

Potential for climate-induced disruption of plant-fungal symbioses in the Rocky Mountains

Melanie Kazenel

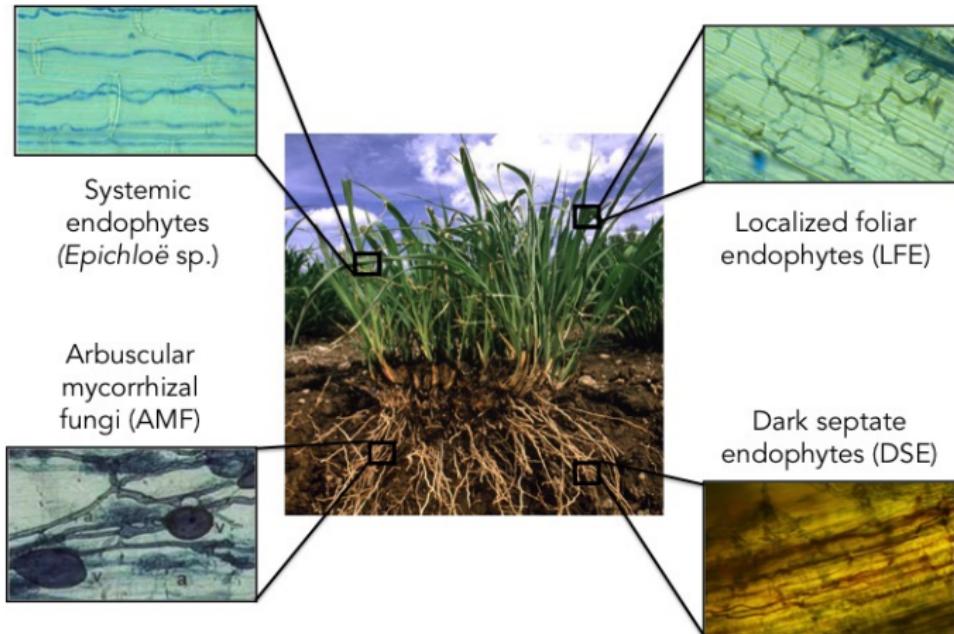
7 April 2016





How will climate change alter plant-symbiont interactions?

Plants and Fungal Symbionts



Symbionts can mediate plant responses to climate change



American Journal of Botany 100(7): 1445–1457. 2013.

SPECIAL INVITED PAPER—GLOBAL BIOLOGICAL CHANGE

FUNGAL Symbionts ALTER PLANT RESPONSES TO GLOBAL CHANGE¹

STEPHANIE N. KIVLIN^{2,5}, SARAH M. EMERY³, AND JENNIFER A. RUDGERS⁴

Symbionts altered plant responses to drought, N deposition, and warming

Climate change may disrupt symbioses as organisms experience range shifts

SCIENCE VOL 336 20 APRIL 2012

Recent Plant Diversity Changes on Europe's Mountain Summits

Harald Pauli,^{2*} Michael Gottfried,^{2†} Stefan Dullinger,^{2,3*} Otari Abdaladze,⁴ Maia Akhalkatsi,⁴ José Luis Benito Alonso,⁵ Gheorghe Coldea,⁶ Jan Dick,⁷ Brigitte Erschbamer,⁸ Rosa Fernández Calzado,⁷ Dany Ghosh,¹⁰ Jarle I. Holten,¹¹ Robert Kanka,¹² George Kazakis,¹⁰ Jozef Kollár,¹² Per Larsson,¹³ Pavel Moiseev,¹⁴ Dmitry Moiseev,¹⁴ Ulf Molau,¹³ Joaquín Molero Mesa,⁹ László Nagy,^{15,16} Giovanni Pelino,¹⁷ Mihai Puscas,¹⁸ Graziano Rossi,¹⁹ Angela Stanisci,²⁷ Anne O. Syverhuset,¹¹ Jean-Paul Theurillat,^{20,21} Marcello Tomaselli,²² Peter Unterluggauer,⁸ Luis Villar,²³ Pascal Vittoz,²⁵ Georg Grabbherr¹

nature
climate change

LETTERS

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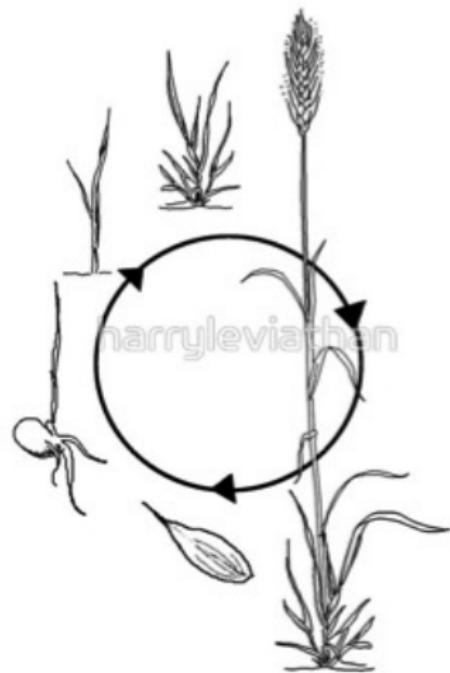
Continent-wide response of mountain vegetation to climate change

Michael Gottfried¹, Harald Pauli^{2*}, Andreas Futschik³, Maia Akhalkatsi⁴, Peter Barančok⁵, José Luis Benito Alonso⁶, Gheorghe Coldea⁷, Jan Dick⁸, Brigitte Erschbamer⁹, María Rosa Fernández Calzado¹⁰, George Kazakis¹¹, Ján Kraječí¹², Per Larsson¹³, Martin Mallau¹³, Ottar Michelsen¹⁴, Dmitry Moiseev¹⁵, Pavel Moiseev¹⁶, Ulf Molau¹⁶, Abderrahmane Merzouki¹⁶, László Nagy^{17,18}, George Nakutnirishvili¹⁹, Bård Pedersen²⁰, Giovanni Pelino²¹, Mihai Puscas²², Graziano Rossi²³, Angela Stanisci²⁷, Jean-Paul Theurillat^{24,25}, Marcello Tomaselli²⁶, Luis Villar⁶, Pascal Vittoz²⁷, Ioannis Vogiatzakis²⁸ and Georg Grabbherr¹

Mechanisms for disruption of plant-symbiont interactions

Plants and symbionts may have different:

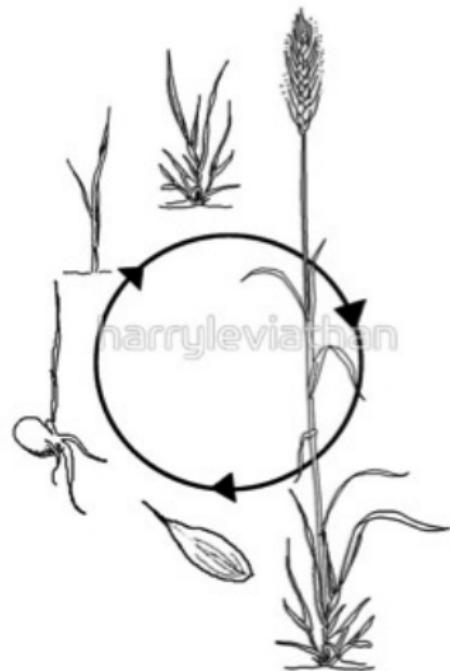
- ▶ Physiological tolerances



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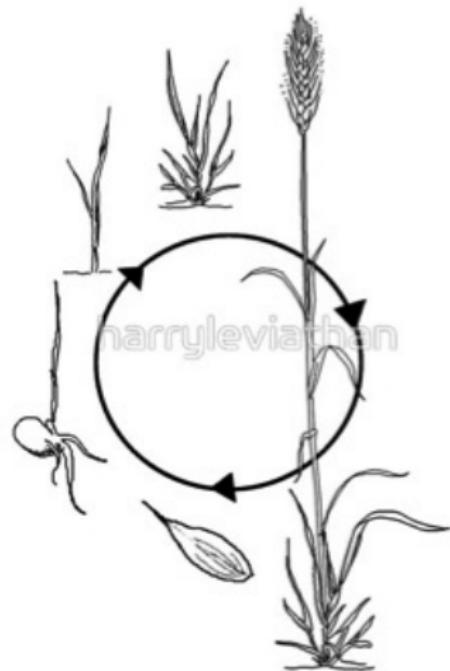
- ▶ Physiological tolerances
- ▶ Dispersal rates



Mechanisms for disruption of plant-symbiont interactions

Plants and symbionts may have different:

- ▶ Physiological tolerances
- ▶ Dispersal rates
- ▶ Phenological responses



Study System



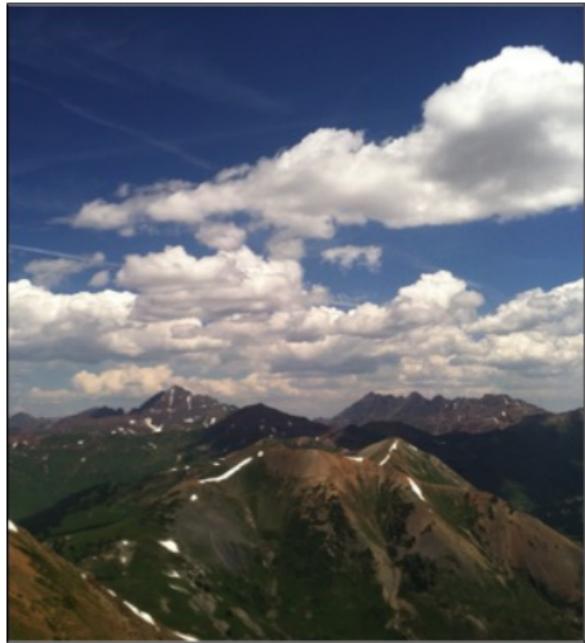
Mountains

- ~25% of land area on Earth
- 50% of the human water supply
- 1/3 of terrestrial plant diversity

Grasses

- Cover 1/3 of land area (>10,000 species)
- Provide the majority of food for humans and domesticated animals
- All have mycorrhizal fungi in roots and fungi in leaves

Altitudinal Gradients and Experimental Warming



Focal Questions

How do symbionts change with altitude and warming?

- ▶ Altitude response?

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- ▶ Are they the same?

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Warming Experiment

Rocky Mountain Biological Laboratory

- ▶ Established in 1991



Photo: Mary Ellen
Harte

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- ▶ Warms top 15 cm of soil by $\sim 2^{\circ}\text{C}$



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- ▶ Dries soil by 10-20%



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Warming Experiment

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- ▶ Established in 1991
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- ▶ Dries soil by 10-20%
- ▶ Extends growing season by ~12 days on each end



Photo: Mary Ellen
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Study Species

*Achnatherum
lettermanii*
(ACLE)



*Festuca
thurberi*
(FETH)

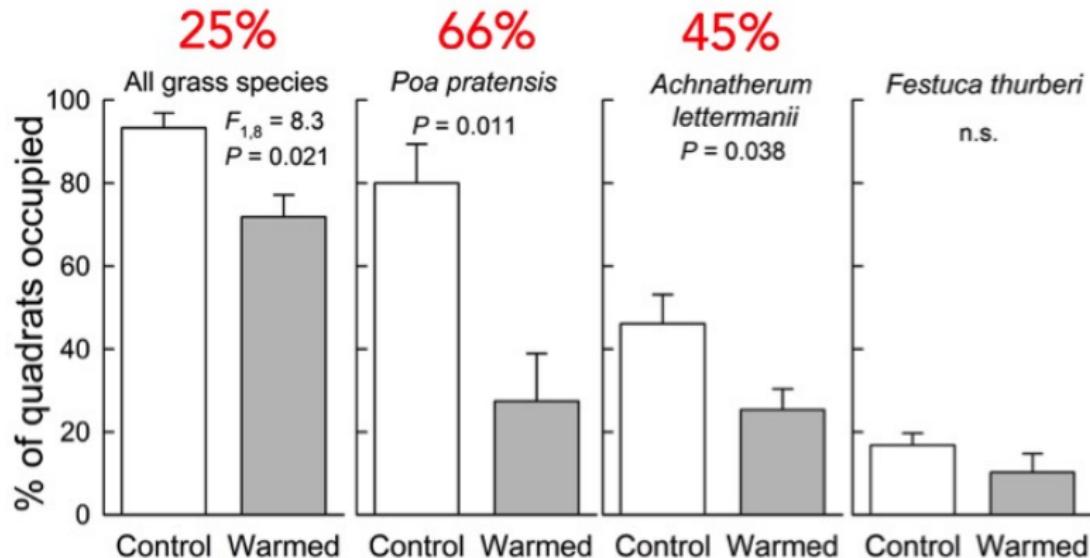


*Poa
pratensis*
(POPR)



ÅNGSTRÖM, POA PRATENSIS L.

Experimental warming reduced grasses (1991 – 2011)



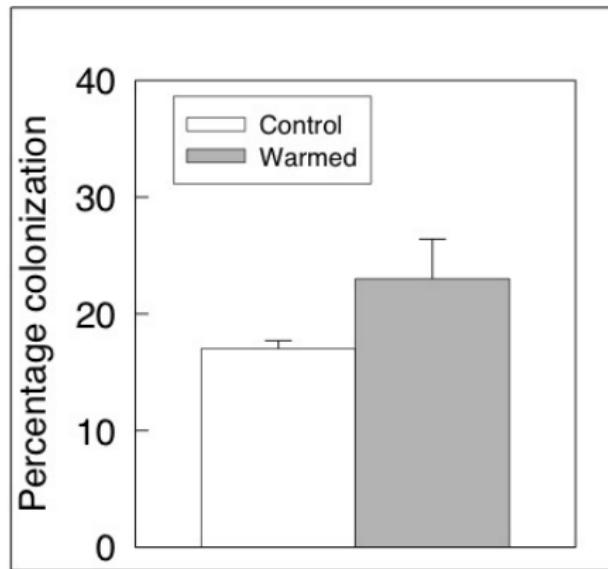
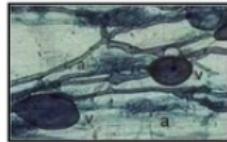
Mean % \pm s.e. of 49 (0.2m \times 0.2m) quadrats surveyed per plot. $n = 5$ plots per warming treatment. Rudgers et al. *Ecology* (2014)

Experimental warming increased mycorrhizal colonization of roots



A. lettermanii

Arbuscular
mycorrhizal fungi



Rudgers et al. *Ecology* (2014).

Field collection methods

- ▶ 3 focal grass species:



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 - ▶ *Achnatherum lettermanii*



Photo: Noah Whiteman

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 - ▶ *Festuca thuerberi*



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Field collection methods

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 - ▶ *Festuca thuerberi*
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- ▶ 6 individual plants collected per species per plot



Photo: Noah Whiteman

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- ▶ Roots and leaves (2014)



Photo: Noah Whiteman

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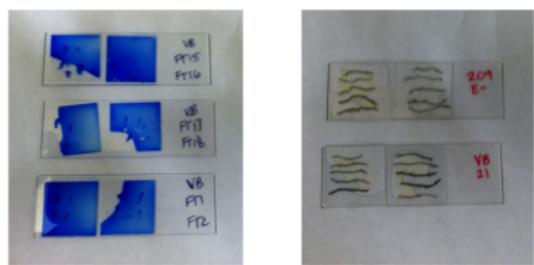
- ▶ 3 focal grass species:
 - ▶ *Achnatherum lettermanii*
 - ▶ *Festuca thuerberi*
 - ▶ *Poa pratensis*
- ▶ 6 individual plants collected per species per plot
- ▶ Roots and leaves (2014)
- ▶ Phenology: June and September



Photo: Noah Whiteman

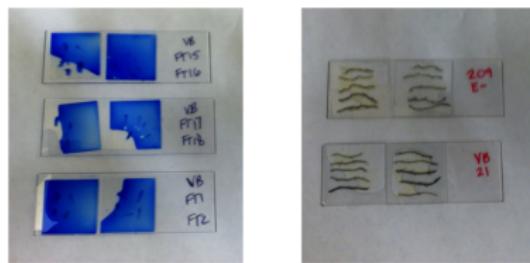
Laboratory methods

- ▶ Staining and microscopy → colonization



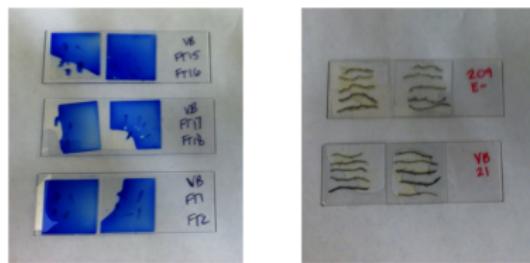
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- ▶ Illumina MiSeq DNA sequencing → composition



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Illumina Sequencing

Paired-end sequencing of fungal nuclear ribosomal DNA using primers targeting:

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Paired-end sequencing of fungal nuclear ribosomal DNA using primers targeting:

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- ▶ ~300bp in the 28S region (for AMF) (FLR3-FLR4 primers)



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- ▶ Conducted analyses on 802 OTUs

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- ▶ Random effect: block (pair of plots)

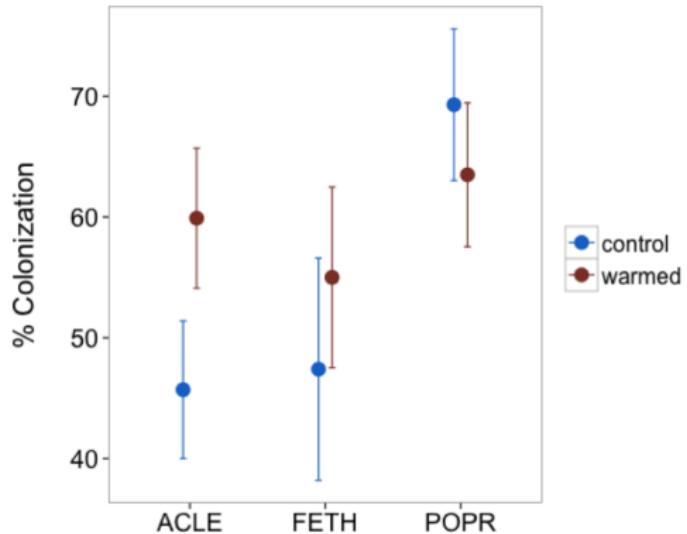
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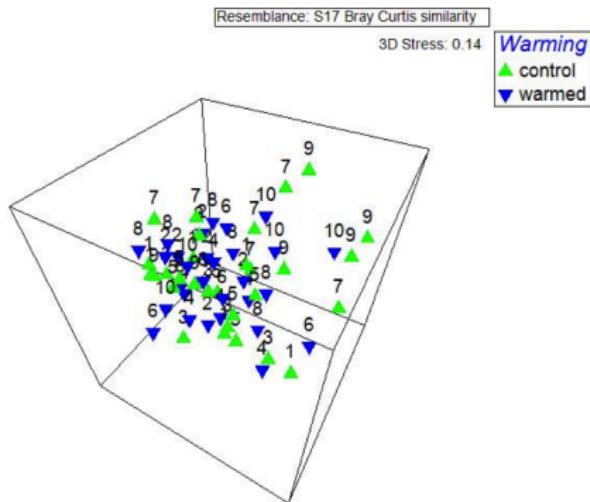
- ▶ NMDS: to visualize OTU composition
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- ▶ Fixed effects: warming treatment, host species, sampling date
- ▶ Random effect: block (pair of plots)
- ▶ PERMDISP: to test for dispersion within groups
- ▶ Indicator species analysis (SIMPER): to identify OTUs that contributed strongly to differences among groups

Arbuscular mycorrhizal fungi



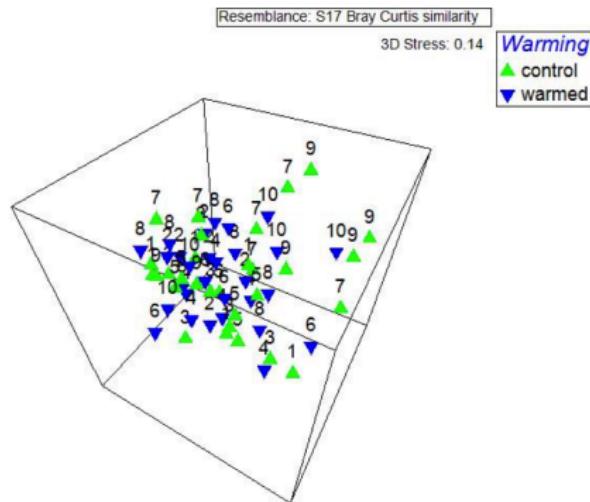
Results: All Species

- ▶ OTU composition did not differ between warming treatments ($\text{df} = 1$, $\text{pseudo-F} = 1.361$, $P = 0.1391$)



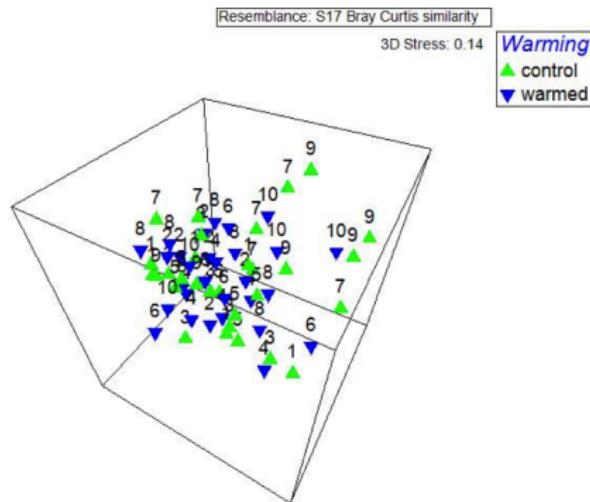
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- ▶ OTU composition did not differ between warming treatments ($\text{df} = 1$, $\text{pseudo-F} = 1.361$, $P = 0.1391$)
- ▶ High stress value

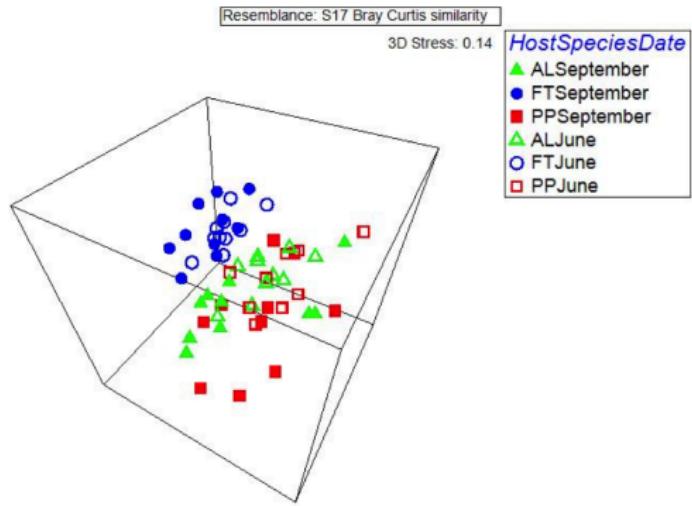


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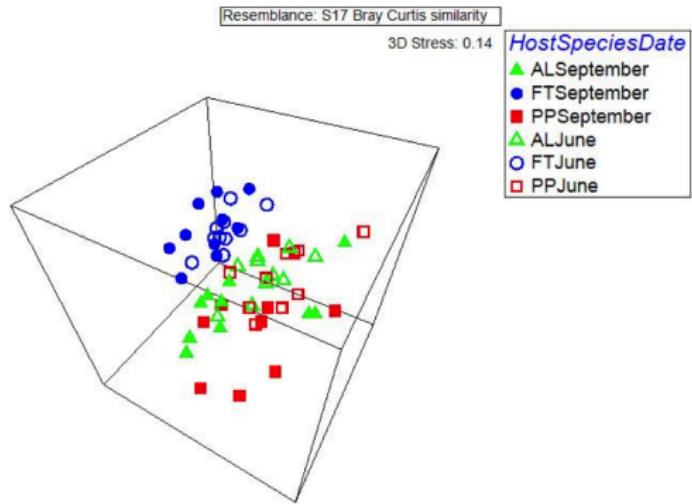
- ▶ OTU composition did not differ between warming treatments ($\text{df} = 1$, $\text{pseudo-F} = 1.361$, $P = 0.1391$)
- ▶ High stress value
- ▶ Spatial heterogeneity (significant effect of block)



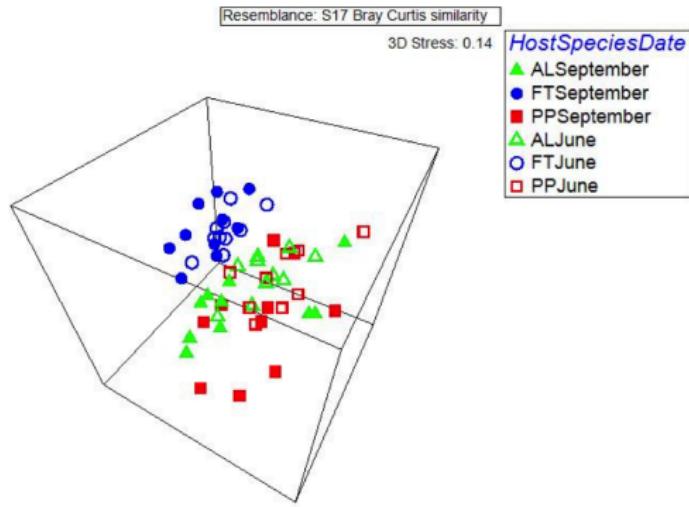
- ▶ OTU composition differed between sampling dates ($df = 1$, $pseudo-F = 2.9483$, $P = 0.0009$) and among host species ($df = 2$, $pseudo-F = 5.4469$, $P = 0.0001$)



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 - ▶ FT differed from AL and PP

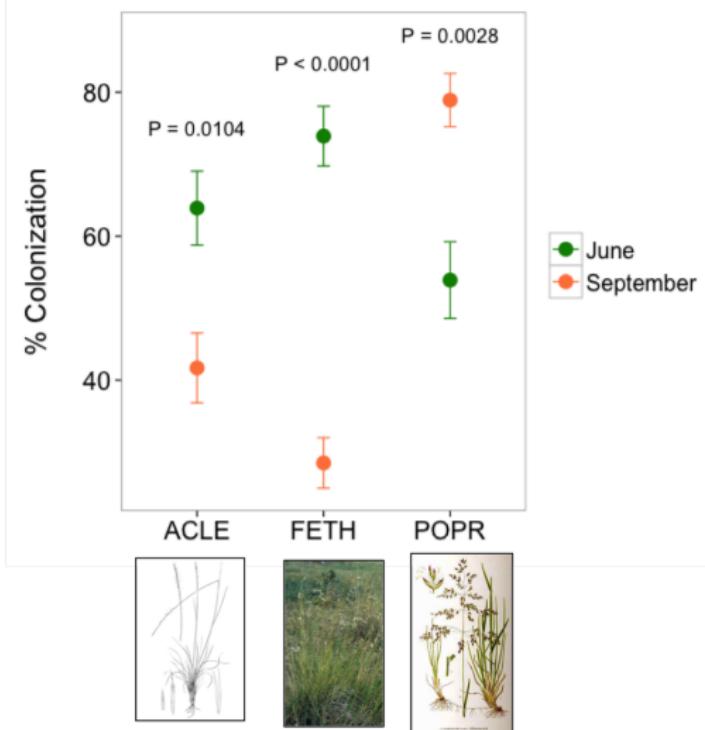
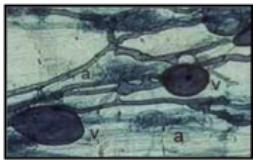


- ▶ OTU composition differed between sampling dates ($df = 1$, $pseudo-F = 2.9483$, $P = 0.0009$) and among host species ($df = 2$, $pseudo-F = 5.4469$, $P = 0.0001$)
 - ▶ FT differed from AL and PP
 - ▶ Communities of AL and PP were significantly more dispersed relative to communities of FT (PERMDISP)

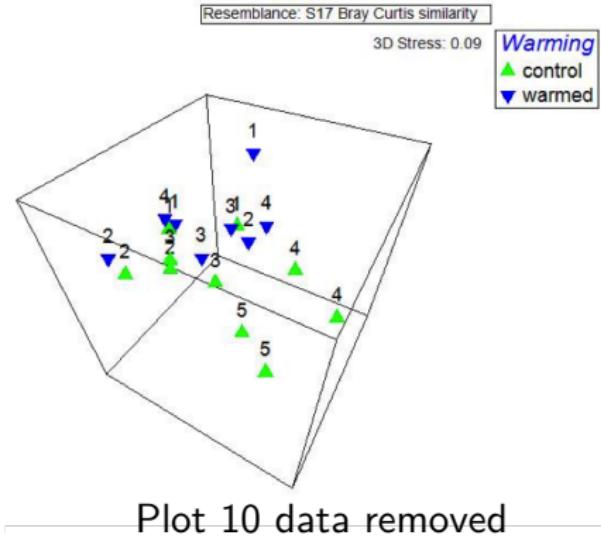
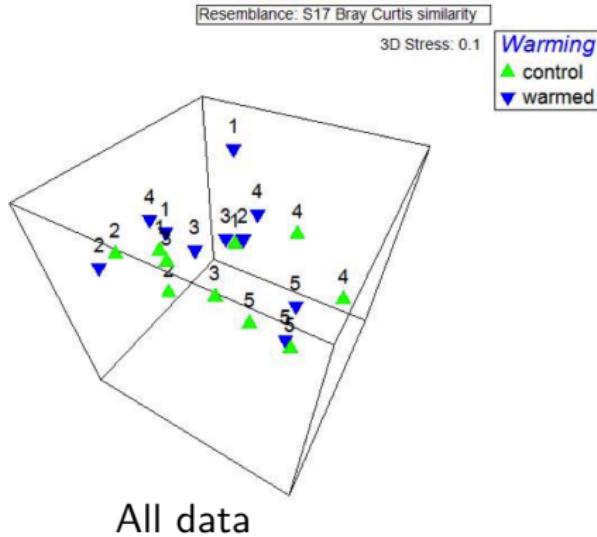


Changes in AMF colonization between June and September for all three grasses

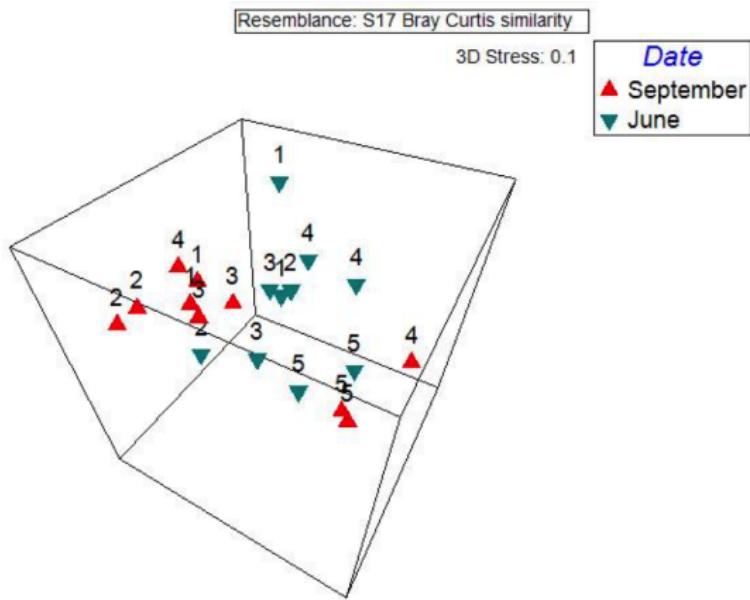
Arbuscular
mycorrhizal
fungi



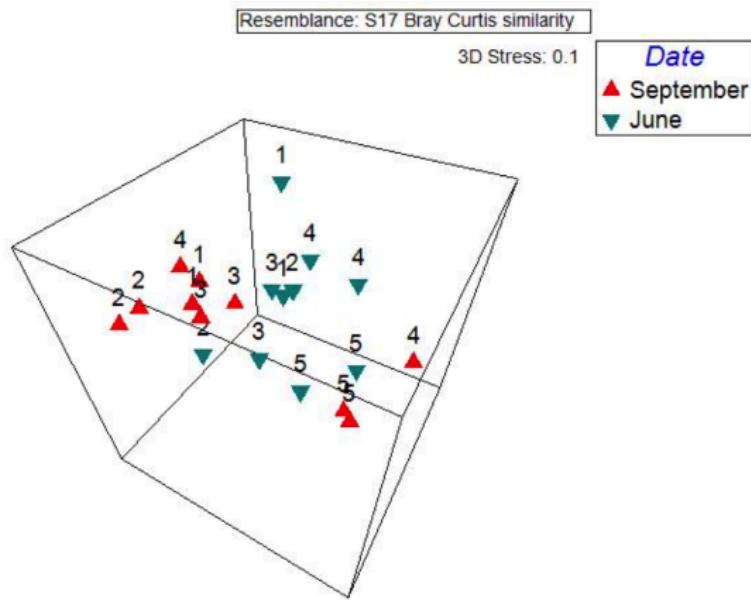
Results: *A. lettermanii*



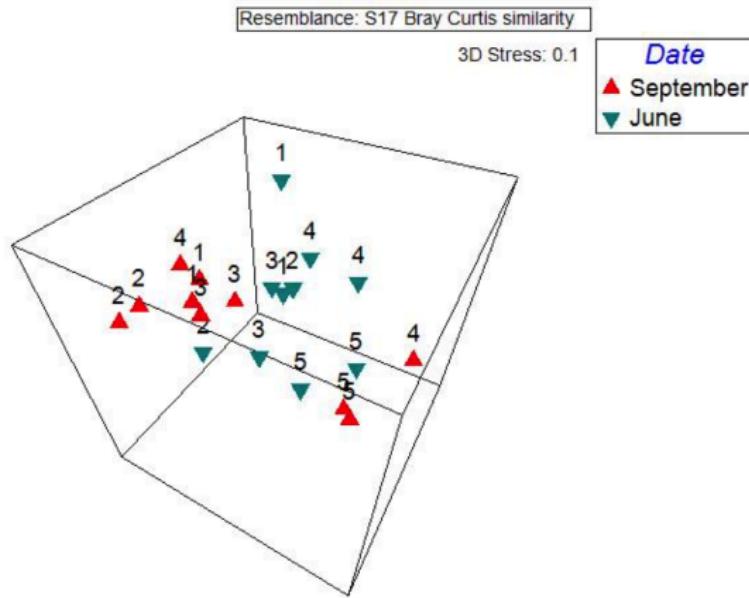
- Sampling date affected OTU composition ($df = 1$, psuedo- $F = 3.1274$, $P = 0.0024$)



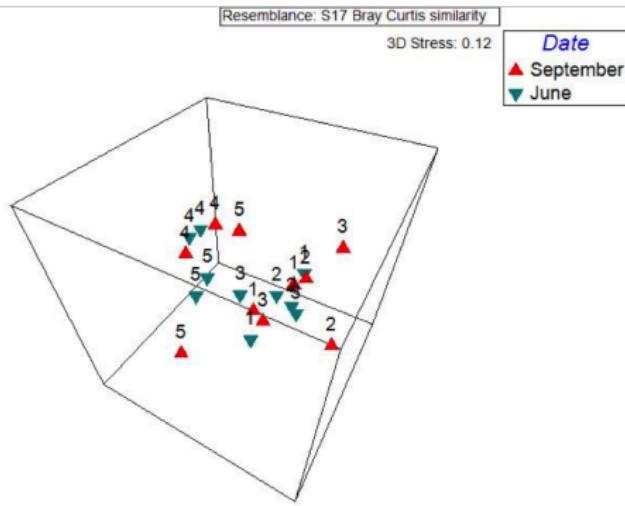
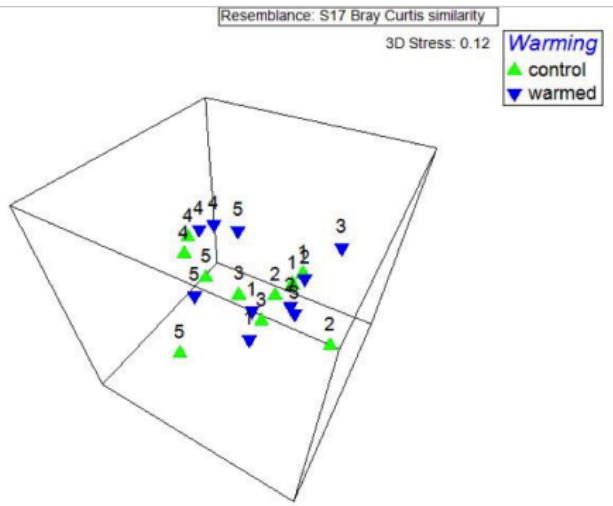
- ▶ Sampling date affected OTU composition ($df = 1$, psuedo- $F = 3.1274$, $P = 0.0024$)
- ▶ No difference in dispersion between two dates



- ▶ Sampling date affected OTU composition ($df = 1$, psuedo- $F = 3.1274$, $P = 0.0024$)
- ▶ No difference in dispersion between two dates
- ▶ Grouping by plot



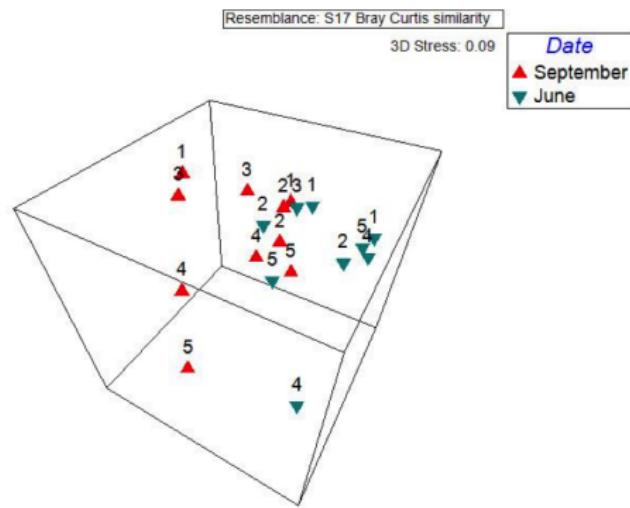
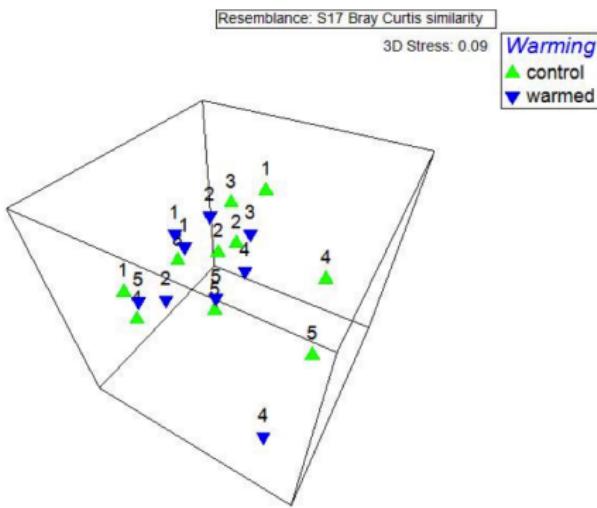
Results: *F. thurberi*



No effect of warming or sampling date

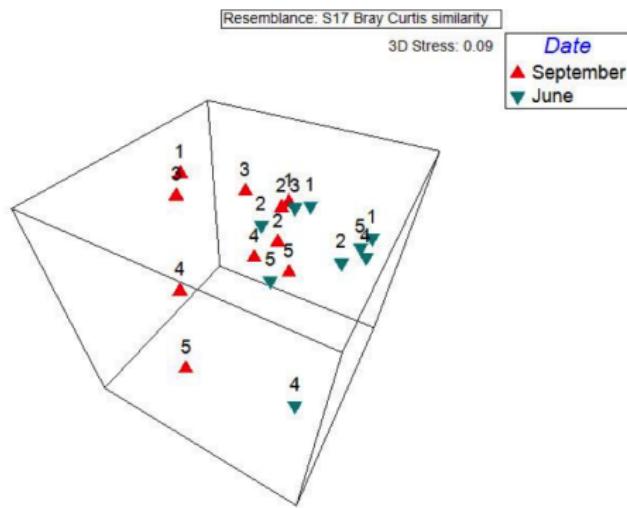
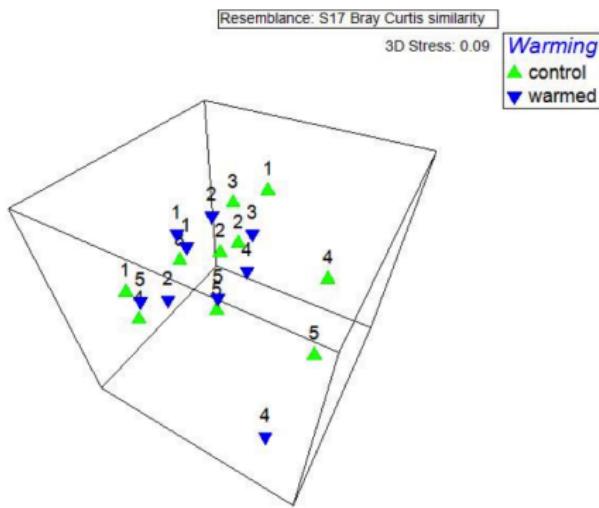
Results: *P. pratensis*

- ▶ No effect of warming



Results: *P. pratensis*

- ▶ No effect of warming
- ▶ Effect of sampling date ($df = 1$, pseudo- $F = 2.6595$, $P = 0.0065$)



Indicator Species Analysis (SIMPER)

Control vs. warmed plots

OTU	Control	Warmed	Dissimilarity	Contribution %
	Avg. Abundance	Avg. Abundance		
OTU5	7.19	6.72	0.47	0.96
OTU4	7.52	7.14	0.42	0.86
OTU6	3.5	1.94	0.42	0.86
OTU15	3.99	4.02	0.42	0.85
OTU12	4.05	4.59	0.42	0.84

F. thurberi vs. *P. pratensis*

OTU	FETH	POPR	Avg. Dissimilarity	Contribution %
	Avg. Abundance	Avg. Abundance		
OTU15	7.17	2.36	0.53	1.02
OTU16	5.39	2.01	0.5	0.96
OTU11	4.39	8.72	0.49	0.95
OTU24	5.04	0.78	0.48	0.93
OTU12	5.96	2.72	0.47	0.91

Resemblance: S17 Bray Curtis similarity

3D Stress: 0.14



OTU	FETH	ACLE	Avg. Dissimilarity	Contribution %
	Avg. Abundance	Avg. Abundance		
OTU15	7.17	2.41	0.54	1.07
OTU5	4.81	8.68	0.53	1.05
OTU24	5.04	0.3	0.5	0.98
OTU16	5.39	1.46	0.47	0.92
OTU11	4.39	7.53	0.43	0.85

F. thurberi vs. *A. lettermanii*

Taxonomy

OTU	Top BLAST Hit	Top BLAST Hit Details			Citation
		Study Location	Study System		
OTU16	Uncultured Glomeromycota	California, USA	Giant sequoia (<i>Sequoiadendron giganteum</i>)	Fahey et al. 2012, <i>Mycologia</i>	
OTU15	Uncultured Glomus	Michigan, USA	Northern hardwood forest dominated by sugar maple (<i>Acer saccharum</i>)	van Diepen et al. 2013, <i>Applied Soil Ecology</i>	
OTU24	Uncultured Glomeromycota	Qinghai-Tibetan Plateau, China	Alpine meadow	Yang et al. 2013, <i>PLOS ONE</i>	
OTU11	Uncultured Glomeromycota	Hungary	Agricultural system (corn, wheat, alfalfa, barley, peas)	Magurno et al. 2014, <i>Open Journal of Ecology</i>	
OTU5	Uncultured Glomeromycota	Montana, USA	Native grassland vs. system dominated by <i>Centaurea maculosa</i> (spotted knapweed)	Mummey and Rillig 2006, <i>Plant and Soil</i>	
OTU12	Uncultured Glomeraceae	Czech Republic	<i>Knautia arvensis</i> (Caprifoliaceae)	Doubková et al. 2013, <i>Mycorrhiza</i>	
OTU4	Uncultured Glomeromycota	Tibetan Plateau, China	Herbaceous plants	Li et al., Unpublished	
OTU6	Uncultured Glomeromycota	Canada	Crested wheatgrass (<i>Agropyron cristatum</i>)	Perez et al. 2008, <i>Agriculture and Agrifood Canada</i>	
OTU10	Uncultured Glomeromycota	Canada	Switchgrass (<i>Panicum virgatum</i>)	Perez et al. 2008, <i>Agriculture and Agrifood Canada</i>	

Questions or comments?

