

Purpose

- Analyze the evolution and spread of influenza viruses by creating a time-calibrated maximum likelihood phylogenetic tree and geospatial map
- Apply new avian influenza functionality in Freyja to investigate ongoing transmission and spread

Introduction

- Zoonotic spillover: pathogen transfer from animals to humans, common through direct animal contact or fomites^[1]
 - Ex. SARS-CoV-2 pandemic (wildlife trade), avian influenza: bird → human emergence^[2]
 - Risks of reassortment and widespread infections in animal populations
- Analyzing pathogen spread and evolution through wastewater and environmental sampling^[3]
 - Monitoring for outbreaks within communities before first case reported
 - Less expensive
 - More representative of the population
- Freyja
 - Tool typically used to analyze virus lineages in mixed SARS-CoV-2 wastewater samples^[4]

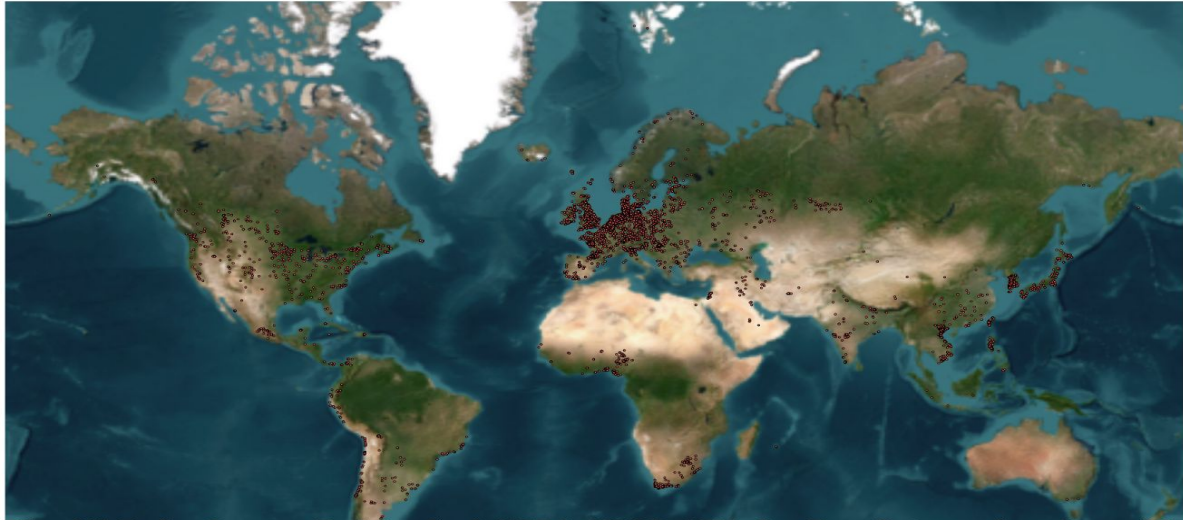


Figure 1. A geospatial map showing avian influenza case distribution between August 2018 to August 2023 was constructed with the Esri World Map basemap layer and plotted using Matplotlib^[5]

Methods and Materials

Materials:

- Phylogenetic and Geospatial Analyses: GISAID data samples, EMPRES-i case data, Geopandas, Matplotlib, Cartopy, Contextily, MAFFT, TimeTree
- Variant Analyses: Freyja, iVar

Methods:

- Obtained 1,689 influenza sequences for phylogeny creation from GISAID and geospatial data from EMPRES-i^{[7][8]}
- Aligned flu sequences and processed temporal data to generate a time-calibrated maximum likelihood phylogenetic tree^[6]
- Processed avian influenza data using various Python packages to create a geospatial map showing case distribution between August 2018-2023^[5]
- Aligned avian influenza environmental samples to plot variant prevalence using Freyja^[4]

Exploring the Spread and Evolution of Avian Influenza through Bioinformatic Analyses

Dr. Yilmaz Systems Lab 2025 Pd 3 - Jasmine Ma

Results

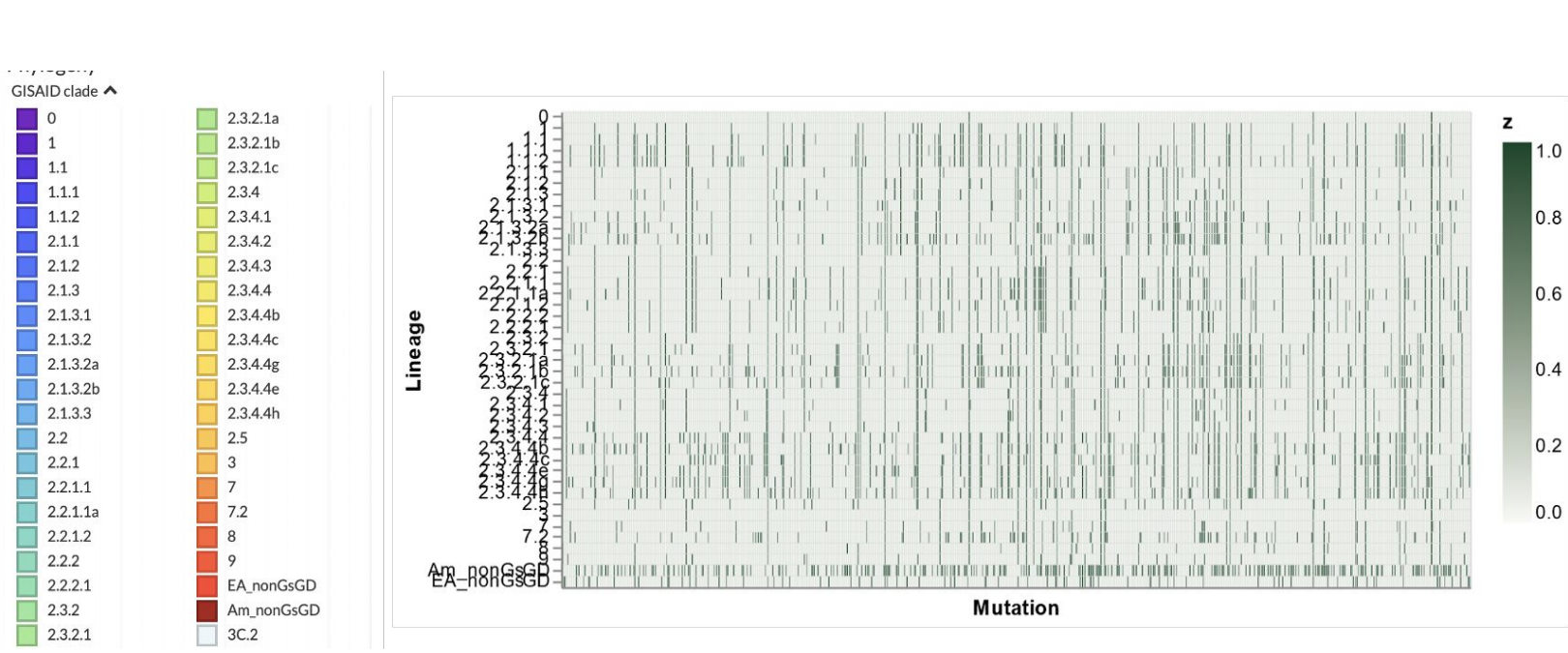


Figure 2. Barcodes used for Freyja analysis^[4]

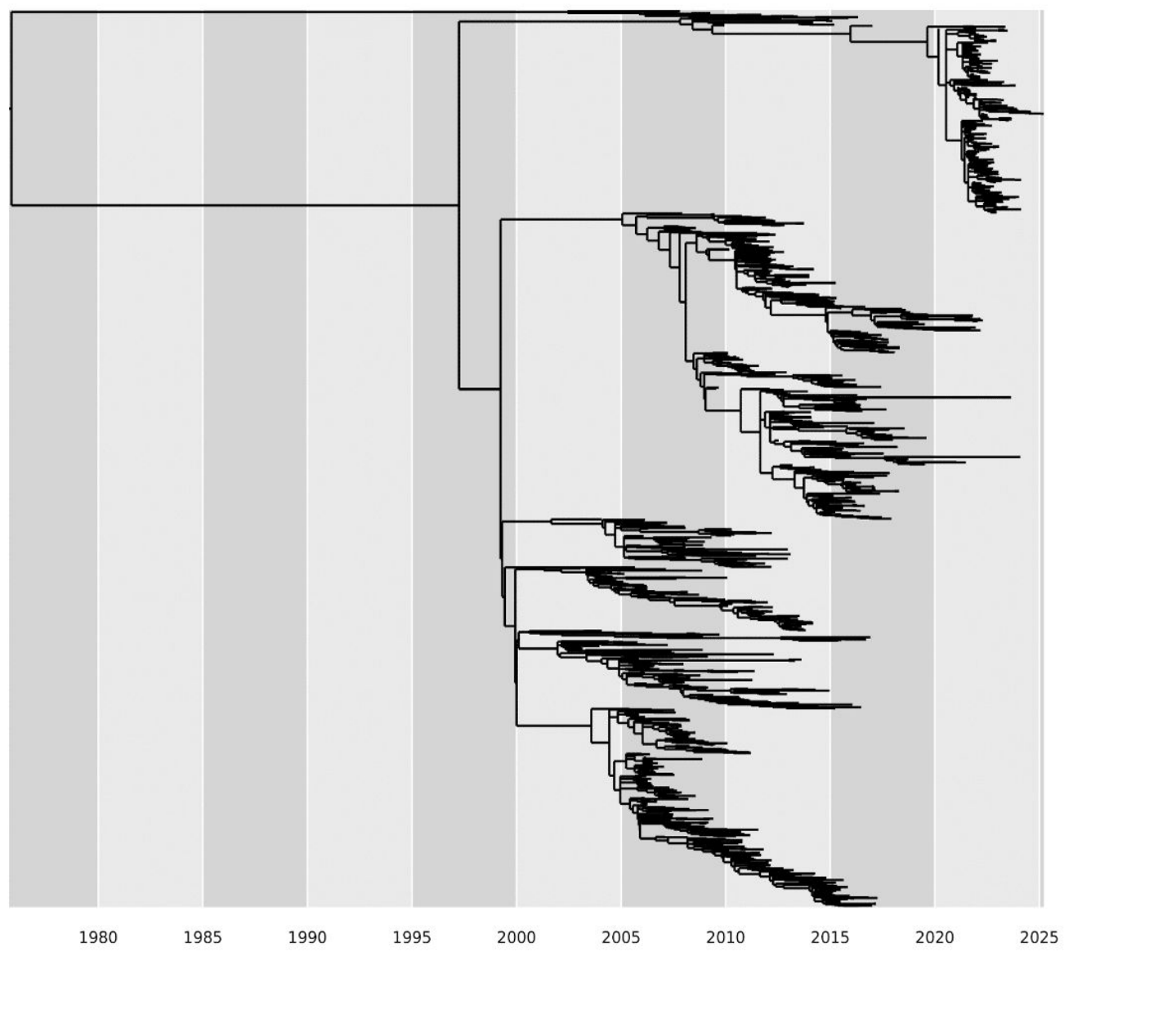


Figure 4. A time-calibrated maximum likelihood phylogenetic tree was constructed using TimeTree based on 1,689 influenza sequence samples^[6]

sample_outputs					
	summarized	lineages	abundances	resid	coverage
SRR7496088.tsv	[[('Other', 0.9978474561560453)]]	h52.3.4.4h h52.3.4.4	0.99572276 0.00212470	16.250032633310500	81.07954545454550
SRR7643082.tsv	[[('Other', 0.9999999999999996)]]	h5AmnonGsGD	1.00000000	0.9947018912430020	0.0
SRR23208411.tsv	[[('Other', 0.99979400000021175)]]	h5AmnonGsGD	0.99979400	17.24638512958310	34.65909090909090

Figure 6. Freyja output after demixing and aggregating 3 mixed environmental avian influenza samples for lineage prevalence. Bar chart shows plotting for lineage prevalence in a mixed environmental sample of avian influenza^[4]
Note: lineage prevalence was also plotted for samples SRR7643082 and SRR23208411 but didn't appear on the plot due to having mostly pure lineage.

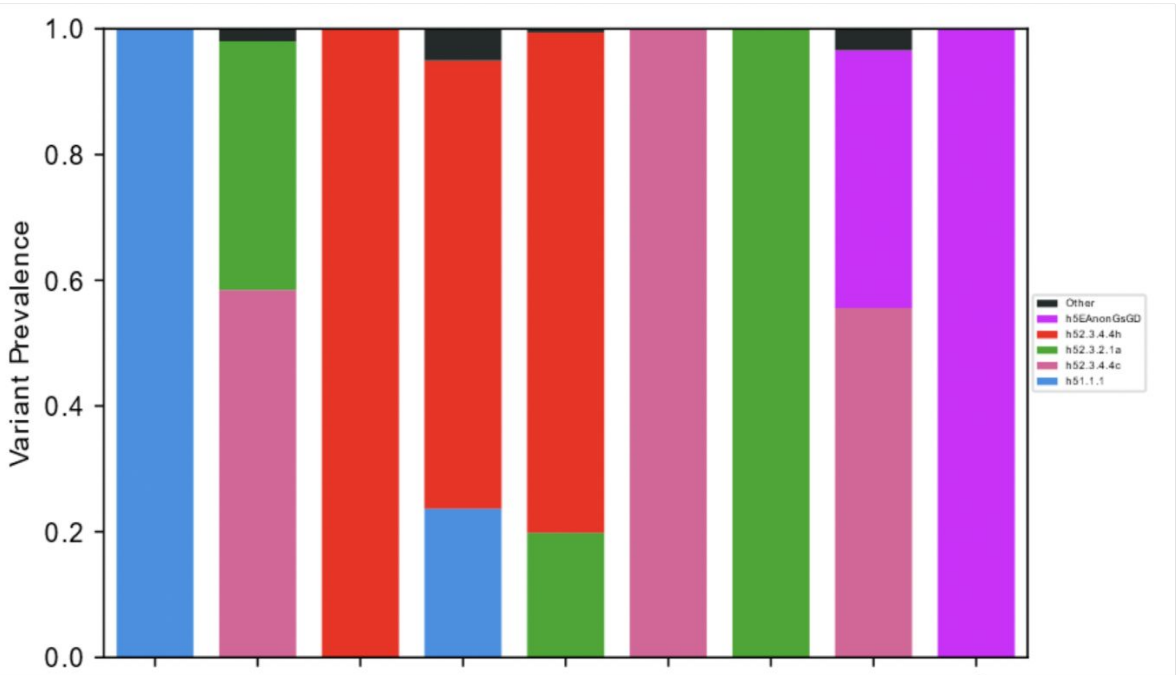


Figure 3. Freyja analyses for a simulated set of flu viruses, showing lineage prevalence in each sample^[4]

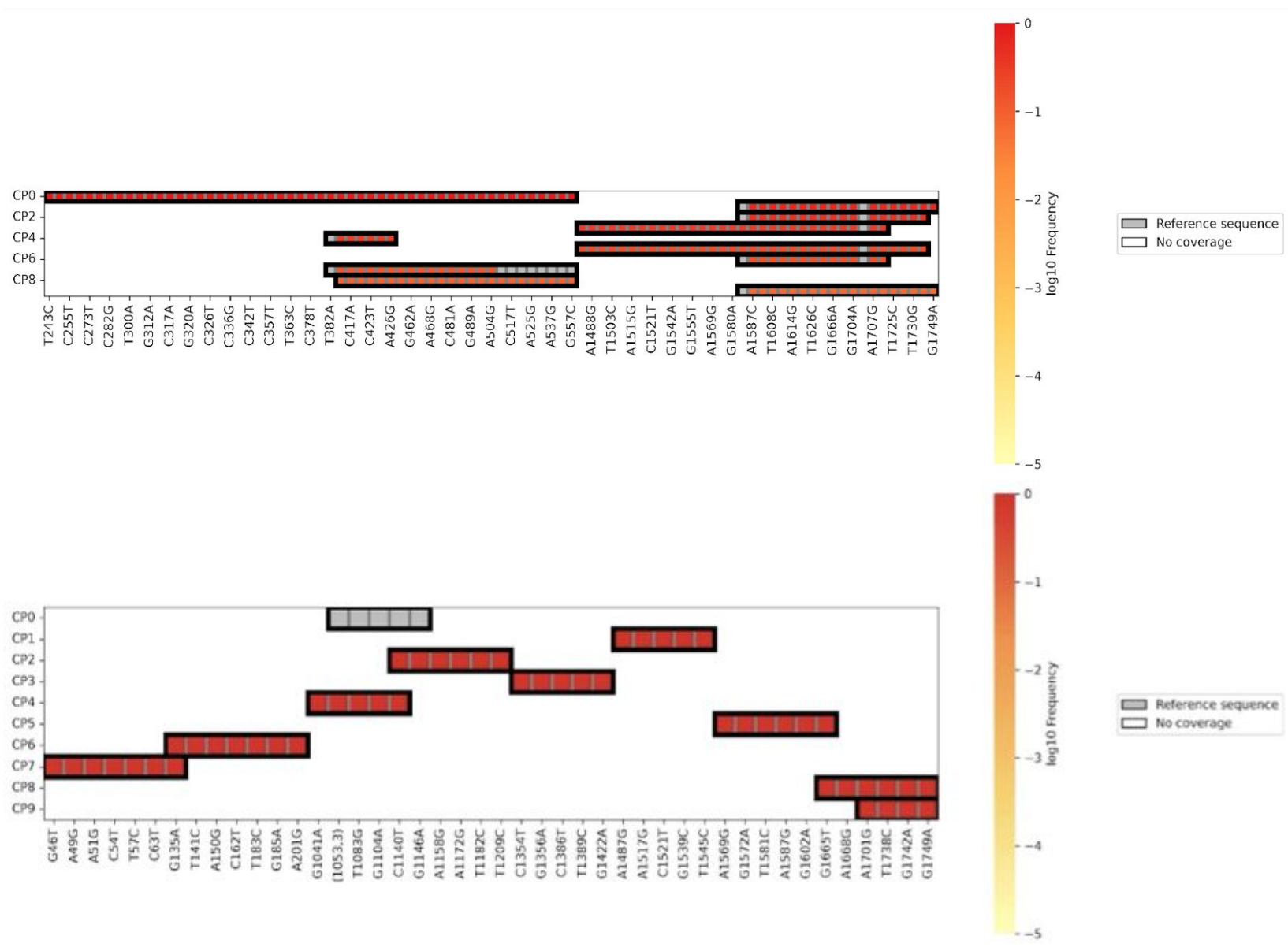
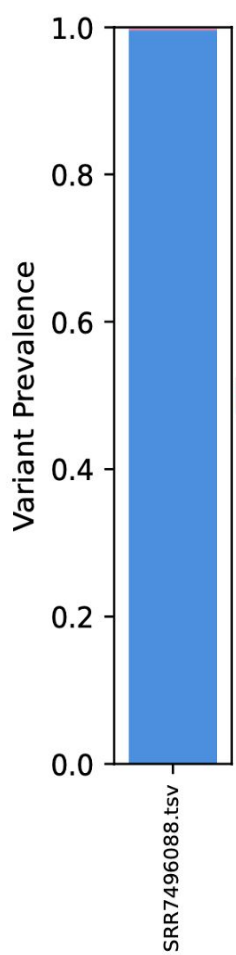


Figure 5. Covariant analysis for SRR7496088 and SRR23208411^[4]



Results

- Freyja covariant and mixed lineage analyses show a high prevalence of mutated lineages within avian influenza environmental samples
- Signs of mixed lineages show risks of zoonotic spillover and genetic reassortment (Figure 6)
- Evolving lineages were detected in environmental samples of avian influenza, showing Freyja's adaptability to avian influenza viral strains
- Phylogenetic analyses of the influenza sequences suggest an evolutionary relationship
- Geospatial analyses using environmental data showed effects of migration on virus spread

Conclusion

- Phylogenetic analyses
 - Analyzes evolutionary relationships
 - Environmental samples can show virus mutation trends over time
- Geospatial mapping
 - Visualizes and tracks the geographic spread of known pathogens
 - Shows patterns and trends through mapping data
- Mixed sample analyses using Freyja
 - Ability to detect variants of influenza virus even at low concentrations
 - Surveillance benefits: tracing the evolution of the pathogen through new strains
- Implications:
 - Monitoring common migratory routes through geospatial mapping can pinpoint emerging regions
 - Sampling from frequent migration routes can prevent human transmission and improve public health

References

[1] Himsworth, C. G., Duan, J., Prystajec, N., Coombe, M., Baticados, W., Jassem, A. N., Tang, P., Sanders, E., & Hsiao, W. (2020). Targeted Resequencing of Wetland Sediment as a Tool for Avian Influenza Virus Surveillance. *Journal of wildlife diseases*, 56(2), 397–408.

[2] Joseph, U., Su, Y. C., Vijaykrishna, D., & Smith, G. J. (2017). The ecology and adaptive evolution of influenza A interspecies transmission. *Influenza and other respiratory viruses*, 11(1), 74–84. <https://doi.org/10.1111/irv.12412>

[3] Yeo, S. J., Hoang, V. T., Duong, T. B., Nguyen, N. M., Tuong, H. T., Azam, M., Sung, H. W., & Park, H. (2022). Emergence of a Novel Reassortant H5N3 Avian Influenza Virus in Korean Mallard Ducks in 2018. *Intervirology*, 65(1), 1–16. <https://doi.org/10.1159/000517057>

[4] Karthikeyan, S., Levy, J. I., De Hoff, P. et al. (2022). Wastewater sequencing reveals early cryptic SARS-CoV-2 variant transmission. *Nature*, 609, 101–108. <https://doi.org/10.1038/s41586-022-05049-6>

[5] ArcGIS Online Basemaps [Map]. (2010).

[6] Sagulenko, P., Puller, V., & Neher, R. A. (2018). TreeTime: Maximum-likelihood phylodynamic analysis. *Virus evolution*, 4(1). <https://doi.org/10.1093/ve/vex042>

[7] Food and Agriculture Organization of the United Nations. (n.d.). Avian Influenza Diseases Map. EMPRES-i. <https://empres-i.apps.fao.org/diseases>

[8] Khare, S., et al. (2021). GISAID's role in pandemic response. *China CDC Weekly*, 3(49), 1049-1051. <https://doi.org/10.46234/ccdcw2021.255>