CODE

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# Part I data exploration
# read data
library(xlsx)
mortality <- read.xlsx("mortality.xls", sheetIndex = 1)
mortality <- mortality[, -8]
plot(mortality)
par(mfrow = c(3, 3))
sapply(colnames(mortality), function(x) hist(mortality[[x]], main = x))
# summary statistics
apply(mortality, 2, summary)
# single regression
single regression <- lapply(colnames(mortality)[-7], function(x) {
fit <- lm(as.formula(sprintf("MORTALITY ~ %s", x)), data = mortality std)
summary(fit)
})
# compare initial, log, square, square root
par(mfrow = c(3, 4))
sapply(colnames(mortality)[1:6], function(x) {
hist(mortality[[x]], main = sprintf("Hist of %s", x), xlab = x)
hist(log(mortality[[x]]), main = sprintf("Hist of log(%s)", x), xlab = sprintf("%s", x))
hist(\mathbf{sqrt}(\mathbf{mortality}[[x]]), \mathbf{main} = \mathbf{sprintf}(\mathbf{''Hist} \text{ of } \mathbf{sqrt}(\%s)\mathbf{''}, x), xlab = \mathbf{sprintf}(\mathbf{''\%s''}, x))
hist(mortality[[x]]^2, main = sprintf("Hist of square(%s)", x), xlab = sprintf("%s", x))
# do the transformation based on our discussion, log for NOX and SO2, sqrt for NONWHITE
mortality new <- mortality
mortality new$NONWHITE <- sqrt(mortality new$NONWHITE)
mortality new$NOX <- log(mortality new$NOX)
mortality new$SO2 <- log(mortality new$SO2)
# standardize
mortality std <- as.data.frame(scale(mortality new))
# see the correlation
plot(mortality std)
cor(mortality std)
library(corrplot)
par(mfrow=c(1,1), mar=c(1,1,1,1))
corrplot(cor(mortality std), method = "circle", pch = 1, tl.cex=0.75)
corrplot(cor(mortality std), method = "number", pch = 1, tl.cex=0.75)
# vif
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vif regression <- sapply(colnames(mortality std)[-7], function(x) {
fit \leftarrow lm(as.formula(sprintf("%s \sim . - MORTALITY", x)), data = mortality std)
return(1/(1 - summary(fit)\$r.squared))
})
# Additional: regress y only on pollution
lm fit0 \leftarrow lm(MORTALITY \sim NOX + SO2, data = mortality std)
summary(lm fit0)
# not significant
#-----
# Part II stepwise regression
# first order initial
lm fit1 <- lm(MORTALITY \sim 0 + ., data = mortality std)
summary(lm fit1)
par(mfrow = c(2, 2))
plot(lm fit1, which = 1:4)
# first order stepwise
library(MASS)
lm fit2 \le stepAIC(lm(MORTALITY \sim 1, data = mortality std), scope = list(upper = lm fit1),
          direction = "both", k = 2)
lm fit2 \le lm(formula = MORTALITY \sim 0 + NONWHITE + EDUC + SO2 + PRECIP, data =
mortality std)
plot(lm fit2, which = 1:4)
# we can see outliers, but to compare this model to subsequent methods easily, we do not
# drop them, also because this is not a severe proble
# second order initial
lm fit3 \leftarrow stepAIC(lm(MORTALITY \sim 1, data = mortality std),
          scope = list(upper = lm(MORTALITY \sim 0 + . ^ 2, data = mortality_std)),
          direction = "both", k = 2)
lm fit3 <- lm(formula = MORTALITY ~ 0 + NONWHITE + EDUC + SO2 + PRECIP +
NONWHITE:SO2 +
        EDUC:SO2 + SO2:PRECIP, data = mortality std)
plot(lm fit3, which = 1:4)
# cross validation statistics
lm fit2 cv \leftarrow 1 / 60 * sum((residuals(lm fit2) / (1 - influence(lm fit2)$hat))^2)
lm fit3 cv < 1 / 60 * sum((residuals(lm fit3) / (1 - influence(lm fit3) $\)hat))^2)
# cross validation 0.384, 0.394, so select linear model
#-----
# Part III PLS
library(pls)
par(mfrow = c(1,1))
set.seed(1)
pls fit1 \leftarrow plsr(MORTALITY \sim 0 + ., 5, data = mortality std, validation = 'CV')
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summary(pls fit1)
# after the random seed is set, I see 4 components
set.seed(1)
pls fit2 \leftarrow plsr(MORTALITY \sim 0 + ... 4, data = mortality std, validation = 'CV')
summary(pls fit2)
pls fit2 coef <- pls fit2$coefficients[,,4]
pls fit2 fitted <- pls fit2\fitted.values[,4]
pls fit2 residual <- mortality std$MORTALITY - pls fit2 fitted
loadings(pls fit2)
# diagnostic plot
plot diagnostic <- function(fitted values, residual values, string) {
par(mfrow = c(2, 2))
# the observed against the fitted values
plot(mortality std$MORTALITY, fitted values,
   main = sprintf("observed value vs fitted values (%s)", string),
   xlab = "mortality", ylab = 'fitted mortality')
abline(a = 0, b = 1)
# ganorm
qqnorm(fitted values, main = sprintf("QQ plot (%s)", string))
qqline(fitted values)
# residuals against fitted
plot(residual values, fitted values,
   main = sprintf("residuals vs fitted values (%s)", string),
   xlab = "residuals", ylab = 'fitted mortality')
abline(h = 0)
# histogram of residuals
hist(residual values, main = sprintf('histogram of residuals (%s)', string),
   xlab = 'residuals')
par(mfrow = c(1, 1))
plot diagnostic(pls fit2 fitted, pls fit2 residual, "pls")
# cross-validation result
summary(pls fit2)
# Part IV ridge
set.seed(1)
ridge fit1 <- cv.glmnet(as.matrix(mortality std[, -7]), mortality std$MORTALITY,
              alpha = 0, intercept = F)
plot(ridge fit1)
ridge lambda <- ridge fit1$lambda.min
ridge coef \leftarrow coef(ridge fit1, s = ridge lambda) ax
ridge fit1 fitted <- as.matrix(mortality std[, -7]) %*% ridge coef
ridge fit1 residual <- mortality std$MORTALITY - ridge fit1 fitted
#ridge vif
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R \leftarrow as.matrix(t(mortality std[, -7])) \%*\% as.matrix(mortality std[, -7])
A <- solve(R + diag(ridge lambda, 6)) %*% R %*% solve(R + diag(ridge lambda, 6))
ridge vif <- diag(solve(A)) * diag(A)
# only a little smaller, since our lambda is very small
# diagnostic plot
plot diagnostic(ridge fit1 fitted, ridge fit1 residual, "ridge")
# cross validation result
ridge fit1 cv <- ridge fit1$cvm[which(ridge fit1$lambda == ridge lambda)]
#-----
# Part V lasso
set.seed(1)
lasso fit1 <- cv.glmnet(as.matrix(mortality std[, -7]), mortality std$MORTALITY,
             alpha = 1, intercept = F)
plot(lasso fit1)
lasso lambda <- lasso fit1$lambda.min
lasso coef \leftarrow coef(lasso fit1, s = lasso lambda) ax
# someone is set to zero, see here
lasso coef \leftarrow c(lasso coef[1:3], 0, lasso coef[4:5])
lasso fit1 fitted <- as.matrix(mortality std[, -7]) %*% lasso coef
lasso fit1 residual <- mortality std$MORTALITY - ridge fit1 fitted
# diagnostic plot
plot diagnostic(lasso fit1 fitted, lasso fit1 residual, "lasso")
# cross validation result
lasso fit1 cv <- lasso fit1$cvm[which(lasso fit1$lambda == lasso lambda)]
#-----
# Part VI linear model revisit (validation)
# we know the outliers may be 37.
lm fit4 \leftarrow lm(formula = MORTALITY \sim 0 + NONWHITE + EDUC + SO2 + PRECIP,
      data = mortality std[-37, ]
par(mfrow = c(2, 2))
plot(lm fit4, which = 1:4)
lm fit4 cv < 1 / 60 * (sum((residuals(lm fit4) / (1 - influence(lm fit4) hat))^2) +
              sum(mortality std[37, 7] - predict(lm fit4, mortality std[37, ])))
```