

# Dealing with noise

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## Where are we so far?

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So far, we've imported and transformed our measures, combined them with our design information, pre-processed, processed, plotted, and analyzed our data. Here, we're going to learn potential strategies for dealing with noise in our growth curve data.

If you haven't already, load the necessary packages.

```

library(gcplyr)

library(dplyr)
library(ggplot2)

#This code was previously explained
#Here we're re-running it so it's available for us to work with
example_design <- make_design(
  pattern_split = ",", n_rows = 8, n_cols = 12,
  "Bacteria_strain" = make_designpattern(
    values = paste("Strain", 1:48),
    rows = 1:8, cols = 1:6, pattern = 1:48, byrow = TRUE),
  "Bacteria_strain" = make_designpattern(
    values = paste("Strain", 1:48),
    rows = 1:8, cols = 7:12, pattern = 1:48, byrow = TRUE),
  "Phage" = make_designpattern(
    values = c("No Phage"), rows = 1:8, cols = 1:6, pattern = "1"),
  "Phage" = make_designpattern(
    values = c("Phage Added"), rows = 1:8, cols = 7:12, pattern = "1"))

sample_wells <- c("A1", "F1", "F10", "E11")

```

## Introduction

Oftentimes, growth curve data produced by a plate reader will have some noise in it. In model-fitting analysis of growth curves implemented by other packages, the effect of this noise is often eliminated by the fitting step. However, since `gcplyr` does model-free analyses, our approach can sometimes be more sensitive to noise, necessitating steps to reduce the effects of noise.

When assessing the effects of noise in our data, one of the first steps is simply to visualize our data. In particular, we want to visualize the raw data, but also any derivatives we'll be using in our analyses. This is especially important because per-capita derivatives are often the most sensitive to noise, especially when bacterial population sizes are small. By visualizing our data, we can assess whether the density, derivative, and per-capita derivative are all changing smoothly, as we would expect. If, instead, we observe spikes and rapid fluctuations, we know that noise is likely to throw off our estimates of maxima and minima of the data or derivatives.

Broadly speaking, there are three strategies we can use to deal with noise:

- Using fitting during derivative calculations
- Smooth the raw data
- Analyze only less-noisy subsets of the data

Let's start by pulling out some example data. Luckily for us, there is a version of the same example data we've been working with but with simulated noise added to it.

```

#This is the data we've been working with previously
noiseless_data <-
  trans_wide_to_tidy(example_widedata_noiseless, id_cols = "Time")
#This is the same data but with simulated noise added
noisy_data <- trans_wide_to_tidy(example_widedata, id_cols = "Time")
#We'll add some identifiers and then merge them together

```

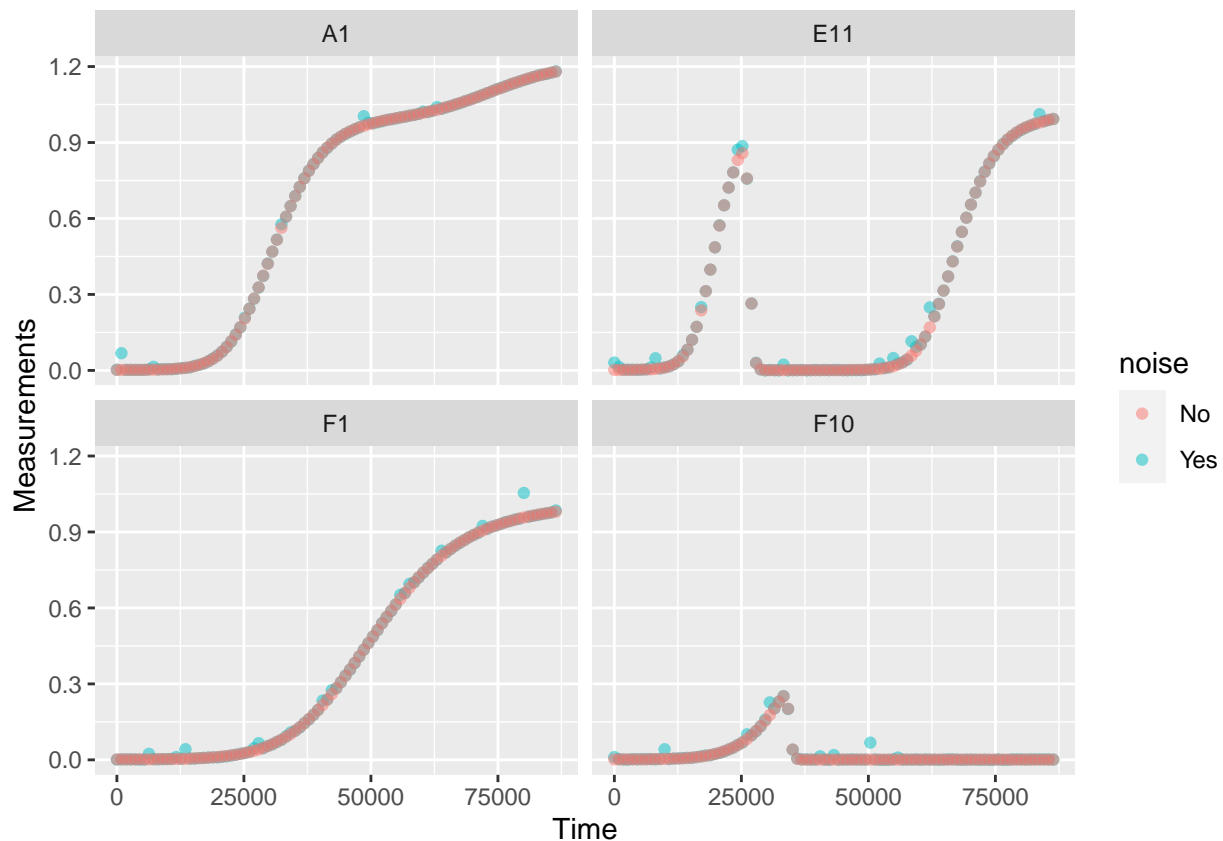
```

noiseless_data <- mutate(noiseless_data, noise = "No")
noisy_data <- mutate(noisy_data, noise = "Yes")
ex_dat_mrg <- merge_dfs(noisy_data, noiseless_data)
#> Joining, by = c("Time", "Well", "Measurements", "noise")
#> Warning in merge_dfs(noisy_data, noiseless_data):
#> merged_df has more rows than x or y, this may indicate
#>      mis-matched values in the shared column(s) used to merge
#>      (e.g. 'Well')
ex_dat_mrg <- merge_dfs(ex_dat_mrg, example_design)
#> Joining, by = "Well"

ex_dat_mrg$Well <-
  factor(ex_dat_mrg$Well,
         levels = paste(rep(LETTERS[1:8], each = 12), 1:12, sep = ""))

#Plot with a linear y-axis
ggplot(data = dplyr::filter(ex_dat_mrg, Well %in% sample_wells),
       aes(x = Time, y = Measurements, color = noise)) +
  geom_point(alpha = 0.5) +
  facet_wrap(~Well)

```

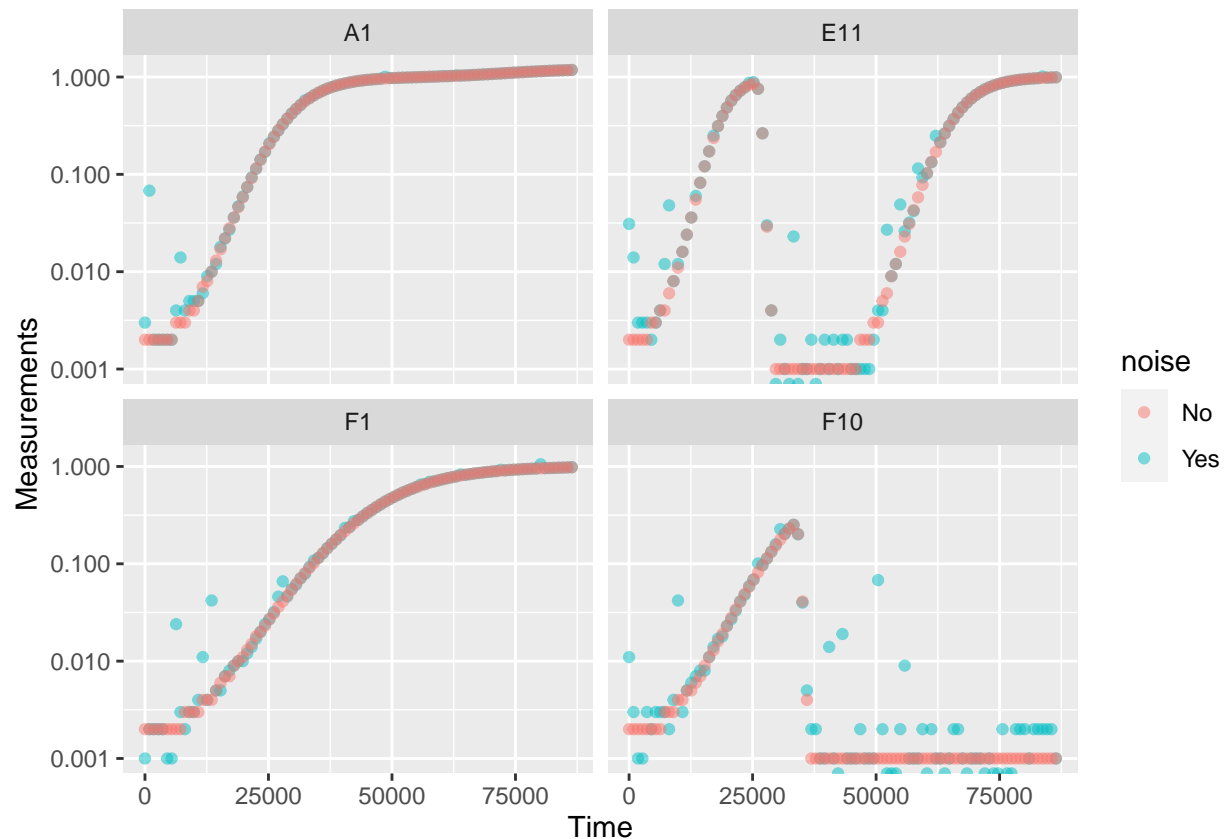


```

#Plot with a log y-axis
ggplot(data = dplyr::filter(ex_dat_mrg, Well %in% sample_wells),
       aes(x = Time, y = Measurements, color = noise)) +
  geom_point(alpha = 0.5) +

```

```
facet_wrap(~Well) +
scale_y_continuous(trans = "log10")
#> Warning: Transformation introduced infinite values in continuous y-axis
```

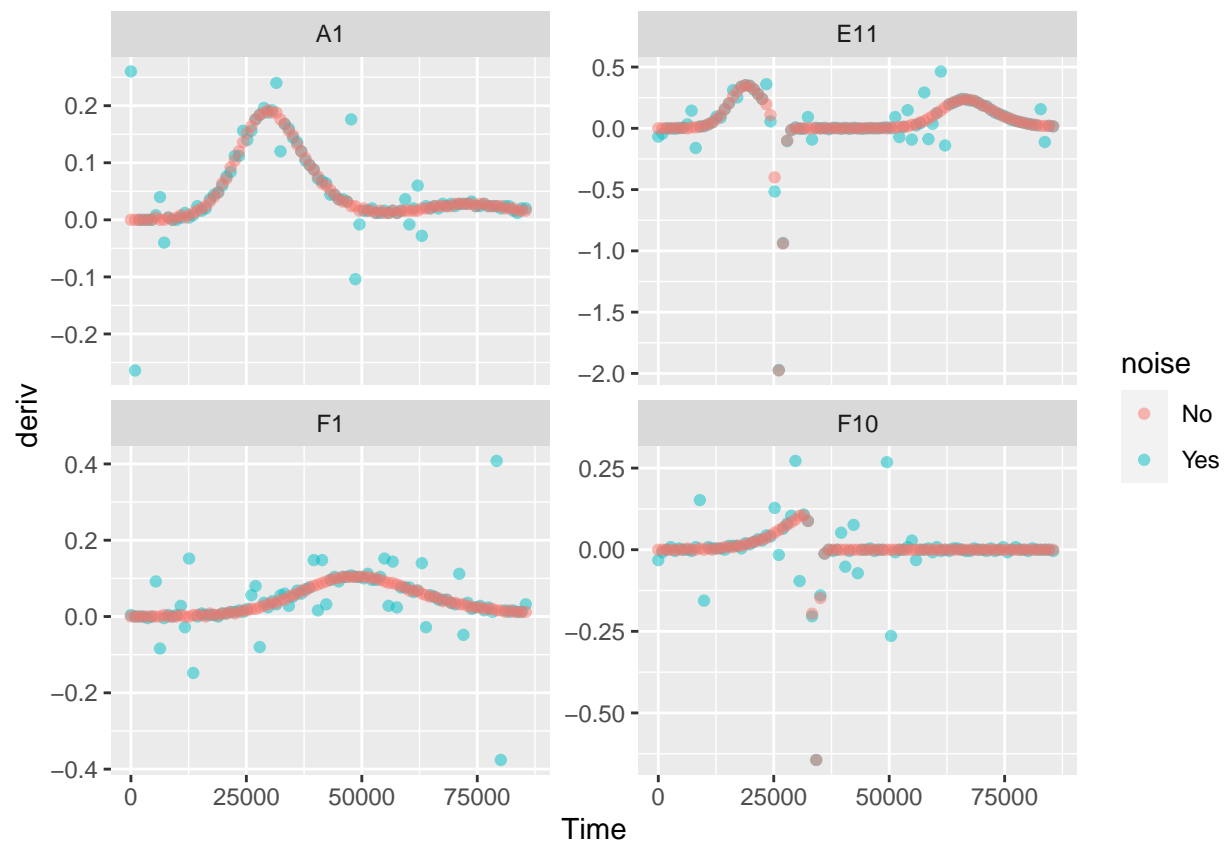


Great! Here we can see how the noisy and noiseless data compare. We've plotted our data both with linear axes and with log-transformed y-axes. log axes are useful because exponential growth is a straight line when plotted on a log scale, but in this case it also helps highlight the higher relative noise at low densities compared to high densities. In fact, this is a common occurrence: **at low densities, random noise tends to have a much larger effect** than at high densities.

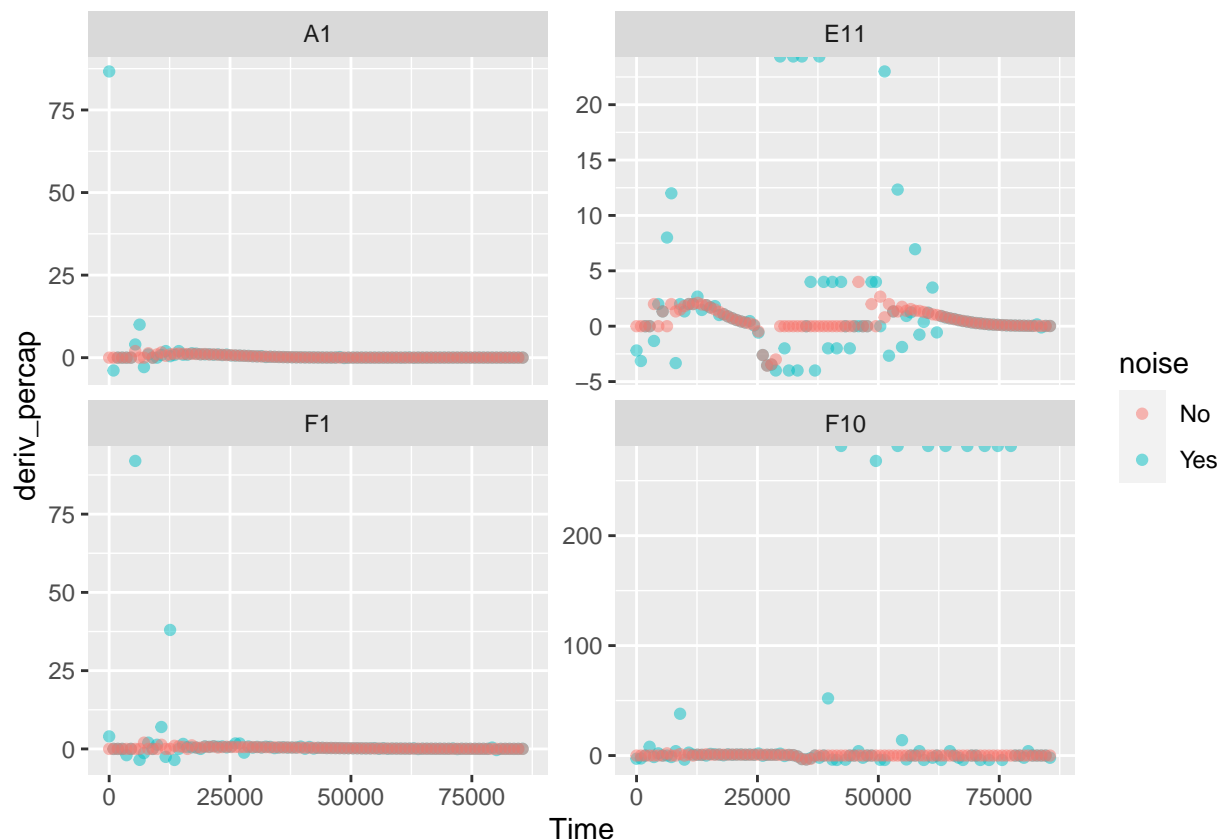
This level of noise doesn't seem like it would mess up calculations of maximum density or area under the curve much, so that's not enough of a reason to smooth. But let's look at what our derivatives look like.

```
ex_dat_mrg <-
  mutate(group_by(ex_dat_mrg, Well, Bacteria_strain, Phage, noise),
    deriv = calc_deriv(x = Time, y = Measurements, x_scale = 3600),
    deriv_percap = calc_deriv(x = Time, y = Measurements, x_scale = 3600,
      percapita = TRUE, blank = 0))

#Plot derivative
ggplot(data = dplyr::filter(ex_dat_mrg, Well %in% sample_wells),
  aes(x = Time, y = deriv, color = noise)) +
  geom_point(alpha = 0.5) +
  facet_wrap(~Well, scales = "free_y")
#> Warning: Removed 8 rows containing missing values (geom_point).
```



```
#Plot per-capita derivative
ggplot(data = dplyr::filter(ex_dat_mrg, Well %in% sample_wells),
  aes(x = Time, y = deriv_percap, color = noise)) +
  geom_point(alpha = 0.5) +
  facet_wrap(~Well, scales = "free_y")
#> Warning: Removed 12 rows containing missing values (geom_point).
```



Those values are jumping all over the place, including some where the growth rate was calculated as infinite! Let's see what we can do to address this.

## Fitting during derivative calculation

One thing we can do is actually something we already did in the Calculating Derivatives article (`vignette("process")`): instead of calculating the derivative of each point relative to the next, we can use a moving window of more than two points and fit a linear regression to this data. In the earlier situation we had used more than two points because of limited resolution at low densities. However, the same solution can apply here. By calculating our derivatives by fitting many points instead of just two, the effect of any single noisy point will be reduced.

To use the fitting functionality of `calc_deriv`, we need to specify either the `window_width` parameter, or the `window_width_n` parameter. `window_width` specifies how wide the window used to include points for the fitting is in units of `x`, while `window_width_n` specifies it in number of data points. Here, we'll demonstrate its use by fitting regressions with more data points. Note that when using `calc_deriv` in this way, you should use *as few* points as is necessary for your analyses to work, so you should visualize different window widths and choose the smallest one that is sufficient for your analyses to succeed.

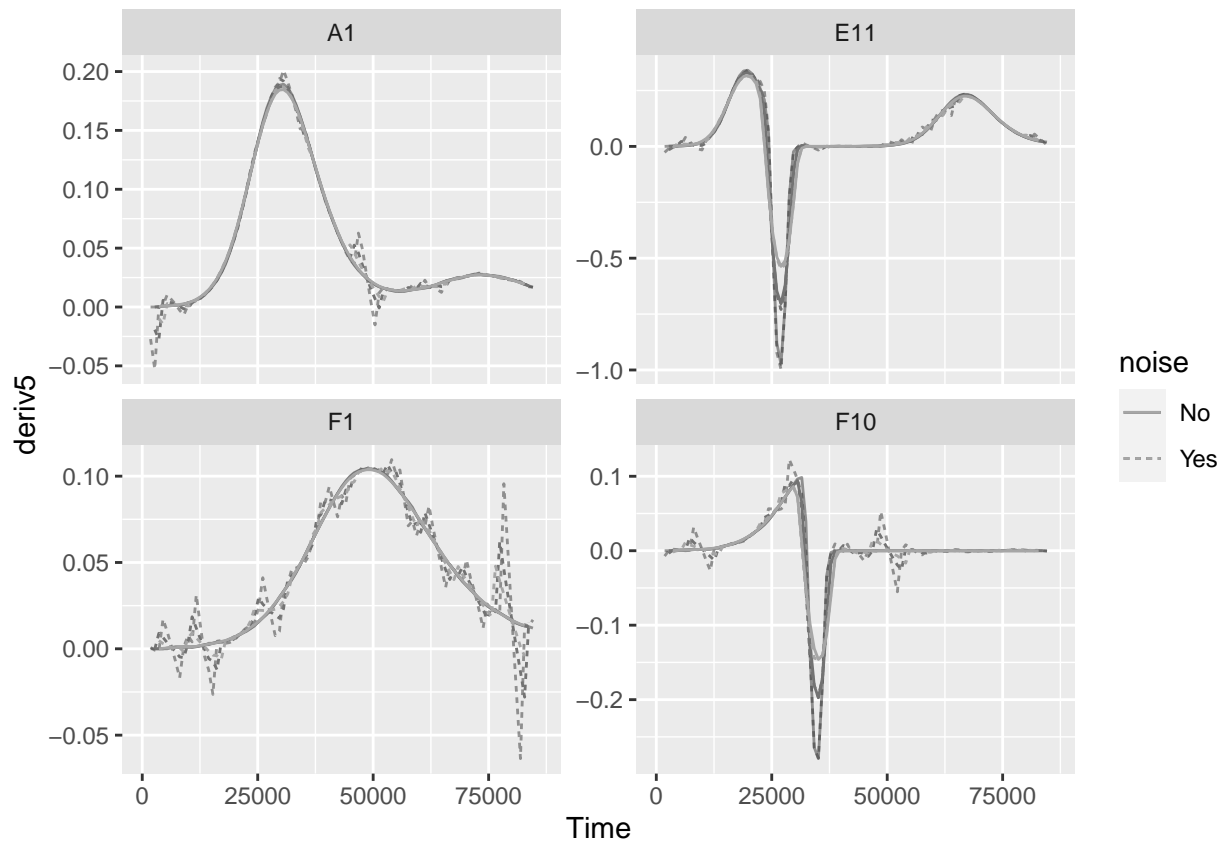
```
ex_dat_mrg <-
  mutate(group_by(ex_dat_mrg, Well, Bacteria_strain, Phage, noise),
    deriv5 = calc_deriv(x = Time, y = Measurements, x_scale = 3600,
      window_width_n = 5),
    deriv_percap5 = calc_deriv(x = Time, y = Measurements, x_scale = 3600,
      percapita = TRUE, blank = 0,
```

```

                                window_width_n = 5),
deriv7 = calc_deriv(x = Time, y = Measurements, x_scale = 3600,
                    window_width_n = 7),
deriv_percap7 = calc_deriv(x = Time, y = Measurements, x_scale = 3600,
                           percapita = TRUE, blank = 0,
                           window_width_n = 7),
deriv9 = calc_deriv(x = Time, y = Measurements, x_scale = 3600,
                    window_width_n = 9),
deriv_percap9 = calc_deriv(x = Time, y = Measurements, x_scale = 3600,
                           percapita = TRUE, blank = 0,
                           window_width_n = 9))

#Plot derivative
ggplot(data = dplyr::filter(ex_dat_mrg, Well %in% sample_wells),
       aes(x = Time, y = deriv5, lty = noise)) +
  geom_line(alpha = 0.5, color = "gray20") +
  geom_line(aes(y = deriv7, color = "gray45") +
  geom_line(aes(y = deriv9, color = "gray65") +
  facet_wrap(~Well, scales = "free_y")
#> Warning: Removed 8 row(s) containing missing values (geom_path).
#> Warning: Removed 12 row(s) containing missing values (geom_path).
#> Warning: Removed 16 row(s) containing missing values (geom_path).

```



```

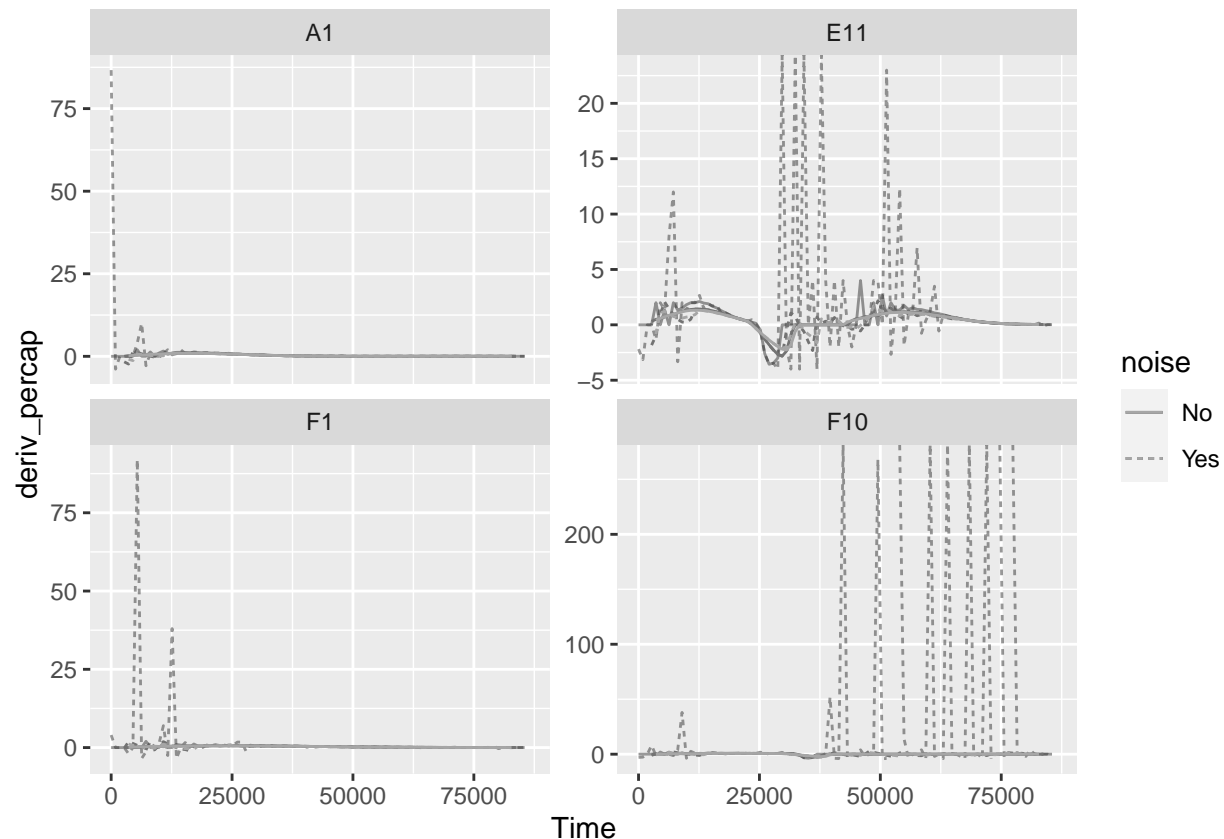
#Plot per-capita derivative
ggplot(data = dplyr::filter(ex_dat_mrg, Well %in% sample_wells),

```

```

aes(x = Time, y = deriv_percap, lty = noise)) +
geom_line(alpha = 0.5, color = "gray20") +
geom_line(aes(y = deriv_percap7), color = "gray45") +
geom_line(aes(y = deriv_percap9), color = "gray65") +
facet_wrap(~Well, scales = "free_y")
#> Warning: Removed 2 row(s) containing missing values (geom_path).
#> Warning: Removed 12 row(s) containing missing values (geom_path).
#> Warning: Removed 16 row(s) containing missing values (geom_path).

```



Great! As we can see, increasing the number of points in the derivative calculation reduces the amount of noise. However, we can also see that it tends to bias our results, making peaks less high and valleys less deep. Moreover, in some of our derivatives, especially the per-capita derivative, some noise remains. In the next two sections, we'll explore how smoothing raw data and analyzing just a subset of our data can further reduce the effects on noise on our analyses.

## Smoothing raw data

One of the most obvious approaches to deal with noise in our raw data is to use a smoothing algorithm. `ggplot2` has a `smooth_data` function that can carry out such smoothing. Note that when using `smooth_data`, you should generally carry out *as little* smoothing as is necessary for your analyses to work, so you should visualize different degrees of smoothing and choose the least smoothed one that is sufficient for your analyses to succeed.

`smooth_data` has four different smoothing algorithms to choose from: `moving-average`, `moving-median`, `loess`, and `gam`.



- **moving-average** is a simple smoothing algorithm that primarily acts to reduce the effects of outliers on the data
- **moving-median** is another simple smoothing algorithm that primarily acts to reduce the effects of outliers on the data
- **loess** is a spline-fitting approach that uses polynomial-like curves, which produces curves with smoothly changing derivatives, but can in some cases create curvature artifacts not present in the original data
- **gam** is also spline-fitting approach that uses polynomial-like curves, which produces curves with smoothly changing derivatives, but can in some cases create curvature artifacts not present in the original data

**Additionally, all four smoothing algorithms have a tuning parameter** that controls how “smoothed” the data are. For whichever smoothing method you’re using, **you should plot smoothing with multiple different tuning parameter values**, then choose the value that smooths the data as little as is necessary to reduce noise. Make sure to plot the smoothing for every well in your data, so that you’re choosing the best setting for all your data and not just one well.

Smoothing data is a step that alters the values you will analyze. Because of that, and because there are so many options for how to smooth your data, it is a step that can be rife with pitfalls. I recommend starting with the simplest and least “smoothed” smoothing, plotting your results, and only increasing your smoothing as much as is needed to enable downstream analyses. Additionally, when sharing your findings, it’s important to be transparent by sharing the raw data and smoothing methods, rather than treating the smoothed data as your source.

To use `smooth_data`, pass your x and y values, your method of choice, and any additional arguments needed for the method. It will return a vector of your smoothed y values.

## Smoothing with moving-average

For **moving-average**, there are two tuning parameters to choose between:

- **window\_width** specifies how wide the moving window used to calculate the average is in units of x.
- **window\_width\_n** specifies how many data points wide the moving window used to calculate the average is.

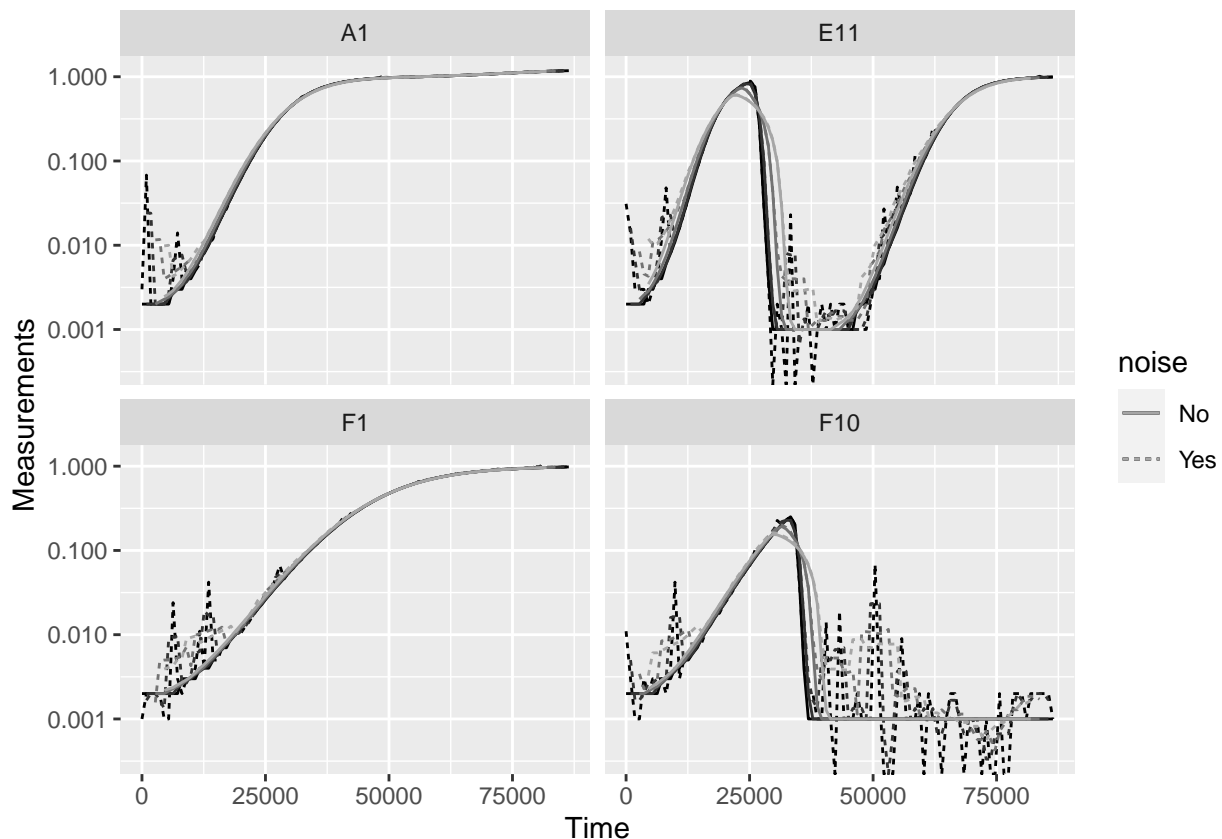
Specifying the **window\_width** or **window\_width\_n** is required, and larger values will be more “smoothed”. Think carefully about whether you want to hold the *amount* of time or the *number* of data points in each window constant (if your data was all collected on constant intervals, then there will be no difference).

Here, we’ll show moving averages with **window\_width\_n** values of 3, 7, or 11 data points wide (because the window is centered on each data point, **window\_width\_n** must be an odd number of data points wide). Note that **moving-average** returns NA for data points at the start and end of your data where the window extends beyond the domain of your data.

```
ex_dat_mrg <-
  mutate(group_by(ex_dat_mrg, Well, Bacteria_strain, Phage, noise),
    smoothed3 = smooth_data(x = Time, y = Measurements,
      sm_method = "moving-average", window_width_n = 3),
    smoothed7 = smooth_data(x = Time, y = Measurements,
      sm_method = "moving-average", window_width_n = 7),
    smoothed11 = smooth_data(x = Time, y = Measurements,
      sm_method = "moving-average", window_width_n = 11))

#What does the smoothed data look like compared to the noisy original?
```

```
#Lighter lines are wider window_width_n's and more "smoothed"
ggplot(data = dplyr::filter(ex_dat_mrg, Well %in% sample_wells),
  aes(x = Time, lty = noise)) +
  geom_line(aes(y = Measurements)) +
  geom_line(aes(y = smoothed3), color = "gray20") +
  geom_line(aes(y = smoothed7), color = "gray45") +
  geom_line(aes(y = smoothed11), color = "gray65") +
  facet_wrap(~Well) +
  scale_y_continuous(trans = "log10")
#> Warning: Transformation introduced infinite values in continuous y-axis
#> Transformation introduced infinite values in continuous y-axis
#> Warning: Removed 4 row(s) containing missing values (geom_path).
#> Warning: Removed 12 row(s) containing missing values (geom_path).
#> Warning: Removed 20 row(s) containing missing values (geom_path).
```



Here we can see that moving-average has helped reduce the effects of some of that early noise. However, with `window_width_n = 11` (the lightest line), the smoothing has started biasing our medium-density data points to be higher than they actually are. Based on this, we'd probably want to use a `window_width_n` less than 11. Unfortunately, with smaller `window_width_n` our early data is still being affected by that early noise, so we should explore other smoothing methods, or try combining multiple smoothing methods.

## Smoothing with moving-median

For moving-median, there are the same two tuning parameters:

- `window_width` specifies how wide the moving window used to calculate the average is in units of `x`.
- `window_width_n` specifies how many data points wide the moving window used to calculate the average is.

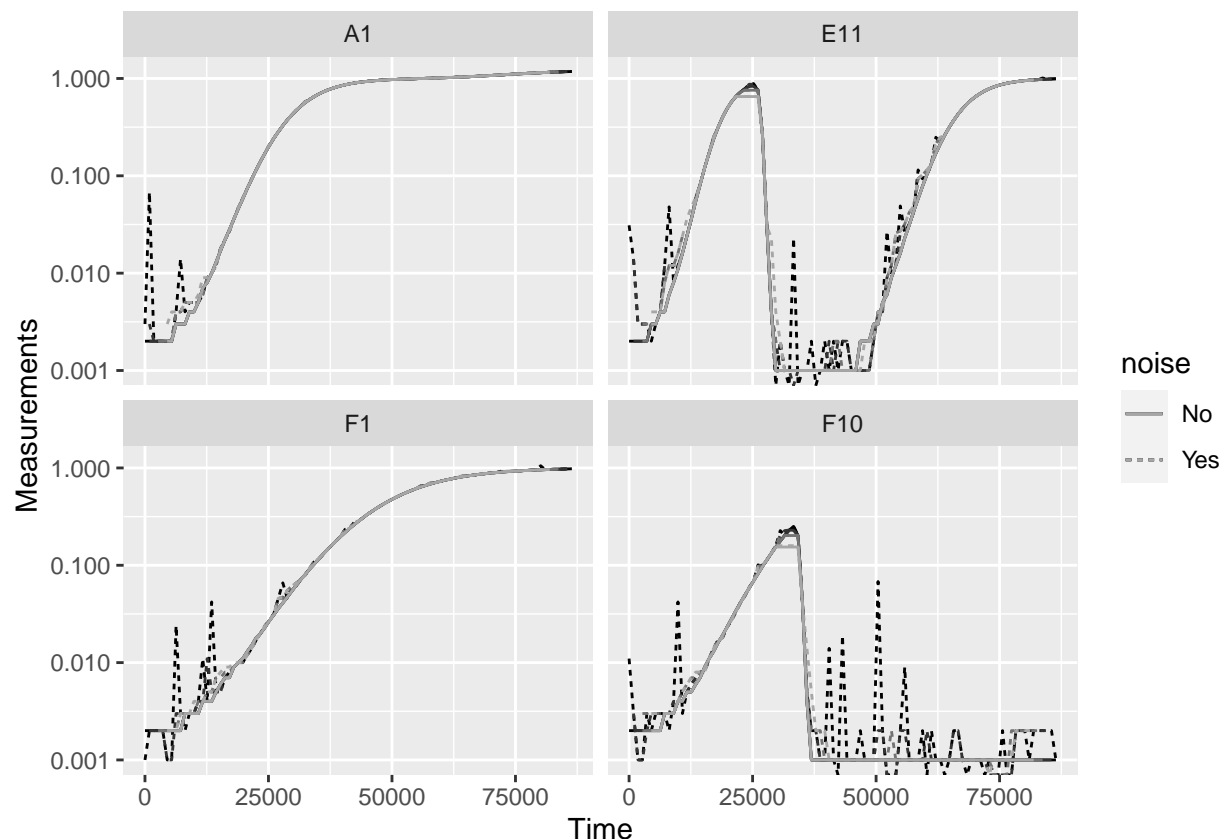
Specifying the `window_width` or `window_width_n` is required, and larger values will be more “smoothed”. Think carefully about whether you want to hold the *amount* of time or the *number* of data points in each window constant (if your data was all collected on constant intervals, then there will be no difference).

Here, we’ll show moving medians with windows that are 3, 7, and 11 data points wide (because the window is centered on each data point, it must be an odd number of data points wide). Note that `moving-median` returns NA for data points at the start and end of your data where the window extends beyond the domain of your data.

```
ex_dat_mrg <-
  mutate(group_by(ex_dat_mrg, Well, Bacteria_strain, Phage, noise),
    smoothed3 =
      smooth_data(x = Time, y = Measurements,
        sm_method = "moving-median", window_width_n = 3),
    smoothed7 =
      smooth_data(x = Time, y = Measurements,
        sm_method = "moving-median", window_width_n = 7),
    smoothed11 =
      smooth_data(x = Time, y = Measurements,
        sm_method = "moving-median", window_width_n = 11))

#What does the smoothed data look like compared to the noisy original?
#Lighter lines are wider window_width_n's and more "smoothed"
ggplot(data = dplyr::filter(ex_dat_mrg, Well %in% sample_wells),
  aes(x = Time, lty = noise)) +
  geom_line(aes(y = Measurements)) +
  geom_line(aes(y = smoothed3, color = "gray20")) +
  geom_line(aes(y = smoothed7, color = "gray45")) +
  geom_line(aes(y = smoothed11, color = "gray65")) +
  facet_wrap(~Well) +
  scale_y_continuous(trans = "log10")

#> Warning: Transformation introduced infinite values in continuous y-axis
#> Transformation introduced infinite values in continuous y-axis
#> Transformation introduced infinite values in continuous y-axis
#> Transformation introduced infinite values in continuous y-axis
#> Warning: Removed 4 row(s) containing missing values (geom_path).
#> Warning: Removed 12 row(s) containing missing values (geom_path).
#> Warning: Removed 20 row(s) containing missing values (geom_path).
```



Here we can see that `moving-median` has really excluded that low-density noise, even with the smallest `window_width_n = 3`. Additionally, `moving-median` did not bias our larger data hardly at all, except with the widest `window_width_n`. However, it has produced a smoothed density that is fairly “jumpy”, something that wider `window_width_n` did not fix. **This is common with `moving-median`**, so often you may need to try other smoothing methods or combining `moving-median` with other methods.

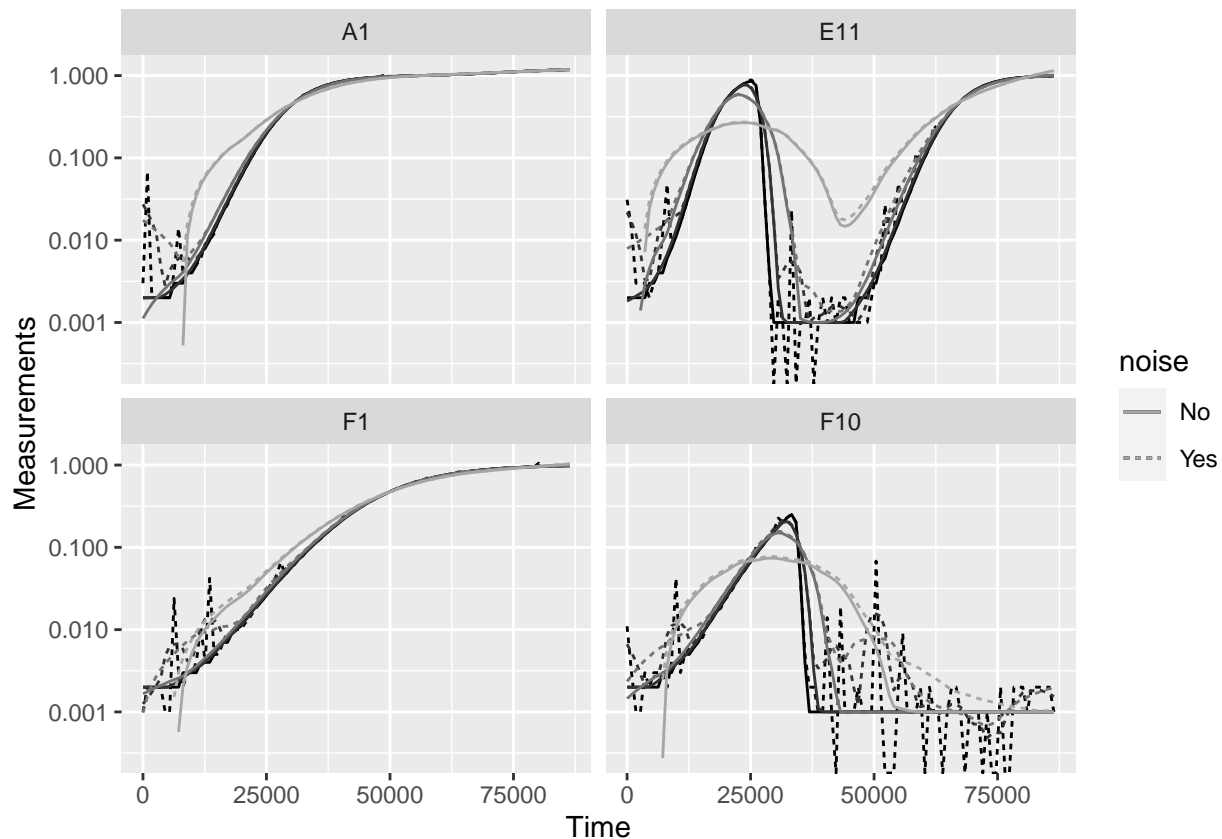
## Smoothing with LOESS

For `loess`, the tuning parameter is the `span` argument. `loess` works by doing fits on subset windows of the data centered at each data point. These fits can be linear (`degree = 1`) or polynomial (typically `degree = 2`). `span` is the width of the window, as a fraction of all data points. For instance, with the default `span` of 0.75, 75% of the data points are included in each window. Thus, `span` values typically are between 0 and 1 (although see `?loess` for use of `span` values greater than 1), and larger values are more “smoothed”. Here, we’ll show `loess` smoothing with spans of 0.1, 0.2, and 0.5 and `degree = 1`.

```
ex_dat_mrg <-
  mutate(group_by(ex_dat_mrg, Well, Bacteria_strain, Phage, noise),
    smoothed1 = smooth_data(x = Time, y = Measurements,
                           sm_method = "loess", span = .1, degree = 1),
    smoothed2 = smooth_data(x = Time, y = Measurements,
                           sm_method = "loess", span = .2, degree = 1),
    smoothed5 = smooth_data(x = Time, y = Measurements,
                           sm_method = "loess", span = .5, degree = 1))

#What does the smoothed data look like compared to the noisy original?
```

```
#Lighter lines are larger span's and more "smoothed"
ggplot(data = dplyr::filter(ex_dat_mrg, Well %in% sample_wells),
  aes(x = Time, lty = noise)) +
  geom_line(aes(y = Measurements)) +
  geom_line(aes(y = smoothed1), color = "gray20") +
  geom_line(aes(y = smoothed2), color = "gray45") +
  geom_line(aes(y = smoothed5), color = "gray65") +
  facet_wrap(~Well) +
  scale_y_continuous(trans = "log10")
#> Warning: Transformation introduced infinite values in continuous y-axis
#> Warning in self$trans$transform(x): NaNs produced
#> Warning: Transformation introduced infinite values in continuous y-axis
#> Warning in self$trans$transform(x): NaNs produced
#> Warning: Transformation introduced infinite values in continuous y-axis
#> Warning: Removed 18 row(s) containing missing values (geom_path).
```



Here we can see that `loess` with smaller spans (darker lines) have smoothed the data somewhat but are still sensitive to outliers. However, `loess` with a larger span (lightest line) has introduced significant bias. To fix this, we might explore other smoothing methods, or combining `loess` with other smoothing methods.

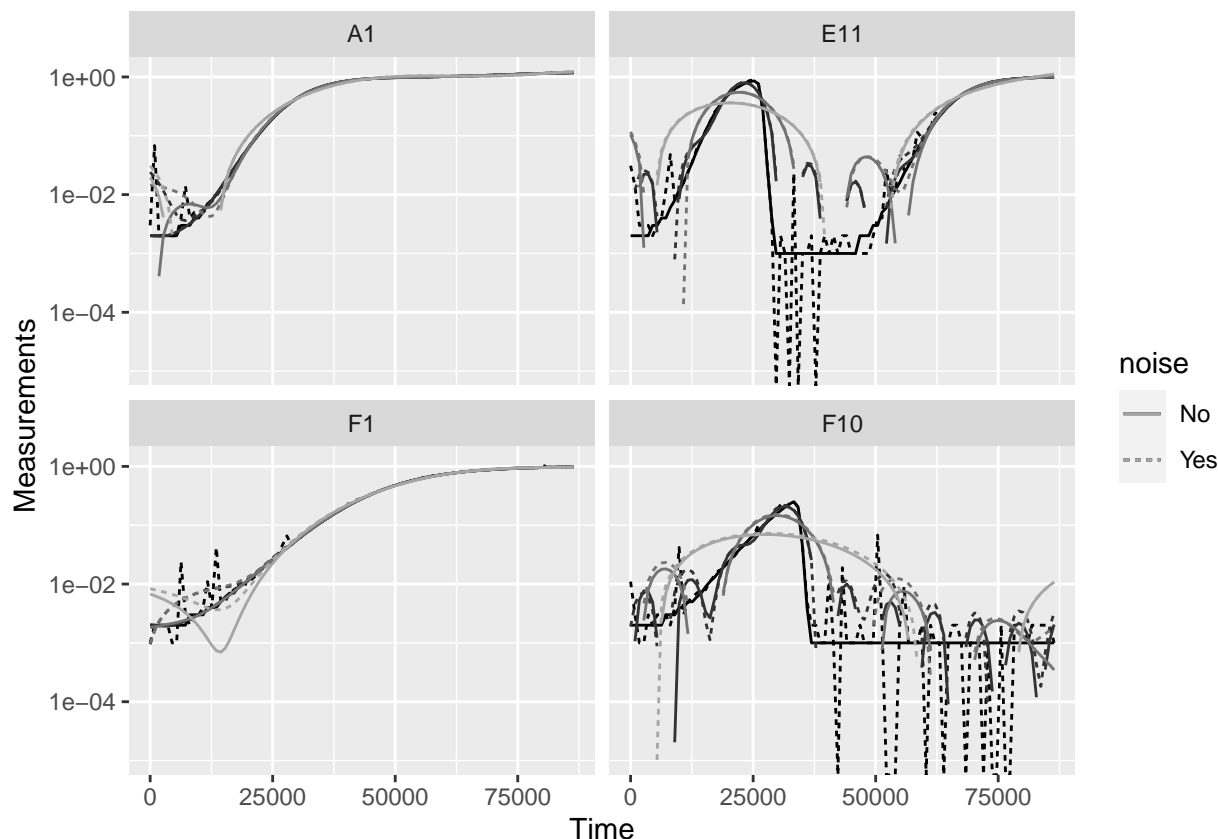
## Smoothing with GAM

For `gam`, the primary tuning parameter is the `k` argument. `gam` works by doing fits on subsets of the data and linking these fits together. `k` determines how many link points (“knots”) it can use. If not specified, the default `k` value for smoothing a time series is 10, with **smaller values being more “smoothed”** (note

this is opposite the trend with other smoothing methods). However, **unlike earlier methods, k values that are too large are also problematic**, as they will tend to ‘overfit’ the data. k cannot be larger than the number of data points, and should usually be substantially smaller than that. Also note that **gam can sometimes create artifacts**, especially oscillations in your density and derivatives. You should check that gam is not doing so before carrying on with your analyses. Here, we’ll show gam smoothing with k values of 5, 10, and 20.

```
ex_dat_mrg <-
  mutate(group_by(ex_dat_mrg, Well, Bacteria_strain, Phage, noise),
    smoothed20 = smooth_data(x = Time, y = Measurements,
                             sm_method = "gam", k = 20),
    smoothed10 = smooth_data(x = Time, y = Measurements,
                             sm_method = "gam", k = 10),
    smoothed5 = smooth_data(x = Time, y = Measurements,
                             sm_method = "gam", k = 5))

#What does the smoothed data look like compared to the noisy original?
#Lighter lines are smaller k and more "smoothed"
ggplot(data = dplyr::filter(ex_dat_mrg, Well %in% sample_wells),
  aes(x = Time, lty = noise)) +
  geom_line(aes(y = Measurements)) +
  geom_line(aes(y = smoothed20), color = "gray20") +
  geom_line(aes(y = smoothed10), color = "gray45") +
  geom_line(aes(y = smoothed5), color = "gray65") +
  facet_wrap(~Well) +
  scale_y_continuous(trans = "log10")
#> Warning: Transformation introduced infinite values in continuous y-axis
#> Warning in self$trans$transform(x): NaNs produced
#> Warning: Transformation introduced infinite values in continuous y-axis
#> Warning in self$trans$transform(x): NaNs produced
#> Warning: Transformation introduced infinite values in continuous y-axis
#> Warning in self$trans$transform(x): NaNs produced
#> Warning: Transformation introduced infinite values in continuous y-axis
#> Warning: Removed 2 row(s) containing missing values (geom_path).
```



Here we can see that `gam` does alright when working with the no phage-added wells (A1 and F1): higher `k` values (darkest line) have smoothed the data but are still sensitive to those early outliers, while lower `k` values (lighter lines) have introduced significant bias. However, `gam` is struggling when phage have been added (E11 and F10). Across all the `k` values it has added many fluctuations and often dips into values of 0 or lower (plotted here as breaks in the line, since the log of numbers  $\leq 0$  are undefined). To fix this, we might explore other smoothing methods or combining `gam` with other smoothing methods.

## Combining multiple smoothing methods

Often, combining multiple smoothing methods can provide improved results. For instance, `moving-median` is particularly good at removing outliers, but not very good at producing continuously smooth data. In contrast, `moving-average`, `loess`, and `gam` work better at producing continuously smooth data, but aren't as good at removing outliers. Here's an example using the strengths of both `moving-median` and `moving-average`. (Note that earlier columns created in `mutate` are available during creation of later columns, so both can be done in one step):

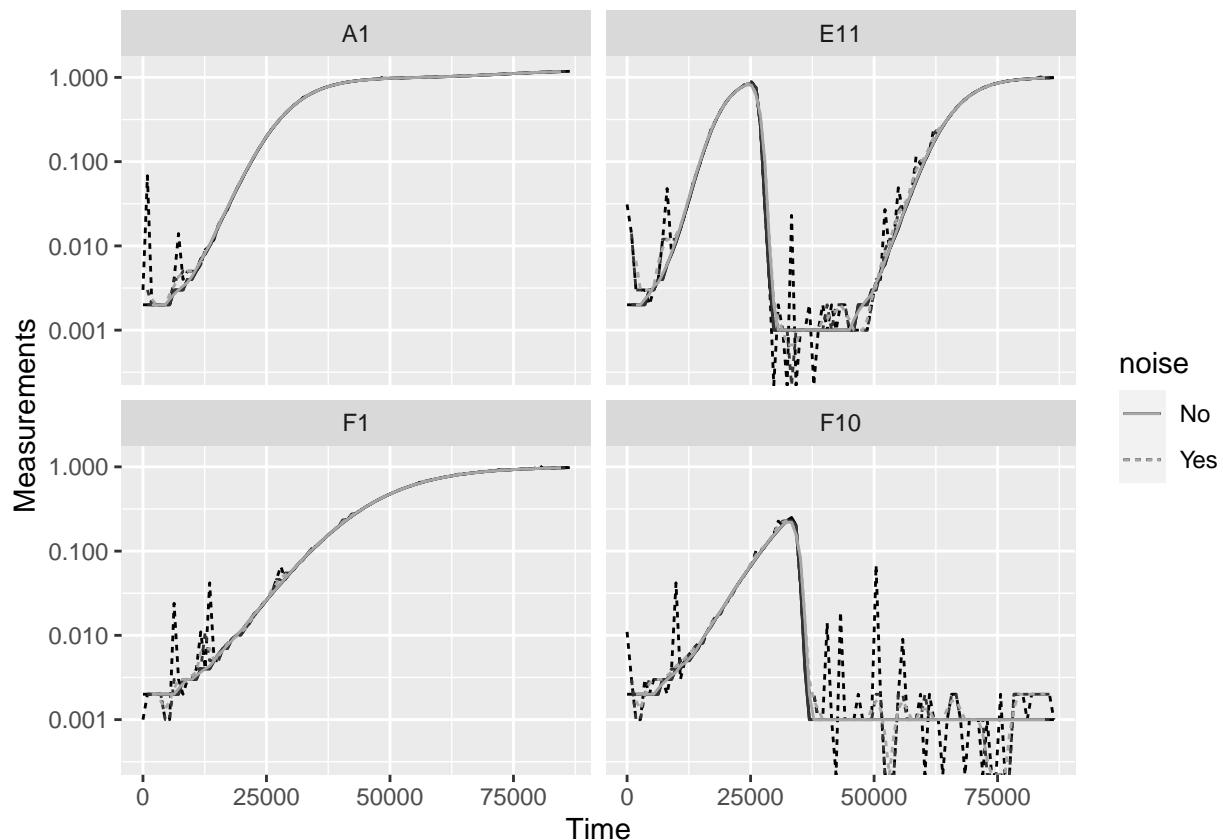
```
ex_dat_mrg <-
  mutate(group_by(ex_dat_mrg, Well, Bacteria_strain, Phage, noise),
    smoothed_med3 =
      smooth_data(x = Time, y = Measurements,
        sm_method = "moving-median", window_width_n = 3),
    #Note that for the second round, we're using the
    #first smoothing as the input y
    smoothed =
      smooth_data(x = Time, y = smoothed_med3,
```

```

sm_method = "moving-average", window_width_n = 3))

#What does the smoothed data look like compared to the noisy original?
#The first round of smoothing with moving-median is plotted in lighter colors
#The second round of smoothing with moving-average is plotted in darker colors
ggplot(data = dplyr::filter(ex_dat_mrg, Well %in% sample_wells),
  aes(x = Time, lty = noise)) +
  geom_line(aes(y = Measurements)) +
  geom_line(aes(y = smoothed_med3), color = "gray20") +
  geom_line(aes(y = smoothed), color = "gray65") +
  facet_wrap(~Well) +
  scale_y_continuous(trans = "log10")
#> Warning: Transformation introduced infinite values in continuous y-axis
#> Transformation introduced infinite values in continuous y-axis
#> Transformation introduced infinite values in continuous y-axis
#> Warning: Removed 4 row(s) containing missing values (geom_path).
#> Warning: Removed 8 row(s) containing missing values (geom_path).

```



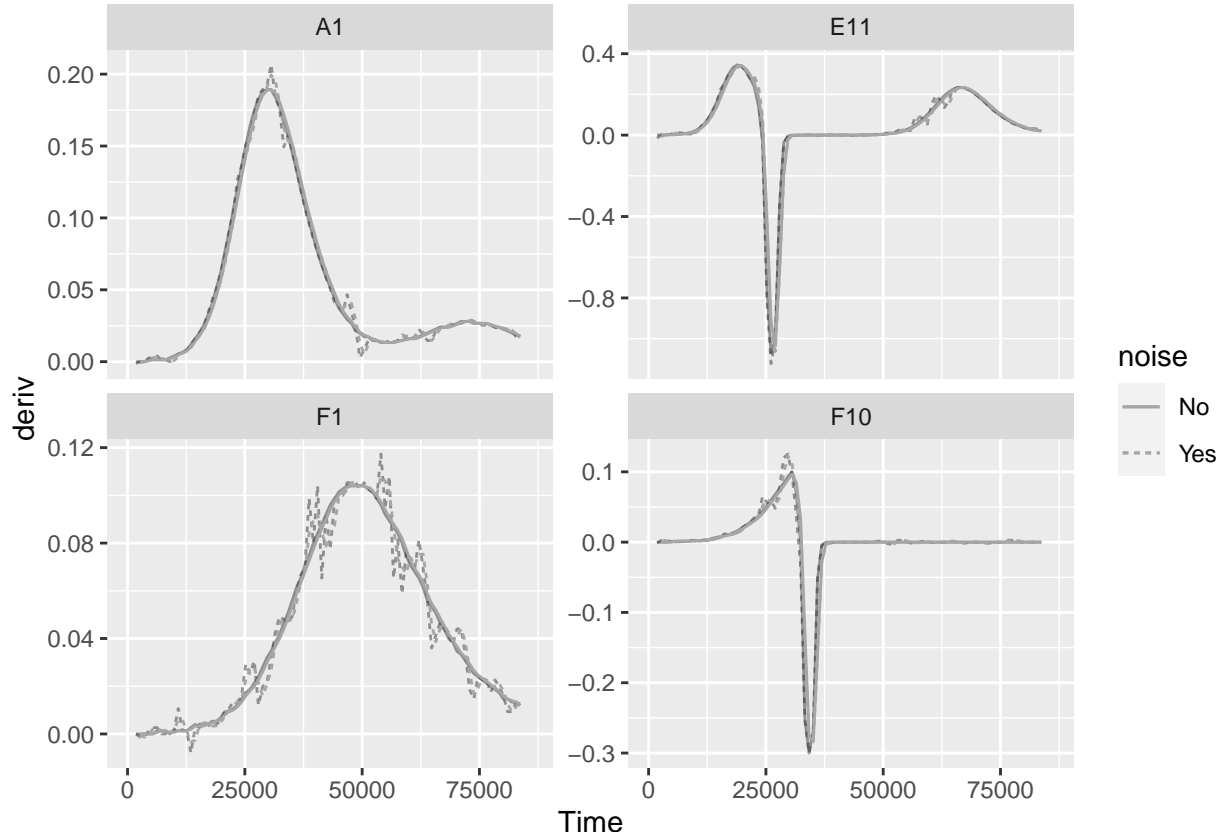
Here we can see that the combination of minimal moving-median and moving-average smoothing has produced a curve that has most of the noise removed with minimal introduction of bias. (Note that the first and last 2 data points are now NA because of the smoothing)



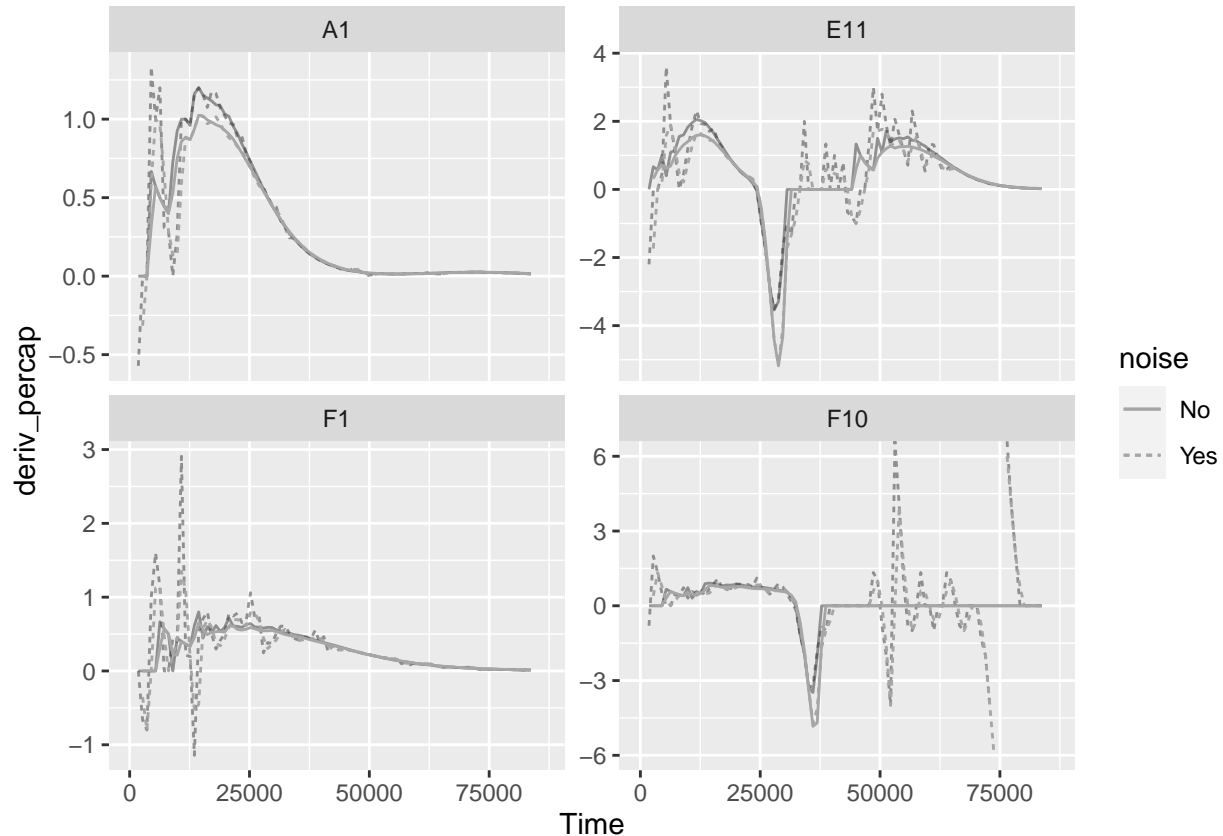
## Calculating derivatives of smoothed data

Once you've smoothed your data, you can calculate derivatives using the smoothed data. Combining smoothing of raw data and fitting using multiple points for calculating derivatives can be a powerful combination for reducing the effects of noise while minimizing the introduction of bias.

```
ex_dat_mrg <-  
  mutate(group_by(ex_dat_mrg, Well, Bacteria_strain, Phage, noise),  
    deriv = calc_deriv(x = Time, y = smoothed, x_scale = 3600),  
    deriv_percap = calc_deriv(x = Time, y = smoothed, x_scale = 3600,  
                             percapita = TRUE, blank = 0),  
    deriv3 = calc_deriv(x = Time, y = smoothed, x_scale = 3600,  
                       window_width_n = 3),  
    deriv_percap3 = calc_deriv(x = Time, y = smoothed, x_scale = 3600,  
                              percapita = TRUE, blank = 0,  
                              window_width_n = 3))  
  
#Plot derivative  
ggplot(data = dplyr::filter(ex_dat_mrg, Well %in% sample_wells),  
  aes(x = Time, y = deriv, lty = noise)) +  
  geom_line(alpha = 0.5, color = "gray20") +  
  geom_line(aes(y = deriv3, color = "gray65") +  
    facet_wrap(~Well, scales = "free_y")  
#> Warning: Removed 10 row(s) containing missing values (geom_path).  
#> Warning: Removed 12 row(s) containing missing values (geom_path).
```



```
#Plot per-capita derivative
ggplot(data = dplyr::filter(ex_dat_mrg, Well %in% sample_wells),
  aes(x = Time, y = deriv_percap, lty = noise)) +
  geom_line(alpha = 0.5, color = "gray20") +
  geom_line(aes(y = deriv_percap3, color = "gray65") +
  facet_wrap(~Well, scales = "free_y")
#> Warning: Removed 10 row(s) containing missing values (geom_path).
#> Removed 12 row(s) containing missing values (geom_path).
```

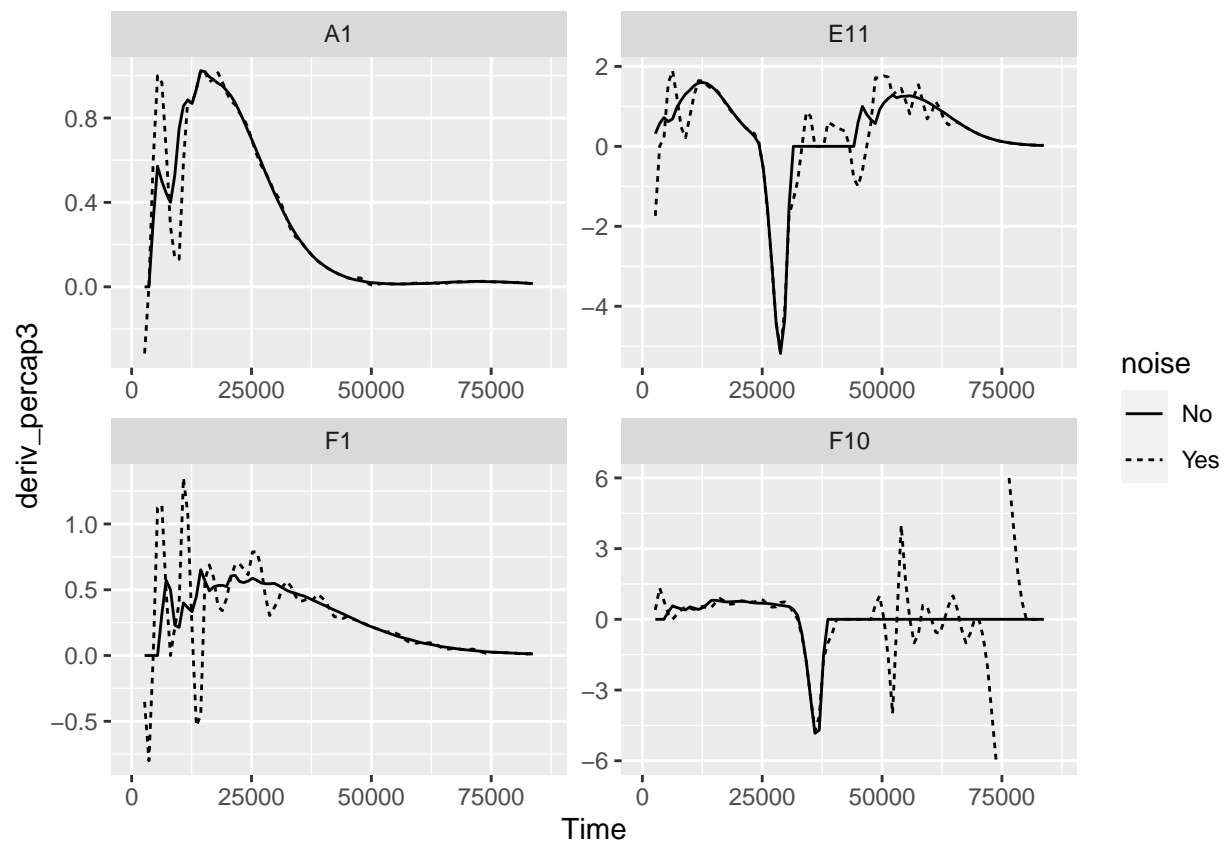


## Summarizing on subsets of derivatives

There is one final strategy we can employ when dealing with noisy data: since noise often has relatively stronger effects when densities are near 0, we can simply exclude data points where the density is near 0.

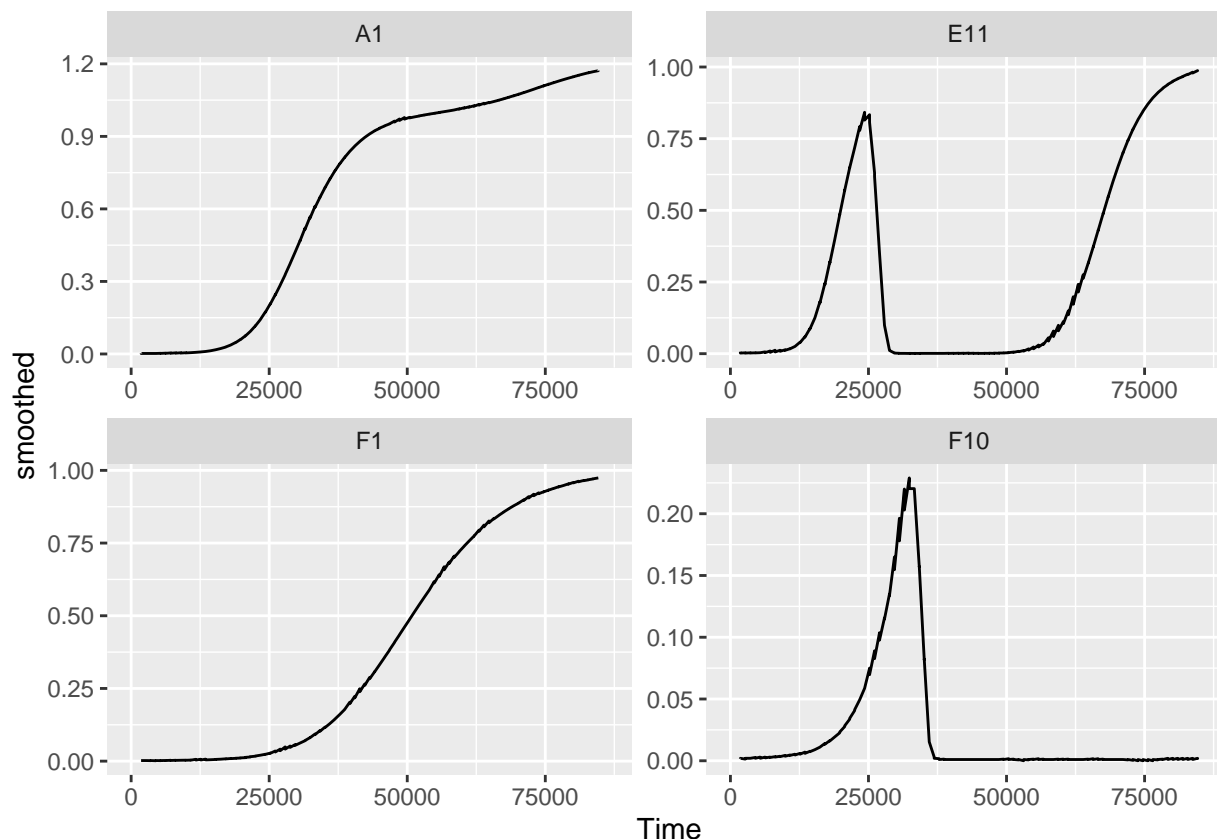
Let's look again at our smoothed per-capita growth rates:

```
ggplot(data = dplyr::filter(ex_dat_mrg, Well %in% sample_wells),
  aes(x = Time, y = deriv_percap3, lty = noise)) +
  geom_line() +
  facet_wrap(~Well, scales = "free")
#> Warning: Removed 12 row(s) containing missing values (geom_path).
```



And now let's compare to the density plots:

```
ggplot(data = dplyr::filter(ex_dat_mrg, Well %in% sample_wells),
  aes(x = Time, y = smoothed)) +
  geom_line() +
  facet_wrap(~Well, scales = "free")
#> Warning: Removed 8 row(s) containing missing values (geom_path).
```



Clearly we can see that most of the noise in the per-capita growth rate occurs when the bacterial population density is very low. Indeed, **this is common with per-capita growth rates, which are very sensitive to noise at low densities**. What can we do about it? We can simply exclude all the values when the *density* is really low.

Let's plot our per-capita growth rate data at different cutoffs for the minimum *density* of bacteria:

```
for (my_well in sample_wells) {
  #Title
  title <- cowplot::ggdraw() +
    cowplot::draw_label(paste("Well", my_well),
                        fontface = "bold", x = 0, hjust = 0) +
    theme(plot.margin = margin(0, 0, 0, 7))

  #Save x and y limits for all plots so they're all on the same axes
  xdat <- dplyr::filter(ex_dat_mrg, Well == my_well)$Time
  ydat <- dplyr::filter(ex_dat_mrg, Well == my_well)$deriv_percap3
  xlims <- c(min(xdat[is.finite(xdat)], na.rm = TRUE),
             max(xdat[is.finite(xdat)], na.rm = TRUE))
  ylims <- c(min(ydat[is.finite(ydat)], na.rm = TRUE),
             max(ydat[is.finite(ydat)], na.rm = TRUE))

  #Plot unfiltered data
  p1 <- ggplot(data = dplyr::filter(ex_dat_mrg, Well == my_well),
              aes(x = Time, y = deriv_percap3, color = noise)) +
    geom_point(alpha = 0.5) + facet_wrap(~Well, scales = "free") +
    ggtitle("all data") +
```

```

xlim(xlims[1], xlims[2]) + ylim(ylims[1], ylims[2])

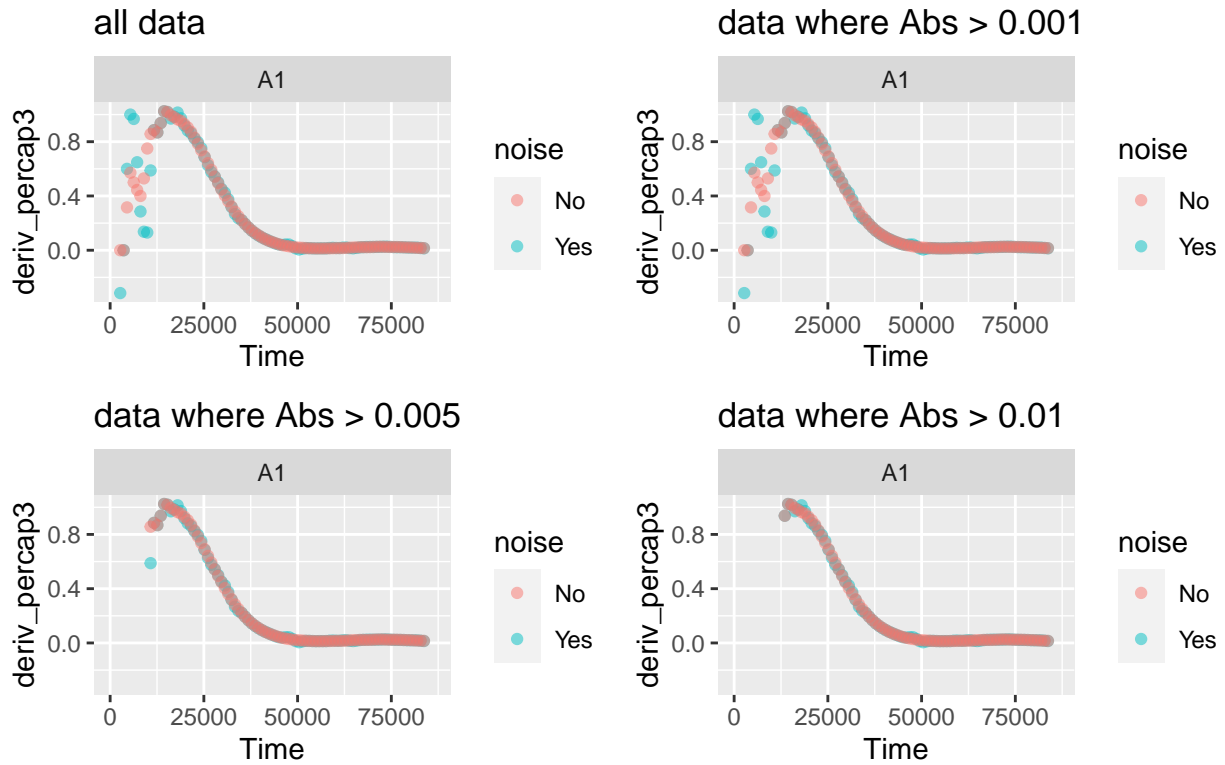
#Plot data with filters for density
p2 <- ggplot(data = dplyr::filter(ex_dat_mrg,
                                Well == my_well, smoothed > 0.001),
             aes(x = Time, y = deriv_percap3, color = noise)) +
  geom_point(alpha = 0.5) + facet_wrap(~Well, scales = "free") +
  ggtitle("data where Abs > 0.001") +
  xlim(xlims[1], xlims[2]) + ylim(ylims[1], ylims[2])
p3 <- ggplot(data = dplyr::filter(ex_dat_mrg,
                                Well == my_well, smoothed > 0.005),
             aes(x = Time, y = deriv_percap3, color = noise)) +
  geom_point(alpha = 0.5) + facet_wrap(~Well, scales = "free") +
  ggtitle("data where Abs > 0.005") +
  xlim(xlims[1], xlims[2]) + ylim(ylims[1], ylims[2])
p4 <- ggplot(data = dplyr::filter(ex_dat_mrg,
                                Well == my_well, smoothed > 0.01),
             aes(x = Time, y = deriv_percap3, color = noise)) +
  geom_point(alpha = 0.5) + facet_wrap(~Well, scales = "free") +
  ggtitle("data where Abs > 0.01") +
  xlim(xlims[1], xlims[2]) + ylim(ylims[1], ylims[2])

print(cowplot::plot_grid(title, cowplot::plot_grid(p1, p2, p3, p4, ncol = 2),
                        ncol = 1, rel_heights = c(0.1, 1)))
}

#> Warning: Removed 12 rows containing missing values (geom_point).
#> Warning: Removed 4 rows containing missing values (geom_point).
#> Warning: Removed 2 rows containing missing values (geom_point).
#> Removed 2 rows containing missing values (geom_point).
#> Warning: Removed 12 rows containing missing values (geom_point).
#> Warning: Removed 4 rows containing missing values (geom_point).
#> Warning: Removed 2 rows containing missing values (geom_point).
#> Removed 2 rows containing missing values (geom_point).

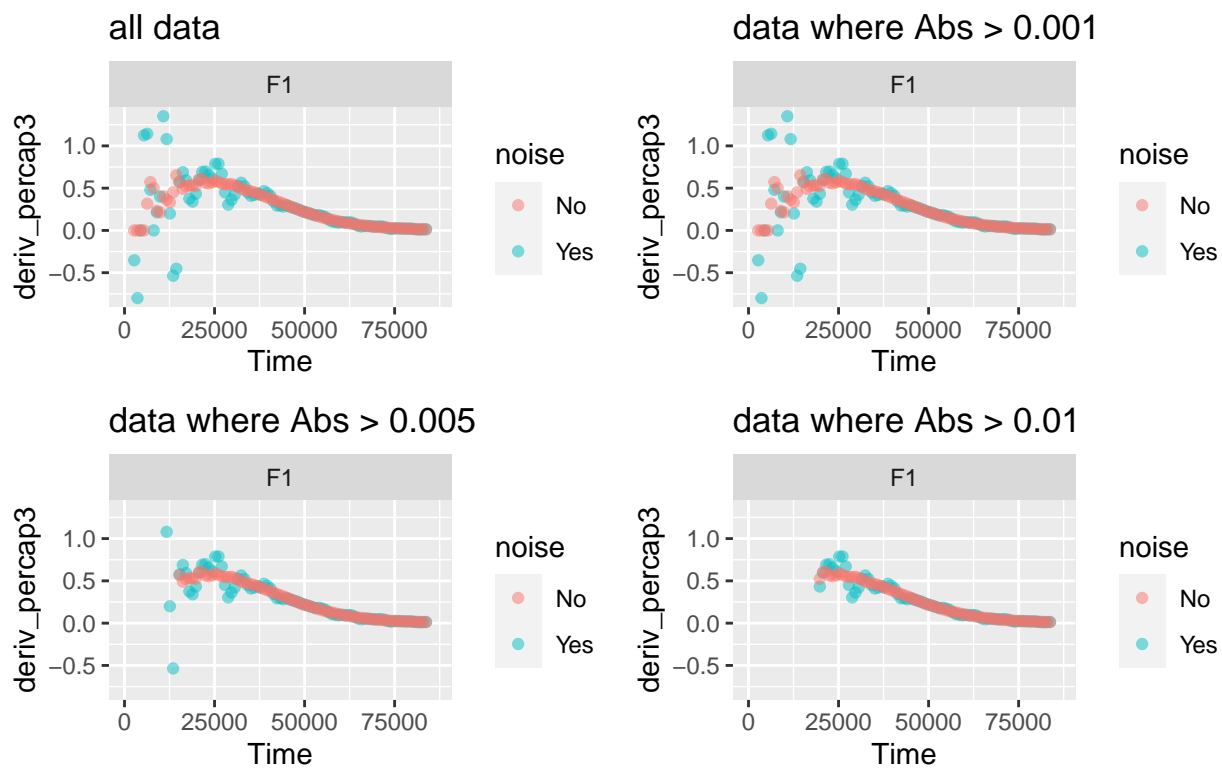
```

## Well A1



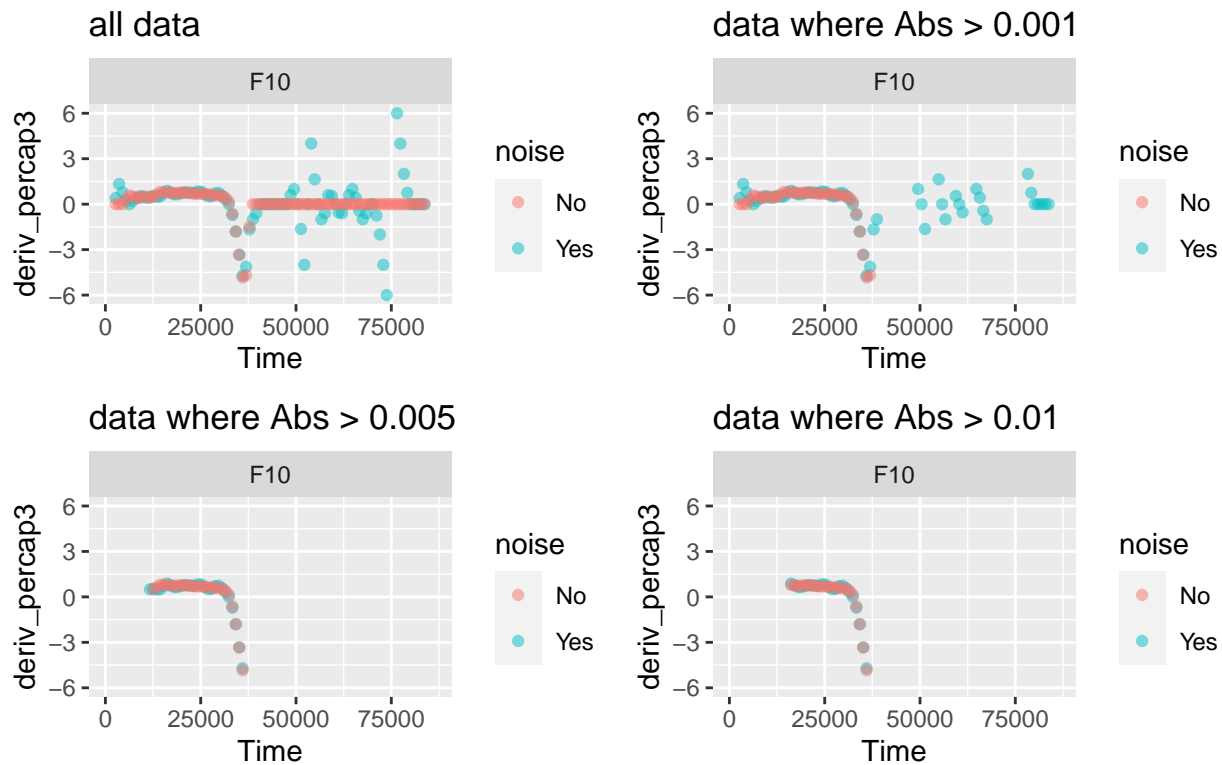
```
#> Warning: Removed 14 rows containing missing values (geom_point).  
#> Warning: Removed 3 rows containing missing values (geom_point).
```

## Well F1



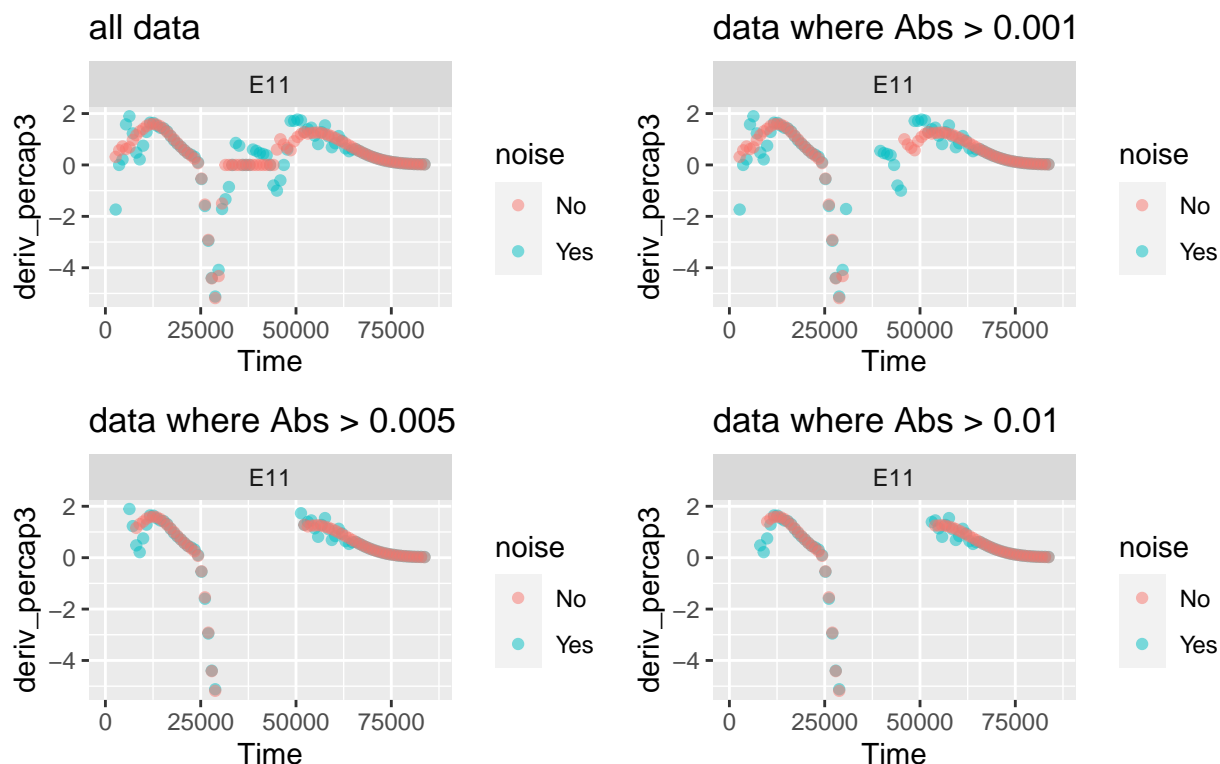
```
#> Warning: Removed 12 rows containing missing values (geom_point).
#> Warning: Removed 4 rows containing missing values (geom_point).
#> Warning: Removed 3 rows containing missing values (geom_point).
#> Warning: Removed 2 rows containing missing values (geom_point).
```

## Well F10





## Well E11



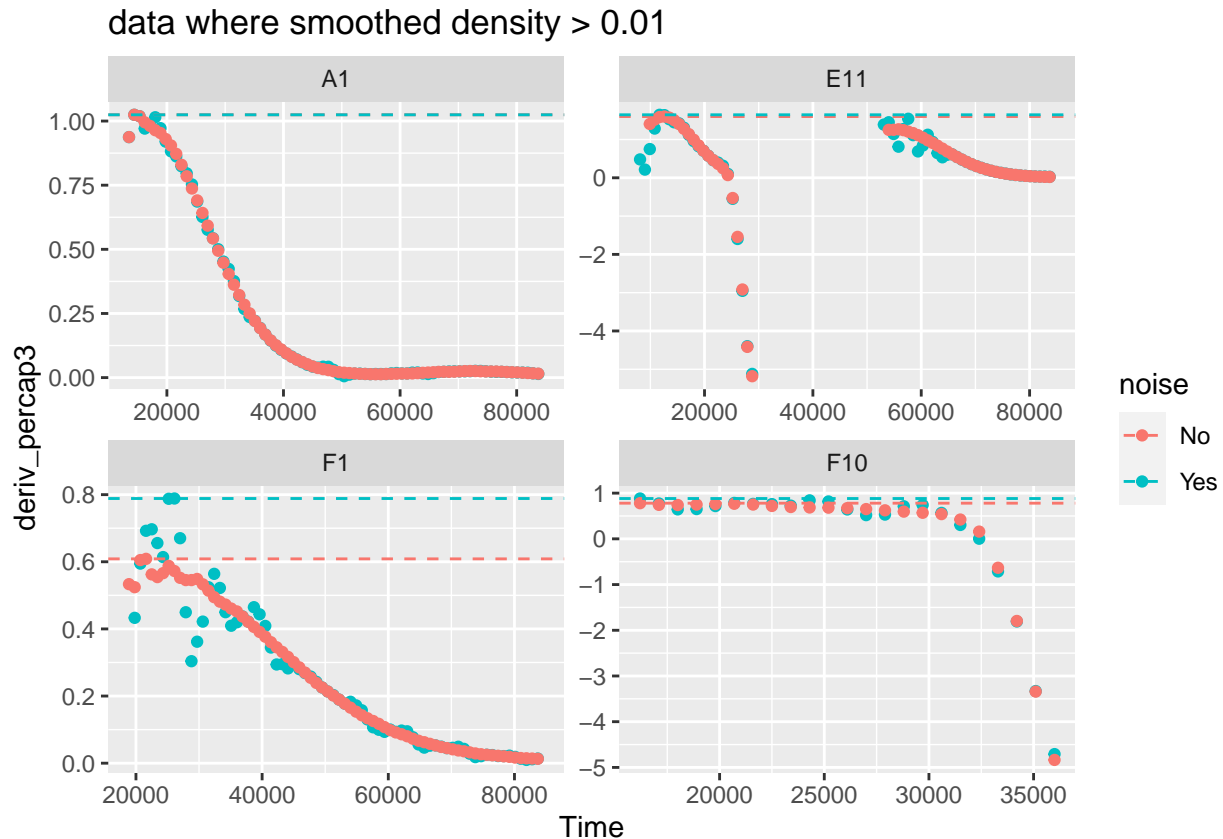
As we can see, as we limit our analyses to derivatives where the bacterial population is above a cutoff density, many of the most noisy points disappear, and the noisy derivative curves look increasingly similar to the noiseless derivative curves.

To take this to the final step, we can use these cutoffs in our `summarize` commands to calculate the maximum growth rate of the bacteria when their density is at least 0.005.

```
ex_dat_mrg_sum <-
  summarize(group_by(ex_dat_mrg, Well, Bacteria_strain, Phage, noise),
            max_growth_rate = max(deriv_percap3[smoothed > 0.01],
                                   na.rm = TRUE))
#> `summarise()` has grouped output by 'Well', 'Bacteria_strain', 'Phage'. You can override
#> using the `.groups` argument.
head(ex_dat_mrg_sum)
#> # A tibble: 6 x 5
#> # Groups:   Well, Bacteria_strain, Phage [3]
#>   Well Bacteria_strain Phage   noise max_growth_rate
#>   <fct> <chr>          <chr>   <chr>          <dbl>
#> 1 A1    Strain 1        No Phage No           1.02
#> 2 A1    Strain 1        No Phage Yes          1.02
#> 3 A2    Strain 2        No Phage No           1.35
#> 4 A2    Strain 2        No Phage Yes          1.49
#> 5 A3    Strain 3        No Phage No           0.935
#> 6 A3    Strain 3        No Phage Yes          1.19
```

And now we can visualize our findings:

```
ggplot(data = dplyr::filter(ex_dat_mrg,
                           Well %in% sample_wells, smoothed >= 0.01),
       aes(x = Time, y = deriv_percap3, color = noise)) +
  geom_point() +
  facet_wrap(~Well, scales = "free") +
  ggtitle("data where smoothed density > 0.01") +
  geom_hline(data = dplyr::filter(ex_dat_mrg_sum, Well %in% sample_wells),
            aes(yintercept = max_growth_rate, color = noise), lty = 2)
#> Warning: Removed 6 rows containing missing values (geom_point).
```



Here we can see that by limiting our analyses to just a subset of the data, the maximum per-capita growth rate is nearly identical in three of the example wells, while in F1 the noise is altering the calculated maximum somewhat. If this happens in your data, continue to try alternate smoothing, derivative calculating, and subset strategies to try to further reduce the effects of noise on your findings.

## What's next?

Now that you've analyzed your data and dealt with any noise, there's just some concluding notes on best practices for running statistics, merging growth curve analyses with other data, and additional resources for analyzing growth curves.

1. Introduction: `vignette("gcplyr")`
2. Importing and transforming data: `vignette("import_transform")`
3. Incorporating design information: `vignette("incorporate_designs")`

4. Pre-processing and plotting your data: `vignette("preprocess_plot")`
5. Processing your data: `vignette("process")`
6. Analyzing your data: `vignette("analyze")`
7. Dealing with noise: `vignette("noise")`
8. **Statistics, merging other data, and other resources:** `vignette("conclusion")`