



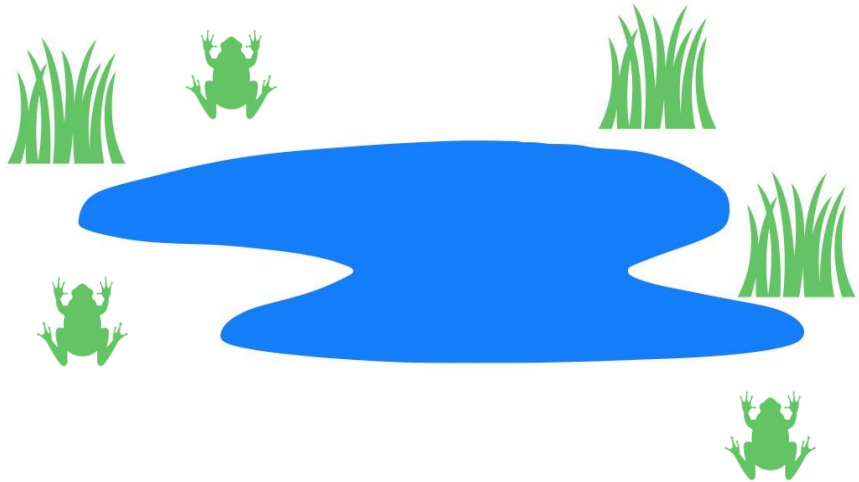
eDNA and qPCR survey data

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eDNA and qPCR survey data



Terminology:

eDNA: traces of DNA left by organisms in environmental samples such as water and soil

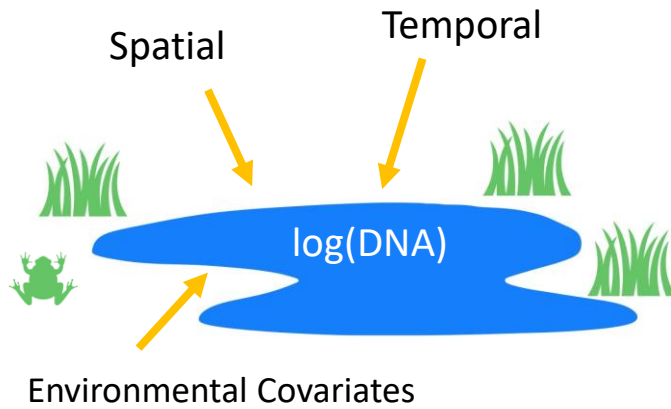
qPCR: Quantitative PCR is a technique used to amplify and quantify a target DNA molecule

eDNA and qPCR survey data

qPCR survey pipeline

1. DNA availability:

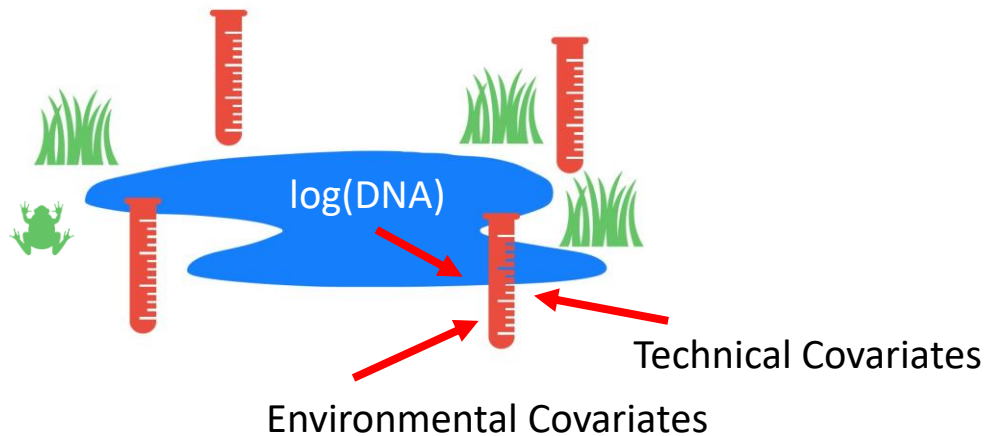
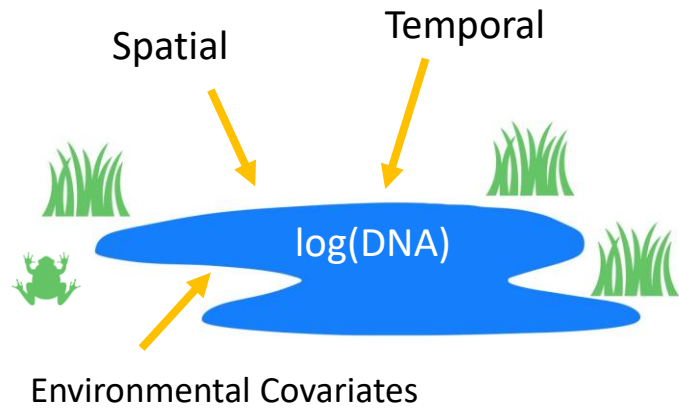
Across sites $i = 1, \dots, S$ and time-points $t = 1, \dots, T$.



eDNA and qPCR survey data

qPCR survey pipeline

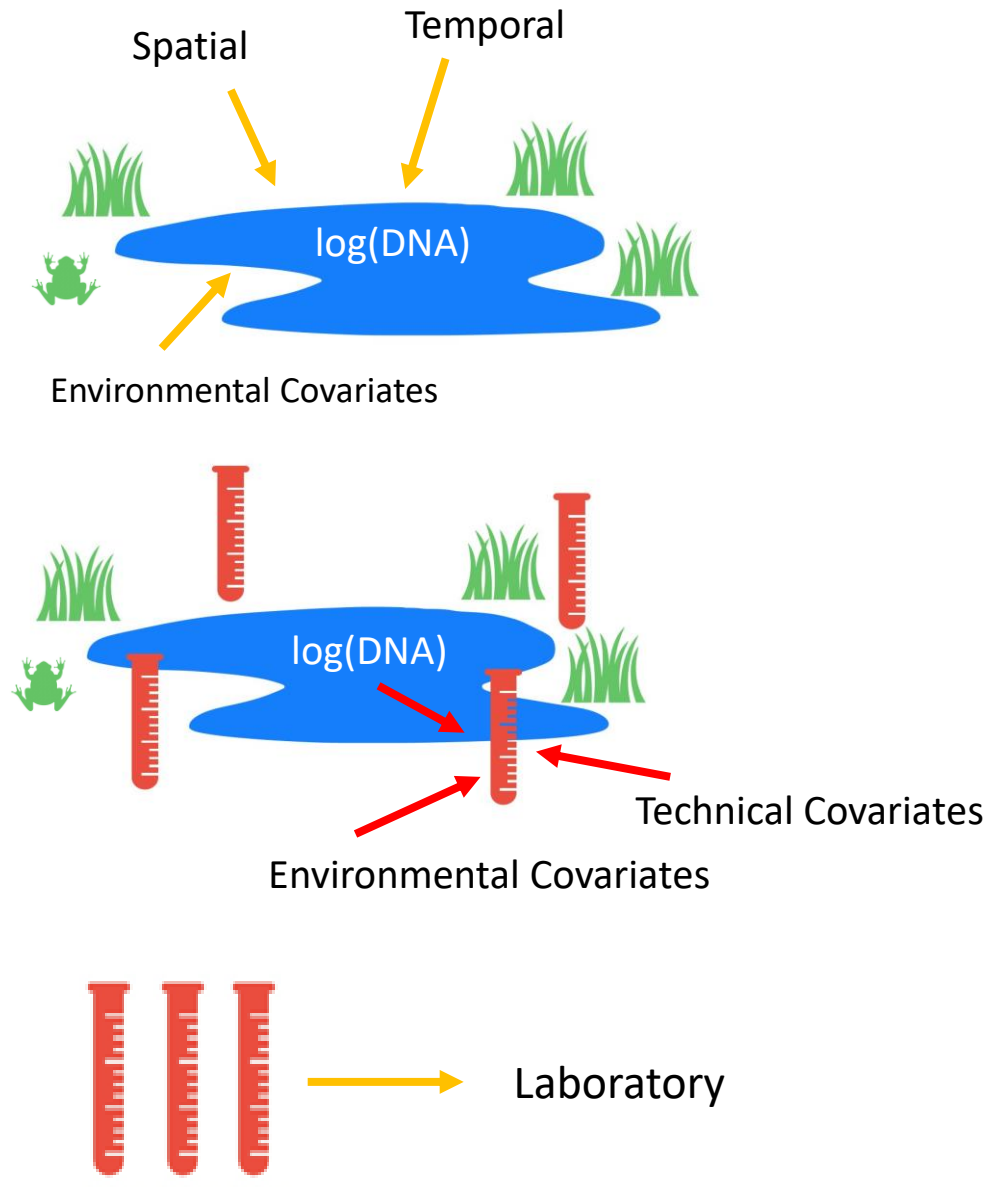
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Across sites $i = 1, \dots, S$ and time-points $t = 1, \dots, T$.
2. DNA collection:
In samples $m = 1, \dots, M$.



eDNA and qPCR survey data

qPCR survey pipeline

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3. DNA analysis:
Model qPCR output on DNA concentration in the sample



eDNA and qPCR survey data

qPCR survey pipeline

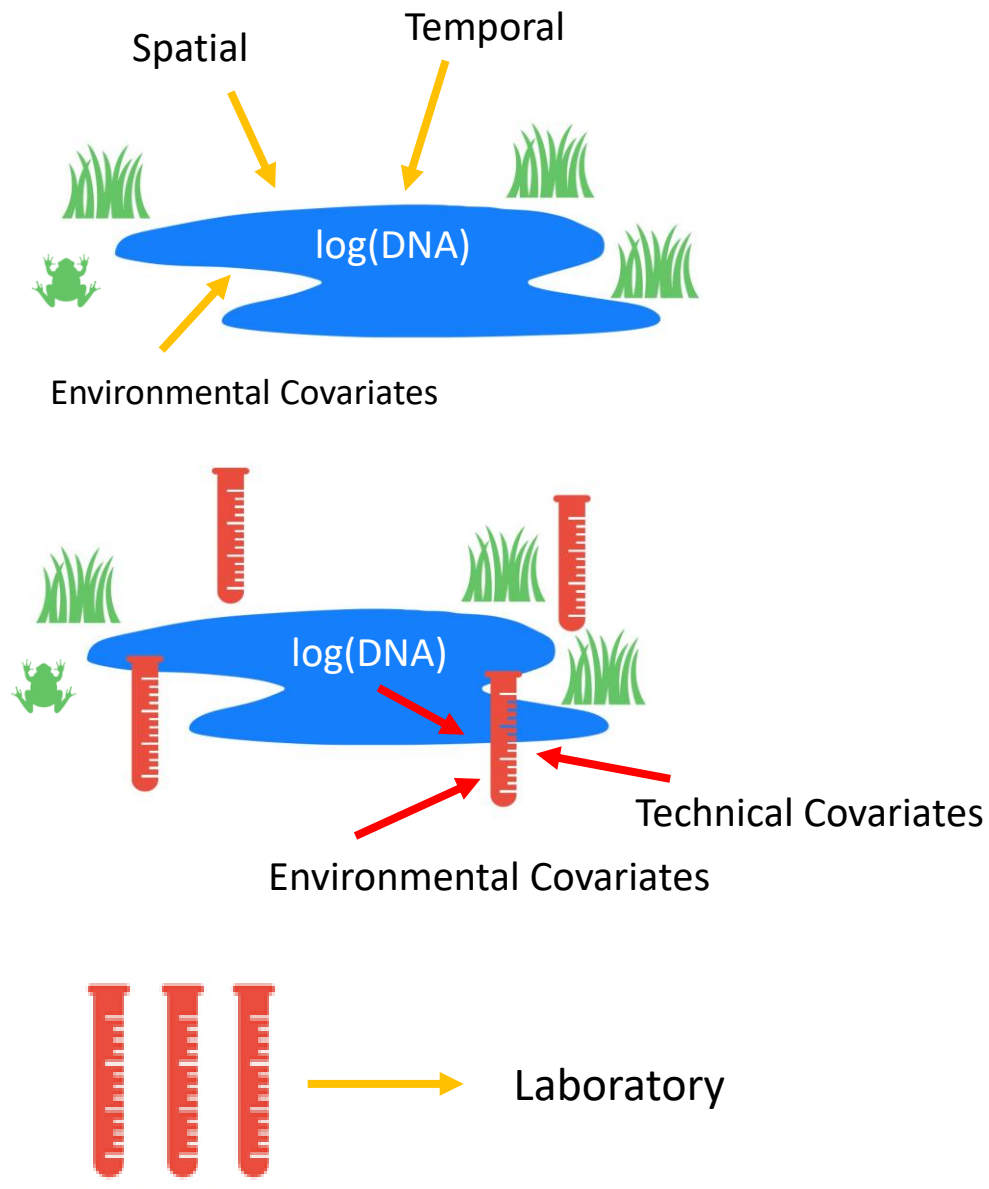
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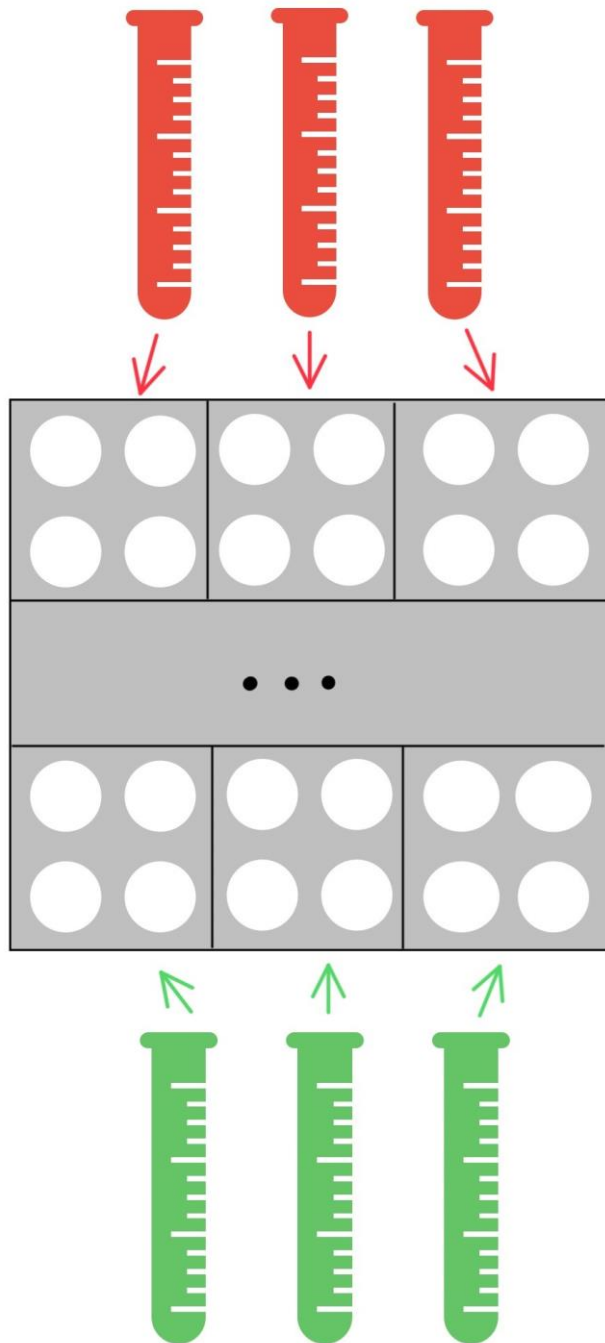
Previous work:

- [Espe et al. 2022] – artemis package
- [Shelton et al. 2022] – spatial model for Pacific Hake

Our focus:

- Modelling contamination and inhibition in the lab
- qPCR output heteroscedasticity





Collected samples
Unknown Concentrations

Plate p

K replicates
per sample

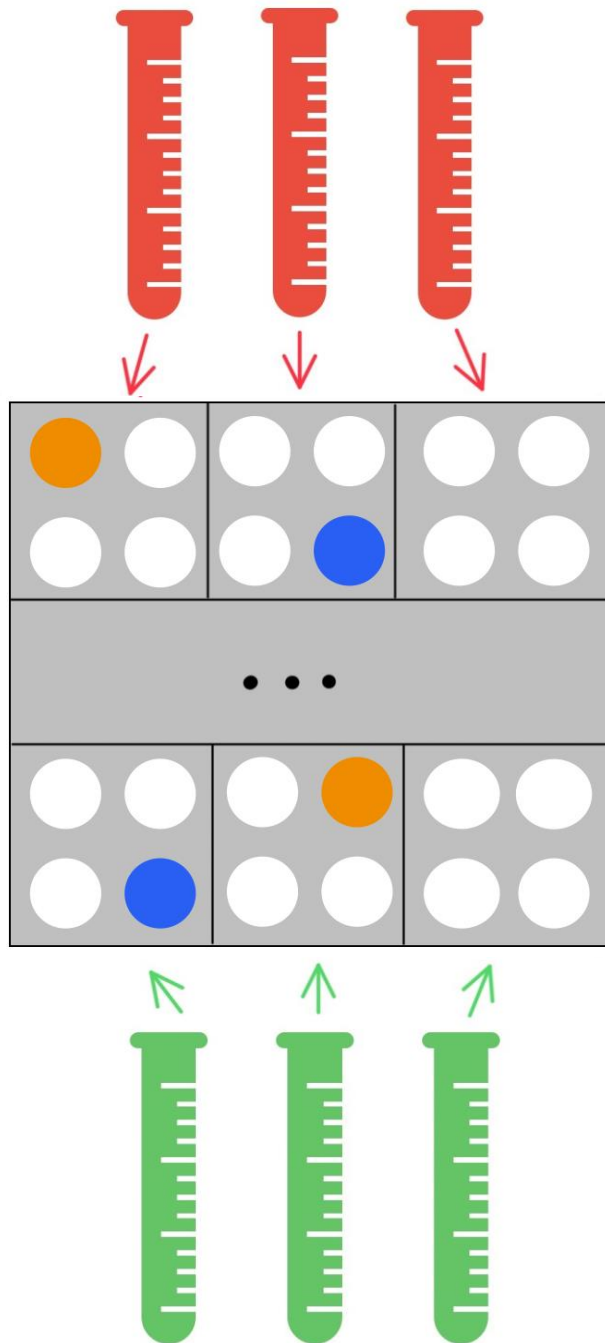
K^* replicates
per standard

Standards
Known Concentrations

DNA analysis

Each plate $p = 1, \dots, P$ contains:

- Samples m each divided into K replicates
- **Standards** (solutions of known DNA concentration) divided into K^* replicates



M samples
Unknown Concentrations

Plate p

● Contaminated
● Inhibited

Standards
Known Concentrations

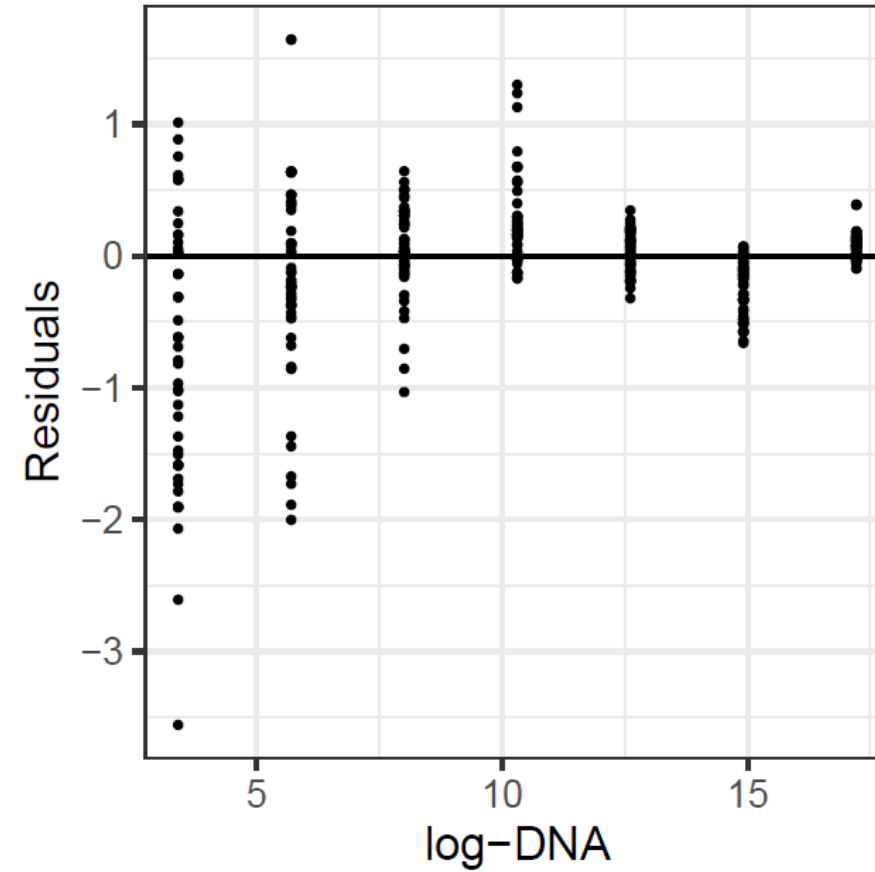
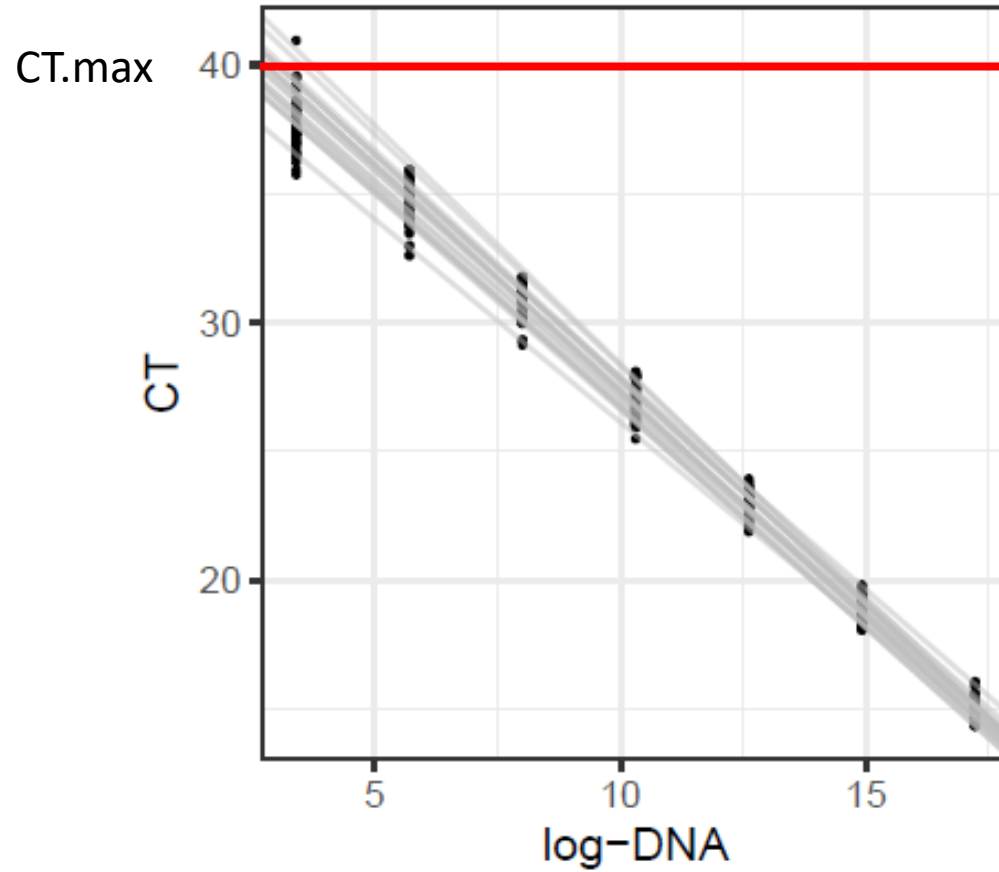
DNA analysis

Contamination: presence of additional DNA in wells from lab through contaminated equipment or clothing

Inhibition: factors (substances or chemicals) interfering with the PCR process delaying DNA amplification

CT value – cycles to threshold, the (fractional) cycle number at which DNA is detected

DNA analysis

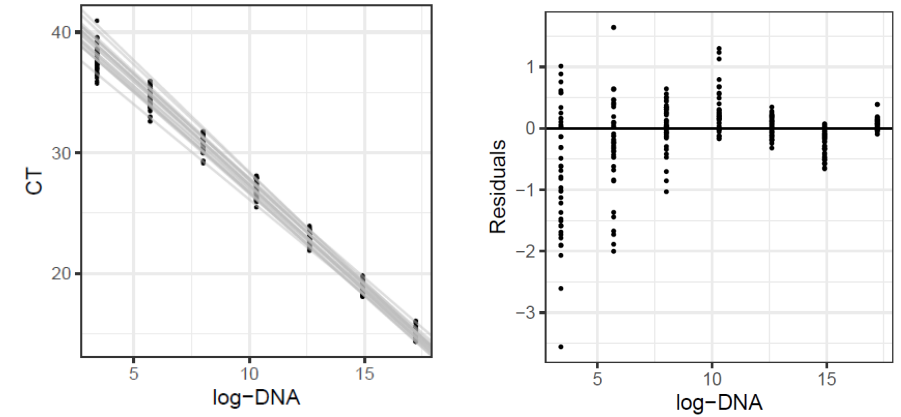


w_{imt} = DNA concentration
 where i is the site, t is the time point,
 m is the sample, and k is the replicate.

CT regression line $\mu_{imtk} = \alpha_p^1 + \alpha_p^2 \log(w_{imt}),$
CT heteroscedasticity $\sigma_y^2(w_{imt}) = \exp(a_1 + a_2 \log(w_{imt})),$

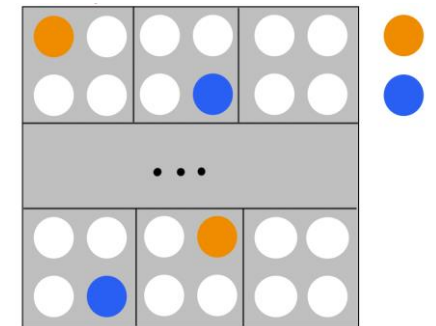
$$\alpha_p^1 \sim N(\alpha_0^1, \sigma_\alpha^2)$$

$$\alpha_p^2 \sim N(\alpha_0^2, \sigma_\alpha^2)$$



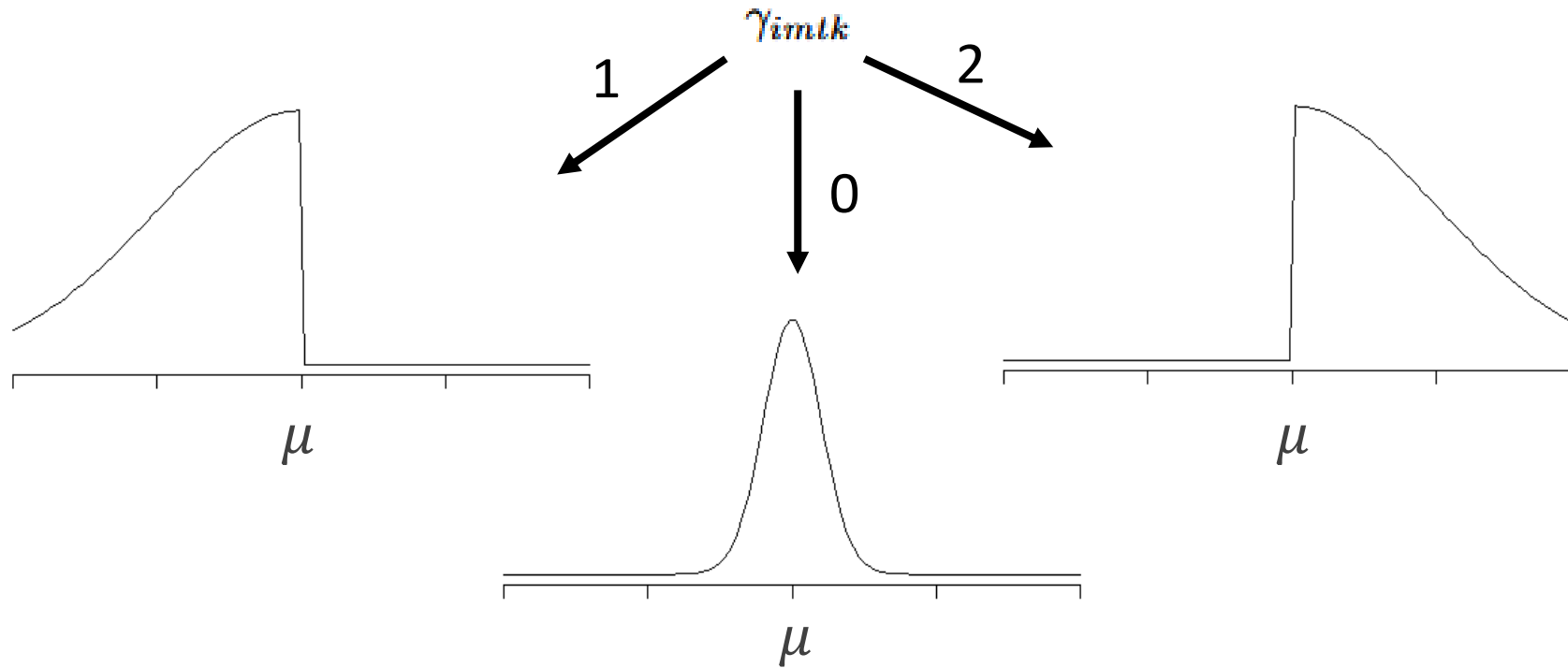
Status of replicate $\gamma_{imtk} = \begin{cases} 1 & \text{contaminated with probability } p_c, \\ 2 & \text{inhibited with probability } p_h, \\ 0 & \text{neither with probability } 1 - p_c - p_h, \end{cases}$

$$(p_c, 1 - p_c - p_h, p_h) \sim \text{Dirichlet}(\pi_1, \pi_2, \pi_3)$$



Uncensored CT value

$$\tilde{C}_{imtk} \sim \begin{cases} N(\mu_{imtk}, \sigma_y^2(w_{imtk})) & \text{if } \gamma_{imtk} = 0, \\ TN_{0, \mu_{imtk}}(\mu_{imtk}, \sigma_c^2) & \text{if } \gamma_{imtk} = 1, \\ TN_{\mu_{imtk}, \infty}(\mu_{imtk}, \sigma_c^2) & \text{if } \gamma_{imtk} = 2, \end{cases}$$



Censored CT value

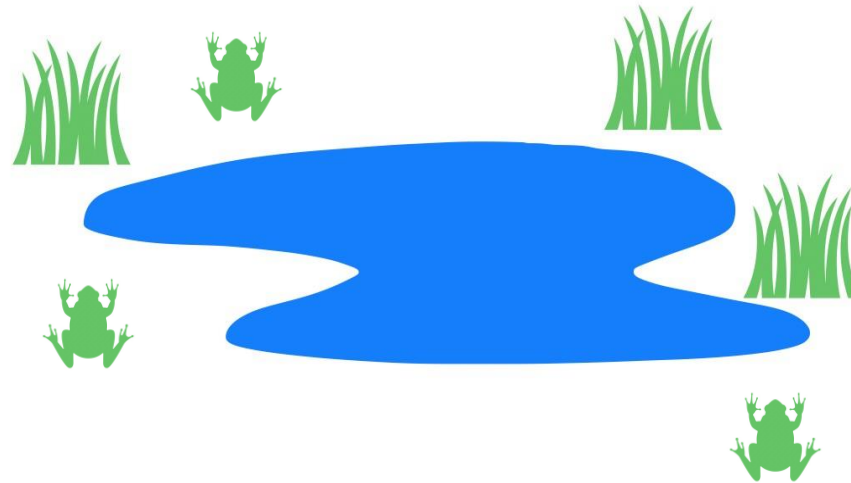
$$C_{imtk} = \begin{cases} \tilde{C}_{imtk} & \text{if } \tilde{C}_{imtk} < \text{CT.max}, \\ \text{NA} & \text{otherwise,} \end{cases}$$

Final Comments

Prior sensitivity analysis on $\alpha_p^1, \alpha_p^2, a_1, a_2$ show these are robust to prior specification.

Ignoring contamination, inhibition, and heteroscedasticity leads to increased bias and uncertainty.

Contamination/inhibition can occur at lab or DNA collection stages. This approach does not account for the collection stage.



References

Espe, M.B., Johnston, M., Blankenship, S.M., Dean, C.A., Bowen, M.D., Schultz, A., Schumer, G.: The artemis package for environmental DNA analysis in R. *Environmental DNA* 4(3), 523–532 (2022) <https://doi.org/10.1002/edn3.277>

Shelton, A.O., Ramon-Laca, A., Wells, A., Clemons, J., Chu, D., Feist, B.E., Kelly, R.P., Parker-Stetter, S.L., Thomas, R., Nichols, K.M., Park, L.: Environmental DNA provides quantitative estimates of pacific hake abundance and distribution in the open ocean. *Proceedings of the Royal Society B* 289(1971), 20212613 (2022) <https://doi.org/10.1098/rspb.2021.2613>

Matz, M.V., Wright, R.M., Scott, J.G.: No control genes required: Bayesian analysis of qRT-PCR data. *PLOS ONE* 8(8), 71448 (2013) <https://doi.org/10.1371/journal.pone.0071448>