## Exercise 4

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We load our data (please use your corresponding path)

```
load("C:/Users/Miriam/Box Sync/Masters/IncompleteDataAnalysis/Assignment1_MF/databp.Rdata")
```

### 4a: Complete case analysis

We create a dataset only of complete observations

```
data_complete <- databp[complete.cases(databp), ]</pre>
```

We calculate the mean and standard error of the complete data For the standard error, we define a function calculating the error.

```
mean(data_complete$recovtime)

## [1] 19.27273

# sterror = st deviation divided by squareroot of length of data
sterror <- function(x) sd(x)/sqrt(length(x))
sterror(data_complete$recovtime)</pre>
```

```
## [1] 2.603013
```

To calculate the pearson correlation, we need to first calculate the correlation matrix (excluding missing cases). We then divide the corresponding entry by the product of the two variables standard deviation. We note than when calculating their standard deviation, we must use only the entries included in our complete case analysis.

```
covcc <- cov(databp, use = "complete")

correlation between recovery and dose

covcc["logdose", "recovtime"]/(sd(data_complete$logdose) * sd(data_complete$recovtime))

## [1] 0.2391256

correlation between time and bpressure

covcc["bloodp", "recovtime"]/(sd(data_complete$bloodp) * sd(data_complete$recovtime))</pre>
```

# 4b: Mean imputation

## [1] -0.01952862

We first observe that the time is the only variable with missing observations, we thus only need to perform imputation on this variable

```
mean_recov <- mean(databp$recovtime, na.rm = TRUE)
recov_mi <- ifelse(is.na(databp$recovtime), mean_recov, databp$recovtime)</pre>
```

Mean and st error of recovtime with mean imputation

```
data_mi <- data.frame(logdose = databp$logdose, bloodp = databp$bloodp, recovtime = recov_mi)
mean(data_mi$recovtime)

## [1] 19.27273

sterror(data_mi$recovtime)

## [1] 2.284135

We again calculate the pearson correlation, this time on our new dataset with the missing values filled in with the mean (and we don't need to say we only want complete observations as the data is complete)

covmi <- cov(data_mi)

correlation between recovery and dose

covmi["logdose", "recovtime"]/(sd(data_mi$logdose) * sd(data_mi$recovtime))

## [1] 0.2150612

correlation between time and bpressure

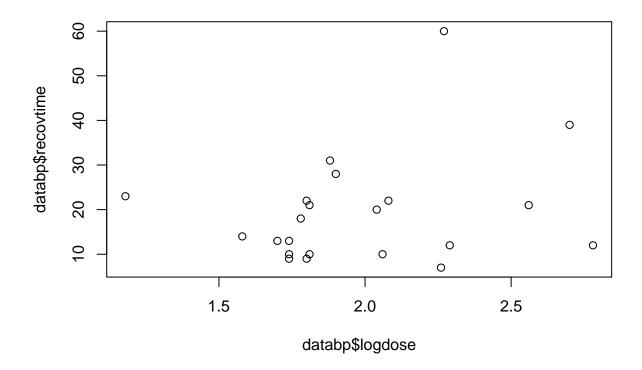
covmi["bloodp", "recovtime"]/(sd(data_mi$bloodp) * sd(data_mi$recovtime))

## [1] -0.01934126</pre>
```

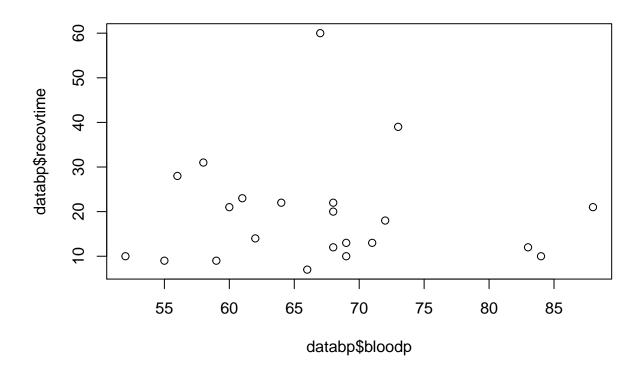
### 4c: Mean regression imputation

We first plot the variables, to see if the assumption of a linear dependence used for our regression is accurate. Whereas I wouldn't be really confident saying there is a linear dependence, the plots give at least no evidence to say the dependence would be for example logarithmic or quadratic. So in my opinion, this data is more telling us there is no reason not to do linear regression, more than they say there is reason to do linear regression.

```
plot(databp$logdose, databp$recovtime)
```



plot(databp\$bloodp, databp\$recovtime)



We include both of our observed variables into our regression. Our regressionmodel is

```
Recovtime_i = \beta_0 + \beta_1 * Logdose_i + \beta_2 * Bloodpressure_i + \varepsilon_i
```

assuming  $\varepsilon_i \stackrel{iid}{\sim} N(0, \sigma^2)$ . We expect  $\beta_1 > 0$  and  $\beta_2 < 0$ . As we have seen a positive correlation between time and dose and a negative correlation between time and byressure.

```
fit_mr <- lm(databp$recovtime ~ databp$logdose + databp$bloodp)
fit_mr$coefficients</pre>
```

```
## (Intercept) databp$logdose databp$bloodp
## 15.2159065 11.4290287 -0.2769265
```

We create a new vector with the predicted times and include it to a new dataset with our new predicted values.

```
predicted_time <- predict(fit_mr, newdata = databp)
recov_predict <- ifelse(is.na(databp$recovtime), predicted_time, databp$recovtime)
data_mr <- data.frame(logdose = databp$logdose, bloodp = databp$bloodp, recovtime = recov_predict)</pre>
```

Mean and standard error of the predicted dataset

```
mean(data_mr$recovtime)
## [1] 19.44428
```

## [1] 2.312845

sterror(data mr\$recovtime)

```
Calculating the correlation matrix
covmr <- cov(data_mr)</pre>
correlation between recovery and dose
covmr["logdose", "recovtime"]/(sd(data mr$logdose) * sd(data mr$recovtime))
## [1] 0.2801835
correlation between time and bpressure
covmr["bloodp", "recovtime"]/(sd(data_mr$bloodp) * sd(data_mr$recovtime))
## [1] -0.0111364
4d:
We use the model of 3c and add a random number
set.seed(49)
predicted_sri <- predict(fit_mr, newdata = databp) + rnorm(nrow(databp), 0, sigma(fit_mr))</pre>
recov_sri <- ifelse(is.na(databp$recovtime), predicted_sri, databp$recovtime)
We calculate the mean and standard error
data_sri <- data.frame(logdose = databp$logdose, bloodp = databp$bloodp, recovtime = recov_sri)</pre>
mean(data_sri$recovtime)
## [1] 18.72278
sterror(data_sri$recovtime)
## [1] 2.341176
Calculating the correlation matrix
covsri <- cov(data_sri)</pre>
correlation between recovery and dose
covsri["logdose", "recovtime"]/(sd(data_sri$logdose) * sd(data_sri$recovtime))
## [1] 0.251039
correlation between time and bpressure
covsri["bloodp", "recovtime"]/(sd(data_sri$bloodp) * sd(data_sri$recovtime))
## [1] -0.01514032
As we include a random error term, we need to make sure that we don't predict negative times. Further, we
might consider to limit the fractional part of the predicted values, thus we might include a floor function to
our predicted values, making them nonfractional. In that case, we would use
recov_sri_int <- ifelse(is.na(databp$recovtime), floor(predicted_sri), databp$recovtime)</pre>
data_sri_int <- data.frame(logdose = databp$logdose, bloodp = databp$bloodp, recovtime = recov_sri_int)</pre>
giving us the following mean, standard error, correlation matrix:
mean(data_sri_int$recovtime)
```

## [1] 18.64

```
sterror(data_sri_int$recovtime)
## [1] 2.347964
covsriint <- cov(data_sri_int)
covsriint["logdose", "recovtime"]/(sd(data_sri_int$logdose) * sd(data_sri_int$recovtime))
## [1] 0.2479823
covsriint["bloodp", "recovtime"]/(sd(data_sri_int$bloodp) * sd(data_sri_int$recovtime))
## [1] -0.01684754

4e: predictive mean
We define a function which will calculate the index of the closest observed value to our prediction.
mis_index <- which(is.na(databp$recovtime))
best_replace <- function(val) {
    min <- .Machine$integer.max
    ind <- -1</pre>
```

```
best_replace <- function(val) {</pre>
    available_data <- which(is.na(databp$recovtime) == FALSE)</pre>
    for (i in available data) {
        squaredis <- (val - databp$recovtime[i])^2</pre>
        if (squaredis < min) {</pre>
             min <- squaredis
             ind <- i
        }
    }
    return(ind)
}
recov_prmtp <- 1:length(databp$recovtime)</pre>
for (inde in mis_index) {
    best <- best_replace(predicted_sri[inde])</pre>
    recov_prmtp[inde] <- databp$recovtime[best]</pre>
    cat("datapoint ", inde, " we take value ", databp$recovtime[best], " of index ",
        best, "\n")
}
```

```
## datapoint 4 we take value 7 of index 1
## datapoint 10 we take value 21 of index 7
## datapoint 22 we take value 18 of index 3

recov_prm <- ifelse(is.na(databp$recovtime), recov_prmtp, databp$recovtime)
data_prm <- data.frame(logdose = databp$logdose, bloodp = databp$bloodp, recovtime = recov_prm)</pre>
```

We calculate the mean and st error

```
mean(data_sri_int$recovtime)
```

## [1] 18.64

```
sterror(data_sri_int$recovtime)

## [1] 2.347964
and the correlation
covprm <- cov(data_prm)
covprm["logdose", "recovtime"]/(sd(data_prm$logdose) * sd(data_prm$recovtime))

## [1] 0.2590259
covprm["bloodp", "recovtime"]/(sd(data_prm$bloodp) * sd(data_prm$recovtime))

## [1] -0.01407195</pre>
```

#### 4f:

One advantage of predictive mean matching is that we can be certain that our predicted values are values which comply with the pattern of the given variable. With that I mean that if our variable is time, and has only integer values, predictive mean matching will also only predict integer values (as it simply takes already given values). Stochastic regression imputation on the other side calculates the value with regression, and might predict non-integer values. However, predictive mean matching can have similar disadvantages to mean imputation: using already existing values as the values for missing data makes the overall data less variable (as we do not add new values to the dataset, but just have already existing data with higher frequency in the data afte the imputation), leading to changes in the standard deviation of the data.