Lab: Logistic Regression for Gene Expression Data

In this lab, we use logistic regression to predict biological characteristics ("phenotypes") from gene expression data. In addition to the concepts in <u>breast cancer demo (./demo04_breast_cancer.ipynb)</u>, you will learn to:

- Handle missing data
- Perform binary classification, and evaluating performance using various metrics
- Perform multi-class logistic classification, and evaluating performance using accuracy and confusion matrix
- Use L1-regularization to promote sparse weights for improved estimation (Grad students only)

Background

Genes are the basic unit in the DNA and encode blueprints for proteins. When proteins are synthesized from a gene, the gene is said to "express". Micro-arrays are devices that measure the expression levels of large numbers of genes in parallel. By finding correlations between expression levels and phenotypes, scientists can identify possible genetic markers for biological characteristics.

The data in this lab comes from:

https://archive.ics.uci.edu/ml/datasets/Mice+Protein+Expression (https://archive.ics.uci.edu/ml/datasets/Mice+Protein+Expression)

In this data, mice were characterized by three properties:

- Whether they had down's syndrome (trisomy) or not
- Whether they were stimulated to learn or not
- Whether they had a drug memantine or a saline control solution.

With these three choices, there are 8 possible classes for each mouse. For each mouse, the expression levels were measured across 77 genes. We will see if the characteristics can be predicted from the gene expression levels. This classification could reveal which genes are potentially involved in Down's syndrome and if drugs and learning have any noticeable effects.

Load the Data

We begin by loading the standard modules.

```
In [121]:
```

```
import pandas as pd
import numpy as np
import matplotlib
import matplotlib.pyplot as plt
%matplotlib inline
from sklearn import linear_model, preprocessing
```

Use the pd.read excel command to read the data from

https://archive.ics.uci.edu/ml/machine-learning-databases/00342/Data Cortex Nuclear.xls (https://archive.ics.uci.edu/ml/machine-learning-databases/00342/Data Cortex Nuclear.xls)

into a dataframe df. Use the index_col option to specify that column 0 is the index. Use the df.head() to print the first few rows.

In [122]:

```
url = "https://archive.ics.uci.edu/ml/machine-learning-databases/00342/Data_Cor
tex_Nuclear.xls"
df = pd.read_excel(url, index_col = 0)
df.head(3)
```

Out[122]:

	DYRK1A_N	ITSN1_N	BDNF_N	NR1_N	NR2A_N	pAKT_N	pBRAF_N	k
MouseID								
309_1	0.503644	0.747193	0.430175	2.816329	5.990152	0.218830	0.177565	2
309_2	0.514617	0.689064	0.411770	2.789514	5.685038	0.211636	0.172817	2
309_3	0.509183	0.730247	0.418309	2.687201	5.622059	0.209011	0.175722	2

3 rows × 81 columns

This data has missing values. The site:

http://pandas.pydata.org/pandas-docs/stable/missing_data.html (http://pandas.pydata.org/pandas-docs/stable/missing_data.html)

has an excellent summary of methods to deal with missing values. Following the techniques there, create a new data frame df1 where the missing values in each column are filled with the mean values from the non-missing values.

```
In [123]:
```

```
df1 = df
df1.iloc[0, 0] = np.nan # test
df1.head(1)
```

Out[123]:

	DYRK1A_N	ITSN1_N	BDNF_N	NR1_N	NR2A_N	pAKT_N	pBRAF_N	р
MouseID								
309_1	NaN	0.747193	0.430175	2.816329	5.990152	0.21883	0.177565	2

1 rows × 81 columns

```
In [124]:
```

```
df1 = df1.fillna(df1.mean())
df1.head(1)
```

Out[124]:

	DYRK1A_N	ITSN1_N	BDNF_N	NR1_N	NR2A_N	pAKT_N	pBRAF_N	р
MouseID								
309_1	0.425738	0.747193	0.430175	2.816329	5.990152	0.21883	0.177565	2

1 rows × 81 columns

Binary Classification for Down's Syndrome

We will first predict the binary class label in df1['Genotype'] which indicates if the mouse has Down's syndrome or not. Get the string values in df1['Genotype'].values and convert this to a numeric vector y with 0 or 1. You may wish to use the np.unique command with the return_inverse=True option.

```
In [125]:
```

```
val = df1["Genotype"].values
print(val)
labels, y = np.unique(val, return_inverse = True)
print(labels)
print(y)

['Control' 'Control' ... 'Ts65Dn' 'Ts65Dn' 'Ts65Dn']
```

As predictors, get all but the last four columns of the dataframes. Standardize the data matrix and call the standardized matrix xs. The predictors are the expression levels of the 77 genes.

```
In [126]:
```

['Control' 'Ts65Dn']
[0 0 0 ... 1 1 1]

```
Xs = df1.iloc[:,:-4]
Xs = preprocessing.scale(Xs)
```

Adding a cell here to split the Xs data into training and test.. without doing this, the acc is output as 1.0 because they're the same:

```
In [127]:
```

```
from sklearn.model_selection import train_test_split
Xtr, Xts, ytr, yts = train_test_split(Xs, y, test_size=0.3, random_state=98)
```

Create a LogisticRegression object logreg and fit the training data. Use C = 1e5.

```
In [128]:
logreg = linear_model.LogisticRegression(C=1e5)
logreg.fit(Xtr, ytr)

/Users/mkarroqe/anaconda3/lib/python3.5/site-packages/sklearn/linea
r_model/logistic.py:432: FutureWarning: Default solver will be chan
ged to 'lbfgs' in 0.22. Specify a solver to silence this warning.
    FutureWarning)
```

```
LogisticRegression(C=100000.0, class_weight=None, dual=False, fit_intercept=True, intercept_scaling=1, l1_rati o=None, max_iter=100, multi_class='warn', n_jobs=None, p enalty='12', random_state=None, solver='warn', tol=0.0001, ve rbose=0, warm_start=False)
```

Measure the accuracy of the classifer. That is, use the logreg.predict function to predict labels yhat and measure the fraction of time that the predictions match the true labels.

```
In [129]:
```

Out[128]:

```
yhat = logreg.predict(Xts)
acc = np.mean(yhat == yts)
print("Accuracy on training data =", acc)
```

Accuracy on training data = 0.9567901234567902

Also, plot the ROC curve, and measure the AUC. Later, we will properly measure the accuracy and AUC on cross-validation data.

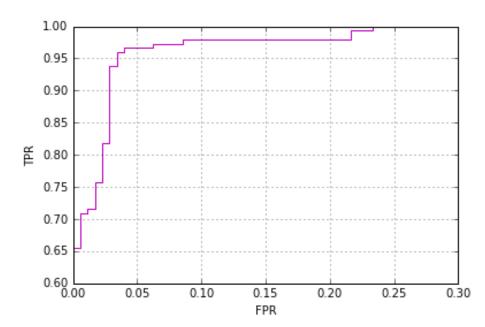
```
In [130]:
```

```
from sklearn import metrics
yprob = logreg.predict_proba(Xts)
fpr, tpr, thresholds = metrics.roc_curve(yts, yprob[:,1])

plt.plot(fpr, tpr, color="m")
plt.grid()
plt.xlabel('FPR')
plt.ylabel('TPR')
plt.ylim([0.6,1])
plt.xlim([0,0.3])
```

Out[130]:

(0, 0.3)

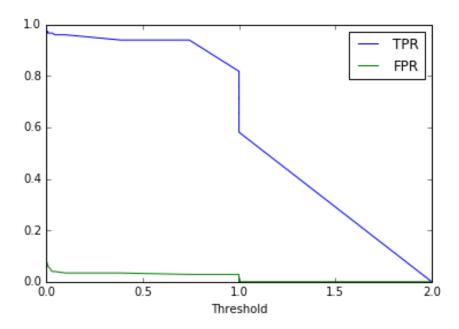


```
In [131]:
```

```
plt.plot(thresholds,tpr, thresholds,fpr)
plt.legend(['TPR','FPR'])
plt.xlabel('Threshold')
```

Out[131]:

<matplotlib.text.Text at 0x12d654128>



```
In [132]:
```

```
auc=metrics.roc_auc_score(yts, yprob[:,1])
print("AUC=%f" % auc)
```

AUC=0.987638

Interpreting the weight vector

Create a stem plot of the coefficients, w in the logistic regression model. You can get the coefficients from logreg.coef_, but you will need to reshape this to a 1D array.

In [133]:

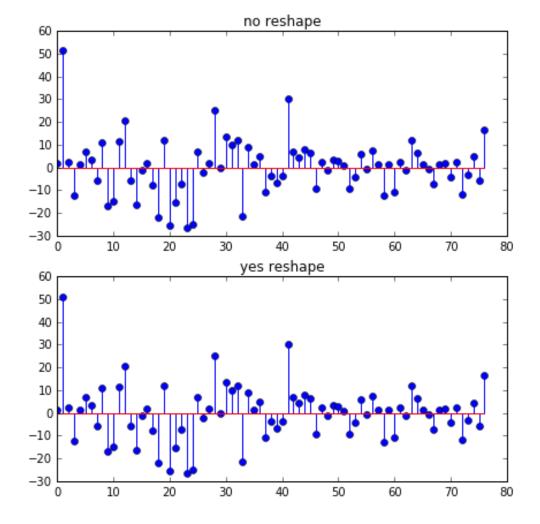
```
W = logreg.coef_
plt.figure(figsize=(7,7))
plt.subplot(2,1,1)
plt.stem(W[0,:])
plt.title("no reshape")

# i played with these values just for myself
plt.subplot(2,1,2)
W = np.reshape(W, 77)
plt.stem(W)
plt.title("yes reshape")

print("these weights show how much the gene matters/is likely to predict down s
yndrome:\n\n W =", W)
```

these weights show how much the gene matters/is likely to predict d own syndrome:

```
1.56920307 51.25417374
                                2.35718544 -12.59994686
                                                            1.47554
M = [
472
   6.98046645
                3.1810072
                            -5.55816374
                                         11.10597539 -16.73222621
-14.72594373
               11.52109419
                            20.41844148
                                        -5.68986468 -16.55929192
                            -7.96143771 -21.78201221
                                                      11.89288996
 -1.03235017
                1.72334188
-25.78000703 -15.48344403
                            -7.30060119 -26.3735751
                                                     -24.90179814
   7.03863687
               -1.99639584
                            1.67007869
                                        25.14859193
                                                     -0.31688052
 13.46447767
               9.69653206
                            12.10526872 -21.62425341
                                                      9.06745152
               4.74191849 -10.65368426
                                        -3.92097904
                                                      -6.58262613
   1.29137533
 -3.6858914
               30.11483819
                            6.95531989
                                         4.5126539
                                                      7.98480837
   6.26325393
               -9.30433885
                           2.54182824
                                        -1.14865016
                                                      3.55476871
                                        -4.05036161
  2.83210424
                0.58920647 -9.26972464
                                                      5.66086053
 -0.57565534
               7.62901951
                            1.29331703 -12.6189467
                                                       1.2803133
-10.631795
               2.50166368
                            -1.27976939
                                        11.93507255
                                                      6.5067559
   1.31455682
              -0.84614559
                           -7.47237798
                                         1.22851759
                                                      1.68667705
 -4.05036161
               2.12898461 -11.87222397
                                        -3.13804444
                                                      4.61942789
 -5.94767934
               16.45663459]
```



You should see that W[i] is very large for a few components i. These are the genes that are likely to be most involved in Down's Syndrome.

Find the names of the genes for two components i where the magnitude of W[i] is largest.

```
In [135]:

Ws = np.argsort(W)
genes = df1.keys()
max2 = genes[Ws[-2:]]
print("The two top predictors are:")
for i in max2:
    print(i)
```

```
The two top predictors are: TIAM1_N
ITSN1_N
```

Cross Validation

The above meaured the accuracy on the training data. It is more accurate to measure the accuracy on the test data. Perform 10-fold cross validation and measure the average precision, recall and f1-score, as well as the AUC. Note, that in performing the cross-validation, you will want to randomly permute the test and training sets using the shuffle option. In this data set, all the samples from each class are bunched together, so shuffling is essential. Print the mean precision, recall and f1-score and error rate across all the folds.

```
In [148]:
```

```
from sklearn.model_selection import KFold
from sklearn.metrics import precision recall fscore support
nfold = 10
kf = KFold(n splits=nfold, shuffle=True)
prec = []
rec = []
f1 = []
acc = []
auc = []
for train, test in kf.split(Xs):
    # Get training and test data
    Xtr = Xs[train,:]
    ytr = y[train]
    Xts = Xs[test,:]
    yts = y[test]
    # Fit a model
    logreg.fit(Xtr, ytr)
    yhat = logreg.predict(Xts)
    # Measure performance
    preci,reci,fli, = precision recall fscore support(yts,yhat,average='binary'
)
    prec.append(preci)
    rec.append(reci)
    fl.append(fli)
    acci = np.mean(yhat == yts)
    acc.append(acci)
    # AUC
    yprob = logreg.predict proba(Xts)
    auci = metrics.roc_auc_score(yts, yprob[:,1])
    auc.append(auci)
# Take average values of the metrics
precm = np.mean(prec)
recm = np.mean(rec)
f1m = np.mean(f1)
accm = np.mean(acc)
aucm = np.mean(auc)
# Compute the standard errors
prec se = np.std(prec)/np.sqrt(nfold-1)
rec se = np.std(rec)/np.sqrt(nfold-1)
f1 se = np.std(f1)/np.sqrt(nfold-1)
acc se = np.std(acc)/np.sqrt(nfold-1)
auc_se = np.std(auc)/np.sqrt(nfold-1)
```

```
/Users/mkarroqe/anaconda3/lib/python3.5/site-packages/sklearn/linea r_model/logistic.py:432: FutureWarning: Default solver will be chan ged to 'lbfgs' in 0.22. Specify a solver to silence this warning. FutureWarning)
```

/Users/mkarroqe/anaconda3/lib/python3.5/site-packages/sklearn/linea r_model/logistic.py:432: FutureWarning: Default solver will be chan ged to 'lbfgs' in 0.22. Specify a solver to silence this warning. FutureWarning)

/Users/mkarroqe/anaconda3/lib/python3.5/site-packages/sklearn/linea r_model/logistic.py:432: FutureWarning: Default solver will be chan ged to 'lbfgs' in 0.22. Specify a solver to silence this warning. FutureWarning)

/Users/mkarroqe/anaconda3/lib/python3.5/site-packages/sklearn/linea r_model/logistic.py:432: FutureWarning: Default solver will be chan ged to 'lbfgs' in 0.22. Specify a solver to silence this warning. FutureWarning)

/Users/mkarroqe/anaconda3/lib/python3.5/site-packages/sklearn/linea r_model/logistic.py:432: FutureWarning: Default solver will be chan ged to 'lbfgs' in 0.22. Specify a solver to silence this warning. FutureWarning)

/Users/mkarroqe/anaconda3/lib/python3.5/site-packages/sklearn/linea r_model/logistic.py:432: FutureWarning: Default solver will be chan ged to 'lbfgs' in 0.22. Specify a solver to silence this warning. FutureWarning)

/Users/mkarroqe/anaconda3/lib/python3.5/site-packages/sklearn/linea r_model/logistic.py:432: FutureWarning: Default solver will be chan ged to 'lbfgs' in 0.22. Specify a solver to silence this warning. FutureWarning)

/Users/mkarroqe/anaconda3/lib/python3.5/site-packages/sklearn/linea r_model/logistic.py:432: FutureWarning: Default solver will be chan ged to 'lbfgs' in 0.22. Specify a solver to silence this warning. FutureWarning)

/Users/mkarroqe/anaconda3/lib/python3.5/site-packages/sklearn/linea r_model/logistic.py:432: FutureWarning: Default solver will be chan ged to 'lbfgs' in 0.22. Specify a solver to silence this warning. FutureWarning)

/Users/mkarroqe/anaconda3/lib/python3.5/site-packages/sklearn/linea r_model/logistic.py:432: FutureWarning: Default solver will be chan ged to 'lbfgs' in 0.22. Specify a solver to silence this warning. FutureWarning)

```
In [151]:
```

```
print('Precision = {0:.4f}, SE={1:.4f}'.format(precm,prec_se))
print('Recall = {0:.4f}, SE={1:.4f}'.format(recm, rec_se))
print('f1 = {0:.4f}, SE={1:.4f}'.format(f1m, f1_se))
print('Accuracy = {0:.4f}, SE={1:.4f}'.format(accm, acc_se))
print('AUC = {0:.4f}, SE={1:.4f}'.format(aucm, auc_se))
```

```
Precision = 0.9532, SE=0.0084

Recall = 0.9489, SE=0.0090

f1 = 0.9509, SE=0.0080

Accuracy = 0.9537, SE=0.0076

AUC = 0.9884, SE=0.0027
```

Multi-Class Classification

Now use the response variable in df1['class']. This has 8 possible classes. Use the np.unique funtion as before to convert this to a vector y with values 0 to 7.

```
In [158]:
```

```
labels, y = np.unique(df1["class"], return_inverse = True)
print(labels)
print(y) # mapping
```

```
['c-CS-m' 'c-CS-s' 'c-SC-m' 'c-SC-s' 't-CS-m' 't-CS-s' 't-SC-m' 't-SC-s']
[0 0 0 ... 7 7 7]
```

Fit a multi-class logistic model by creating a LogisticRegression object, logreg and then calling the logreg.fit method. In general, you could either use the 'one over rest (ovr)' option or the 'multinomial' option. In this exercise use the default 'ovr' and C=1. As an optional exercise, you could also compare the results obtained with these two options.

```
In [171]:

Xtr, Xts, ytr, yts = train_test_split(Xs, y, test_size=0.3, random_state=98)
logreg = linear_model.LogisticRegression(multi_class="ovr", C=1)
logreg.fit(Xtr, ytr)
```

Measure the accuracy on the training data.

```
In [173]:
```

```
yhat = logreg.predict(Xts)
acc = np.mean(yhat == yts)
print("Accuracy on training data = %f" % acc)
```

Accuracy on training data = 0.984568

Now perform 10-fold cross validation, and measure the confusion matrix C on the test data in each fold. You can use the confustion_matrix method in the sklearn package. Add the confusion matrix counts across all folds and then normalize the rows of the confusion matrix so that they sum to one. Thus, each element C[i,j] will represent the fraction of samples where yhat==j given ytrue==i. Print the confusion matrix. You can use the command

```
print(np.array str(C, precision=4, suppress small=True))
```

to create a nicely formatted print. Also print the overall mean and SE of the test accuracy across the folds.

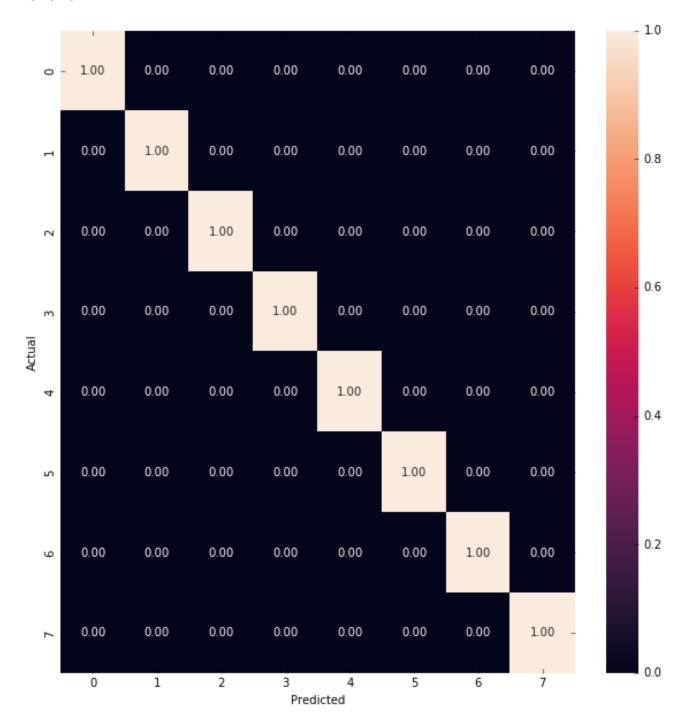
```
In [188]:
```

```
import warnings
warnings.filterwarnings('ignore') # :-)
from sklearn.metrics import confusion matrix
from sklearn.model_selection import KFold
import seaborn as sns
npen = 20
C test = np.logspace(-2, 2, npen)
# Create the cross-validation object and error rate matrix
nfold = 10
kf = KFold(n splits=nfold,shuffle=True)
err rate = np.zeros((npen,nfold))
num nonzerocoef = np.zeros((npen,nfold))
# Create the logistic regression object
logreg = linear model.LogisticRegression(penalty='11',warm start=True)
# Loop over the folds in the cross-validation
for ifold, Ind in enumerate(kf.split(Xs)):
    # Get training and test data
    Itr, Its = Ind
    Xtr = Xs[Itr,:]
    ytr = y[Itr]
    Xts = Xs[Its,:]
    yts = y[Its]
    # Loop over penalty levels
    for ipen, c in enumerate(C test):
        # Set the penalty level
        logreg.C= c
        # Fit a model on the training data
        logreg.fit(Xtr, ytr)
        # Predict the labels on the test set.
        yhat = logreg.predict(Xts)
        # Measure the accuracy
        err rate[ipen,ifold] = np.mean(yhat != yts)
        num nonzerocoef[ipen,ifold]=np.sum(abs(logreg.coef )>0.001)
    print("Fold %d" % ifold)
    # s/o to https://stackoverflow.com/a/57452776 for cool viz
    C = preprocessing.normalize(confusion matrix(yts, yhat))
    # normalizes:
    cmn = C.astype('float') / C.sum(axis=1)[:, np.newaxis]
```

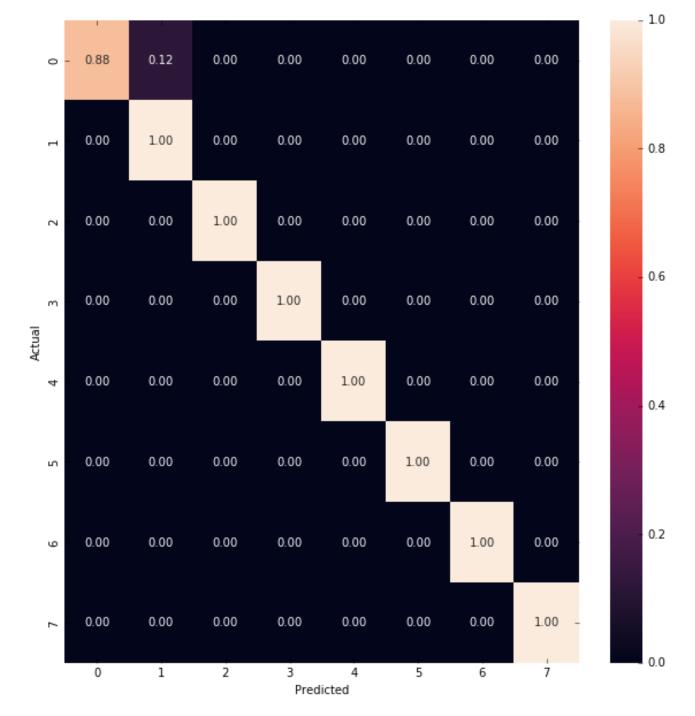
```
fig, ax = plt.subplots(figsize=(10,10))
    sns.heatmap(cmn, annot=True, fmt='.2f')#, xticklabels=target_names, ytickla
bels=target_names)
    plt.ylabel('Actual')
    plt.xlabel('Predicted')
    plt.show(block=False)

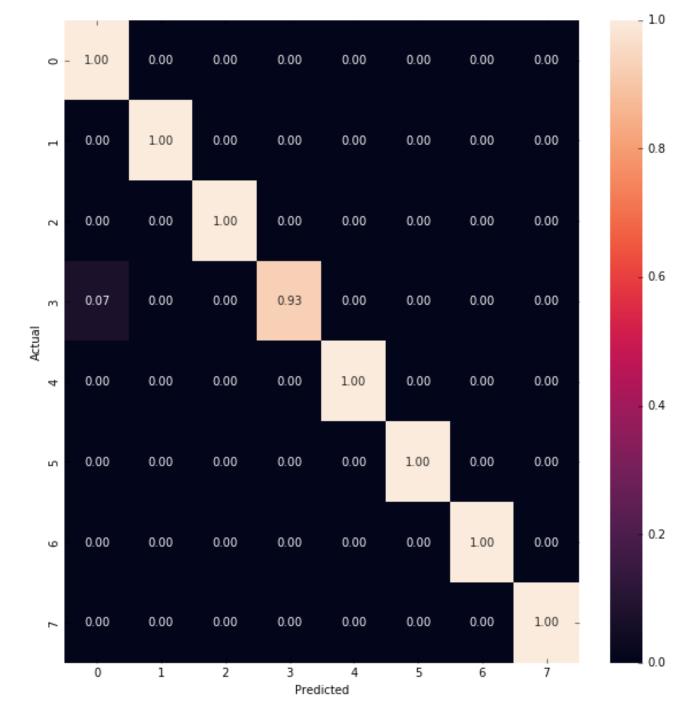
# print(np.array_str(C, precision=4, suppress_small=True))

err_mean = np.mean(err_rate, axis=1)
num_nonzerocoef_mean = np.mean(num_nonzerocoef, axis=1)
err_se = np.std(err_rate,axis=1)/np.sqrt(nfold-1)
imin = np.argmin(err_mean)
```

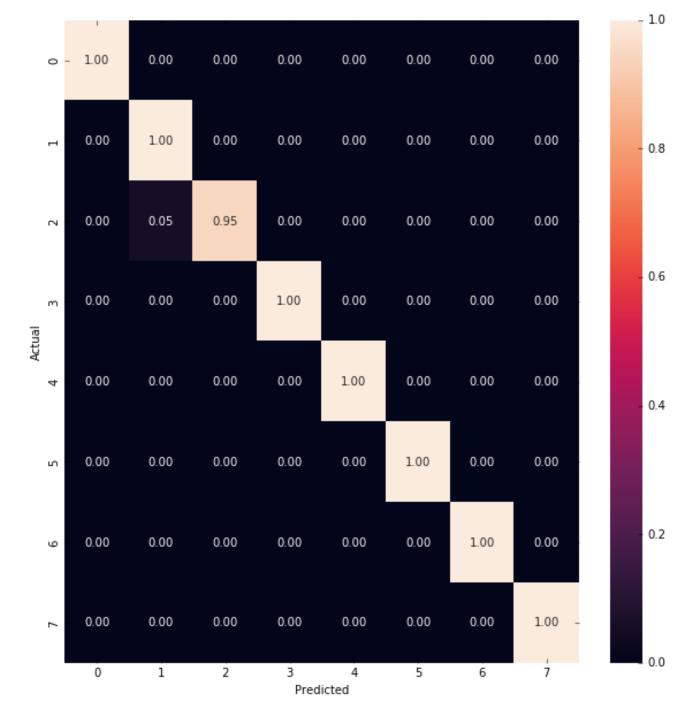


Fold 1

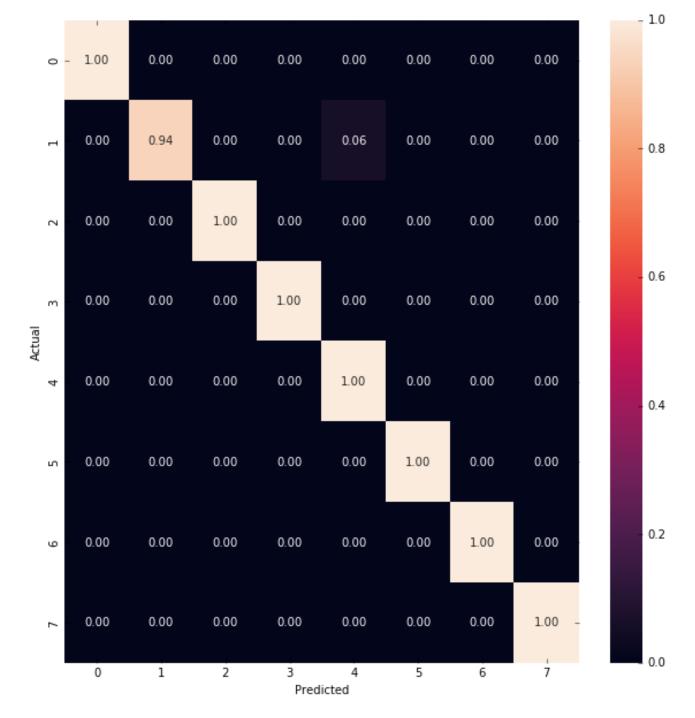




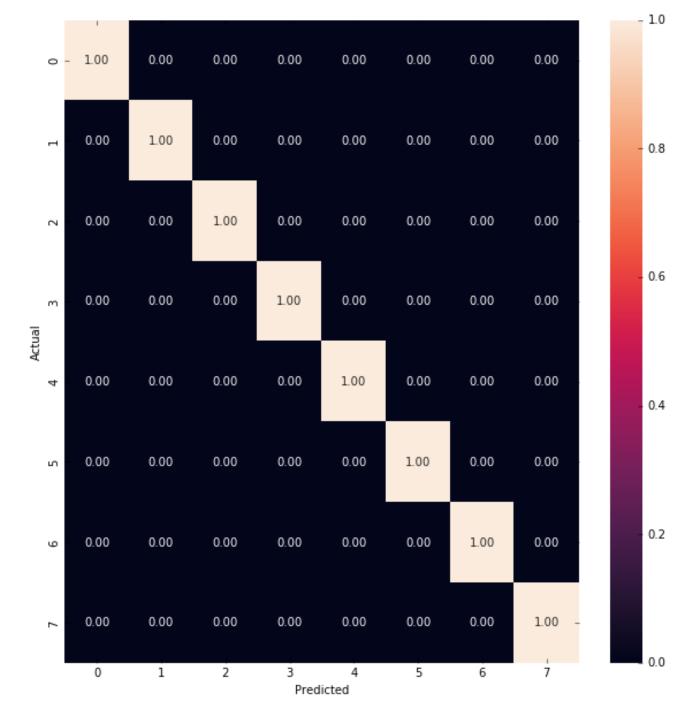
Fold 3

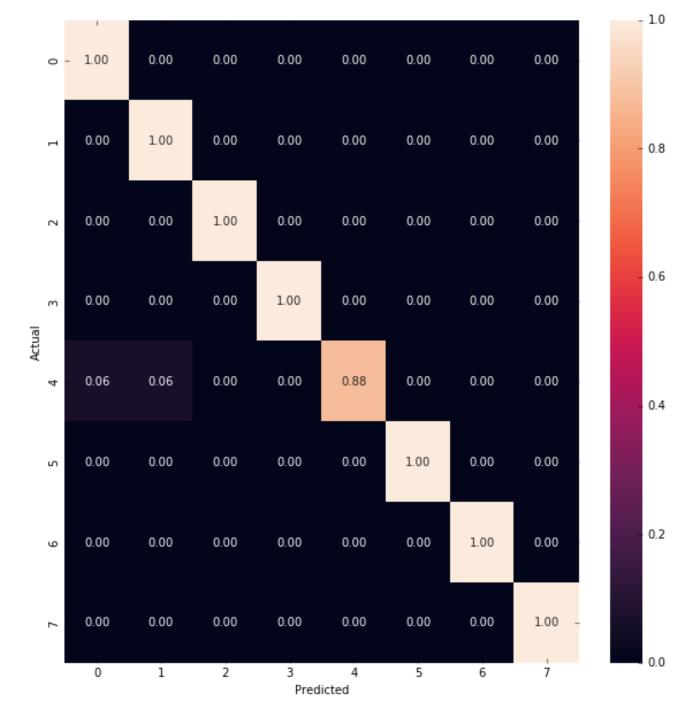


Fold 4

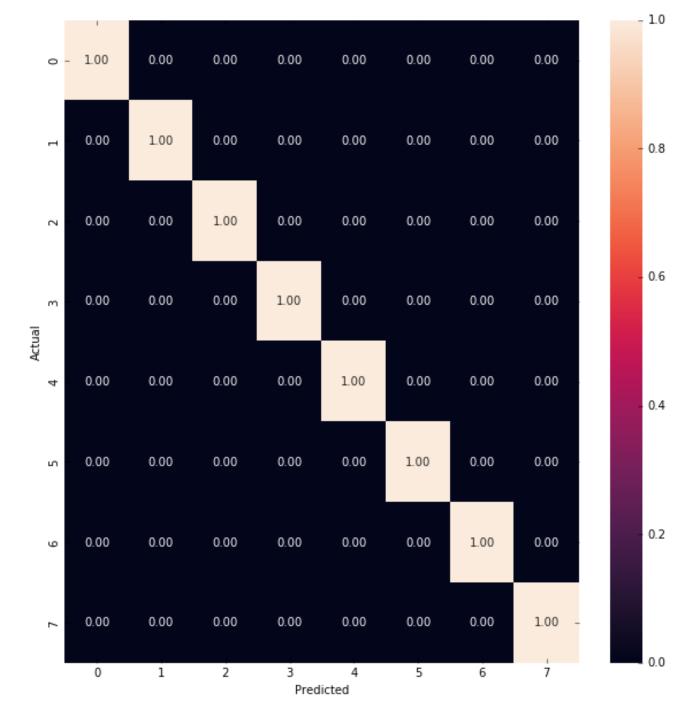


Fold 5

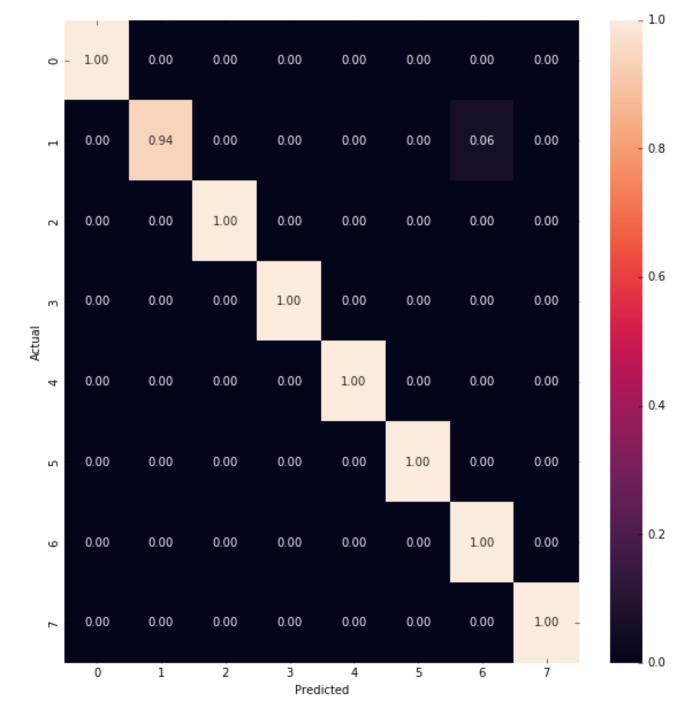




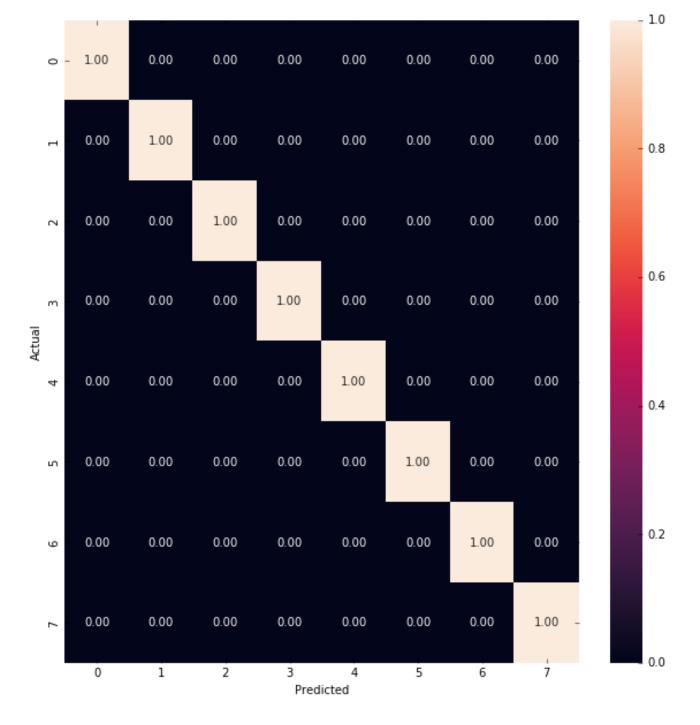
Fold 7



Fold 8



Fold 9



In [183]:

```
print("The minimum test error rate = %12.4e, SE=%12.4e" % (err_mean[imin], err_
se[imin]))
print("The C value corresponding to minimum error = %12.4e" % (C_test[imin]))
```

```
The minimum test error rate = 9.2593e-03, SE= 3.0864e-03
The C value corresponding to minimum error = 6.1585e+01
```

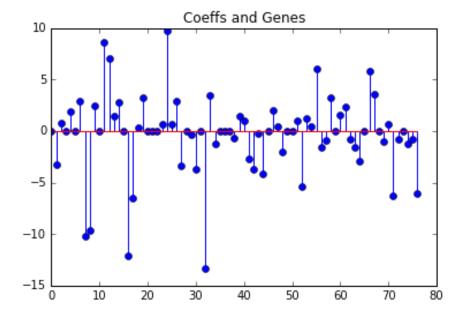
Re-run the logistic regression on the entire training data and get the weight coefficients. This should be a 8 x 77 matrix. Create a stem plot of the first row of this matrix to see the coefficients on each of the genes for the first class.

In [189]:

```
logreg.fit(Xs, y)
W = logreg.coef_
plt.title("Coeffs and Genes")
plt.stem(W[0,:])
```

Out[189]:

<Container object of 3 artists>



L1-Regularization

Graduate students only complete this section. :-)

In most genetic problems, only a limited number of the tested genes are likely influence any particular attribute. Hence, we would expect that the weight coefficients in the logistic regression model should be sparse. That is, they should be zero on any gene that plays no role in the particular attribute of interest. Genetic analysis commonly imposes sparsity by adding an I1-penalty term. Read the sklearn documentation (http://scikit-

<u>learn.org/stable/modules/generated/sklearn.linear_model.LogisticRegression.html)</u> on the LogisticRegression class to see how to set the I1-penalty and the inverse regularization strength, C.

Using the model selection strategies from the <u>prostate cancer analysis demo</u> (../unit03 model sel/demo03 2 prostate.ipynb), use K-fold cross validation to select an appropriate inverse regularization strength.

- Use 10-fold cross validation
- You should select around 20 values of C. It is up to you to find a good range.
- For each C and each fold, you should compute the classification error rate
- For each C and each fold, you should also determine the nubmer of non-zero coefficients for the first class. For this purpse, you can assume coefficient with magnitude <0.01 as zero.

```
In [16]:
# TODO
```

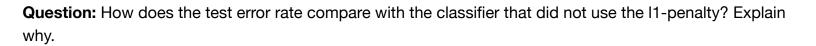
Now compute the mean and standard error on the error rate for each C and plot the results (Use errorbar() method). Also determine and print the minimum test error rate and corresponding C value.

```
In [ ]:
# TODO
```

We see that the minimum error rate is significantly below the classifier that did not use the I1-penalty. Use the one-standard error rule to determine the optimal C and the corresponding test error rate. Note that because C is inversely proportional to the regularization strength, you want to select a C as *small* as possible while meeting the error target!

```
In [17]:

# TODO
# C_opt =
```



Type Answer Here:

Now plot the nubmer of non-zero coefficients for the first class for different C values. Also determine and print the number of non-zero coefficients corresponding to C_opt.

In [18]:
TODO

For the optimal C, fit the model on the entire training data with I1 regularization. Find the resulting weight matrix, W_11 . Plot the first row of this weight matrix and compare it to the first row of the weight matrix without the regularization. You should see that, with I1-regularization, the weight matrix is much more sparse and hence the roles of particular genes are more clearly visible. Please also compare the accuracy for the training data using optimal C with the previous results not using LASSO regularization. Do you expect the accuracy to improve?

In [19]:
TODO
In []: