



University of  
Zurich<sup>UZH</sup>



URPP Evolution  
in Action

# URPP tutorial

## R basic tutorial I

Dr. Heidi E.L. Tschanz-Lischer  
University of Zurich  
Switzerland

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# Outline

## 1. Repeating things:

Introduction to ways of automating repetitive tasks

## 2. Control structures

## 3. Missing data

## 4. Bad habits in coding

## 5. R graphics

# Test data set

- We will work with a test data set of R:

- To see all test data sets of R:

```
data()
```

- Iris dataset (Edgar Anderson's Iris Data):

```
data(iris)
```

- Summary of the dataset:

```
summary(iris)
```

```
summary(iris)
```

| Sepal.Length  | Sepal.Width   | Petal.Length  | Petal.Width   | Species       |
|---------------|---------------|---------------|---------------|---------------|
| Min. :4.300   | Min. :2.000   | Min. :1.000   | Min. :0.100   | setosa :50    |
| 1st Qu.:5.100 | 1st Qu.:2.800 | 1st Qu.:1.600 | 1st Qu.:0.300 | versicolor:50 |
| Median :5.800 | Median :3.000 | Median :4.350 | Median :1.300 | virginica :50 |
| Mean :5.843   | Mean :3.057   | Mean :3.758   | Mean :1.199   |               |
| 3rd Qu.:6.400 | 3rd Qu.:3.300 | 3rd Qu.:5.100 | 3rd Qu.:1.800 |               |
| Max. :7.900   | Max. :4.400   | Max. :6.900   | Max. :2.500   |               |

Quantify the morphologic variation of three *Iris* flowers:

- setosa
- versicolor
- virginica

Sepal and petal length/width



# Ways of automating repetitive tasks

- Many times you have to repeat things
  - Common problem:
    - copy-and-pasted some piece of code a bunch of times
    - changing one thing each time
- Script gets very long  
→ Hard to read  
→ Bugs accumulate

# Apply a function over and over again

- Use the iris data set:

```
head(iris)
```

|   | Sepal.Length | Sepal.Width | Petal.Length | Petal.Width | Species |
|---|--------------|-------------|--------------|-------------|---------|
| 1 | 5.1          | 3.5         | 1.4          | 0.2         | setosa  |
| 2 | 4.9          | 3.0         | 1.4          | 0.2         | setosa  |
| 3 | 4.7          | 3.2         | 1.3          | 0.2         | setosa  |
| 4 | 4.6          | 3.1         | 1.5          | 0.2         | setosa  |
| 5 | 5.0          | 3.6         | 1.4          | 0.2         | setosa  |
| 6 | 5.4          | 3.9         | 1.7          | 0.4         | setosa  |

- We want to get the mean of each column:

```
mean(iris$Sepal.Length)  
mean(iris$Sepal.Width)  
mean(iris$Petal.Length)  
mean(iris$Petal.Width)
```

→ Very repetitive!

# sapply

- We can use **sapply**: apply a Function over a List or Vector:
  - first argument: a list (or vector)
  - applies a function to each element
- Example:

```
obj <- list(a=1:5, b=c(1, 5, 6))
obj
$a
[1] 1 2 3 4 5
```

```
$b
[1] 1 5 6
```

```
sapply(obj, length)
a b
5 3
```

← computes the length of each element in obj  
(this works for any function!)

# sapply



Apply it to estimate means of each column in the iris data set.  
Additionally estimate variance and standard deviation.  
(hint: use `iris[,1:4]` to select first 4 columns)

```
mean(iris$Sepal.Length)
mean(iris$Sepal.Width)
mean(iris$Petal.Length)
mean(iris$Petal.Width)
```

# sapply

- `sapply` can also be used with an own function:

```
range <- function(x) {  
  max(x)-min(x)  
}  
sapply(iris[,1:4],range)  
Sepal.Length Sepal.Width Petal.Length Petal.Width  
      3.6          2.4          5.9          2.4
```

- Also possible to supply more than one argument:

```
getQuantile <- function(x,probability) {  
  quantile(x,probs=probability)  
}  
sapply(iris[,1:4],getQuantile,0.3)  
Sepal.Length.30% Sepal.Width.30% Petal.Length.30% Petal.Width.30%  
      5.27          2.80          1.70          0.40
```

Additional arguments are put behind function name

# lapply

- **sapply:**
  - apply a Function over a List or Vector
  - returns a vector or matrix
- **lapply:**
  - apply a Function over a List or Vector
  - returns a list

```
obj <- list(a=1:5, b=c(1, 5, 6))
sapply(obj, length)
a b
5 3

lapply(obj, length)
$a
[1] 5

$b
[1] 3
```

# apply

- To apply a function over an array or matrix use `apply`:
  - Second argument:
    - 1: apply function over rows
    - 2: apply function over columns
  - Third argument: function to apply

```
matrix.irisSe <- as.matrix(iris[iris$Species=="setosa",1:4])
apply(matrix.irisSe[,1:4],1,mean)
  1      2      3      4      5      6      7      8      9      10     11 
2.550 2.375 2.350 2.350 2.550 2.850 2.425 2.525 2.225 2.400 2.700 ... 

apply(matrix.irisSe[,1:4],2,mean)
Sepal.Length Sepal.Width Petal.Length Petal.Width
      5.006       3.428       1.462       0.246
```

# Apply a function over each treatment

- Often one wants to apply a function over each treatment/sample
- Example: mean sepal length for each iris species:

```
mean(iris$Sepal.Length[iris$Species=="setosa"])
mean(iris$Sepal.Length[iris$Species=="versicolor"])
mean(iris$Sepal.Length[iris$Species=="virginica"])
```

- **tapply**:
  - apply a Function over a List or Vector
  - Second argument: is the grouping variable (list of one or more factor)
  - Third argument: function to apply

```
tapply(iris$Sepal.Length,iris$Species,mean)
      setosa  versicolor  virginica
      5.006       5.936       6.588
```

# Summary: apply functions

| function | apply to            | returns                       | special                                  |
|----------|---------------------|-------------------------------|--|
| supply   | - List<br>- vector  | - Vector<br>- matrix          |  |
| lapply   | - List<br>- vector  | - List                        |  |
| apply    | - Array<br>- Matrix | - Vector<br>- Array<br>- List |  |
| tapply   | - Vector            | - Array                       | Apply function to each factor separately |

# loops

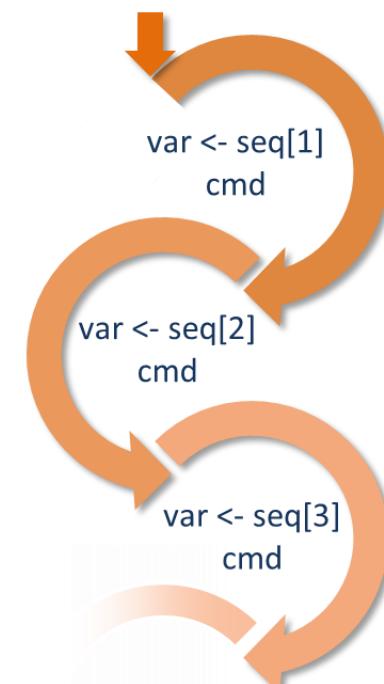
You can also use **loops** to repeat things

- **for:** If you know in advance the values that the loop variable will have each time it goes through the loop
- **while:** If you do not know in advance how often the instructions will be executed

## For-loop:

- **controlled by a looping vector**  
→ do something for all items in the vector
- In every iteration of the loop one value in the looping vector is assigned to a variable

```
for(var in seq) {  
  cmd  
}
```



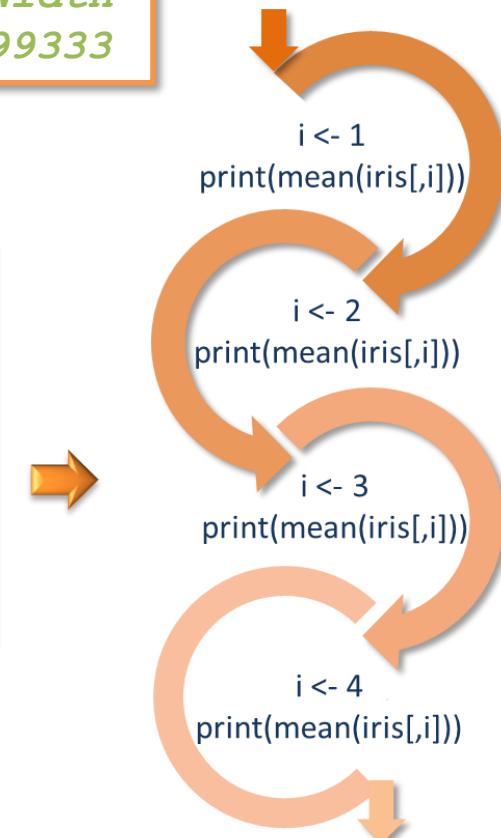
# For-loop

- Everything we did before can also be done with a for-loop:

```
sapply(iris[,1:4],mean)
Sepal.Length  Sepal.Width  Petal.Length  Petal.Width
      5.843333    3.057333    3.758000    1.199333
```

```
for(i in 1:4){
  print(mean(iris[,i]))
}
[1] 5.843333
[1] 3.057333
[1] 3.758
[1] 1.199333
```

In a loop `print` is needed to make results visible



# For-loop

- You can also store results of a for-loop in a vector:

```
out <- numeric(4)
for(i in 1:4){
  out[i] <- mean(iris[,i])
}
out
[1] 5.843333 3.057333 3.758000 1.199333
```

```
names(out) <- names(iris[,1:4])
Sepal.Length Sepal.Width Petal.Length Petal.Width
      5.843333      3.057333      3.758000      1.199333
```

Add names to the out vector

# For-loop

- There are times where loops are absolutely the best (or the only way) of doing something
- Example: if one wants to get cumulative sum over the elements

```
out <- numeric(4)
total <- 0      # running total
for(i in 1:4) {
  total <- total + sum(iris[,i])
  out[i] <- total
}
names(out) <- names(iris[,1:4])
out
Sepal.Length  Sepal.Width  Petal.Length  Petal.Width
  876.5        1335.1       1898.8       2078.7

sapply(iris[,1:4],sum)
Sepal.Length  Sepal.Width  Petal.Length  Petal.Width
  876.5        458.6       563.7        179.9
```

# Apply/for-loop exercises

Use the data set Orange (`data(Orange)`):

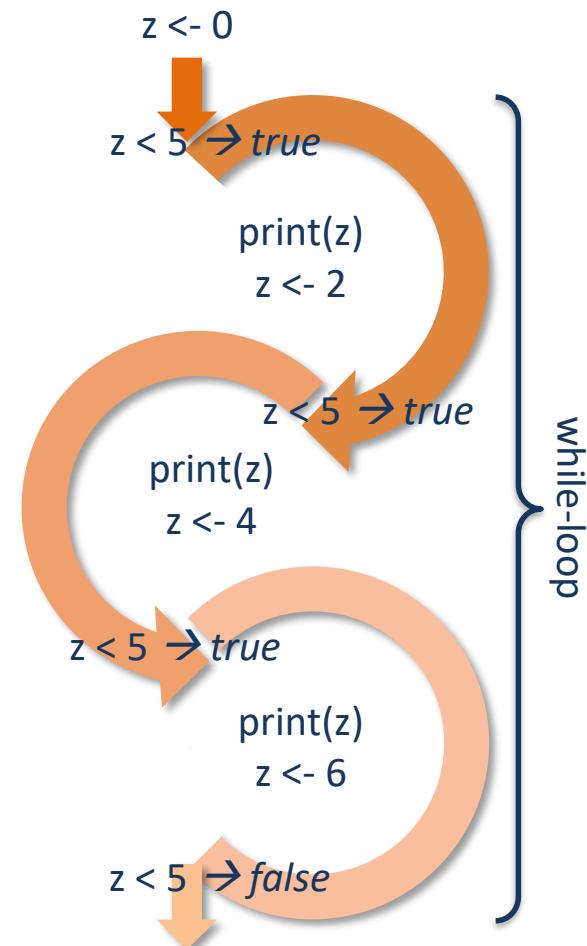
- ① Estimate mean and sd of the age and circumference across all trees
- ② Estimate mean and sd of circumference for each tree separately
- ③ Use for-loop to estimate the minimum and maximum of the age and circumference across all trees

# while loop

- controlled by a conditional statement  
→ do something while condition is true

```
while (cond1=true) {  
    cmd  
}
```

```
z <- 0  
while(z < 5) {  
    print(z)  
    z <- z + 2  
}  
[1] 0  
[1] 2  
[1] 4
```



# Control structures

- **Control structures:**  
a block of programming that analyzes variables and chooses a direction in which to go based on given parameters
- **basic decision-making process:** choose' which lines of code to execute depending on the state of the variables inside the control structure
  - could make the code jump to a completely different part of the program
  - could make it re-run a certain piece again
  - just skip a bunch of code
- immensely important for programming  
→ they make the programs function properly
- without control structures the program would only flow in one way  
→ **Changing what the code does based on a variable is what makes programs useful!**

# If-else statement

- **If-else statements:**  
can be used to check for something and do something else depending on the outcome of the check
- **if-else** can be combined unlimited

```
if(cond1=true) {  
    cmd1  
} else if(cond2=true) {  
    cmd2  
} else {  
    cmd3  
}
```

→ Avoid inserting newlines  
between '} else'

```
number <- 0  
if(number < 0){  
    print("negative")  
} else if (number == 0) {  
    print("zero")  
} else {  
    print("positive")  
}  
[1] "zero"
```

# Conditional statements

Comparison operators:

| Operator           | Meaning                  |
|--------------------|--------------------------|
| <code>==</code>    | Equality                 |
| <code>&lt;</code>  | Less than                |
| <code>&gt;</code>  | Greater than             |
| <code>&lt;=</code> | Less than or equal to    |
| <code>&gt;=</code> | Greater than or equal to |

Logical operators:

| Operator                | Meaning                             |
|-------------------------|-------------------------------------|
| <code>&amp;&amp;</code> | AND<br>(both statements true)       |
| <code>  </code>         | OR<br>(at least one statement true) |
| <code>!</code>          | NOT                                 |

```
if(a < 4 && b < 4) {  
    print("both < 4")  
} else {  
    print("min one >= 4")  
}
```

```
if(a < 4 || b < 4) {  
    print("min one < 4")  
} else {  
    print("both >= 4")  
}
```

```
if(a != 4) {  
    print("a not 4")  
} else {  
    print("a is 4")  
}
```

# Control structures exercises



## while-loop:

Write some lines of code that will figure out how many terms in the sum  $1+2+3+\dots$  it requires for the sum to exceed one million

→ Hint: Create a variable that will store the current sum and another variable that keeps track of what number you are adding to the sum



## Condition:

Write a short program that lists all numbers from 1 to 100 that are divisible by 2 and 3

→ Hint: use  $5\%2$  to get the rest of a division = 1

# Missing data

- Missing data are quite common in biology  
(Individuals die, equipment breaks, you forget to measure something,...)
- If you load in data with blank cells, they will appear as: NA
- NA is not a string ("NA")! → number that could stand in for anything
- Artificially insert an missing value in our iris data set:

```
iris[2,1] <- NA
```

```
head(iris)
```

|   | Sepal.Length | Sepal.Width | Petal.Length | Petal.Width | Species |
|---|--------------|-------------|--------------|-------------|---------|
| 1 | 5.1          | 3.5         | 1.4          | 0.2         | setosa  |
| 2 | NA           | 3.0         | 1.4          | 0.2         | setosa  |
| 3 | 4.7          | 3.2         | 1.3          | 0.2         | setosa  |
| 4 | 4.6          | 3.1         | 1.5          | 0.2         | setosa  |
| 5 | 5.0          | 3.6         | 1.4          | 0.2         | setosa  |
| 6 | 5.4          | 3.9         | 1.7          | 0.4         | setosa  |

# Missing data

- If we estimate the mean of something which contains missing values, the result will also be missing (mean of anything will be anything)

```
iris[2,1] <- NA  
mean(iris[,1])  
[1] NA
```

- A lot of functions therefore include an option to remove missing values:

```
mean(iris[,1],na.rm=TRUE)  
[1] 5.849664
```

```
sum(iris[,1],na.rm=TRUE)  
[1] 871.6
```

# Missing data

- To check for missing data: `is.na()`
- To remove missing data from the data set: `na.omit()`

```
iris[2,1] <- NA
is.na(iris[2,])
  Sepal.Length Sepal.Width Petal.Length Petal.Width Species
2           TRUE      FALSE      FALSE      FALSE   FALSE

iris.na <- na.omit(iris)
head(iris.na)
  Sepal.Length Sepal.Width Petal.Length Petal.Width Species
1          5.1       3.5       1.4       0.2  setosa
3          4.7       3.2       1.3       0.2  setosa
4          4.6       3.1       1.5       0.2  setosa
5          5.0       3.6       1.4       0.2  setosa
6          5.4       3.9       1.7       0.4  setosa
7          4.6       3.4       1.4       0.3  setosa
```

→ removes whole second row

# Missing data – Exercise

- This standard error function doesn't deal with missing values:

```
standard.error <- function(x) {  
  v <- var(x)  
  n <- length(x)  
  sqrt(v/n)  
}  
standard.error(iris$Sepal.Length)  
[1] NA
```



Rewrite it that it filters missing data

# Bad habits in coding

## 1. Avoid indexing by location:

```
#avoid:  
iris[,1]  
#use instead:  
iris$Sepal.Length
```

- If you change the order, add or delete a columns, the index may change
- More robust and easier to read

## 2. Don't use **T** and **F** as shortcuts for **TRUE** and **FALSE**

- R allows to overwrite **T** but not **TRUE**

```
T <- "hello"  
T  
[1] "hello"  
TRUE <- "hello"  
Error in TRUE <- "hello" : invalid (do_set) left-hand side  
to assignment
```

# Bad habits in coding

3. Don't be inconsistent with assignment symbols, variable names,...  
→ easier to revisit the code in the future
4. Avoid copy and pasting large chunks of code between projects  
→ Make functions and copy those
5. Don't write enormously long functions  
→ difficult to read  
→ each function should do just one thing
6. Don't write functions that depend on global variables  
→ hard to use those functions elsewhere  
→ All data should be given as arguments to the function

# Bad habits in coding

## 7. Document your work

- You probably cannot have enough describing where data files came from, what scripts do what, how particular pieces of code work
- Avoid writing comments that just repeat what the code says
- Write comments that describe your intent, reasons for approaches, sources of data / code / algorithm
- Will make your life easier when you come back to a project

## 8. Keep things cleaned up

- Tidy computing workspaces are as important as tidy lab workspaces

# Bad habits in coding

## 9. Indent your code and do it consistently

- Makes the difference between easy and hard to read code
- Indent depth implies something about program structure

```
standard.error <- function(x) {  
  v <- var(x)  
  n <- length(x)  
  sqrt(v/n)  
}  
standard.error(iris$Sepal.Length)
```

```
standard.error <- function(x) {  
  v <- var(x)  
  n <- length(x)  
  sqrt(v/n)  
}  
standard.error(iris$Sepal.Length)
```

→ more obvious that the body of the function is three lines long

# Graphics within R



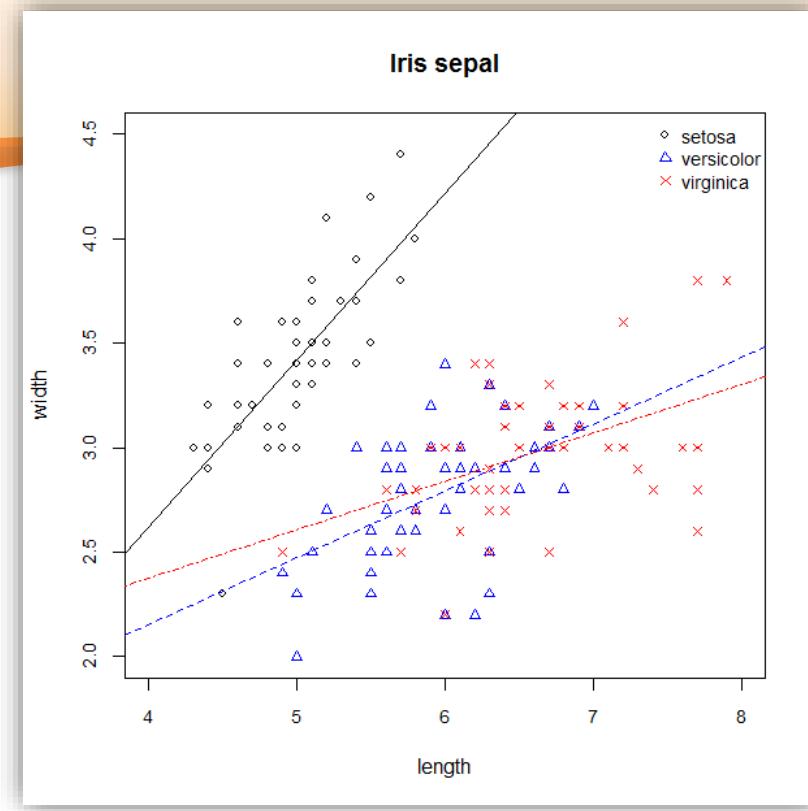
# R graphic functions

- 3 types of R graphic functions:
  - High level: create more or less complete graphs  
Examples: *plot, hist, boxplot, ...*
  - Low level: allow additional information to be added to a graph  
Examples: *points, lines, text, axis, ...*
  - Interactive graphic functions:  
allow to extract information from an existing graph

# Produce Graphic: Steps

## 1. Produce **main graphic** → high-level plotting functions

1. R Reference Card
2. “**R Graph Gallery**” (website)  
→ download code  
→ modify and apply it
3. Google



## 2. **Modify graphic parameters** (line style, colour, font size, ...) 1. Use help!

## 3. **Add additional** lines, points, title, legends, text, ... 1. R Reference Card 2. Google

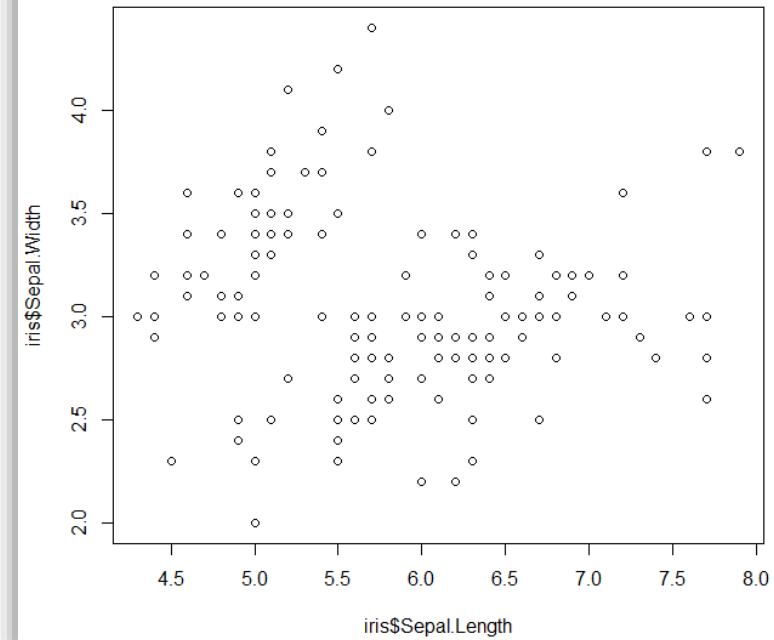
# plot()

- Basic high level plotting function:
- Plot x-coordinates against y-coordinates:

`plot()`

`plot(x, y)`

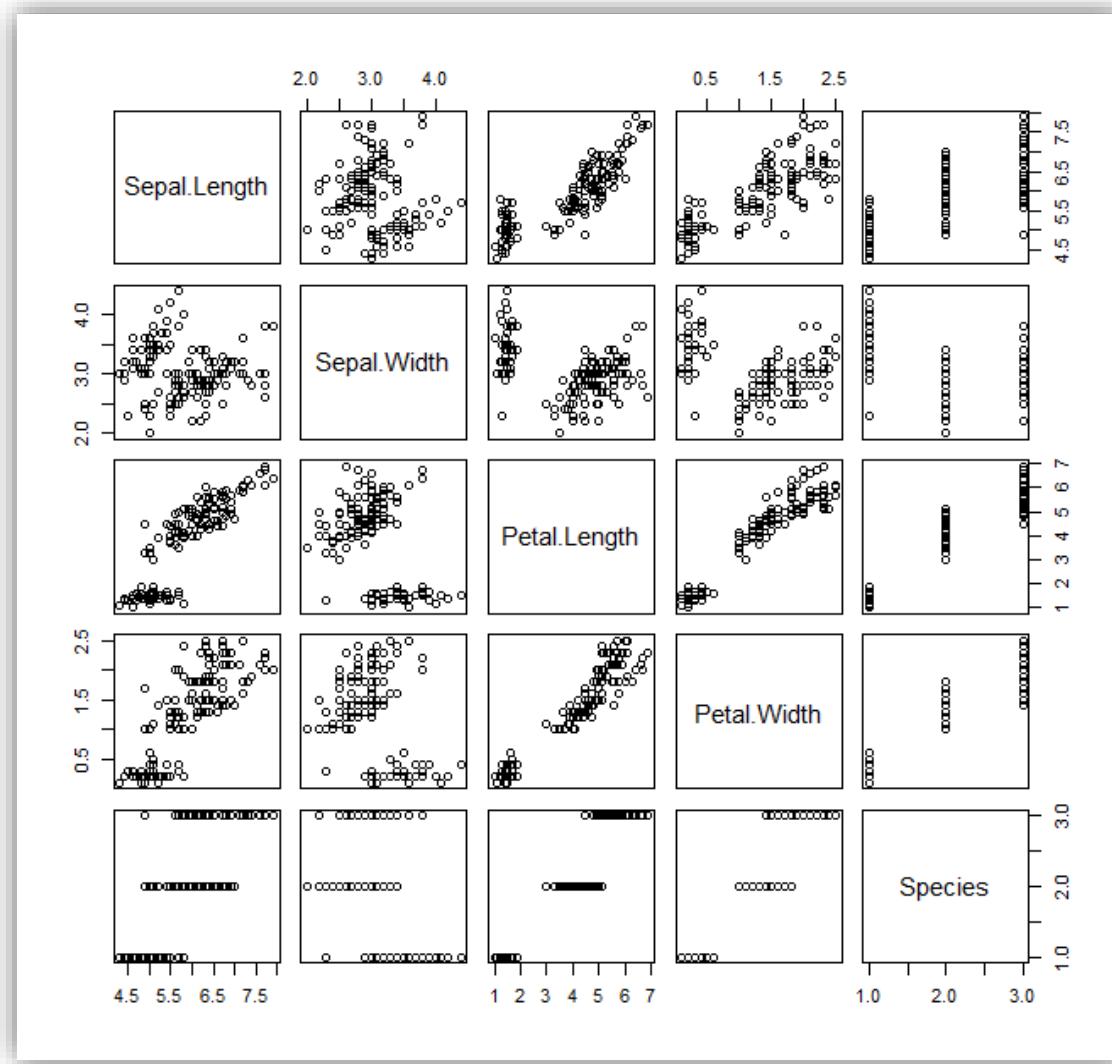
```
plot(iris$Sepal.Length,  
     iris$Sepal.Width)
```



- But, `plot()` is more powerful:
  - What happens if you feed `plot` a whole dataframe?

```
plot(iris)
```

# plot(iris)



# plot() – add low level plotting functions

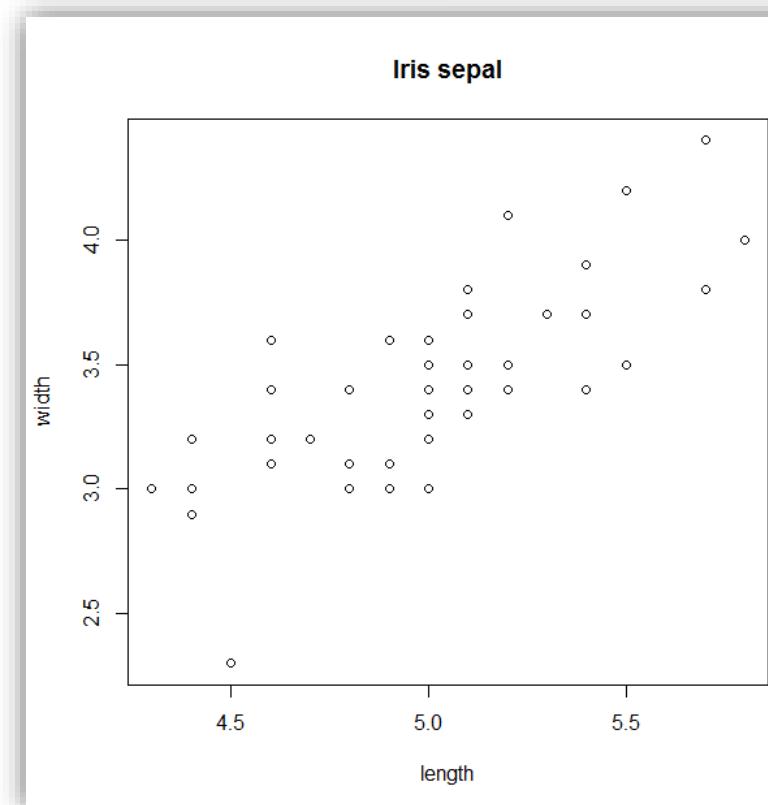
1. Plot sepal length against width of only one species (setosa):

```
irisSe <- iris[iris$Species == "setosa",]  
plot(irisSe$Sepal.Length,irisSe$Sepal.Width)
```

Make graphic nicer with low level plotting functions:

2. Add nicer xy-labels (`xlab`/`ylab`) and title (`main`):

```
plot(irisSe$Sepal.Length,  
     irisSe$Sepal.Width,  
     xlab="length", ylab="width",  
     main="Iris sepal")
```



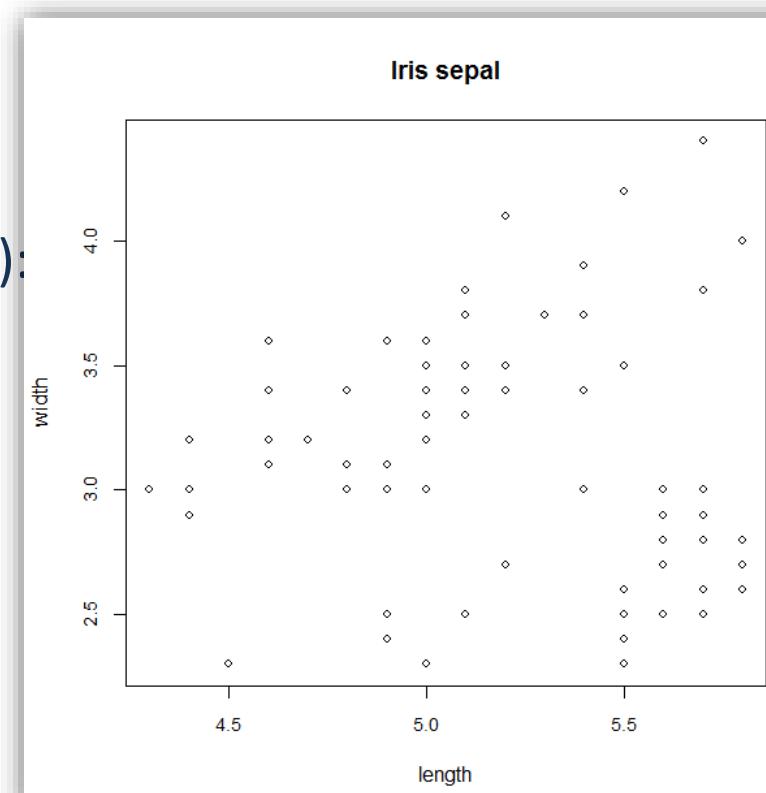
# plot() – add low level plotting functions

3. Reduce size of points (`cex`) and axis (`cex.axis`):

```
plot(iris$Sepal.Length,  
     iris$Sepal.Width,  
     xlab="length", ylab="width",  
     main="Iris sepal",  
     cex=0.8, cex.axis=0.9)
```

4. Add data of other species (`points()`):

```
irisVe <- iris[iris$Species  
              == "versicolor",]  
points(irisVe$Sepal.Length,  
       irisVe$Sepal.Width, cex=0.8)  
  
irisVi <- iris[iris$Species  
              == "virginica",]  
points(irisVi$Sepal.Length,  
       irisVi$Sepal.Width, cex=0.8)
```



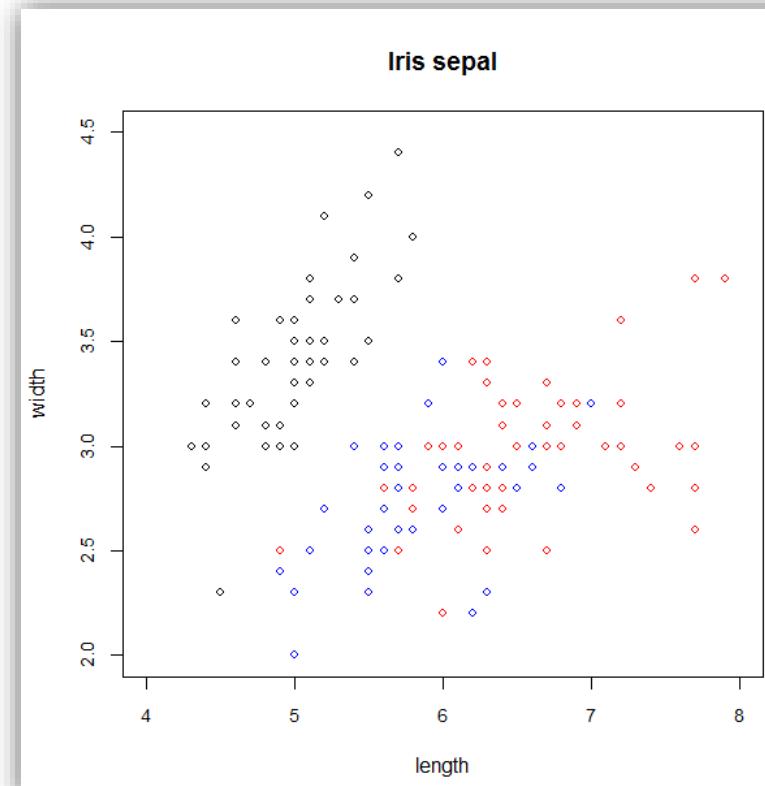
# plot() – add low level plotting functions

5. Adjust axis range (ylim/xlim):

```
plot(irisSe$Sepal.Length,  
     irisSe$Sepal.Width,  
     xlab="length", ylab="width",  
     main="Iris sepal",  
     cex=0.8, cex.axis=0.9,  
     xlim=c(4,8), ylim=c(2,4.5))
```

6. Color points according species (col):

```
points(irisVe$Sepal.Length,  
       irisVe$Sepal.Width, cex=0.8,  
       col="blue")  
  
points(irisVi$Sepal.Length,  
       irisVi$Sepal.Width, cex=0.8,  
       col="red")
```

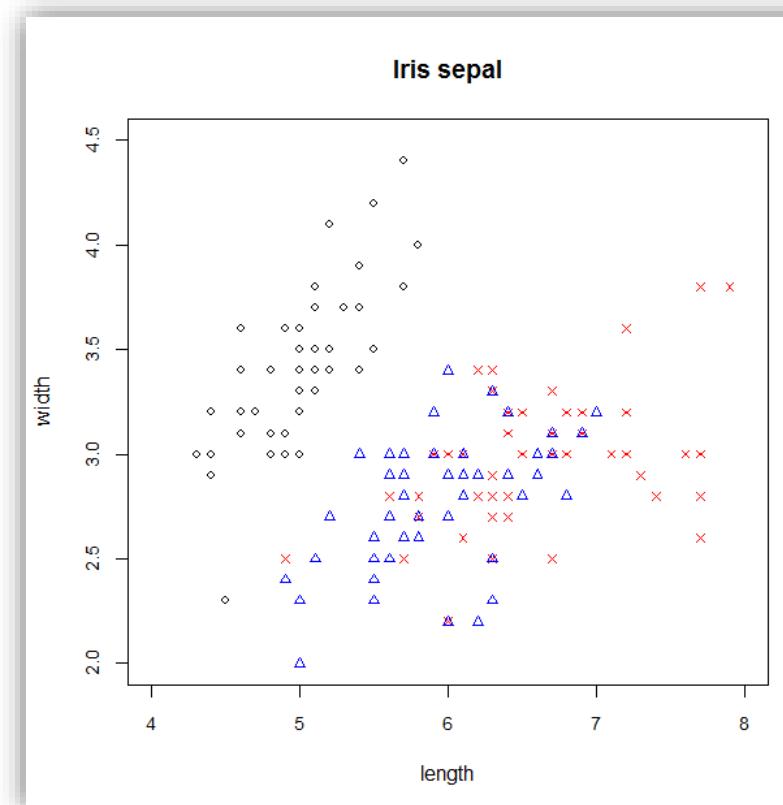


# plot() – add low level plotting functions

7. Adapt symbol of the points according species (pch):

|   |   |   |   |   |   |   |   |   |   |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |
|---|---|---|---|---|---|---|---|---|---|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|
| 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 | 18 | 19 | 20 | 21 | 22 | 23 | 24 | 25 |
| □ | ○ | △ | + | × | ◊ | ▽ | ◻ | * | ◆ | ⊕  | ⊗  | 田  | ▣  | ▣  | ■  | ●  | ▲  | ◆  | ●  | ●  | ○  | □  | ◊  | △  | ▽  |

```
points(irisVe$Sepal.Length,  
       irisVe$Sepal.Width, cex=0.8,  
       col="blue", pch=2)  
  
points(irisVi$Sepal.Length,  
       irisVi$Sepal.Width, cex=0.8,  
       col="red", pch=4)
```

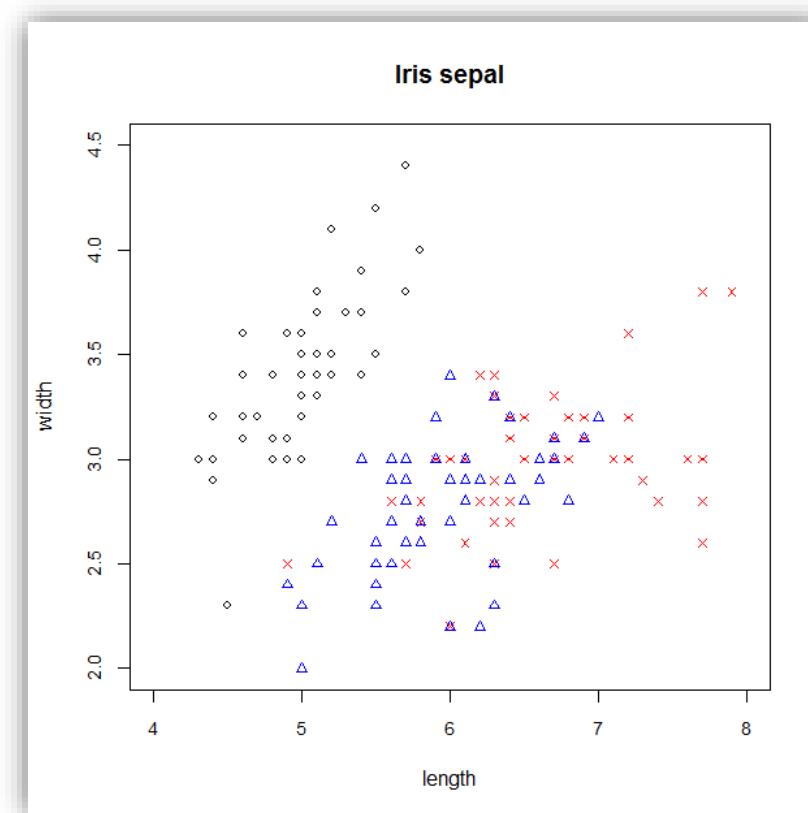


# plot() – add low level plotting functions

## 8. Add legend (`legend()`):



Try to add a legend to the plot, use `?legend` as help

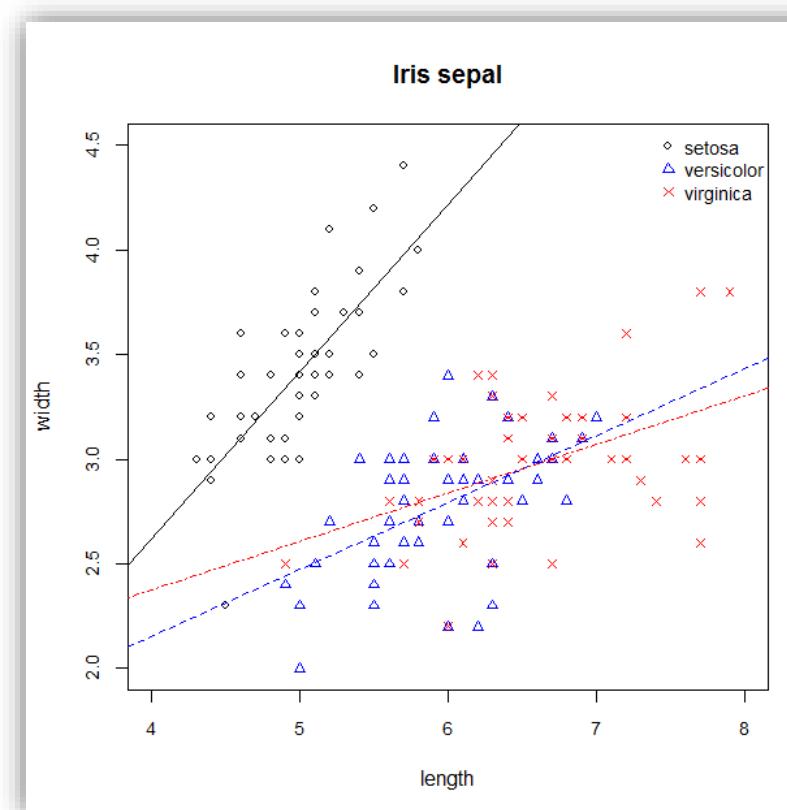


# plot() – add low level plotting functions

## 9. Add regression lines:

- lm(): fit a linear model
- abline(): adds a straight line to the plot
- Adjust line type (lty):  
1:solid, 2:dashed, 3:dotted,  
4:dotdash, 5:longdas, 6:twodash

```
abline(lm(irisSe$Sepal.Width  
~irisSe$Sepal.Length))  
  
abline(lm(irisVe$Sepal.Width  
~irisVe$Sepal.Length),  
col="blue", lty=2)  
  
abline(lm(irisVi$Sepal.Width  
~irisVi$Sepal.Length),  
col="red", lty=4)
```



# Structure of R objects

Side note: Display the structure of an R object      **str(object)**

```
regLine <- lm(irisSe$Sepal.Width~irisSe$Sepal.Length)
```

```
regLine
```

*Call:*

```
lm(formula = irisSe$Sepal.Width ~ irisSe$Sepal.Length)
```

*Coefficients:*

| (Intercept) | irisSe\$Sepal.Length |
|-------------|----------------------|
| -0.5694     | 0.7985               |

```
str(regLine)
```

*List of 12*

```
$ coefficients : Named num [1:2] -0.569 0.799
  ..- attr(*, "names")= chr [1:2] "(Intercept)" "irisSe$Sepal.Length"
$ residuals     : Named num [1:50] -0.00306 -0.34336 0.01635 -0.0038 ...
  ..- attr(*, "names")= chr [1:50] "1" "2" "3" "4" ...
...
Access intercept: regLine$coefficients[1]
```



# Save plot

- Saves a plot in png format (see `?png` for additional options):

```
png("irisPlot.png")  
plot(...)  
points(...)  
...  
dev.off()
```

← opens graphic device  
← closes graphic device

- Saves a plot as pdf (see `?pdf` for additional options):

```
pdf("irisPlot.pdf")  
plot(...)  
points(...)  
...  
dev.off()
```

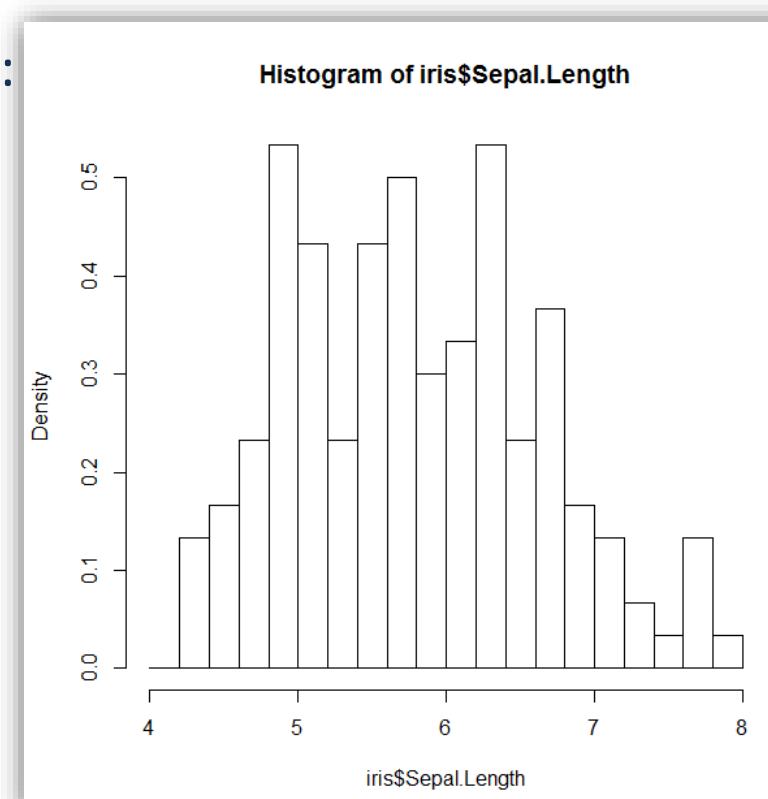


Try to save your plot

# Histogram

- Histogram: `hist()`  
`hist(iris$Sepal.Length)`
- Divide data into more bins by ‘breaks’: e.g.: `breaks=20` or  
`breaks=c(4, 4.5, 5, 5.5, 6)`
- Plotting density instead of frequency: frequency divided by the width of the interval (plot area sums to 1)

```
hist(iris$Sepal.Length,  
     breaks=seq(4,8,by=0.2),  
     freq=FALSE)
```



# Histogram



Plot only data of setosa and change axis labels and title

# Histogram



Overlay data of other species:

- use argument: `add=TRUE`
- Don't forget to color it and add a legend

# Histogram

## 3. Make colors transparent:

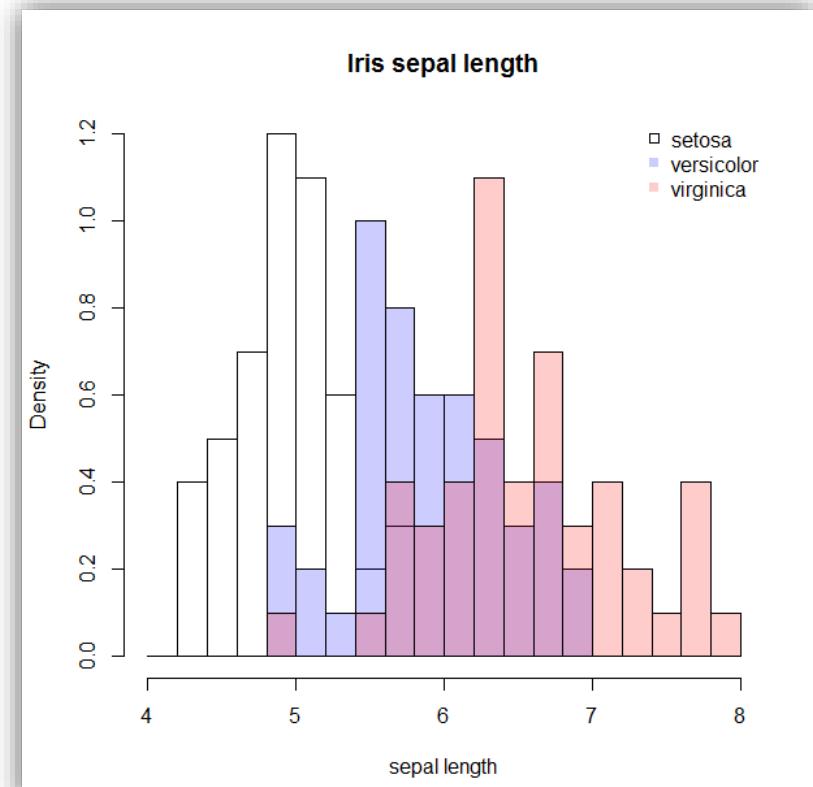
- Use rgb function:  $\text{rgb}(\underbrace{\text{red}, \text{green}, \text{blue}}_{\text{intensity}}, \underbrace{\text{alpha}}_{\text{transparency}})$

```
blueT <- rgb(0,0,1,0.4)
redT <- rgb(1,0,0,0.4)

hist(irisVi$Sepal.Length,
  breaks=seq(4,8,by=0.2),
  freq=FALSE, add=TRUE, col=blueT)

hist(irisVe$Sepal.Length,
  breaks=seq(4,8,by=0.2),
  freq=FALSE, add=TRUE, col=redT)

legend("topright", c("setosa",
  "versicolor", "virginica"),
  col=c("black", blueT, redT),
  pch=c(0,15,15), bty="n")
```

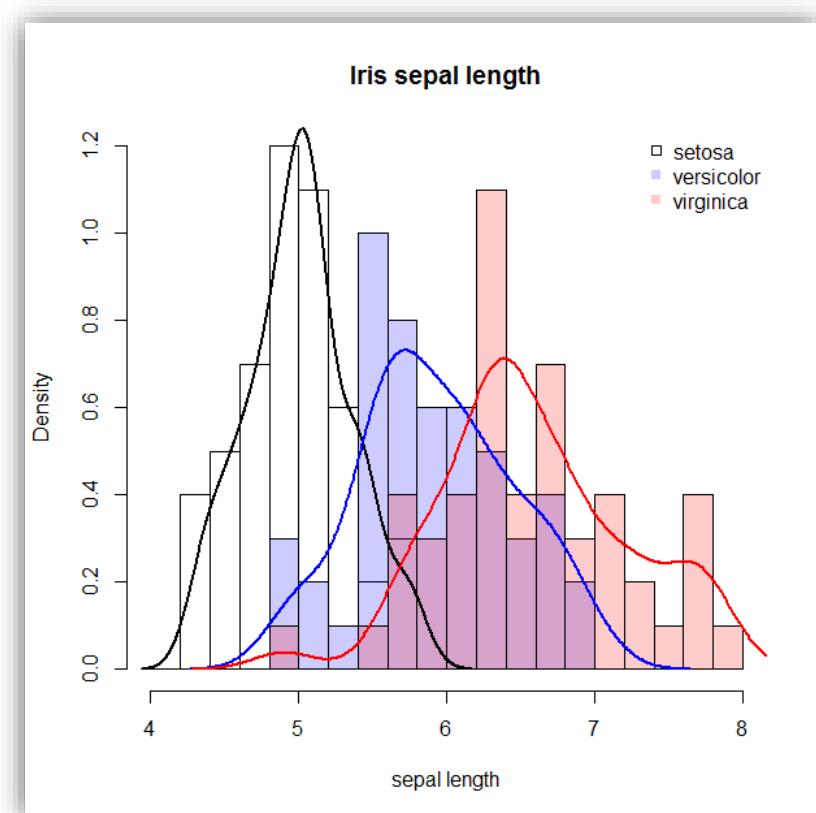


# Histogram

## 4. Add a fitted density curve

- `density()`: computes kernel density estimates
- `lines()`: adds a connected line

```
lines(density(irisSe$Sepal.Length), lwd=2)  
lines(density(irisVe$Sepal.Length), lwd=2, col="blue")  
lines(density(irisVi$Sepal.Length), lwd=2, col="red")
```

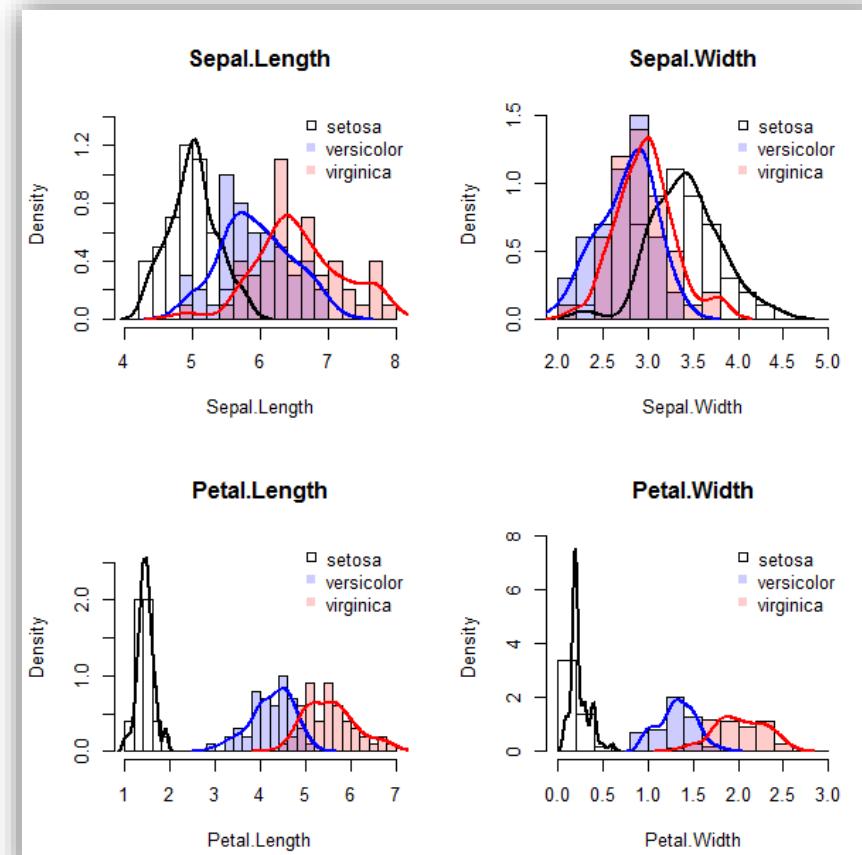


# Multiple plots

5. Put all 4 measurements in one graphic → 4 plots in one graphic
  - Change `mfrow` within graphic parameters (`par`)

```
store old settings  
old.par <- par(mfrow=c(2, 2))  
hist(irisSe$Sepal.Width, ...  
...  
hist(irisSe$Sepal.Length, ...  
...  
hist(irisSe$Petal.Length, ...  
...  
hist(irisSe$Petal.Width, ...  
...  
par(old.par)
```

reset old settings



# Multiple plots

## 5. Put all 4 measurements in one plot → by using **for-loop**

```
old.par <- par(mfrow=c(2, 2))
for(i in 1:(ncol(irisSe)-1)){
  colName <- colnames(iris)[i]
  xmin <- trunc(min(iris[,i]))
  xmax <- trunc(max(iris[,i]))+1
  ymax <- max(density(irisSe[,i])$y, density(irisVe[,i])$y,
               density(irisVi[,i])$y)+0.2
  hist(irisSe[,i],breaks=seq(xmin,xmax,by=0.2),ylim=c(0,ymax),
        freq=FALSE,main=colName,xlab=colName)
  hist(irisVe[,i],breaks=seq(xmin,xmax,by=0.2),freq=FALSE,add=TRUE,
        col=blueT)
  hist(irisVi[,i],breaks=seq(xmin,xmax,by=0.2),freq=FALSE,add=TRUE,
        col=redT)
  legend("topright",c("setosa","versicolor","virginica"),
         col=c("black",blueT,redT),pch=c(0,15,15),bty="n")
  lines(density(irisSe[,i]),lwd=2)
  lines(density(irisVe[,i]),lwd=2,col="blue")
  lines(density(irisVi[,i]),lwd=2,col="red")
}
par(old.par)
```

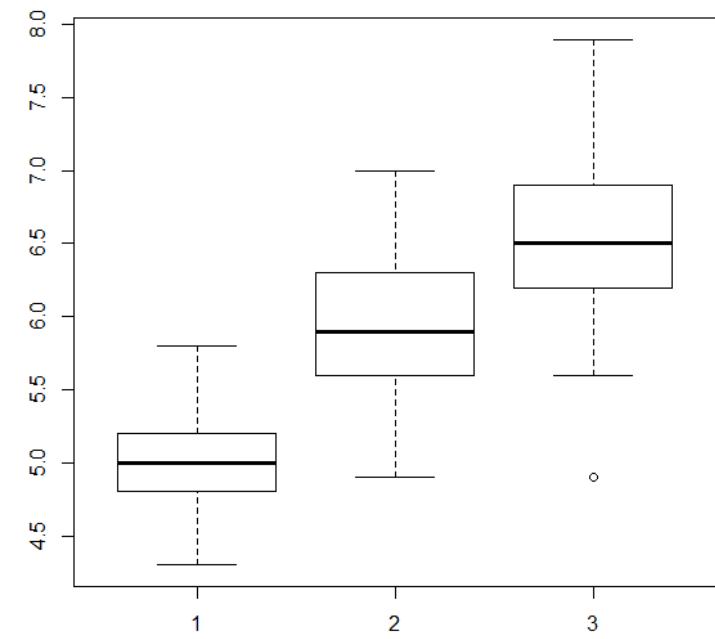
# boxplot

- Boxplots: `boxplot()`

```
boxplot(iris$Sepal.Length)
```

- More than one boxplot in the graphic:

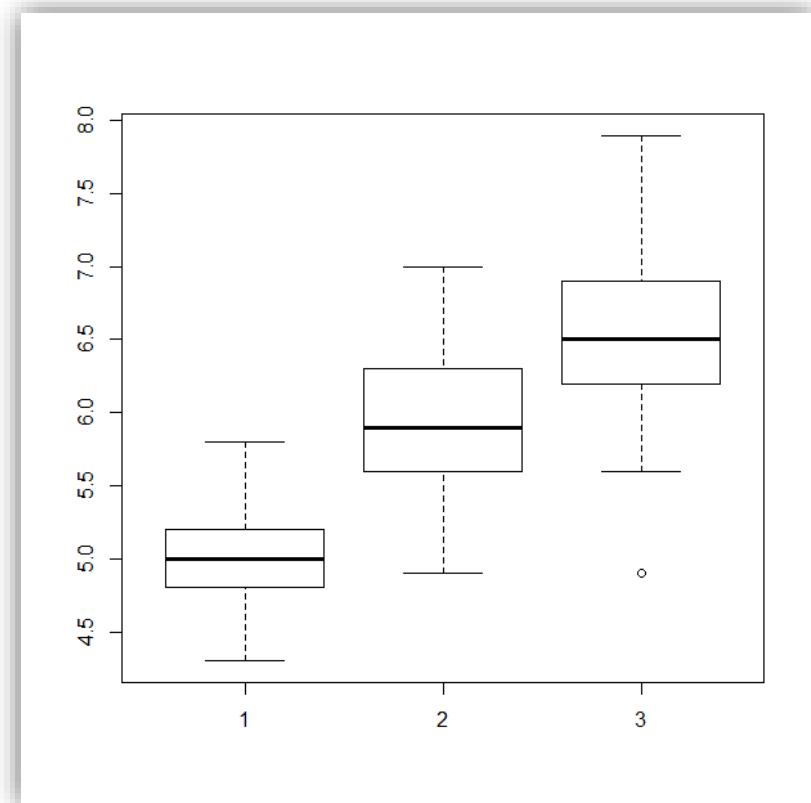
```
boxplot(irisSe$Sepal.Length,  
irisVe$Sepal.Length,  
irisVi$Sepal.Length)
```



# boxplot



Change axis labels, title and color boxplots  
(use `?boxplot` if necessary)



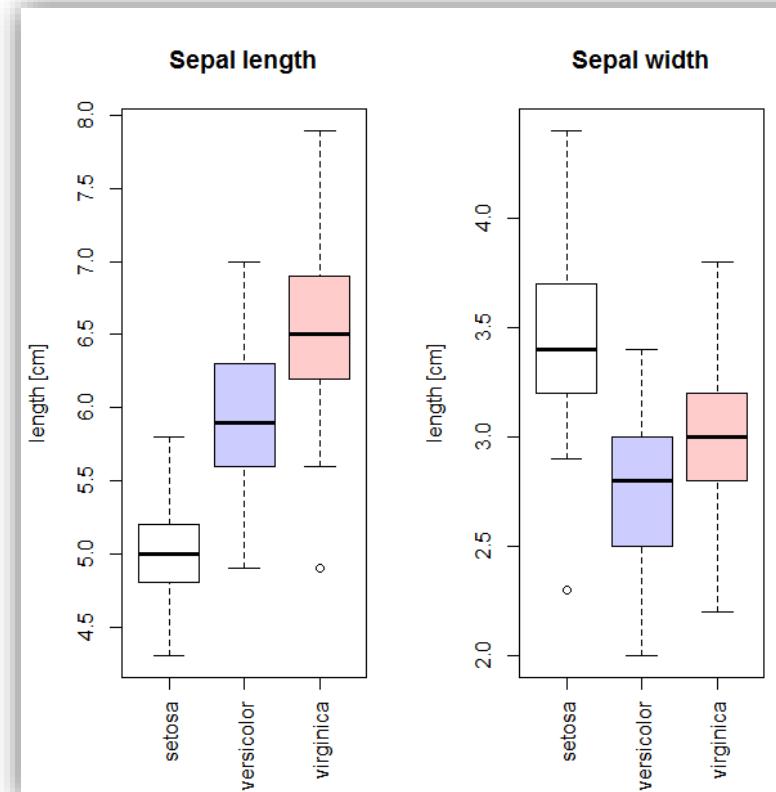
# boxplot

 Try to put two boxplot graphics (sepal length/width) into one graphic

# boxplot

- Write axis labels always vertical: `las=3` (see `?par` for details)

```
old.par <- par(mfrow=c(1,2))
boxplot(irisSe$Sepal.Length,
        irisVe$Sepal.Length,
        irisVi$Sepal.Length,
        ylab="length [cm]",
        names=c("setosa","versicolor",
        "virginica"),main="Sepal length",
        col=c("white",blueT,redT),las=3)
boxplot(irisSe$Sepal.Width,
        irisVe$Sepal.Width,
        irisVi$Sepal.Width,
        ylab="length [cm]",
        names=c("setosa","versicolor",
        "virginica"),main="Sepal width",
        col=c("white",blueT,redT),las=3)
par(old.par)
```



# barplot

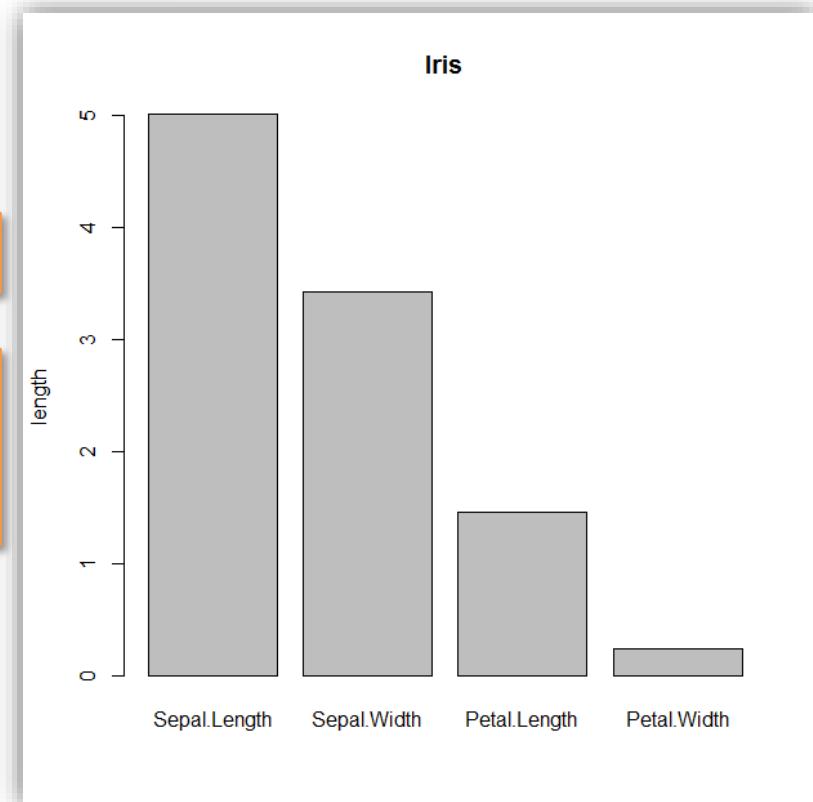
- Barplots: `barplot()`

```
barplot(c(mean(irisSe$Sepal.Length),  
         mean(irisSe$Sepal.Width)))
```

- Use `sapply()` to estimate mean for each columns:

```
a <- sapply(irisSe[,1:4],mean)
```

```
barplot(a,ylab="length",  
        names=colnames(irisSe[,1:4]),  
        main="Iris setosa")
```



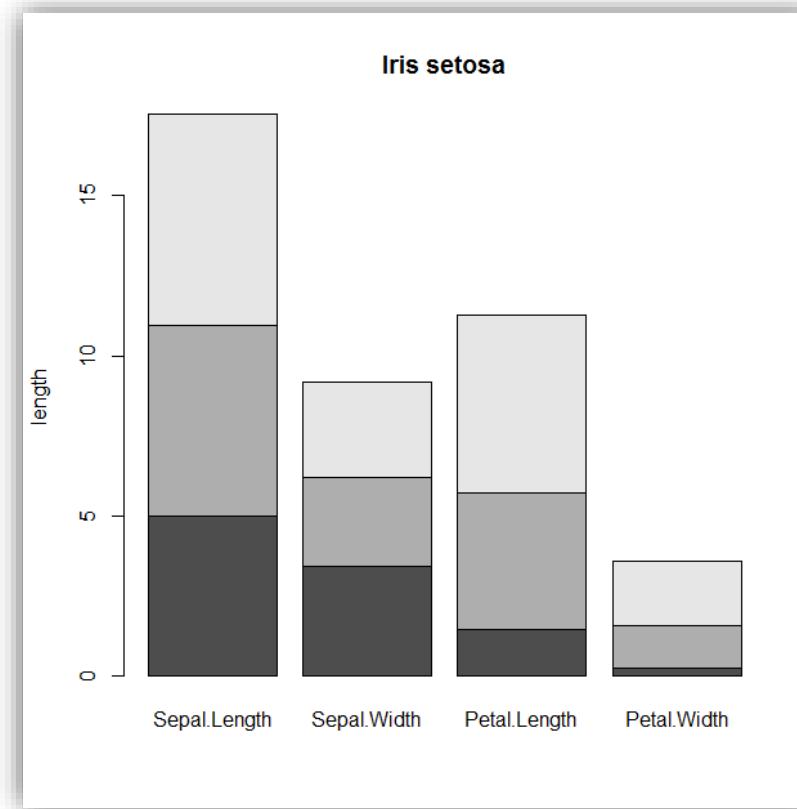
# barplot

- Add data of other species:
    - Feed matrix instead vector: columns: mean measurements  
rows: species

```
means <- matrix(  
  c(sapply(irisSe[,1:4],mean),  
    sapply(irisVe[,1:4],mean),  
    sapply(irisVi[,1:4],mean)),  
  ncol=4, byrow=TRUE)
```

```
[,1] [,2] [,3] [,4]
[1,] 5.006 3.428 1.462 0.246
[2,] 5.936 2.770 4.260 1.326
[3,] 6.588 2.974 5.552 2.026
```

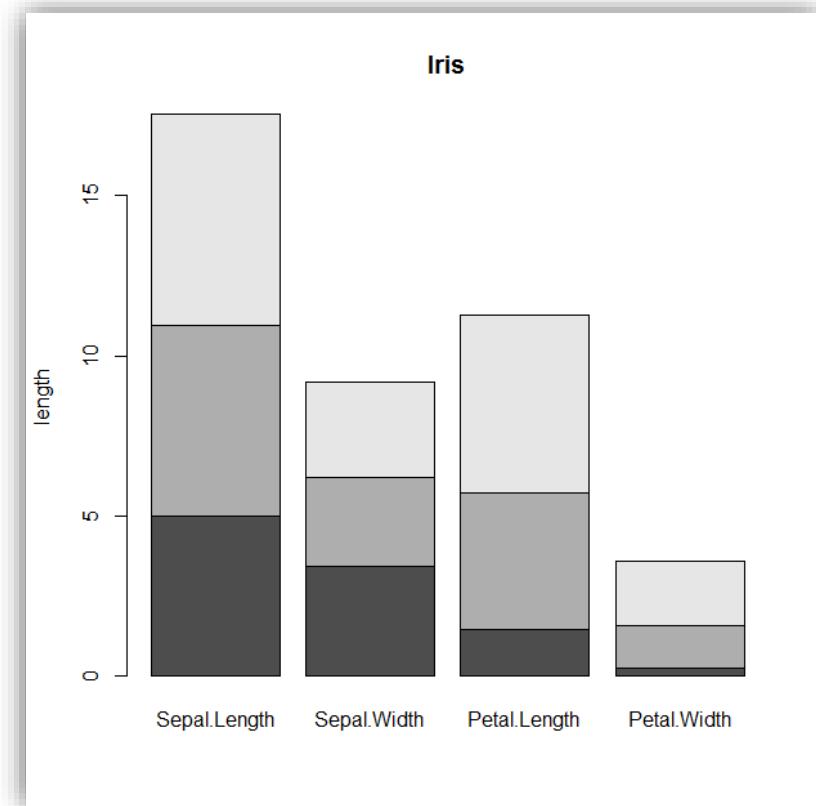
```
barplot(means, ylab="length",
        names=colnames(irisSe[,1:4]),
        main="Iris setosa")
```



# barplot



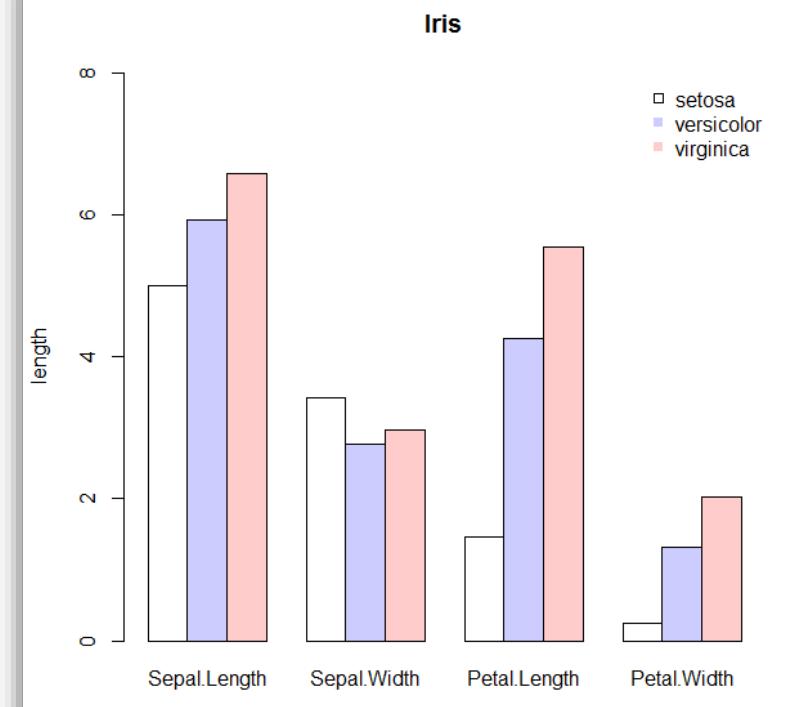
Use `beside=TRUE` to plot bars beside each other, color bars according the color scheme and add legend



# barplot

- Add standard deviations to the bars:
  1. Create matrix with sd:

```
stDev <- matrix(  
  c(sapply(irisSe[,1:4],sd),  
    sapply(irisVe[,1:4],sd),  
    sapply(irisVi[,1:4],sd)),  
  ncol=4, byrow=TRUE)
```



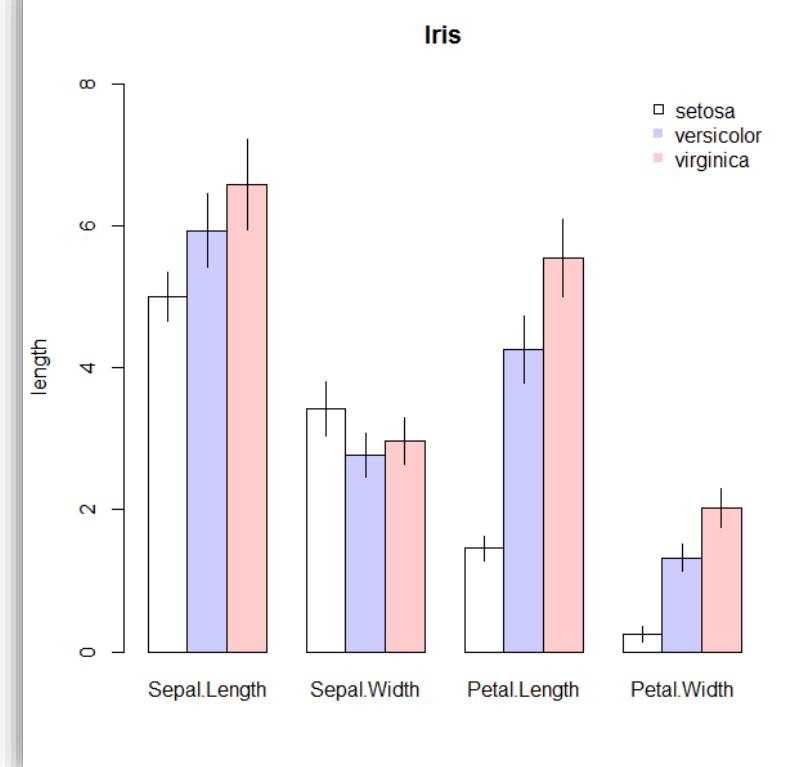
# barplot

## 2. Save “bar positions” in a variable

```
bplot <- barplot(means,  
  ylab="length", names=colnames(  
  irisSe[,1:4]),  
  main="Iris", beside=TRUE,  
  col=c("white",blueT,redT),  
  ylim=c(0,8))
```

## 3. Add the sd bars: segments()

```
segments(bplot,means-stDevs,  
         bplot,means+stDevs)
```



# heatmap

- heatmaps:
  - `image()`
  - `levelplot()` → creates automatically a legend
- We need a matrix data set: relative sepal length across each sample:

```
matrixSe <- matrix(NA,nrow(irisSe),nrow(irisSe))
for(i in 1:nrow(matrixSe)) {
  matrixSe[i,] <- irisSe$Sepal.Length[i]/irisSe$Sepal.Length
}
```

# heatmap



- Try to create an `image()` and a `levelplot()` (use help if necessary)
  - don't forget to add a nice title and axis labels
  - use `colRmp <- colorRampPalette(c("color1", "color2", ...))` (`nbBreaks`) to create your own color ramp

# Acknowledgment

- **Tutorial with solutions:**  
[https://www.dropbox.com/s/kath5t9q1p8cyj1/URPP\\_Tutorial\\_RBasics\\_1\\_v2\\_solutions.pdf](https://www.dropbox.com/s/kath5t9q1p8cyj1/URPP_Tutorial_RBasics_1_v2_solutions.pdf)
- Sources:
  - <http://nicercode.github.io/intro/>
  - <http://www.cyclismo.org/tutorial/R/>
  - <http://manuals.bioinformatics.ucr.edu/home/programming-in-r#TOC-Control-Structures>
  - <http://streaming.stat.iastate.edu/workshops/r-intro/lectures/5-Rprogramming.pdf>
- R reference cards:
  - <http://cran.r-project.org/doc/contrib/Short-refcard.pdf>
  - <http://cran.r-project.org/doc/contrib/Baggott-refcard-v2.pdf>
- Graphic tutorials:
  - <http://ww2.coastal.edu/kingw/statistics/R-tutorials/igraphically.html>
  - <http://www.statmethods.net/graphs/creating.html>
  - [http://bio.fsu.edu/miller/docs/Tutorials/Tutorial3\\_Graphics.pdf](http://bio.fsu.edu/miller/docs/Tutorials/Tutorial3_Graphics.pdf)
  - <http://www.harding.edu/fmccown/r/>
  - [http://manuals.bioinformatics.ucr.edu/home/R\\_BioCondManual#TOC-Graphical-Procedures](http://manuals.bioinformatics.ucr.edu/home/R_BioCondManual#TOC-Graphical-Procedures)
- R graph gallery:
  - <http://rgraphgallery.blogspot.ch/>
  - <http://www.sr.bham.ac.uk/~ajrs/R/r-gallery.html>