One patient died post HCT secondary to treatment complications. HCT-naïve patients had a significantly improved leukemia-free survival compared to non-HCT-naïve patients. Regardless of prior HCT status, patients with early loss of BCA ( $\leq 63$  days post infusion) who underwent consolidative HCT had improved leukemia-free survival compared to those who did not proceed to HCT.<sup>18</sup> Another study with an institutional CD19/41BB-CAR T-cell product demonstrated CRs in 9 of 12 patients treated. Post CR, 5 patients (all HCT naïve, including clinical case 1) went on to consolidative HCT at a median of 2.7 months post-CAR infusion. All these patients remain in remission, with 1 patient dying secondary to transplant-related complications. Conversely, the 4 who did not proceed with HCT all subsequently relapsed. Three of these patients were not considered good candidates for HCT due to prior HCT status or a history of extramedullary disease, and 1 relapsed prior to planned HCT.<sup>8</sup>

The benefit of consolidative transplant in pediatric patients treated with T-cell products that express CD19 CARs with a CD28.zeta signaling domain (CD19/CD28) has been demonstrated by several groups. In one study, 15 of 18 responding

**Table 1. Selected studies reporting outcomes of CD19 CAR T-cell therapy +/- consolidative HCT**

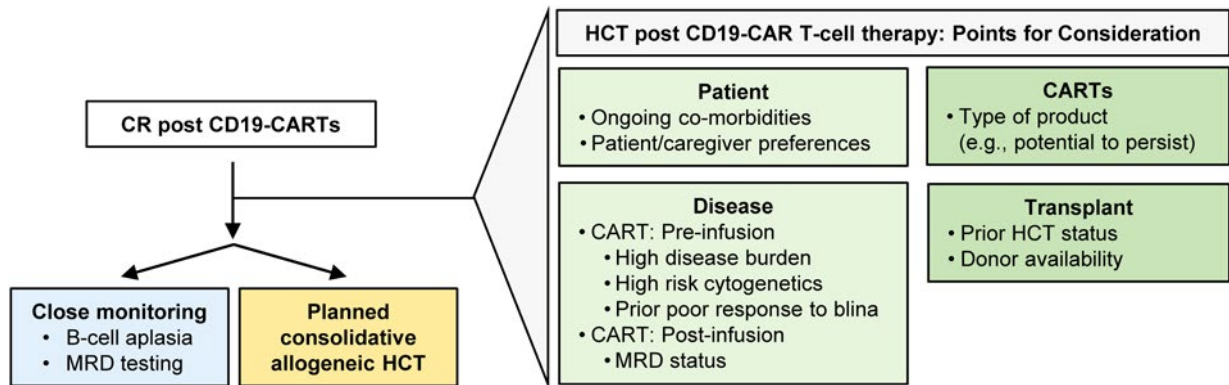
Center/consortium/study	Trial phase	Costim	Patients (no.)	Initial CR (%)	HCT in CR# <sup>a</sup>	Relapse post HCT vs no HCT
CHOP <sup>3</sup>	1	41BB	30	90	3	Relapse: NR vs 8/23 no HCT
ELIANA <sup>4,16</sup>	2		79	81	11	Relapse: 0/8 <sup>b</sup> vs NR
Seattle <sup>5</sup>	1/2		64		23	Relapse: 5/23 vs 19/27 no HCT
St Jude <sup>6</sup>	1		12	75	5	Relapse: 0/5 vs 4/4 no HCT
PRWCC <sup>10</sup>	N/a		185	85	20	NR
CIBMTR <sup>17</sup>	N/a		255	86	34	NR
NCI <sup>6</sup>	1	CD28	50	62	21	Relapse: 2/21 vs 7/7 no HCT
MSKCC <sup>15</sup>	post HCT		15 <sup>c</sup>	N/a	15	Relapse: 3/15
SMC <sup>7</sup>	2		30	86	25	Relapse: 8/25 vs 4/5 no HCT

<sup>a</sup>HCT while in CR.

<sup>b</sup>Data available for 8 out of 11 patients.

<sup>c</sup>One patient received a CAR/41BB T-cell product.

CHOP, Children's Hospital of Philadelphia; CIBMTR, Center for International Blood and Marrow Transplant Research; costim, costimulatory domain; hu, human; St Jude, St Jude Children's Research Hospital; MSKCC, Memorial Sloan Kettering Cancer Center; mur, murine; N/A, not applicable; NCI, National Cancer Institute; NR, not reported; Pat, patients; PRWCC, Pediatric Real World CAR Consortium; Seattle, Seattle Children's Hospital; SMC, Sheba Medical Center.



**Figure 1. Management approach for patients who achieve remission after CD19 CAR T-cell therapy.** The scheme highlights key factors to consider. blina, blinatumomab; CARTs, CAR T cells; MRD, minimal residual disease.

patients proceeded to consolidative HCT at a median of 57 days post infusion. Post HCT, 2 patients suffered disease relapse, and 3 died secondary to HCT complications. Investigators found that patients who received a CD34-selected, T-cell depleted graft or proceeded to HCT fewer than 80 days from CAR infusion had better post-HCT outcomes.<sup>15</sup> In a second phase 1 trial in which 28 patients achieved a CR after CAR T-cell therapy, 21 proceeded to a consolidative HCT, at a median of 54 days post CAR (second HCT, n=4). After transplant, 2 patients experienced subsequent disease relapse, compared to 7 of 7 patients in the nonconsolidative HCT cohort.<sup>6</sup> Additionally, outcomes of a third early-phase clinical study with a CD19/CD28 CAR T-cell product highlighted the benefit of consolidative HCT post CD19 CAR T-cell therapy. Among 30 patients who achieved a CR post CAR T-cell therapy, 25 (17 HCT naive) proceeded to consolidative HCT (median, 71 days). Of these, 8 patients relapsed post HCT, and 2 died secondary to treatment toxicity, with all but 1 patient relapsing in the non-HCT group.<sup>19</sup> In the setting of loss of BCA and rising NGS post CD19 CAR T-cell therapy, a bridging therapy prior to HCT that is readily available (eg, blinatumomab for CD19+ leukemia) might be critical prior to HCT to ensure a disease-free long-term outcome, as illustrated by clinical case 2.

These data collectively indicate that regardless of the utilized CD19 CAR T-cell product, patients who are HCT naive benefit from a consolidative HCT in the setting of limited CAR T-cell persistence. The benefit of a consolidative second HCT is less well established and, given the risk of significant toxicities, should be considered carefully and individualized based on the patient's risk of relapse and ability to tolerate this therapy. In conclusion, based on current literature, it is difficult to give clear recommendations regarding which patients should be considered for HCT post CD19 CAR T-cell therapy. Points for consideration are discussed in the next section and summarized in Figure 1.

### HCT post CD19 CAR T-cell therapy: points for consideration

Prospective studies, albeit difficult to conduct, are ultimately needed to inform the critical question on the role of consolidative HCT after CD19 CAR T-cell therapy, identify high-risk pediatric patients, and support the development of evidence-based clinical-decision algorithms. In lieu of such studies, current approaches to this question weigh the risks and benefits

of pursuing HCT, considering i) risk factors prior to CD19 CAR T-cell therapy and ii) monitoring for persistence and response post infusion (Figure 1).

### Risk factors pre CD19 CAR T-cell therapy

Across several studies, the presence of a high disease burden prior to CD19 CAR T-cell therapy has been associated with a higher risk of relapse after treatment. While a universal cutoff of high burden has not yet been determined, some data suggest a cutoff of as little as greater than or equal to 5% blasts.<sup>6,8,9,11,19</sup> Prior poor response to blinatumomab has also been associated with a higher relapse risk post CAR.<sup>8</sup> Other traditional risk factors for treatment failure have not been recapitulated after treatment with CD19 CAR T-cell therapy, including subgroups with high-risk cytogenetics,<sup>20</sup> Down syndrome,<sup>21</sup> infants,<sup>22,23</sup> and extramedullary disease.<sup>24,25</sup> Thus, CD19 CAR T-cell therapy has the potential to redefine treatments for patients who were historically high risk.

### Monitoring post CD19 CAR T-cell therapy

Longer CD19 CAR T-cell persistence is associated with improved relapse-free survival. Post CAR, close monitoring of ongoing BCA and detection of recurrent and/or persistent disease by NGS, polymerase chain reaction, and/or flow cytometry serves as a surrogate of CAR persistence.<sup>5,8,18,21,26</sup> Notably, relapse risk decreases with time postCAR T-cell therapy, and most relapses occur within the initial year after infusion.<sup>6,10,17,21</sup> Even after treatment with CD19/41-BB CAR T-cell products, the loss of BCA within 6 months or less and/or detectable disease post CAR T-cell therapy, including by NGS testing, place patients at high risk of relapse, and these patients may be considered for HCT prior to disease progression.<sup>26</sup> It is important to note that BCA is not a perfect surrogate for relapse risk, as patients may relapse with antigen loss variants or antigen-positive disease concurrent to findings of loss of BCA. Additional considerations include patient/caregiver preferences, provider experience, and often provider assessment of the availability of additional viable treatment options if the patient were to relapse post CAR T-cell therapy. Importantly, as CAR T-cell therapies continue to evolve and the number of patients treated with such therapies increases, continued investigation and reevaluation of such predictors will be necessary. Finally, besides patient selection the preferred HCT approach remains elusive. Ideally, the attainment of deep

remission with CD19 CAR T-cell therapy would potentially allow for conditioning regimens that do not use total-body irradiation.<sup>27</sup>

### Enhanced CAR T cells

The limited persistence and emergence of antigen loss variants (ie, CD19<sup>-</sup> disease) have emerged as the major limitations of CD19 CAR T-cell therapies,<sup>3-8,14</sup> and both roadblocks will be discussed in this part of the review.

### Prevention of CD19<sup>-</sup> relapse post CAR T-cell therapy

Multiple mechanisms of CD19<sup>-</sup> relapse have been described, including mutations in CD19 that lead to shedding of the extracellular domain, lineage switch with the recurring leukemia having an acute myeloid leukemia phenotype, and the emergence of a preexisting CD19<sup>-</sup> clone (Figure 2A).<sup>28-31</sup> Likewise, the transduction of contaminating ALL blasts in the T-cell products with the viral vector encoding the CD19 CAR can lead to masking of CD19 on the cell surface, resulting in ALL blasts that are resistant to CD19 CAR T cells.<sup>32</sup> The incidence of CD19<sup>-</sup> ALL relapse varies post CD19 CAR T-cell therapy and has been reported to be between 18% and 25%.<sup>3-8</sup>

Targeting additional antigens is actively being pursued to prevent the emergence of CD19<sup>-</sup> ALL blasts. Most efforts are focused on targeting CD22 based on the encouraging results of CD22-CAR T cells as monotherapy for pediatric ALL.<sup>33,34</sup> Conceptually, dual targeting of CD19 and CD22 can be achieved with 3 approaches (Figure 2B): i) sequential or coadministration of 2 CAR T-cell products, 1 expressing a CD19 and the other a CD22 CAR, ii) engineering T cells to simultaneously express a CD19 CAR and a CD22 CAR, and iii) engineering T cells to express a bispecific CAR that recognizes CD19 and CD22. All 3 approaches have been evaluated in early-phase clinical studies, with the largest cohort of patients receiving 2 CAR T-cell products.<sup>35-37</sup> The results of these studies indicate that all 3 approaches are safe; however, it remains to be determined which approach is best to prevent the emergence of antigen loss variants. In addition

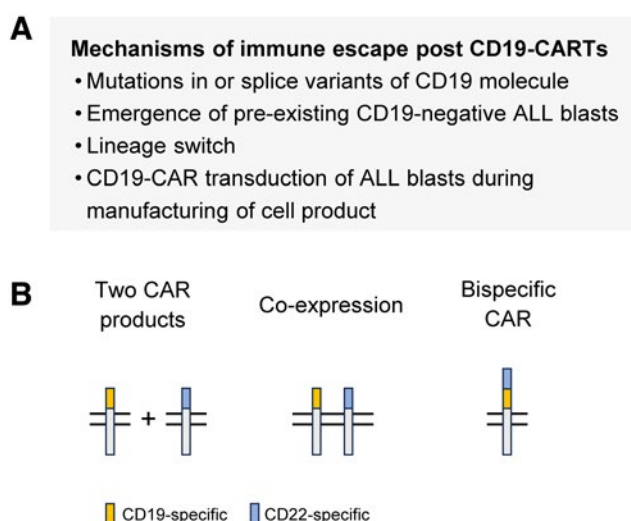
to CD22, other targets are actively being pursued to develop bispecific or trispecific CAR T-cell products for pediatric ALL.<sup>38-40</sup>

### Enhancing persistence of functional CD19 CAR T cells

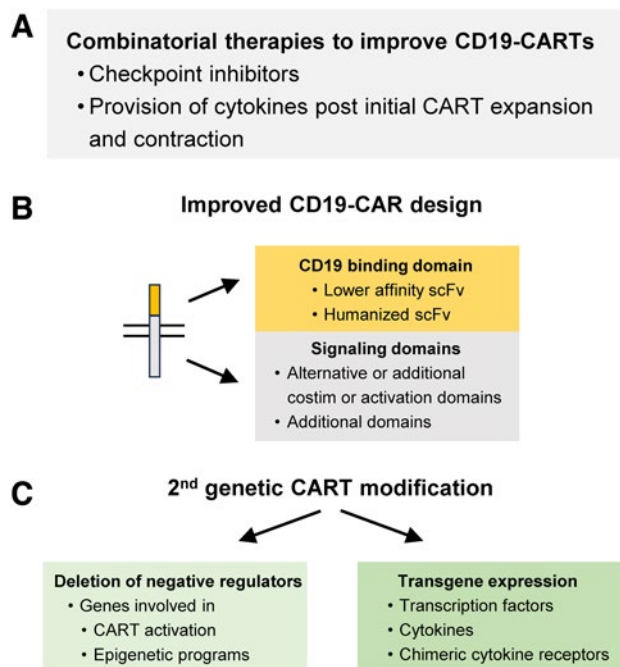
The functional persistence of CD19 CAR T cells is routinely tracked by enumerating normal CD19<sup>+</sup> B cells in the peripheral blood post CD19 CAR T-cell infusion, with the loss of BCA either indicating the limited persistence of CD19 CAR T cells or the persistence of dysfunctional CD19 CAR T cells. Several studies have shed light on the desired characteristics of the leukapheresis product used for CD19 CAR T-cell manufacturing and the product itself associated with the functional persistence of CD19 CAR T cells.<sup>41-44</sup> These include infusing CD19 CAR T cells that are derived from naive T cells and are less differentiated at the end of CD19 CAR T-cell production. However, none of the identified characteristics have been validated prospectively. Likewise, 2 recent studies have highlighted that long-term persisting CD19 CAR T cells have a unique transcriptional profile,<sup>45,46</sup> opening up the opportunity to develop CD19 CAR T-cell products that promote the development of these gene signatures.

### Combinatorial therapies

CD19 CAR T cells might routinely undergo exhaustion reprogramming as highlighted for 1 investigational CD19 CAR T-cell product, and combinatorial therapies represent 1 approach to counteract this (Figure 3A). Combining checkpoint inhibitors with CD19 CAR T cells is actively being pursued for pediatric R/R ALL, and early clinical data suggest that this approach is safe and may augment the effector function and persistence of CD19 CAR T cells.<sup>47</sup> In addition to checkpoint inhibitors, the provision of cytokines after initial CD19 CAR T-cell expansion and contrac-



**Figure 2. Mechanism and prevention of antigen loss variants post CD19 CAR T-cell therapy.** (A) Mechanism of CD19-targeted immune escape. (B) CART T-cell products to enable dual targeting of CD19 and CD22.



**Figure 3. Strategies to enhance the effector function of CD19 CAR T cells.** Examples of (A) combinatorial therapies, (B) strategies to improve CAR design, and (C) second genetic modifications of CAR T cells. costim, costimulation.



tion has demonstrated benefit in preclinical studies, and clinical studies in adults with B-cell lymphoma post CD19 CAR T-cell therapy are ongoing.

### CAR design

Tisagenlecleucel as well as most investigator-initiated CD19 CAR T cells have employed a single-chain Fv that is derived from the monoclonal antibody FMC63. It has a high affinity, and studies have indicated that CAR T cells expressing a CAR that utilizes a CD19-specific scFv with a lower affinity have improved effector function.<sup>48,49</sup> In addition, utilizing a humanized CD19-specific scFv has the potential to reduce CAR-specific immune responses, improving persistence.<sup>50</sup> Currently, the choice of costimulatory domain is the most well-established factor that determines CAR T-cell persistence, with CD19/41BB CARs having longer persistence than CD19/CD28 CARs (see the section "HCT post CD19 CAR T-cell therapy: experience to date"). Finally, the incorporation of novel signaling domains holds the promise to generate CARs that endow CAR T cells with improved effector function (Figure 3B).<sup>51-53</sup>

### Additional genetic modification to enhance the effector function of CD19 CAR T cells

Conceptually, there are 2 main approaches to enhance the effector function, including the persistence of CD19 CAR T cells (Figure 3C). One relies on deleting negative regulators and the other on transgenic expression of transcription factors, cytokines, and chimeric cytokine receptors.<sup>54-57</sup> Examples of deleting negative regulators include molecules that enhance CAR T-cell activation, including RASA2 and Regnase-1,<sup>58,59</sup> and enzymes that are critical for the epigenetic reprogramming of CAR T cells, including TET2 and DNMT3A.<sup>60-62</sup> These approaches are reviewed in detail in recently published articles.<sup>55,56,63</sup>

### Conclusions

Since the infusion of the first pediatric patient with CD19 CAR T cells in 2012, CD19 CAR T-cell therapy has become an integral part of our treatment armamentarium for pediatric patients with R/R ALL. Currently, there are 2 major, complementary efforts ongoing. One focuses on increasing our understanding of how to best use tisagenlecleucel, the only US Food and Drug Administration–approved CD19 CAR T-cell product for R/R pediatric ALL, and the other focuses on developing second-generation ALL-specific CAR T-cell products with enhanced effector function. Based on our current knowledge, subsets of patients who have received tisagenlecleucel will most likely benefit from a consolidative HCT, and further studies are needed to identify these patients. Likewise, the efficacy of tisagenlecleucel might be improved with combinatorial therapies. While we know the desired characteristics of enhanced ALL-specific CAR T-cell products—namely, resistance to antigen loss variants paired with durable persistence—further preclinical and clinical studies are needed to delineate the genetic engineering approach to accomplish this.

### Acknowledgment

Aimee C. Talleur was supported by a scholar grant from the American Society of Hematology.

### Conflict-of-interest disclosure

Aimee C. Talleur: no competing financial interests to declare.  
Swati Naik: no competing financial interests to declare.

Stephen Gottschalk: scientific advisory board: Be Biopharma, CARGO, Immatics; honoraria: TESSA Therapeutics.

### Off-label drug use

Aimee C. Talleur: none are discussed.  
Swati Naik: none are discussed.  
Stephen Gottschalk: none are discussed.

### Correspondence

Swati Naik or Stephen Gottschalk, St Jude Children's Research Hospital, 262 Danny Thomas Pl, MS321, Memphis, TN 38105; e-mail: swati.naik@stjude.org or stephen.gottschalk@stjude.org.

### References

1. Hunger SP, Raetz EA. How I treat relapsed acute lymphoblastic leukemia in the pediatric population. *Blood*. 2020;136(16):1803-1812.
2. Sun W, Malvar J, Spoto R, et al. Outcome of children with multiply relapsed B-cell acute lymphoblastic leukemia: a therapeutic advances in childhood leukemia & lymphoma study. *Leukemia*. 2018;32(11):2316-2325.
3. Maude SL, Frey N, Shaw PA, et al. Chimeric antigen receptor T cells for sustained remissions in leukemia. *N Engl J Med*. 2014;371(16):1507-1517.
4. Maude SL, Laetsch TW, Buechner J, et al. Tisagenlecleucel in children and young adults with B-cell lymphoblastic leukemia. *N Engl J Med*. 2018;378(5):439-448.
5. Gardner RA, Finney O, Annesley C, et al. Intent-to-treat leukemia remission by CD19 CAR T cells of defined formulation and dose in children and young adults. *Blood*. 2017;129(25):3322-3331.
6. Shah NN, Lee DW, Yates B, et al. Long-term follow-up of CD19-CAR T-cell therapy in children and young adults with B-ALL. *J Clin Oncol*. 2021;39(15):1650-1659.
7. Jacoby E, Bielorai B, Hutt D, et al. Parameters of long-term response with CD28-based CD19 chimeric antigen receptor-modified T cells in children and young adults with B-acute lymphoblastic leukaemia. *Br J Haematol*. 2022;197(4):475-481.
8. Talleur AC, Qudeimat A, Métais JY, et al. Preferential expansion of CD8+ CD19-CAR T cells postinfusion and the role of disease burden on outcome in pediatric B-ALL. *Blood Adv*. 2022;6(21):5737-5749.
9. Myers RM, Taraseviciute A, Steinberg SM, et al. Blinatumomab nonresponse and high-disease burden are associated with inferior outcomes after CD19-CAR for B-ALL. *J Clin Oncol*. 2022;40(9):932-944.
10. Schultz LM, Baggott C, Prabhu S, et al. Disease burden affects outcomes in pediatric and young adult B-cell lymphoblastic leukemia after commercial tisagenlecleucel: a pediatric real-world chimeric antigen receptor consortium report. *J Clin Oncol*. 2022;40(9):945-955.
11. Ravich JW, Huang S, Zhou Y, et al. Impact of high disease burden on survival in pediatric patients with B-ALL treated with tisagenlecleucel. *Transplant Cell Ther*. 2022;28(2):73.e1-73.73.e9.
12. Dourthe ME, Rabian F, Yakouben K, et al. Determinants of CD19-positive vs CD19-negative relapse after tisagenlecleucel for B-cell acute lymphoblastic leukemia. *Leukemia*. 2021;35(12):3383-3393.
13. Schultz LM, Eaton A, Baggott C, et al. Outcomes after nonresponse and relapse post-tisagenlecleucel in children, adolescents, and young adults with B-cell acute lymphoblastic leukemia. *J Clin Oncol*. 2023;41(2):354-363.
14. Schultz L, Mackall CL. The future of CAR T-cell therapy for B-cell acute lymphoblastic leukemia in pediatrics and adolescents. *Expert Opin Biol Ther*. 2023;23(7):633-640.
15. Fabrizio VA, Kernan NA, Boulard F, et al. Low toxicity and favorable overall survival in relapsed/refractory B-ALL following CAR T cells and CD34-selected T-cell depleted allogeneic hematopoietic cell transplant. *Bone Marrow Transpl*. 2020;55(11):2160-2169.
16. Laetsch TW, Maude SL, Rives S, et al. Three-year update of tisagenlecleucel in pediatric and young adult patients with relapsed/refractory acute lymphoblastic leukemia in the ELIANA trial. *J Clin Oncol*. 2023;41(9):1664-1669.
17. Pasquini MC, Hu ZH, Curran K, et al. Real-world evidence of tisagenlecleucel for pediatric acute lymphoblastic leukemia and non-Hodgkin lymphoma. *Blood Adv*. 2020;4(21):5414-5424.
18. Summers C, Wu QY, Annesley C, et al. Hematopoietic cell transplantation after CD19 chimeric antigen receptor T cell-induced acute lymphoblastic lymphoma remission confers a leukemia-free survival advantage. *Transplant Cell Ther*. 2022;28(1):21-29.

19. Jacoby E, Ghorashian S, Vormoor B, et al. CD19 CAR T-cells for pediatric relapsed acute lymphoblastic leukemia with active CNS involvement: a retrospective international study. *Leukemia*. 2022;36(6):1525-1532.
20. Leahy AB, Devine KJ, Li Y, et al. Impact of high-risk cytogenetics on outcomes for children and young adults receiving CD19-directed CAR T-cell therapy. *Blood*. 2022;139(14):2173-2185.
21. Laetsch TW, Maude SL, Balduzzi A, et al. Tisagenlecleucel in pediatric and young adult patients with Down syndrome-associated relapsed/refractory acute lymphoblastic leukemia. *Leukemia*. 2022;36(6):1508-1515.
22. Moskop A, Pommert L, Baggott C, et al. Real-world use of tisagenlecleucel in infant acute lymphoblastic leukemia. *Blood Adv*. 2022;6(14):4251-4255.
23. Ghorashian S, Jacoby E, De Moerloose B, et al. Tisagenlecleucel therapy for relapsed or refractory B-cell acute lymphoblastic leukaemia in infants and children younger than 3 years of age at screening: an international, multicentre, retrospective cohort study. *Lancet Haematol*. 2022;9(10):e766-e775.
24. Fabrizio VA, Phillips CL, Lane A, et al. Tisagenlecleucel outcomes in relapsed/refractory extramedullary ALL: a pediatric real world CAR consortium report. *Blood Adv*. 2022;6(2):600-610.
25. Leahy AB, Newman H, Li Y, et al. CD19-targeted chimeric antigen receptor T-cell therapy for CNS relapsed or refractory acute lymphocytic leukaemia: a post-hoc analysis of pooled data from five clinical trials. *Lancet Haematol*. 2021;8(10):e711-e722.
26. Pulsipher MA, Han X, Maude SL, et al. Next-generation sequencing of minimal residual disease for predicting relapse after tisagenlecleucel in children and young adults with acute lymphoblastic leukemia. *Blood Cancer Discov*. 2022;3(1):66-81.
27. Abdel-Azim H, Quigg TC, Malvar J, et al. Excellent relapse-free and overall survival in pre-HCT next-generation sequencing (NGS-MRD) negative B-ALL patients with or without TBI-based conditioning: outcome of the observational arm of the pediatric transplantation and cellular therapy consortium (PTCTC) ONC1701 endrad study. *Transplant Cell Ther*. 2023;29(2):S94-S96.
28. Orlando EJ, Han X, Tribouley C, et al. Genetic mechanisms of target antigen loss in CAR19 therapy of acute lymphoblastic leukemia. *Nat Med*. 2018;24(10):1504-1506.
29. Sotillo E, Barrett DM, Black KL, et al. Convergence of acquired mutations and alternative splicing of CD19 enables resistance to CART-19 immunotherapy. *Cancer Discov*. 2015;5(12):1282-1295.
30. Gardner R, Wu D, Cherian S, et al. Acquisition of a CD19-negative myeloid phenotype allows immune escape of MLL-rearranged B-ALL from CD19 CAR-T-cell therapy. *Blood*. 2016;127(20):2406-2410.
31. Lamble AJ, Myers RM, Taraseviciute A, et al. Preinfusion factors impacting relapse immunophenotype following CD19 CAR T cells. *Blood Adv*. 2023;7(4):575-585.
32. Ruella M, Xu J, Barrett DM, et al. Induction of resistance to chimeric antigen receptor T cell therapy by transduction of a single leukemic B cell. *Nat Med*. 2018;24(10):1499-1503.
33. Fry TJ, Shah NN, Orentas RJ, et al. CD22-targeted CAR T cells induce remission in B-ALL that is naive or resistant to CD19-targeted CAR immunotherapy. *Nat Med*. 2018;24(1):20-28.
34. Shah NN, Highfill SL, Shalabi H, et al. CD4/CD8 T-cell selection affects chimeric antigen receptor (CAR) T-cell potency and toxicity: updated results from a phase I anti-CD22 CAR T-cell trial. *J Clin Oncol*. 2020;38(17):1938-1950.
35. Spiegel JY, Patel S, Muffy L, et al. CAR T cells with dual targeting of CD19 and CD22 in adult patients with recurrent or refractory B cell malignancies: a phase 1 trial. *Nat Med*. 2021;27(8):1419-1431.
36. Cordoba S, Onuoha S, Thomas S, et al. CAR T cells with dual targeting of CD19 and CD22 in pediatric and young adult patients with relapsed or refractory B cell acute lymphoblastic leukemia: a phase 1 trial. *Nat Med*. 2021;27(10):1797-1805.
37. Wang T, Tang Y, Cai J, et al. Coadministration of CD19- and CD22-directed chimeric antigen receptor T-cell therapy in childhood B-cell acute lymphoblastic leukemia: a single-arm, multicenter, phase II trial. *J Clin Oncol*. 2023;41(9):1670-1683.
38. Qin H, Cho M, Haso W, et al. Eradication of B-ALL using chimeric antigen receptor-expressing T cells targeting the TSLPR oncoprotein. *Blood*. 2015;126(5):629-639.
39. Niswander LM, Graff ZT, Chien CD, et al. Potent preclinical activity of FLT3-directed chimeric antigen receptor T-cell immunotherapy against FLT3-mutant acute myeloid leukemia and KMT2A-rearranged acute lymphoblastic leukemia. *Haematologica*. 2023;108(2):457-471.
40. Fousek K, Watanabe J, Joseph SK, et al. CAR T-cells that target acute B-lineage leukemia irrespective of CD19 expression. *Leukemia*. 2021;35(1):75-89.
41. Wilson TL, Kim H, Chou CH, et al. Common trajectories of highly effective CD19-specific CAR T cells identified by endogenous T-cell receptor lineages. *Cancer Discov*. 2022;12(9):2098-2119.
42. Fraietta JA, Lacey SF, Orlando EJ, et al. Determinants of response and resistance to CD19 chimeric antigen receptor (CAR) T cell therapy of chronic lymphocytic leukemia. *Nat Med*. 2018;24(5):563-571.
43. Finney OC, Brakke H, Rawlings-Rhea S, et al. CD19 CAR T cell product and disease attributes predict leukemia remission durability. *J Clin Invest*. 2019;129(5):2123-2132.
44. Chen GM, Chen C, Das RK, et al. Integrative bulk and single-cell profiling of premanufacture T-cell populations reveals factors mediating long-term persistence of CAR T-cell therapy. *Cancer Discov*. 2021;11(9):2186-2199.
45. Anderson ND, Birch J, Accogli T, et al. Transcriptional signatures associated with persisting CD19 CAR-T cells in children with leukemia. *Nat Med*. 2023;29(7):1700-1709.
46. Melenhorst JJ, Chen GM, Wang M, et al. Decade-long leukaemia remissions with persistence of CD4+ CAR T cells. *Nature*. 2022;602(7897):503-509.
47. Li AM, Hucks GE, Dinofia AM, et al. Checkpoint inhibitors augment CD19-directed chimeric antigen receptor (CAR) T cell therapy in relapsed B-cell acute lymphoblastic leukemia. *Blood*. 2018;132(suppl 1):556.
48. Michelozzi IM, Gomez-Castaneda E, Pohle RVC, et al. Activation priming and cytokine polyfunctionality modulate the enhanced functionality of low-affinity CD19 CAR T cells. *Blood Adv*. 2023;7(9):1725-1738.
49. Ghorashian S, Kramer AM, Onuoha S, et al. Enhanced CAR T cell expansion and prolonged persistence in pediatric patients with ALL treated with a low-affinity CD19 CAR. *Nat Med*. 2019;25(9):1408-1414.
50. Myers RM, Li Y, Barz Leahy A, et al. Humanized CD19-targeted chimeric antigen receptor (CAR) T cells in CAR-naïve and CAR-exposed children and young adults with relapsed or refractory acute lymphoblastic leukemia. *J Clin Oncol*. 2021;39(27):3044-3055.
51. Guedan S, Posey AD Jr, Shaw C, et al. Enhancing CAR T cell persistence through ICOS and 4-1BB costimulation. *JCI Insight*. 2018;3(1).
52. Chockley PJ, Ibanez-Vega J, Krenciute G, Talbot LJ, Gottschalk S. Synapse-tuned CARs enhance immune cell anti-tumor activity [published online ahead of print 2 February 2023]. *Nat Biotechnol*.
53. Tousley AM, Rotiroti MC, Labanieh L, et al. Co-opting signalling molecules enables logic-gated control of CAR T cells. *Nature*. 2023;615(7952):507-516.
54. Lynn RC, Weber EW, Sotillo E, et al. c-Jun overexpression in CAR T cells induces exhaustion resistance. *Nature*. 2019;576(7786):293-300.
55. Wagner J, Wickman E, DeRenzo C, Gottschalk S. CAR T cell therapy for solid tumors: bright future or dark reality? *Mol Ther*. 2020;28(11):2320-2339.
56. Labanieh L, Mackall CL. CAR immune cells: design principles, resistance and the next generation. *Nature*. 2023;614(7949):635-648.
57. Hoyos V, Savoldo B, Quintarelli C, et al. Engineering CD19-specific T lymphocytes with interleukin-15 and a suicide gene to enhance their anti-lymphoma/leukemia effects and safety. *Leukemia*. 2010;24(6):1160-1170.
58. Wei J, Long L, Zheng W, et al. Targeting REGNASE-1 programs long-lived effector T cells for cancer therapy. *Nature*. 2019;576(7787):471-476.
59. Carnevale J, Shifrut E, Kale N, et al. RASA2 ablation in T cells boosts antigen sensitivity and long-term function. *Nature*. 2022;609(7925):174-182.
60. Fraietta JA, Nobles CL, Sammons MA, et al. Disruption of TET2 promotes the therapeutic efficacy of CD19-targeted T cells. *Nature*. 2018;558(7709):307-312.
61. Prinzing B, Zebley CC, Petersen CT, et al. Deleting DNMT3A in CAR T cells prevents exhaustion and enhances antitumor activity. *Sci Transl Med*. 2021;13(620):eab0272.
62. Jain N, Zhao Z, Feucht J, et al. TET2 guards against unchecked BATF3-induced CAR T cell expansion. *Nature*. 2023;615(7951):315-322.
63. Rafiq S, Hackett CS, Brentjens RJ. Engineering strategies to overcome the current roadblocks in CAR T cell therapy. *Nat Rev Clin Oncol*. 2020;17(3):147-167.