

Reproducible Research practical

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Choose **File** and then **R Markdown** give *Title*, *Author* and press **OK**.

A Rmarkdown template file (*Untitled1*) is generated use the **Knit** button to create your first reproducible document (you will be requested to give the file a name and to store it somewhere).

Adding code chunks

Now add the following code chunks and type a small description of what kind of analysis is performed.

describe code ...

```
library(airway)
library(DESeq2)
data("airway")
airway$dex <- releve1(airway$dex, "untrt")
dds <- DESeqDataSet(airway, design = ~ cell + dex) #add formula
nrow(dds)
```

```
## [1] 64102
```

describe results...

Filtering un- or lowly expressed genes using counts per million is advocated by the developers of edgeR[@] another package for the differential expression analysis (see section 2.6 Filtering).

```
cpm <- 1e6*counts(dds)/colSums(counts(dds))
keep <- rowSums(cpm>1) >= 4
dds <- dds[keep, ]
nrow(dds)
```

```
## [1] 14360
```

describe results...

describe code ...

```
dds <- DESeq(dds)
```

```
## estimating size factors
```

```
## estimating dispersions
```

```
## gene-wise dispersion estimates
```

```
## mean-dispersion relationship
```

```
## final dispersion estimates
```

```
## fitting model and testing
```

```
res <- results(dds, alpha = 0.05)
res$Symbol <- mapIds(org.Hs.eg.db, rownames(res), "SYMBOL", "ENSEMBL")
```

```
## 'select()' returned 1:many mapping between keys and columns
```

```
res[order(res$padj),]
```

```
## log2 fold change (MAP): dex trt vs untrt
```

```
## Wald test p-value: dex trt vs untrt
```

```
## DataFrame with 14360 rows and 7 columns
```

```
##           baseMean log2FoldChange      lfcSE      stat
##           <numeric>      <numeric> <numeric> <numeric>
## ENSG00000152583    997.5202      4.293616 0.1721530    24.94071
## ENSG00000165995    495.4311      3.174093 0.1274643    24.90182
## ENSG00000101347   12708.7527      3.604035 0.1489683    24.19329
## ENSG00000120129    3411.4330      2.858802 0.1185387    24.11704
## ENSG00000189221    2342.8234      3.216087 0.1366041    23.54312
## ...           ...           ...           ...           ...
## ENSG00000009307   15828.50403 -4.327685e-05 0.07700615 -5.619921e-04
## ENSG00000123728    546.27594 -4.533937e-05 0.10468659 -4.330962e-04
## ENSG00000135722     75.27579  9.042885e-05 0.19991225  4.523427e-04
## ENSG00000173531    228.88746 -7.440328e-05 0.15275451 -4.870775e-04
## ENSG00000180673     23.73033  2.341427e-06 0.30301019  7.727221e-06
##           pvalue      padj      Symbol
##           <numeric> <numeric> <character>
## ENSG00000152583 2.693499e-137 3.867864e-133    SPARCL1
## ENSG00000165995 7.110821e-137 5.105569e-133    CACNB2
## ENSG00000101347 2.617388e-129 1.252856e-125    SAMHD1
## ENSG00000120129 1.656548e-128 5.947008e-125    DUSP1
## ENSG00000189221 1.476513e-122 4.240545e-119    MAOA
## ...           ...           ...           ...
## ENSG00000009307  0.9995516    0.9997241    CSDE1
## ENSG00000123728  0.9996544    0.9997241    RAP2C
## ENSG00000135722  0.9996391    0.9997241    FBXL8
## ENSG00000173531  0.9996114    0.9997241    MST1
## ENSG00000180673  0.9999938    0.9999938    NA
```

```
describe results...
```

Adding a figure

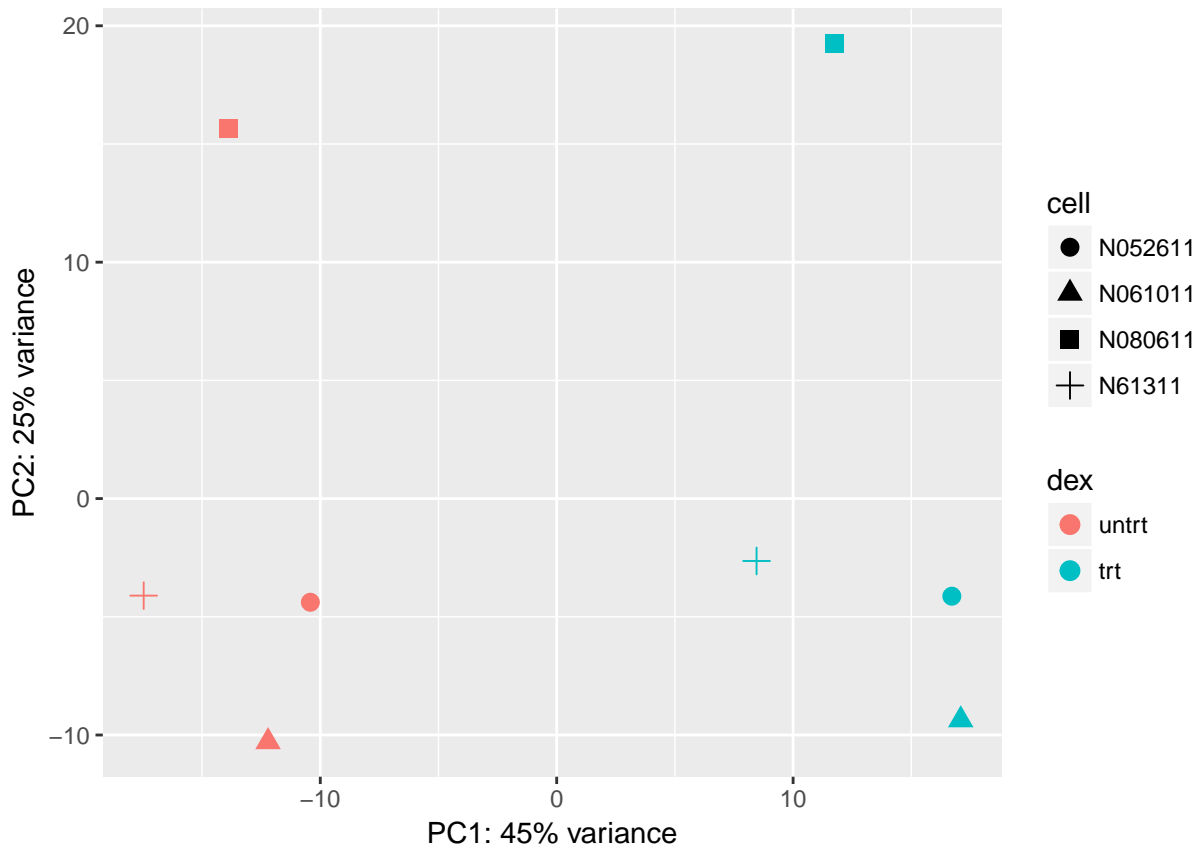
Adding a figure is as easy as a code chunk!

describe code ...

```
library(vsn)
library(ggplot2)
rld <- rlog(dds, blind = FALSE)
pcaData <- plotPCA(rld, intgroup = c("dex", "cell"), returnData = TRUE)
pcaData
```

##		PC1	PC2		group	dex	cell	name
##	SRR1039508	-17.470328	-4.104236	untrt :	N61311	untrt	N61311	SRR1039508
##	SRR1039509	8.454183	-2.638643	trt :	N61311	trt	N61311	SRR1039509
##	SRR1039512	-10.419877	-4.384220	untrt :	N052611	untrt	N052611	SRR1039512
##	SRR1039513	16.715160	-4.128324	trt :	N052611	trt	N052611	SRR1039513
##	SRR1039516	-13.893510	15.656023	untrt :	N080611	untrt	N080611	SRR1039516
##	SRR1039517	11.732798	19.249447	trt :	N080611	trt	N080611	SRR1039517
##	SRR1039520	-12.209613	-10.287692	untrt :	N061011	untrt	N061011	SRR1039520
##	SRR1039521	17.091186	-9.362354	trt :	N061011	trt	N061011	SRR1039521

```
percentVar <- round(100 * attr(pcaData, "percentVar"))
ggplot(pcaData, aes(x = PC1, y = PC2, color = dex, shape = cell)) +
  geom_point(size = 3) +
  xlab(paste0("PC1: ", percentVar[1], "% variance")) +
  ylab(paste0("PC2: ", percentVar[2], "% variance")) +
  coord_fixed()
```



describe results...

Tip of iceberg

This is really the *tip of the iceberg* check out the `rmarkdown` website or the `knitr` website that show many applications of `rmarkdown` i.e. how to write complete books, website, scientific manuscripts and much more!