OCN 750 - HW 4

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```
#Set wd and load data table
#setwd("C:/Users/mkapur/Dropbox/2015 Fall/OCN 750")
roadkill = read.table("RoadKills.txt", header = TRUE, colClasses = "numeric")
```

Plot TOT.N vs. the other variables I mentioned. TOT.N, D.PARK, URBAN, D.WAT.RES. URBAN is highly skewed.

```
#set up graphing device
par(mfrow = c(2,2))

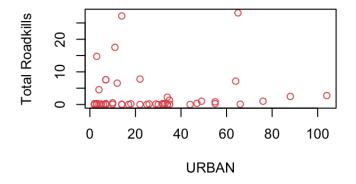
#create index of which columns we want to plot against TOT.N

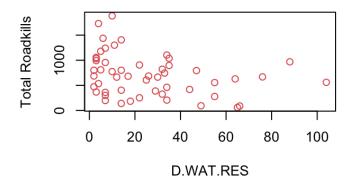
colnums = c(13,18,20)

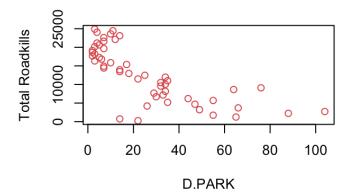
#a for-loop to plot each value quickly. URBAN is, indeed, quite skewed.

for (v in colnums) {
    plot(roadkill$TOT.N, roadkill[,v], xlab = names(roadkill[v]) , ylab = "Total Roadk ills", col = "indianred3")
    }

#dev.off()
```







Make a column where this predictors is square-root transformed, and plot the relationships with the new predictors.

```
#Create three new columns for the square-root transformations
roadkill[24:26] = NA

#designate their names and replace "V24" etc
newcols = c("urb_sqrt", "dwat_sqrt", "dpark_sqrt")
names(roadkill)[24:26] = newcols
sqrtnums = c(24:26)

#do it manually...
sqrt(roadkill$URBAN) -> roadkill[24]
sqrt(roadkill$D.WAT.RES) -> roadkill[25]
sqrt(roadkill$D.PARK) -> roadkill[26]
head(roadkill)[24:26]
```

```
## urb_sqrt dwat_sqrt dpark_sqrt
## 1 2.790520 15.878067 15.81815
## 2 5.210566 11.814102 27.22460
## 3 5.299623 7.692074 35.21477
## 4 0.911592 16.668593 41.71193
## 5 1.565886 31.109613 47.24542
## 6 1.652271 23.664319 52.19281
```

```
#a for-loop to plot the new transformed values against TOT.N. dev.off()
```

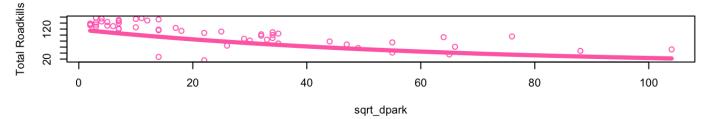
```
## null device
## 1
```

```
par(mfrow = c(2,2))
for (k in sqrtnums){
plot(roadkill$TOT.N, roadkill[,k], xlab = names(roadkill[k]) , ylab = "Total Roadkills", col = "hotpink4")
}
#dev.off()
```

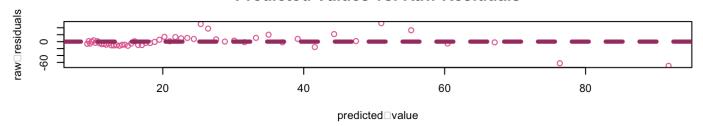
Fit a Poisson GLM to test if total number of roadkills is predicted by D.PARK. Plot the raw relationship and plot the fitted curve on top of it. Plot residuals vs. the predictor: use both the raw (response) residuals and the deviance residuals.

```
require(stats)
#create your Poisson GLM using the square-root transformed values
#I've noticed that for the Effects package to work, you need to separate the data
from the field names.
#(No $ signs!)
poipark = glm(TOT.N ~ dpark sqrt, data = roadkill, family = poisson)
#Plot a curve using generated coefficients against the data
par(mfrow = c(3,1))
plot(roadkill$TOT.N, roadkill$dpark_sqrt, xlab = "sqrt_dpark" , ylab = "Total Road
kills", col = "hotpink1", main = "Total Kills vs DPark", cex.main = 1.5)
curve(exp(coef(poipark)[1]+coef(poipark)[2]*x), col = 'hotpink2', lwd = 4, x
lab = "D.PARK", ylab = "Total Kills", add = TRUE)
#Plot of raw residuals against predicted values based on regression (these are ext
racted via fitted())
plot(residuals(poipark, type = "response") ~ fitted(poipark), ylab
residuals", xlab = "predicted value", col = "hotpink3", main = "Predicted Va
lues vs. Raw Residuals", cex.main = 1.5)
           = 0,
                               lwd = 4,
                                        col = 'hotpink4')
abline(h
                   lty = 2,
#Same as above, but with deviance (Pearson) residuals
plot(residuals(poipark, type
                             = "pearson")
                                             ~ fitted(poipark), ylab
rson residuals", xlab = "predicted
                                       value", main = "Predicted Values vs. Pea
rson (Deviance) Residuals", cex.main = 1.5, col = "indianred1")
abline(h
                               lwd = 4, col = "indianred2")
           = 0,
                  lty = 2,
```

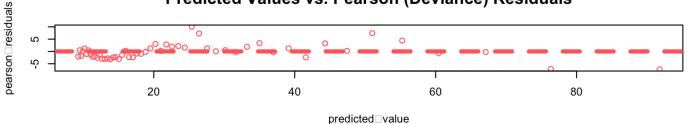




Predicted Values vs. Raw Residuals



Predicted Values vs. Pearson (Deviance) Residuals



dev.off()

null device
1

Are there any strong patterns? How do the raw and deviance residuals differ, and why?

#The deviance residuals are more constrained, though they don't seem to differ gre atly from the pattern. There is a clear negative, potentially logarithmic relation ship between the predictor and response variable, with variance appearing to incre ase with D.Park. This would explain the tighter relationship between values and re siduals at lower values of D.Park.

Do a likelihood ratio test to test for the significance of the predictor.

require(car)
require(effects)
Anova(poipark)

```
## Analysis of Deviance Table (Type II tests)
##
## Response: TOT.N
## LR Chisq Df Pr(>Chisq)
## dpark_sqrt 554.55 1 < 2.2e-16 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1</pre>
```

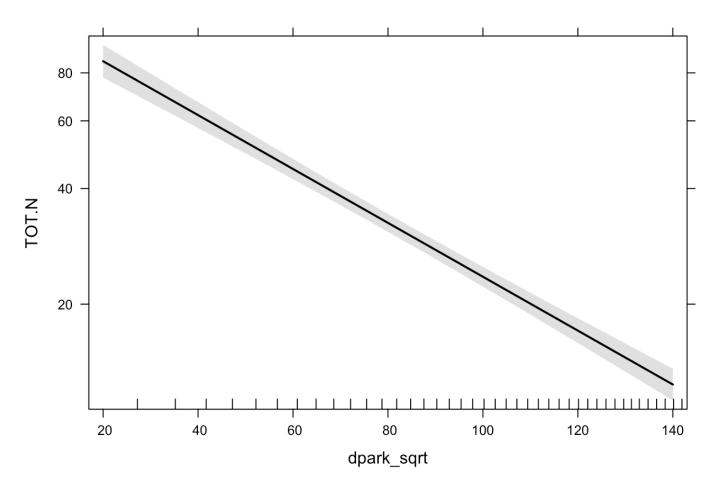
Fit the same model with quasipoisson, and with a negative binomial distribution.

```
#create your Quasi - Poisson GLM using the square-root transformed values
qpoipark = glm(TOT.N ~ dpark_sqrt, data = roadkill, family = quasipoisson)

#Create the negative binomial model
require(MASS)
negbpark = glm.nb(TOT.N ~ dpark_sqrt, data = roadkill)

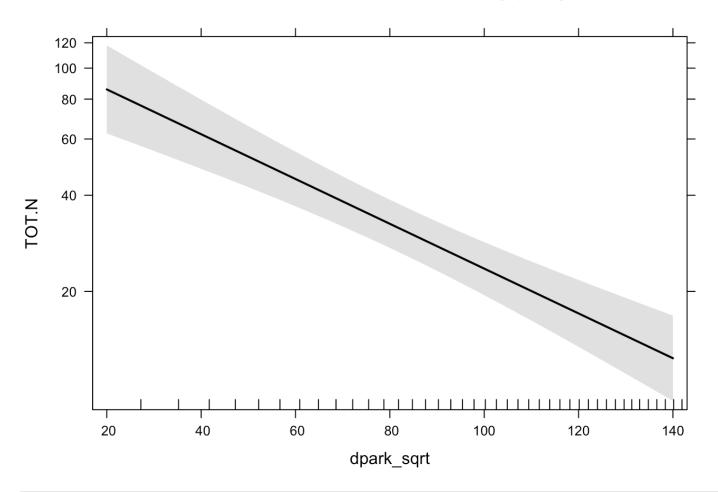
#Plot them all together
par(mfrow = c(3,1))
#This looks very different from the example in class.
plot(allEffects(poipark), main = "Poisson-Distributed Model (dpark)", col = "darks eagreen1")
```

Poisson-Distributed Model (dpark)



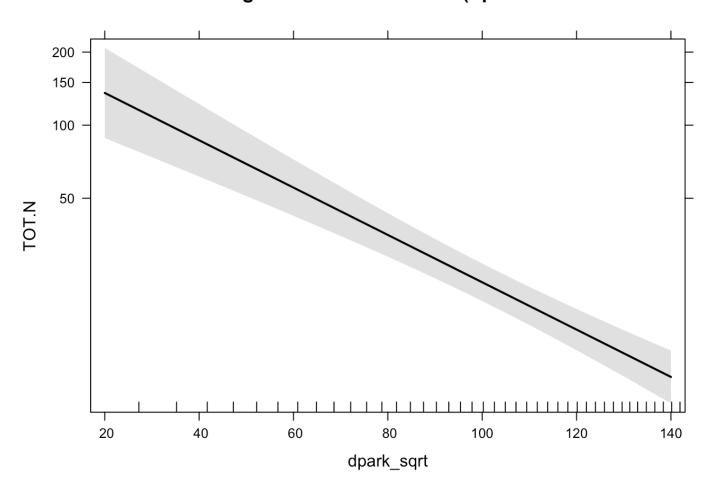
plot(allEffects(qpoipark), main = "QuasiPoisson-Distributed Model (dpark)", col =
"darlseagreen2")

QuasiPoisson-Distributed Model (dpark)



plot(allEffects(negbpark), main = "Negative Binomial Model (dpark", col = "darksea
green3")

Negative Binomial Model (dpark



Do appropriate hypothesis tests on these models.

```
#A bootstrapping may be more approriate, but here we do a Likelhihood Ratio Test
(X2 distributed).
#A t-test would assume normal error (analogous to Wald test).
#The LRT is quickly summarized by Anova()

#Here I run the test and pin it to a new data frame. All three indicate significan
ce.
LRT = cbind(Anova(poipark)[1],
Anova(qpoipark)[1],
Anova(negbpark)[1])
names(LRT)[1:3] = c("poisson", "qpoisson", "negbinom")
LRT
```

```
## poisson qpoisson negbinom
## dpark_sqrt 554.551 55.43311 75.54486
```

How big is overdispersion based on the quasipoisson?

```
#dispersion parameter function - this way you can quickly quantify Phi based on
whatever glm you used.
overdis = function(model) {
        sum(residuals(model, type = "pearson")^2)/(length(model$y) - lengt
h(model$coefficients))
}
overdis(qpoipark) #Phi is quite high
```

```
## [1] 10.00397
```

What is the theta parameter for the negative binomial?

```
theta = overdis(negbpark)
theta #much smaller
```

```
## [1] 0.9288178
```

Does overdispersion affect the conclusions you would draw from this analysis?

#Our Phi value was pretty high even for the QPoisson.
#Looks like the negative binomial model successfully accounts for the overdispersi on, as the Theta value is well under 1.5.

Now fit a Poisson model that adds in the other two predictors, URBAN (squareroot transformed) and D.WAT.RES.

```
poi.urwapa = glm(TOT.N ~ urb_sqrt + dwat_sqrt + dpark_sqrt, data = roadkill, famil
y = "poisson")
summary(poi.urwapa)
```

```
##
## Call:
## glm(formula = TOT.N ~ urb sqrt + dwat sqrt + dpark sqrt, family = "poisson",
##
       data = roadkill)
##
## Deviance Residuals:
##
       Min
                 10
                      Median
                                   30
                                           Max
## -8.7245 -2.1097 -0.2433
                               1.4958
                                        7.9986
##
## Coefficients:
##
                 Estimate Std. Error z value Pr(>|z|)
## (Intercept)
                4.9478538
                          0.1132939 43.673 < 2e-16 ***
               -0.1139244 0.0232024 -4.910 9.11e-07 ***
## urb sqrt
                                       3.048
## dwat sqrt
                0.0127638 0.0041871
                                               0.0023 **
## dpark sqrt -0.0200720
                          0.0009392 -21.372 < 2e-16 ***
## ---
                   0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
## Signif. codes:
##
## (Dispersion parameter for poisson family taken to be 1)
##
##
       Null deviance: 1071.4 on 51 degrees of freedom
## Residual deviance: 472.6 on 48 degrees of freedom
## AIC: 719.99
##
## Number of Fisher Scoring iterations: 5
```

What is effect size of the 3 different predictors, i.e. how much does # roadkills change as these predictors vary? How do residuals vs. fitted values and residuals vs. predictors look (you can just use deviance or pearson residuals, as they are more appropriate for GLMs)? Do appropriate (marginal) likelihood ratio tests for each of the three predictors.

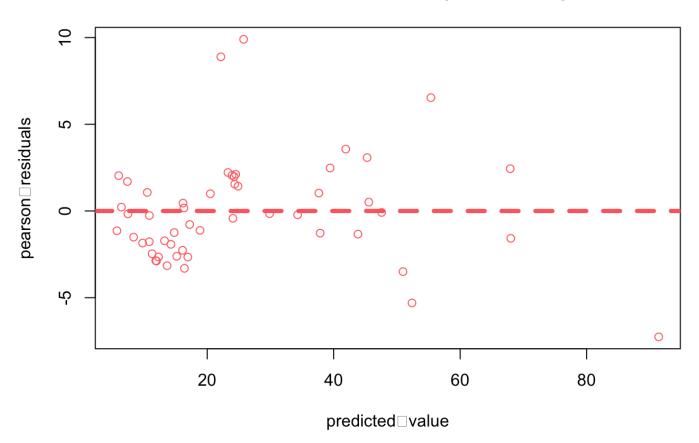
#The effect of each of these predictors is given by the ANOVA.

Anova(poi.urwapa) #it appears that Dpark is still the highest influence upon the m odel.

```
## Analysis of Deviance Table (Type II tests)
##
## Response: TOT.N
##
              LR Chisq Df Pr(>Chisq)
## urb_sqrt
                 25.81 1 3.774e-07 ***
## dwat sqrt
                  9.33
                       1
                            0.002257 **
## dpark sqrt
                494.23 1 < 2.2e-16 ***
## ---
                  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
## Signif. codes:
```

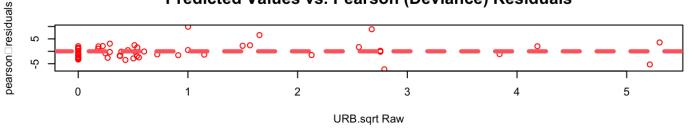
```
#Plot of fitted values vs. deviance (Pearson) residuals
plot(residuals(poi.urwapa, type = "pearson") ~ fitted(poi.urwapa), ylab
= "pearson residuals", xlab = "predicted value", main = "Predicted Values v
s. Pearson (Deviance) Residuals", cex.main = 1.5, col = "indianred1")
abline(h = 0, lty = 2, lwd = 4, col = "indianred2")
```

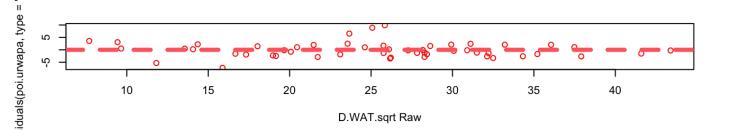
Predicted Values vs. Pearson (Deviance) Residuals

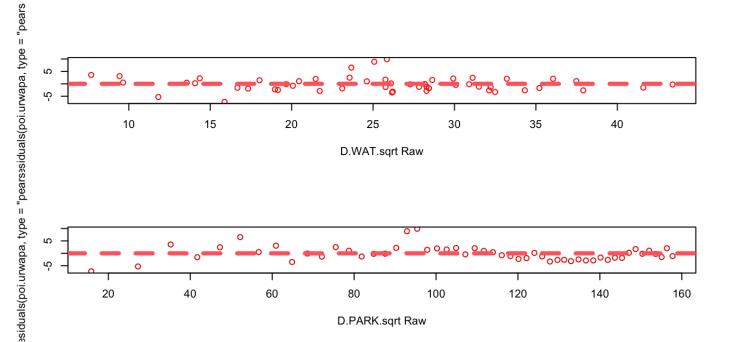


```
#Plot of sqrt-transformed predictor values vs. deviance (Pearson) residuals for th
ree variables
par(mfrow = c(3,1))
#raw urban values
plot(residuals(poi.urwapa, type = "pearson") ~ roadkill$urb_sqrt,
                                                                       ylab
                             = "URB.sqrt Raw", main = "Predicted Values vs. Pears
"pearson residuals", xlab
on (Deviance) Residuals", cex.main = 1.5, col = "firebrick1")
abline(h
           = 0,
                   lty = 2,
                               lwd = 4,
                                           col = "indianred2")
#raw dwat values
plot(residuals(poi.urwapa, type = "pearson") ~ roadkill$dwat_sqrt, xlab = "D.WA
T.sqrt Raw", col = "firebrick2")
                                       col = "indianred2")
abline(h = 0, lty = 2,
                           lwd = 4,
#raw dpark values
plot(residuals(poi.urwapa, type = "pearson") ~ roadkill$dpark_sqrt, xlab = "D.PA
RK.sqrt Raw", col = "firebrick3")
abline(h = 0, lty = 2,
                           lwd = 4,
                                       col = "indianred2")
```

Predicted Values vs. Pearson (Deviance) Residuals







dev.off()

```
## null device
## 1
```

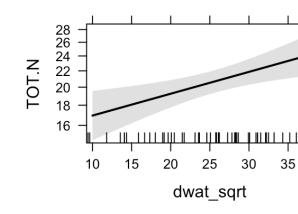
Plot the fitted effects using the 'effects' package. How do you interpret these results?

```
par(mfrow = c(1,2))
#This looks very different from the example in class.
plot(allEffects(poi.urwapa), main = "Poisson-Distributed Model", col = "darkseagre en1")
```

Poisson-Distributed Model

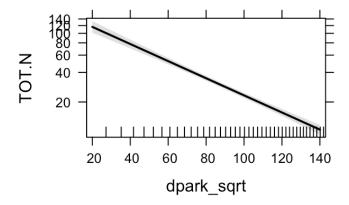
TOT.N

Poisson-Distributed Model



Poisson-Distributed Model

urb_sqrt



Now fit the same model with quasi-Poisson and negative binomial. Plot the fittedeffects using the 'effects' package. How similar are the parameter estimates between Poisson, quasi-Poisson, and negative binomial model?

```
#create your Quasi - Poisson GLM using all square-root transformed values
qpoi.urwapa = glm(TOT.N ~ urb_sqrt + dwat_sqrt + dpark_sqrt, data = roadkill, fami
ly = quasipoisson)

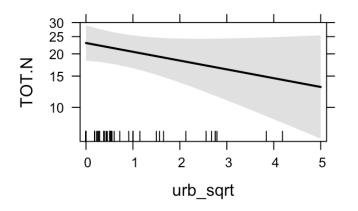
#Create the negative binomial model using all transformed values
require(MASS)
negb.urwapa = glm.nb(TOT.N ~ urb_sqrt + dwat_sqrt + dpark_sqrt, data = roadkill)

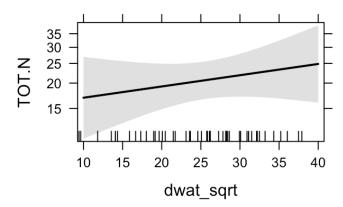
par(mfrow = c(4,3))

#plot for quasi-poisson using "all effects"
plot(allEffects(qpoi.urwapa), main = "QuasiPoisson-Distributed Model", col = "darl seagreen2")
```

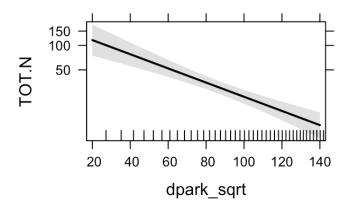
QuasiPoisson-Distributed Model

QuasiPoisson-Distributed Model





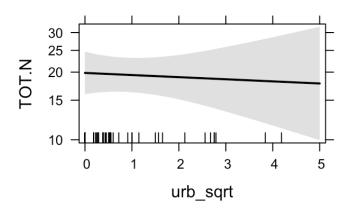
QuasiPoisson-Distributed Model

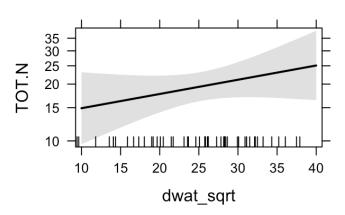


#same as above for negative bionamial
plot(allEffects(negb.urwapa), main = "Negative Binomial Model", col = "darkseagree
n3")

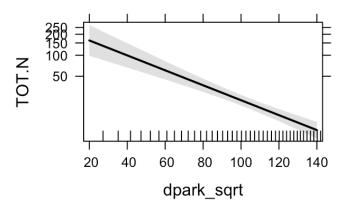
Negative Binomial Model

Negative Binomial Model





Negative Binomial Model



#dev.off()

#The parameter estimates appear to vary greatly amongst model families as well as variables.

How similar are hypothesis test results for the three models? Why do you think these results would differ from what you found for #3?

#This table shows the results from the Anova function for all three variables acro
ss all three models.
#As suggested by the narrowness of the CI for the above plots, D.Park is the only
significant predictor for Tot.n
LRT = cbind(Anova(poi.urwapa)[1],
Anova(qpoi.urwapa)[1],
Anova(negb.urwapa)[1])
names(LRT)[1:3] = c("poisson", "qpoisson", "negbinom")
LRT

```
## poisson qpoisson negbinom
## urb_sqrt 25.806458 2.6031985 0.09092258
## dwat_sqrt 9.327817 0.9409334 1.75216819
## dpark_sqrt 494.233385 49.8552587 67.92663130
```

Let's go back to a model with just TOT.N vs D.PARK. Fit a standard linear regression for this relationship, i.e. with normally distributed error. Plot the data and the fitted relationship.

```
#a standard linear model of the two variables
totpark = lm(TOT.N ~ dpark_sqrt, data = roadkill)
dev.off()
```

```
## null device
## 1
```

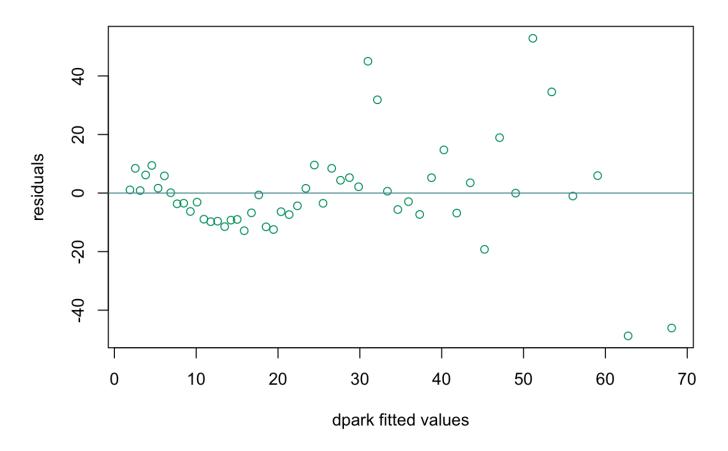
```
par(mfrow = c(2,1))

plot(TOT.N ~ dpark_sqrt, data = roadkill, col = "chartreuse4", main = "Standard LM for Total Kills vs D.Park", xlab = "D.PARK.sqrt", ylab = "Total Kills")
abline(coef= coefficients(totpark), col = "cadetblue4")
```

Plot residuals vs. fitted values. Based on these plots, do you think this model is a good alternative for this data? Explain why you think yes/no.

```
plot(residuals(totpark, type = "response") ~ fitted(totpark), xlab = "dpark f
itted values", ylab = "residuals", col = "seagreen4", main = "Standard LM - residu
als vs fitted values")
abline(h=0, col = "cadetblue4")
```

Standard LM - residuals vs fitted values



#While the general LM seems to fit alright for higher levels of DPark, the residua ls have a "funnel" shape which indicates the variance is not normally distributed. It is likely a bad choice to select this model type while the other tests don't re tain an assumption of normality.