



BIAŁOWIESKA SZKOŁA STATYSTYKI

# Regresja – uogólnione modele liniowe i addytywne

# Krótkie przypomnienie

modele liniowe  $y=ax+b$  `lm(y~x, data=dataset)`

modele mogą mieć wiele czynników

modele mogą być funkcjami liniowymi innymi wielomian 1. stopnia, np. parabole:

$y=ax^2+bx+c$  `lm(y~poly(x,2), dataset)`,  $y=a*\log(x)+b$  `lm(y~log(x), dataset)`

modele mogą mieć wiele zmiennych

$y=ax+by+cz+d$  `lm(y~a+b+c, dataset)`

# GLM

uogólniony model liniowy - generalized linear model

dlaczego uogólniony?

model liniowy zakłada rozkład normalny, GLM uogólnia metodę na inne rozkłady

po co? do innych rozkładów

(przypominamy sobie przykłady)

# GLM z innymi rozkładami

`glm(formula, data, family='rozkład')`

`family='poisson'` - rozkład Poissona (dyskretny - liczby naturalne - liczba osobników, liczba gatunków)

`family=binomial(link='logit')` - regresja logistyczna (0/1 - np. przeżycie, występowanie lub brak - klasyfikacja binarna)

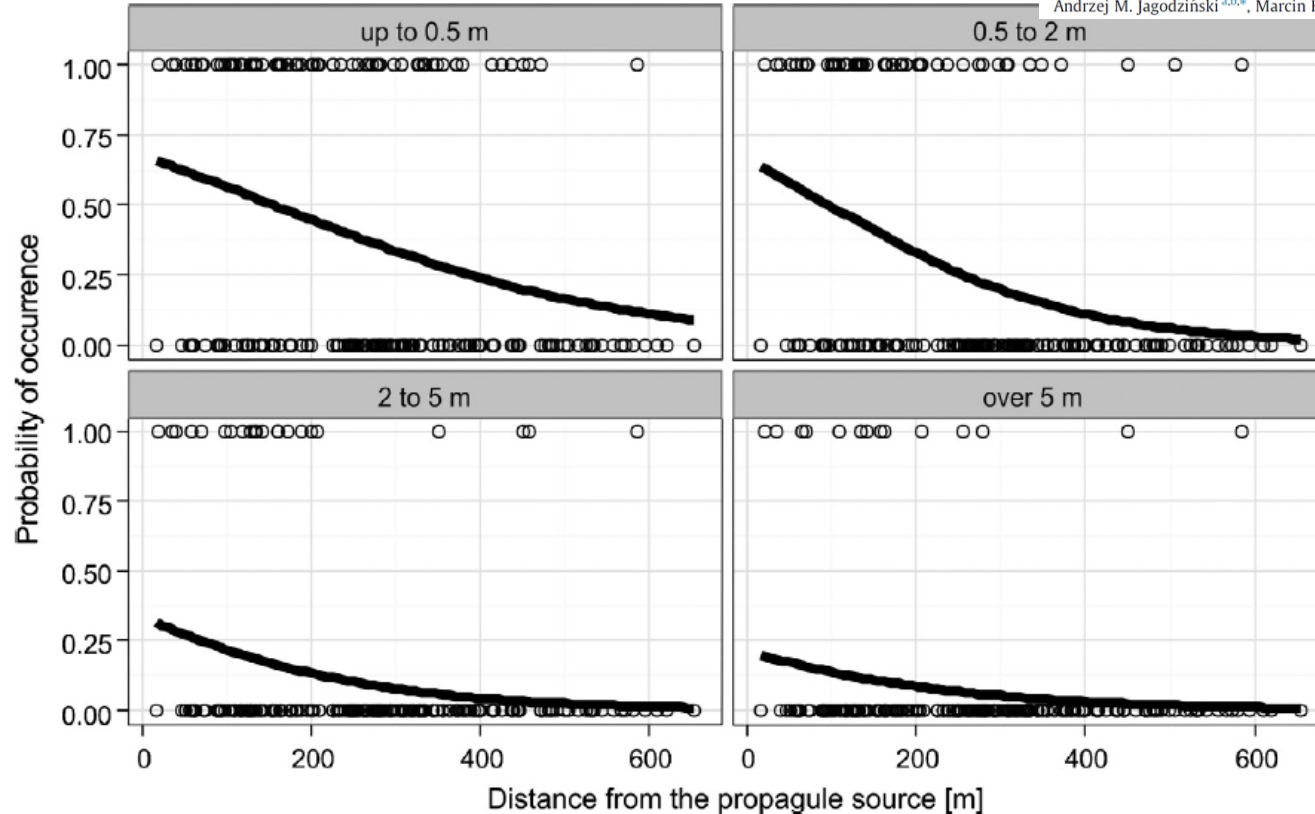
# Rozkład dwumianowy

- Dane zero-jedynkowe
- Dwa stany – prawdopodobieństwo osiągnięcia jednego
- Model rozmieszczenia gatunku  $\Leftrightarrow$  presence i true absence!
- Przeżywalność, sukces lęgowy, obecność

# regresja logistyczna

Plantation of coniferous trees modifies risk and size of *Padus serotina* (Ehrh.) Borkh. invasion – Evidence from a Rogów Arboretum case study

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# GLM binomial (regresja logistyczna)

```
model<-glm(szens~odl, family=binomial(link=logit'), dane)
```

```
summary(model)
```

```
call:
glm(formula = szens ~ odl, family = binomial(link = "logit"))

Deviance Residuals:
    Min       1Q   Median       3Q      Max
-1.4374  -0.9748  -0.7137   1.1419   1.9901

Coefficients:
              Estimate Std. Error z value Pr(>|z|)
(Intercept)  0.661478   0.318846   2.075 0.038024 *
odl          -0.004263   0.001120  -3.806 0.000141 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

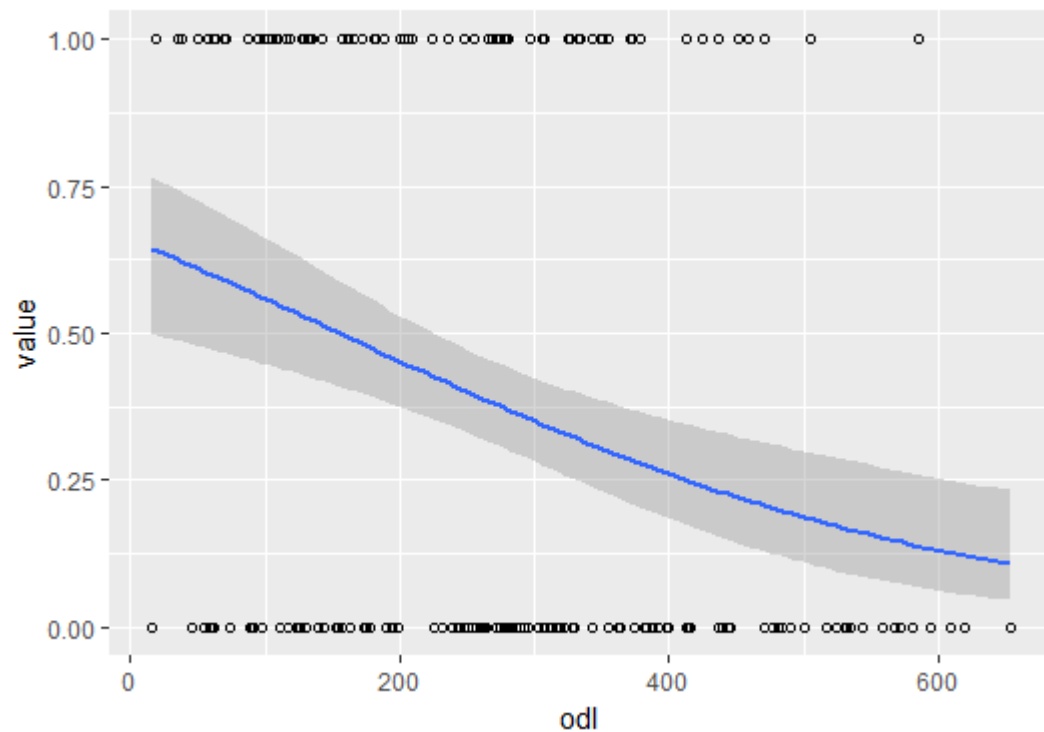
(Dispersion parameter for binomial family taken to be 1)

    Null deviance: 268.48  on 200  degrees of freedom
Residual deviance: 252.23  on 199  degrees of freedom
AIC: 256.23

Number of Fisher Scoring iterations: 4
```



```
ggplot(data=fig4df, aes(x=odl, y=value))+geom_point(shape=1)  
+geom_smooth(method="glm", method.args=c(family="binomial"))
```



Dwie zmienne: typ lasu i odległość `glm(szans~odl+typ, family=binomial(link=logit'), dane)`

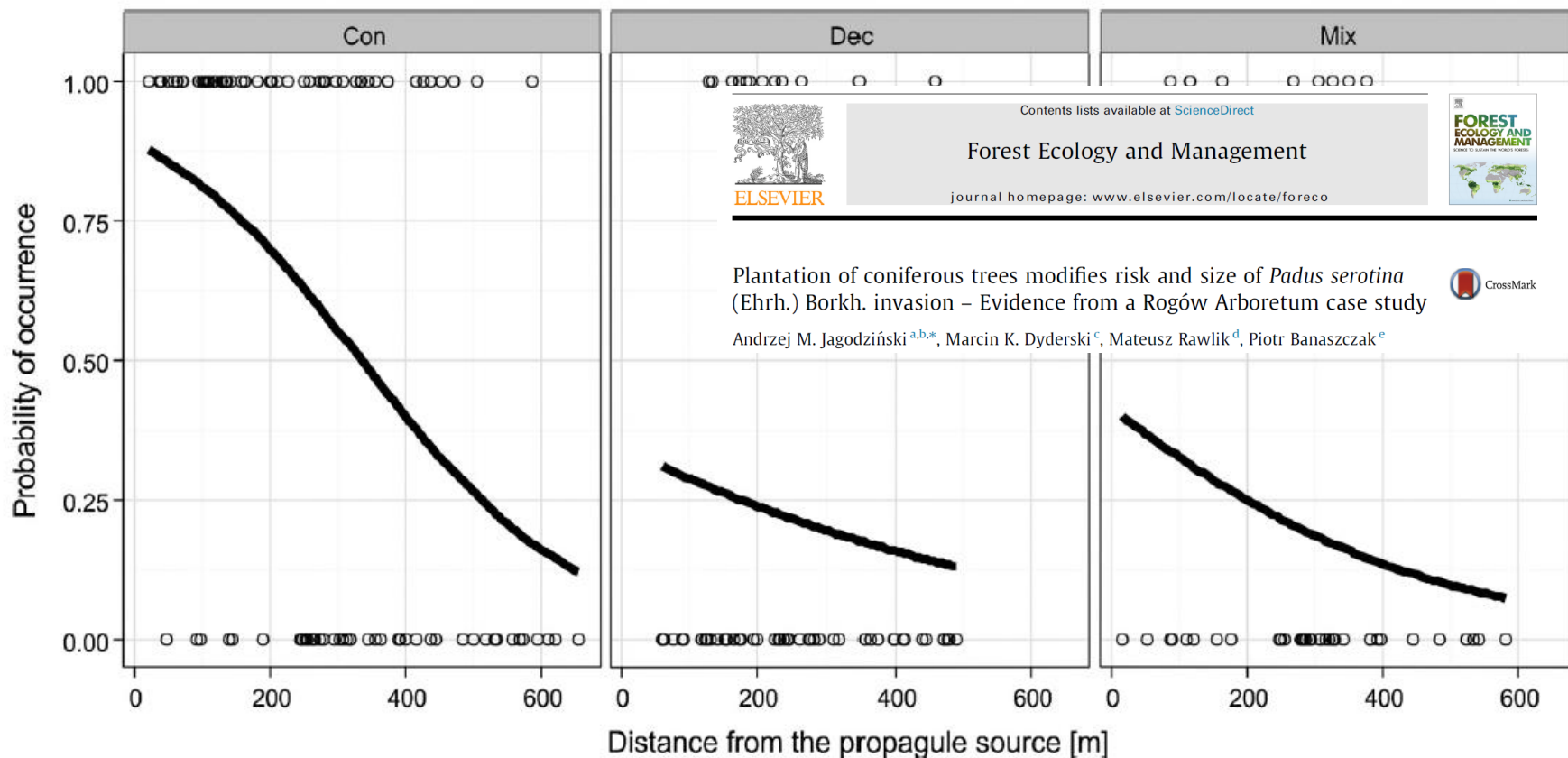


Fig. 4. Effect of coniferous (Con), deciduous (Dec) and mixed (Mix) tree stands on the probability of colonisation by black cherry ( $p < 0.001$ ).

# Inny przykład – gatunki starych lasów

- Zbiór danych z poniedziałku (eks)

```
table(eks$stare.lasy)
```

0 1

263 49

Czy gatunek jest wskaźnikowy dla starych lasów?

# Budujemy model globalny

```
mod.sl<-glm(stare.lasy~SLA+seed_mass+L, family=binomial(link='logit'),eks)
```

```
car::vif(mod.sl)
```

```
      SLA seed_mass      L  
1.108886 1.151267 1.111789
```

```
summary(mod.sl)
```

Call:

```
glm(formula = stare.lasy ~ SLA + seed_mass + L, family = binomial(link = "logit"),  
    data = eks)
```

Deviance Residuals:

```
      Min      1Q  Median      3Q      Max  
-1.6239 -0.2989 -0.1587 -0.0660  3.7668
```

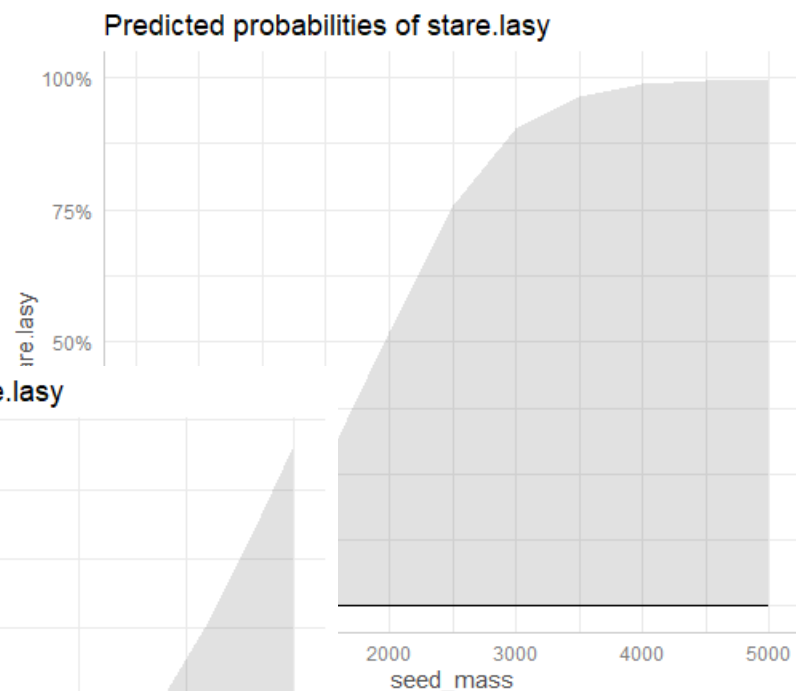
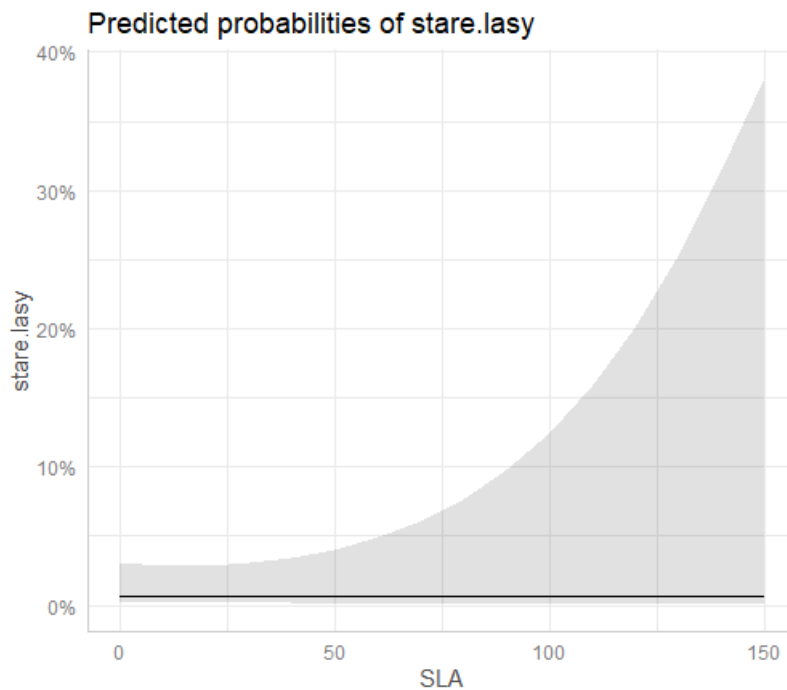
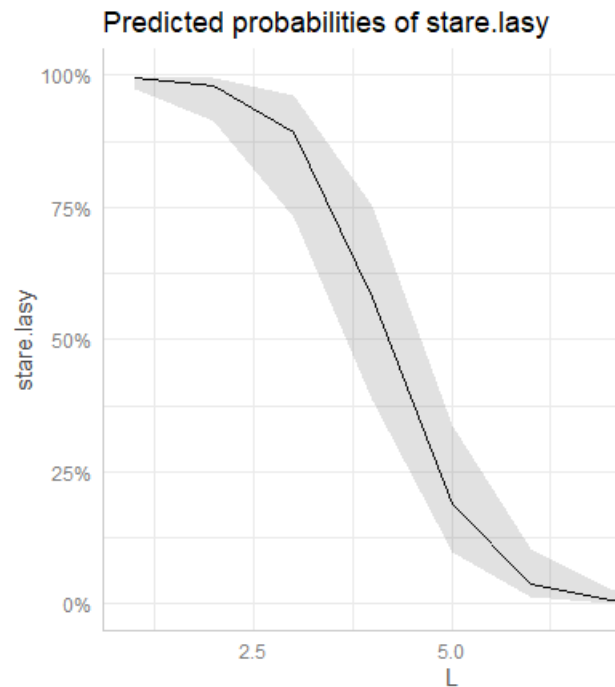
Coefficients:

```
      Estimate Std. Error z value Pr(>|z|)  
(Intercept) 8.1168346 1.5525297  5.228 1.71e-07 ***  
SLA          0.0002634 0.0161591  0.016  0.987  
seed_mass   -0.0104889 0.0064559 -1.625  0.104  
L           -1.7781285 0.2723928 -6.528 6.67e-11 ***
```

# Jak rozumieć effect sizes?

- Względnie – im większy estimate tym większy effect size
- Bezwzględnie – trzeba przeliczyć w oparciu o funkcję łączącą
- Najłatwiej – `ggpredict()`

# ggpredict(mod.sl)



# Dane zliczeniowe

- Liczby naturalne ( $>0$ , całkowite)
  - Bogactwo gatunkowe na powierzchni badawczej
  - Zagęszczenie odnowienia naturalnego
  - Liczba saren na polanie
  - Liczba piskląt w gnieździe
- 
- Rozkład Poissona – pierwszy wybór – założenie braku nadyspersji (overdispersion)
  - Rozkład dwumianowy ujemny – przy naddyspersji

# GLM z rozkładem Poissona –zobaczmy najpierw lm

```
model<-lm(prunusc~richness,data=prunus)
```

```
summary(model)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-4.5654	-1.7546	-0.7967	0.2663	15.6279

Coefficients:

	Estimate	Std. Error	t value	Pr(> t )
(Intercept)	-1.61911	1.70456	-0.950	0.3482
richness	0.19327	0.08731	2.214	0.0329 *

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for gaussian family taken to be 17.97445)

Null deviance: 771.10 on 39 degrees of freedom

Residual deviance: 683.03 on 38 degrees of freedom

AIC: 233.02



```
model<-glm(prunusc~richness,data=prunus, family=poisson)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-3.3062	-1.5175	-1.1999	-0.7244	8.0114

Coefficients:

	Estimate	Std. Error	z value	Pr(> z )
(Intercept)	-1.46686	0.38989	-3.762	0.000168 ***
richness	0.09892	0.01544	6.408	1.48e-10 ***

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for poisson family taken to be 1)

Null deviance: 248.48 on 39 degrees of freedom  
Residual deviance: 203.69 on 38 degrees of freedom  
AIC: 244.38

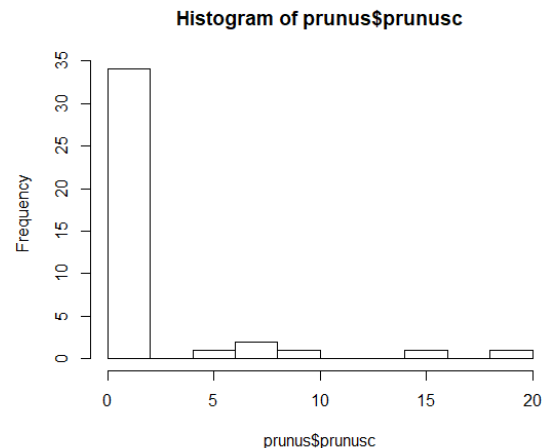
Number of Fisher Scoring iterations: 7

zwiększyło się AIC

teoretycznie jest to gorszy model, ale!

do wykonania GLM jesteśmy uprawnieni  
mając dane o rozkładzie normalnym

logika i założenia modelu > cyferki



# Interpretacja?

```
ggpredict(model)
```

```
$richness
```

```
# Predicted counts of prunusc
```

```
richness | Predicted | 95% CI
```

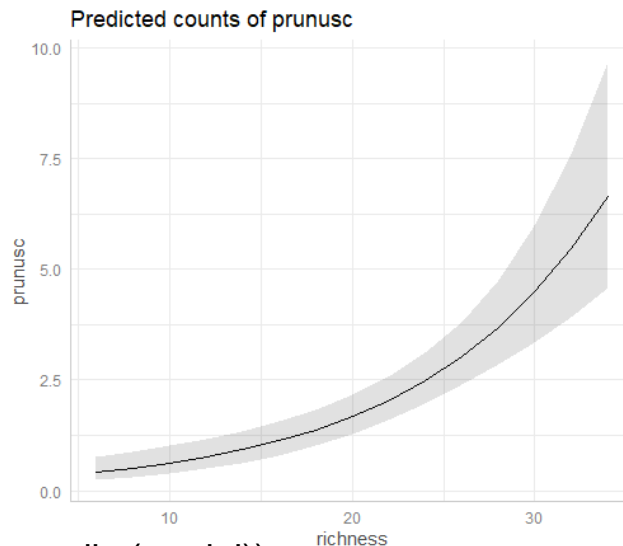
```
-----  
6 | 0.42 | [0.23, 0.76]  
10 | 0.62 | [0.38, 1.01]  
12 | 0.76 | [0.49, 1.16]  
16 | 1.12 | [0.80, 1.57]  
20 | 1.67 | [1.29, 2.16]  
24 | 2.48 | [1.97, 3.11]  
28 | 3.68 | [2.85, 4.76]  
34 | 6.66 | [4.57, 9.70]
```

Co to znaczy?

W d-stanach z 6 gatunkami w runie oczekiwane zagęszczenie czeremchy to 0,42 szt.

W d-stanach z 28 oczekujemy 3,68 szt. na poletko

przydaje się do opisu wyników



```
plot(ggpredict(model))
```

# Czy model jest poprawny?

Przy danych zliczeniowych mamy dwa potencjalne problemy:

- dyspersja (stosunek zmienności w danych do zmienności oczekiwanej w modelu)
- wartości zerowe

Rozkład Poissona - rozkład teoretyczny

Rzeczywistość - puste próby, brak występowania

Usunąć? informacja biologiczna - jak wnioskować o korniku nie badając miejsc gdzie go nie ma?

Zostawić - inflacja zer - model mając wiele zer “idzie na łatwiznę” - częściej przewiduje zero niż by wynikało z rzeczywistości

# Sprawdzamy – pakiet DHARMa

```
>library(DHARMa)
> model<-glm(prunusc~richness,data=prunus, family=poisson)
> simres<-simulateResiduals(model)
> testDispersion(simres)
```

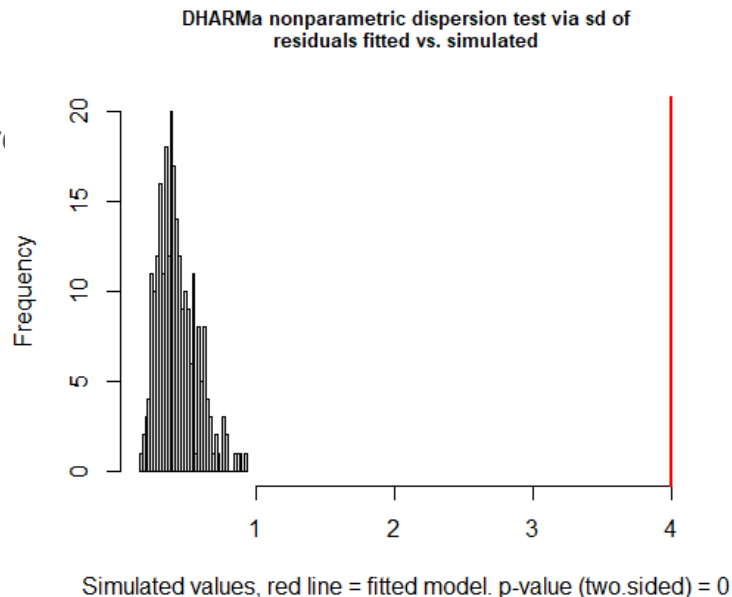
DHARMa nonparametric dispersion test via sd of residuals fitted vs. simulated

data: simulationOutput

**dispersion = 9.2819**, p-value < 2.2e-16

alternative hypothesis: two.sided

**Mamy problem - naddyseprsj**



# Sprawdzamy – pakiet DHARMa

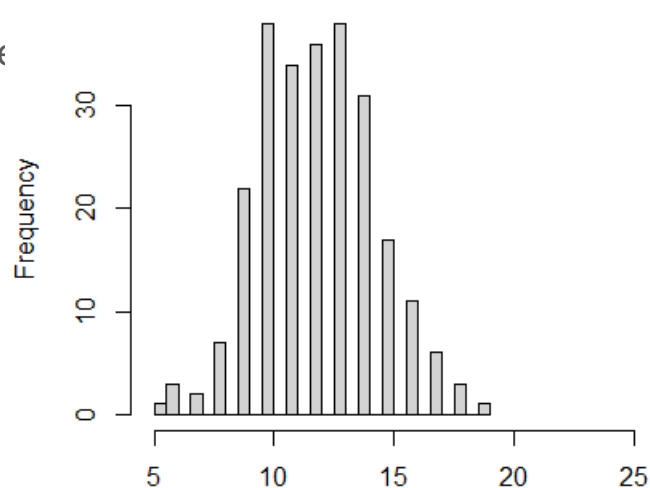
```
>library(DHARMa)
> model<-glm(prunusc~richness,data=prunus, family=poisson)
> simres<-simulateResiduals(model)
> testZeroInflation(simres)
```

DHARMa zero-inflation test via comparison to expected simulation under  $H_0$  = fitted model

data: simulationOutput  
ratioObsSim = 2.4078, p-value < 2.2e-16  
alternative hypothesis: two.sided

**Mamy problem – za dużo zer!**

DHARMa zero-inflation test via comparison to expected zeros with simulation under  $H_0$  = fitted model



Simulated values, red line = fitted model. p-value (two.sided) = 0

# Problem nr 1 - naddyspersja

- Zamiast Poissona wybieramy rozkład negative binomial, lepiej przejść na :

```
library(glmmTMB)
```

```
> model.nb<-glmmTMB(prunusc~richness,data=prunus, family=nbinom1())
```

```
> summary(model.nb)
```

Family: nbinom1 ( log )

Formula: prunusc ~ richness

Data: prunus

AIC	BIC	logLik	deviance	df.resid
111.2	116.3	-52.6	105.2	37

Dispersion parameter for nbinom1 family (): 13.2

Conditional model:

	Estimate	Std. Error	z value	Pr(> z )
(Intercept)	-1.11077	1.00707	-1.103	0.2700
richness	0.08387	0.03905	2.148	0.0317 *

# Sprawdzamy problem z dyspersją

```
model.nb<-glmmTMB(prunusc~richness,data=prunus, family=nbinom1())
```

```
simres<-simulateResiduals(model.nb)
```

```
testDispersion(simres)
```

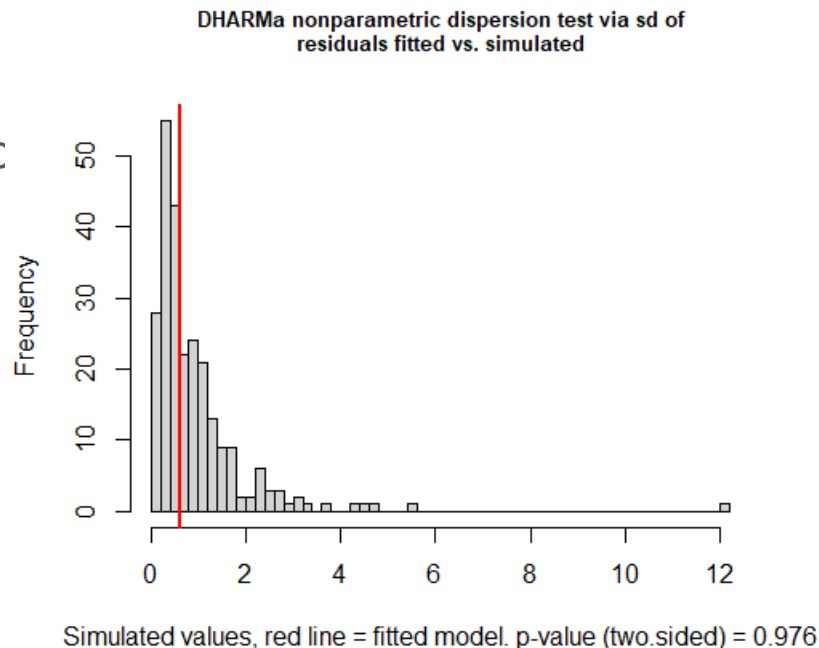
DHARMa nonparametric dispersion  
vs. simulated

data: simulationOutput

dispersion = 0.67144, p-value = 0.976

alternative hypothesis: two.sided

Jest dobrze, nie ma problemu

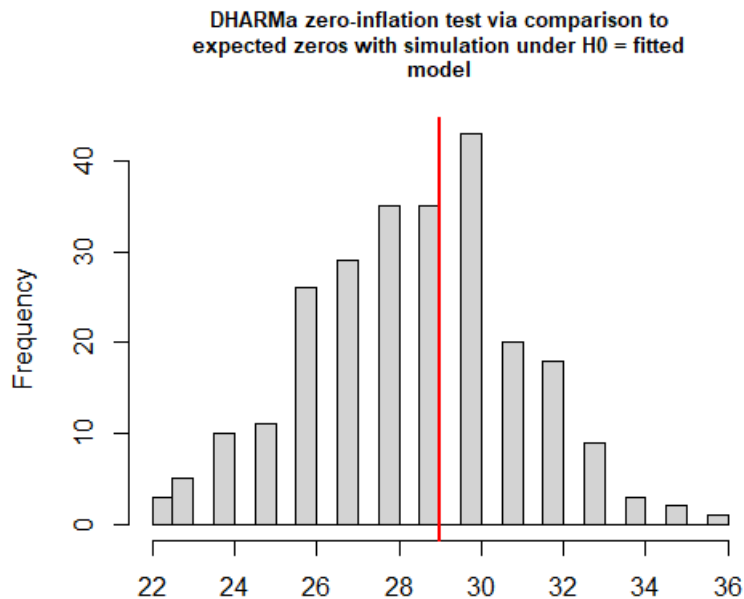


testZeroInflation(simres)

DHARMA zero-inflation test via comparison  
to expected zeros with  
simulation under  $H_0$  = fitted model

data: simulationOutput  
ratioObsSim = 1.0153, p-value = 1  
alternative hypothesis: two.sided

Mimo iż nic nie zrobiliśmy z  
zerami, to inny rozkład pozwolił też  
ominąć ten problem!



Simulated values, red line = fitted model. p-value (two.sided) = 1

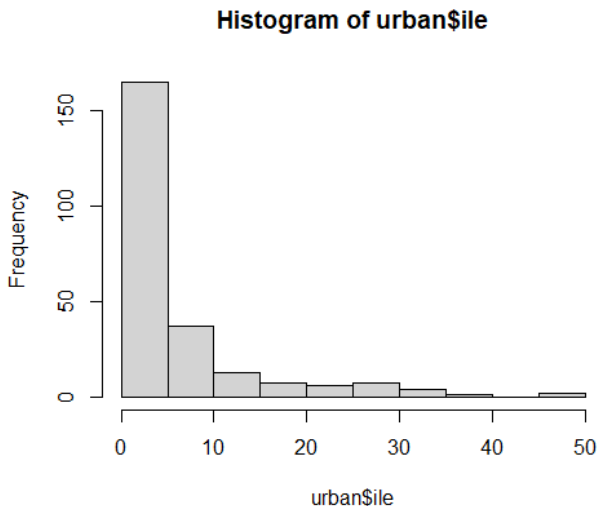


# Problem z zerami – potrzeba modelu zero-inflated

Tzw. modele złożone (hurdle models) - składają się z dwóch części:

1. modeluje prawdopodobieństwo wystąpienia (binomial) - czy w ogóle coś będzie
2. modeluje liczebność - czyli właściwy Poisson (lub negative binomial albo beta)

```
hist(urban$ile)
length(which(urban$ile>0))
187 #na 242
```



```
mod.u<-glmmTMB(ile~Forests+Water, ziformula = ~Forests, urban, family=poisson())
> summary(mod.u)
Family: poisson ( log )
Formula:      ile ~ Forests + Water
Zero inflation: ~Forests
Data: urban
```

ziformula – definiujemy zmienne wpływające na  
0/1  
działa na Poissona, beta, negative binomial

```
AIC    BIC  logLik deviance df.resid
2053.4 2070.8 -1021.7 2043.4    237
```

Conditional model:

```
      Estimate Std. Error z value Pr(>|z|)
(Intercept) 1.5408407  0.0411004  37.49 < 2e-16 ***
Forests      0.0160141  0.0008578  18.67 < 2e-16 ***
Water        0.0164942  0.0028167   5.86 4.74e-09 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

Zero-inflation model:

```
      Estimate Std. Error z value Pr(>|z|)
(Intercept) -0.86517    0.17745 -4.876 1.08e-06 ***
Forests     -0.03498    0.01099 -3.182 0.00146 **
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
ggpredict(mod.u)
$Forests
# Predicted counts of ile
```

Forests	Predicted	95% CI
0	4.91	[ 4.54, 5.30]
20	6.76	[ 6.38, 7.16]
40	9.31	[ 8.82, 9.83]
60	12.83	[11.96, 13.75]
80	17.67	[16.07, 19.43]
100	24.34	[21.50, 27.56]

```
Adjusted for:
* Water = 3.03
```

```
$Water
# Predicted counts of ile
```

Water	Predicted	95% CI
0	6.04	[ 5.66, 6.45]
10	7.12	[ 6.66, 7.62]
20	8.40	[ 7.56, 9.33]
30	9.90	[ 8.50, 11.55]
50	13.78	[10.64, 17.84]
60	16.25	[11.89, 22.20]
70	19.16	[13.28, 27.65]
90	26.65	[16.56, 42.89]

```
Adjusted for:
* Forests = 16.07
```

```
attr(,"class")
[1] "ggalleffects" "list"
attr(,"model.name")
[1] "mod.u"
```

# Raportowanie w publikacji

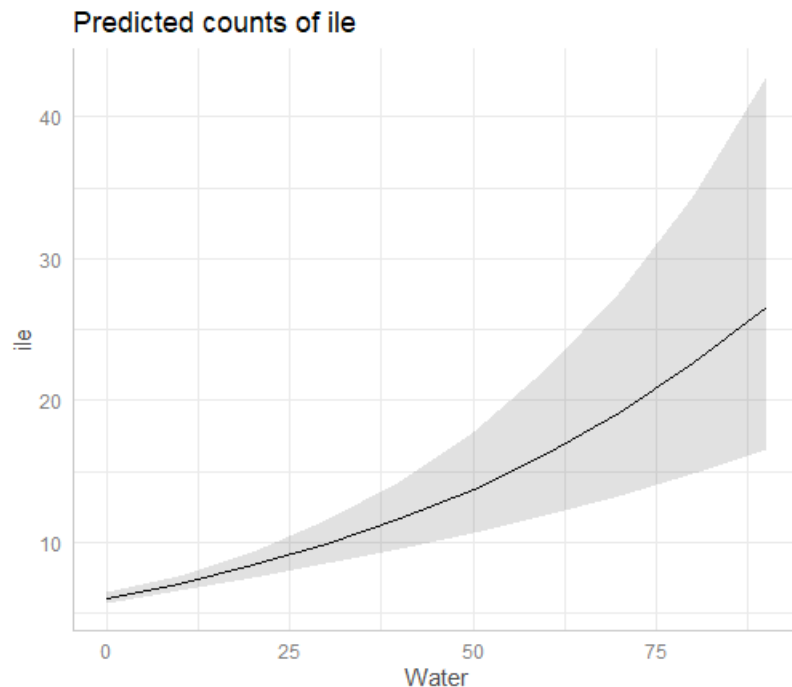
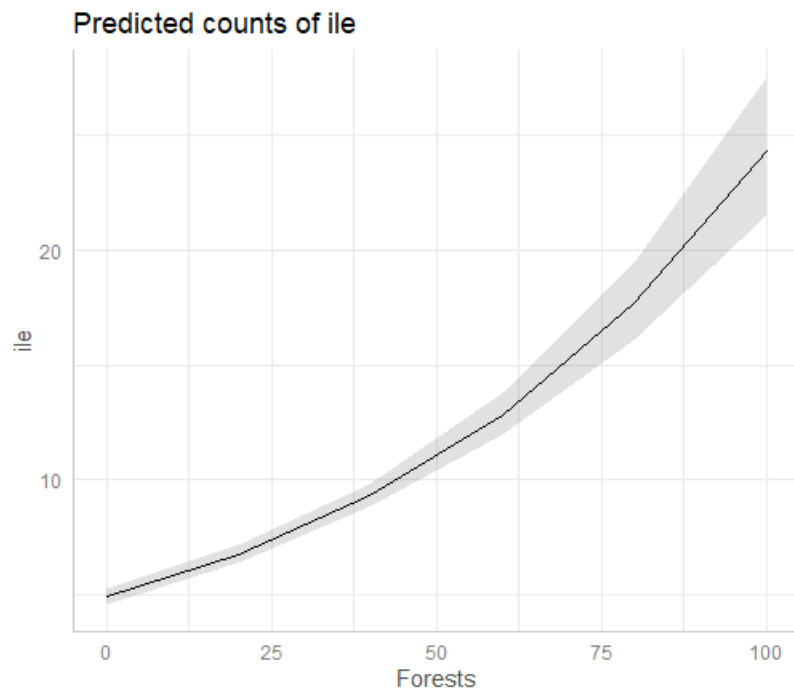
**Table 2**

Generalized Linear Mixed-Effects Models explaining proportion of browsed and damaged trees proportion and density of *F. excelsior* natural regeneration within study plots ( $n = 32$ ).

Response	Variable	Estimate	SE	z	Pr(> z )	Random effect and AICc
Proportion of browsed trees	(Intercept)	-0.2723	0.1315	-2.071	0.0384	Random effect SD = 0.0452
	Density	0.0038	0.0013	2.907	0.0037	AICc = -73.2
	Shrub cover	-0.5302	0.2453	-2.161	0.0307	AICc <sub>0</sub> = -50.1, AICc <sub>g</sub> = -61.7
	CaCO <sub>3</sub> content	-0.0251	0.0100	-2.516	0.0119	
Proportion of damaged trees	Count: (Intercept)	-9.0370	1.9436	-4.650	<0.0001	Random effect (count) SD = 0.2644
	Count: Summer groundwater table level	-0.3997	0.2160	-1.850	0.0643	Random effect (zero-inflation) SD = 0.0512
	Count: Soil pH	1.0719	0.2356	4.550	<0.0001	AICc = -96.9
	Zero-inflation: (Intercept)	-3.4350	1.0350	-3.318	0.0009	AICc <sub>0</sub> = -52.0, AICc <sub>g</sub> = -83.7
Proportion of damaged trees	Count: (Intercept)	-2.8985	0.2811	-10.310	<0.0001	Random effect (count) SD = 0.2502
Alternative model	Count: Defoliation	0.0286	0.0072	4.000	0.0001	Random effect (zero-inflation) SD = 7.063
	Zero-inflation: (Intercept)	-8.431	4.503	-1.872	0.0612	AICc = -96.9, AICc <sub>0</sub> = -52.0,
	(Intercept)	4.1021	0.5673	7.231	<0.0001	Random effect SD = 0.4503
Density	Proportion of damaged trees	-4.4382	1.5805	-2.808	0.0050	AICc = 237.8
	Canopy cover	-1.8231	0.2834	-6.432	<0.0001	AICc <sub>0</sub> = 352.9, AICc <sub>g</sub> = 250.4
	Summer groundwater table level	0.5399	0.2058	2.623	0.0087	

SE – standard error, z – test statistic, Pr(>|z|) – p-value, AICc – Akaike's Information Criterion, with correction for small sample size, AICc<sub>0</sub> – AICc of null model (intercept-only), AICc<sub>g</sub> – AICc of global model (covering all hypothesized variables after excluding intercorrelated variables), SD – standard deviation.

# Efekty brzegowe `plot(ggpredict(mod.u))`



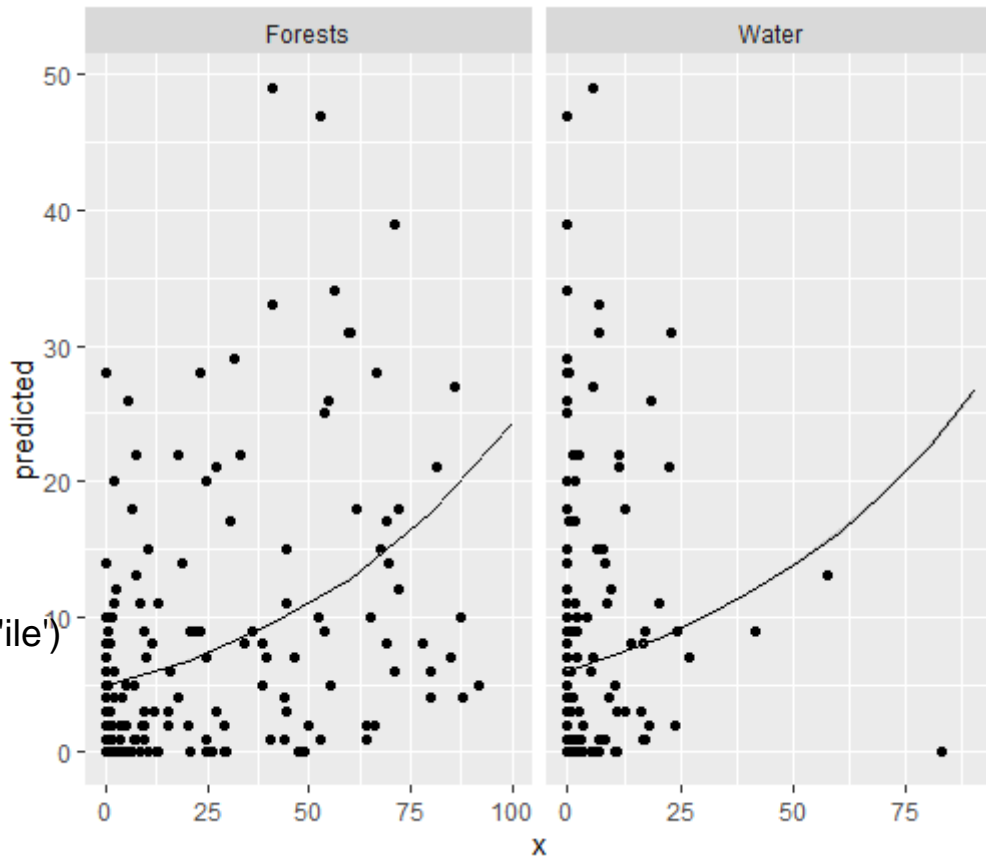
Tylko część zliczeniowa (count), bez zero-inflated!

# Wykres z punktami

```
ggp1<-as.data.frame(ggpredict(mod.u, 'Forests'))
ggp1$group<-'Forests'
ggp2<-as.data.frame(ggpredict(mod.u, 'Water'))
ggp2$group<-'Water'
ggp<-bind_rows(list(ggp1, ggp2))

gg.points<-reshape2::melt(urban[,c(6,14,16)], id.vars='ile')
colnames(gg.points)<-c('predicted','group','x')

ggplot(ggp, aes(x=x, y=predicted))
+geom_point(data=gg.points)
+geom_ribbon(aes(ymin=predicted-std.error
,ymax=predicted+std.error ),fill='gray80')
+geom_line()+facet_wrap(~group, scales='free_x')
```



# Dane zliczeniowe – kolejność wykonywania działań

- Start: wiemy że mamy dane zliczeniowe
- Sprawdzamy model globalny pod kątem VIF
- Po ewentualnej redukcji sprawdzamy zero inflation i overdispersion funkcjami *DHARMA::testDisepersion()* i *DHARMA::testZeroInflation()*
- selekcja zmiennych: *step()* lub *dredge()* – działają z *glmmTMB()*
- summary, wykresy, decyzja

# Proporcje – rozkład Beta

- Do danych dotyczących udziału lub procentów
- $0 < \text{zmienna zależna} < 1$ , choć możliwy rozkład zero inflated beta
- przykłady:
  - udział gatunków wskaźnikowych starych lasów
  - pokrycie koron drzew
  - udział zgryzionych pędów
  - proporcja piskląt które przeżyły pierwszy rok

Co zrobić jak są wartości 1? Jak dużo – iść w kierunku binomial (stracić informację), jak niedużo – zmienić 1 na 0.99 – model wtedy to przepuści

# Przykład – procent zgryzanych drzewek

br to liczba zgryzanych drzewek/liczba wszystkich drzewek

```
summary(prop$br)
```

Min.	1st Qu.	Median	Mean	3rd Qu.	Max.
------	---------	--------	------	---------	------

0.0000	0.2935	0.3649	0.3454	0.4223	0.5556
--------	--------	--------	--------	--------	--------



# Model 1

- `mod.br<-glmmTMB(br~pH+CN+b+c, data = prop, family=beta_family())`
- Error in `eval(family$initialize)` : y values must be  $0 < y < 1$
- Zero-inflation – trzeba dać ziformula
- `mod.br<-glmmTMB(br~pH+CN+b+c, ziformula = ~b+c, data = prop, family=beta_family() , na.action = na.fail) #ostatnie dla dredge`
- `MuMIn::dredge(mod.br)`

```
> MuMin::dredge(mod.br)
Fixed terms are "cond((Int))", "zi((Int))" and "disp((Int))"
Global model call: glmmTMB(formula = br ~ pH + CN + b + c, data = prop, family = beta_family(),
  ziformula = ~b + c, na.action = na.fail, dispformula = ~1)
---
Model selection table
```

	cnd((Int))	zi((Int))	dsp((Int))	cnd(b)	cnd(c)	cnd(CN)	cnd(pH)	zi(b)	zi(c)	df	logLik	AICc
35	-1.5010	-8.404	+		0.01593				0.08317	5	31.119	-50.5
51	-1.5010	-13.330	+		0.01593		0.09288	0.11550		6	32.453	-50.4
3	-1.5010	-2.944	+		0.01593					4	29.744	-50.3
4	-1.2950	-2.944	+	-0.005876	0.01459					5	30.665	-49.6

cnd – conditional (liczebność), zi – zero-inflated

Najlepszy model: c jako cnd i zi

```
mod.br<-glmmTMB(br~c, ziformula = ~c, data = prop,
family=beta_family(), na.action = na.fail)
```

```
> summary(mod.br)
```

Family: beta (logit)

Formula: br ~ c

Zero inflation: ~c

Data: prop

AIC	BIC	logLik	deviance	df.resid
-52.2	-43.8	31.1	-62.2	35

Dispersion parameter for beta family (): 27.2

Conditional model:

	Estimate	Std. Error	z value	Pr(> z )
(Intercept)	-1.50121	0.37774	-3.974	7.06e-05 ***
c	0.01593	0.00631	2.525	0.0116 *

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

Zero-inflation model:

	Estimate	Std. Error	z value	Pr(> z )
(Intercept)	-8.40446	3.71108	-2.265	0.0235 *
c	0.08317	0.05067	1.642	0.1007

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

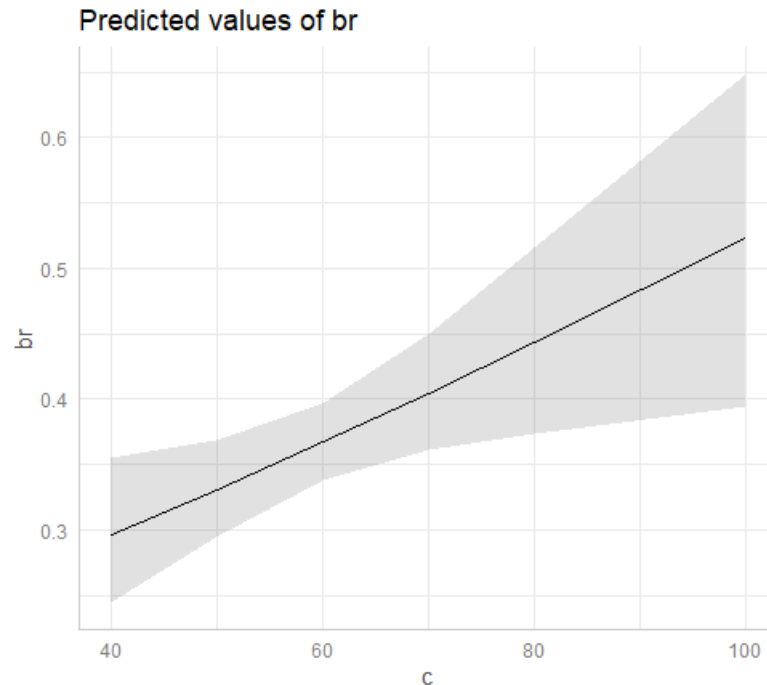
# Wielkość efektu?

```
ggpredict(mod.br)
```

```
$c
```

```
# Predicted values of br
```

c	Predicted	95% CI
40	0.30	[0.24, 0.35]
50	0.33	[0.29, 0.37]
60	0.37	[0.34, 0.40]
70	0.40	[0.36, 0.45]
80	0.44	[0.37, 0.52]
100	0.52	[0.39, 0.65]



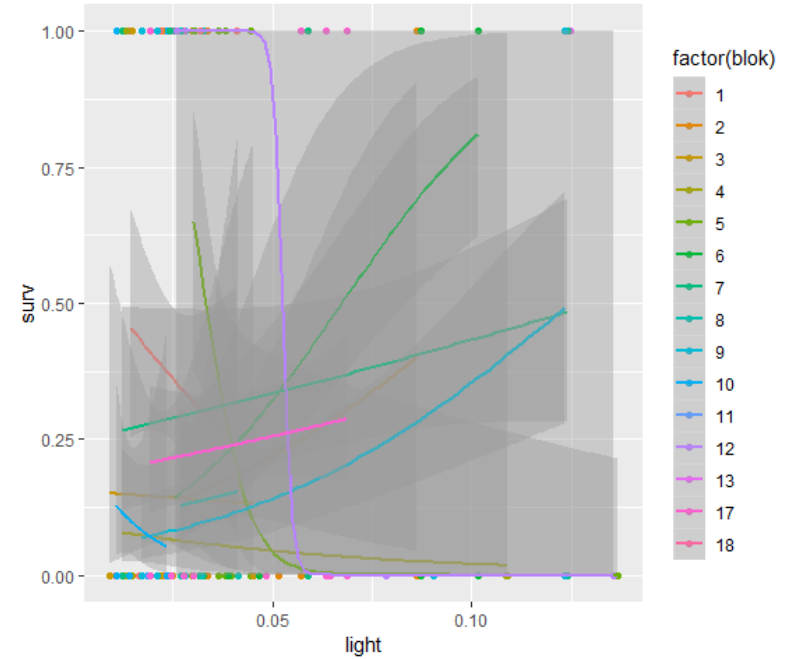
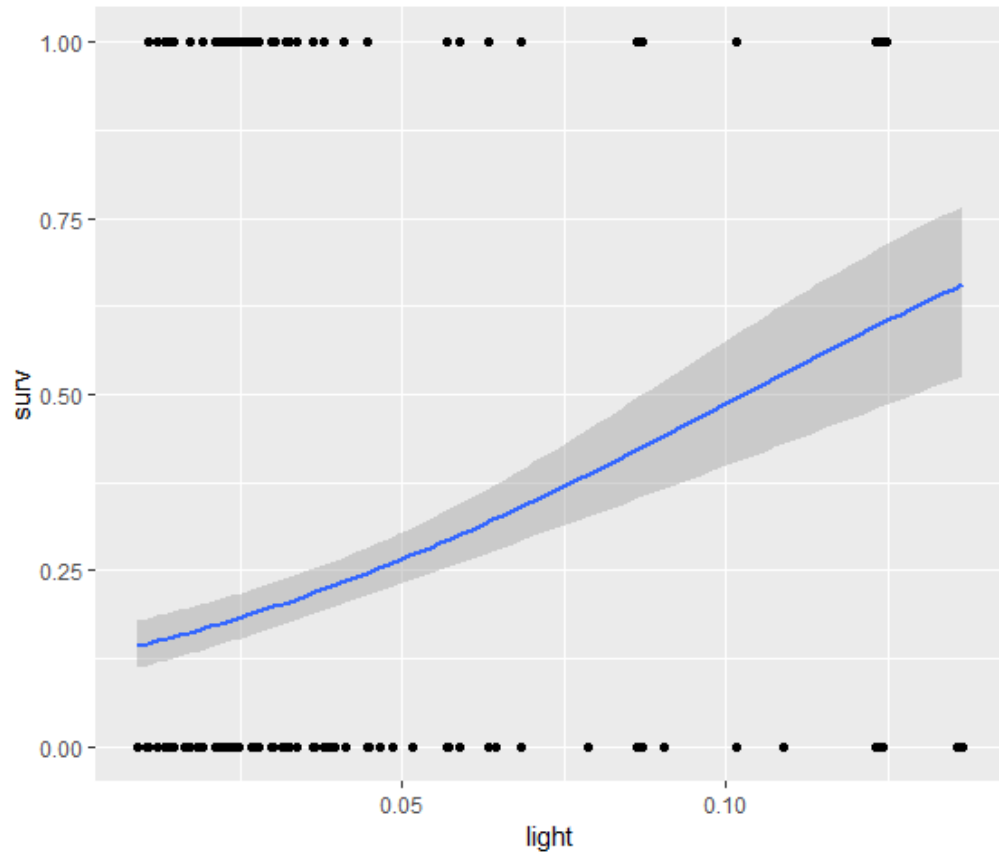
Zwiększenie pokrycia c z 40 do 50% zwiększa proporcję zgryzionych drzewek o 3%,  
zwiększenie z 80 na 100% o 8%

# Uogólnione LMM - Generalized Linear Mixed-effects Models (GLMM)

Ta sama zasada co w przypadku GLM - możemy uogólnić LMM na inne rozkłady

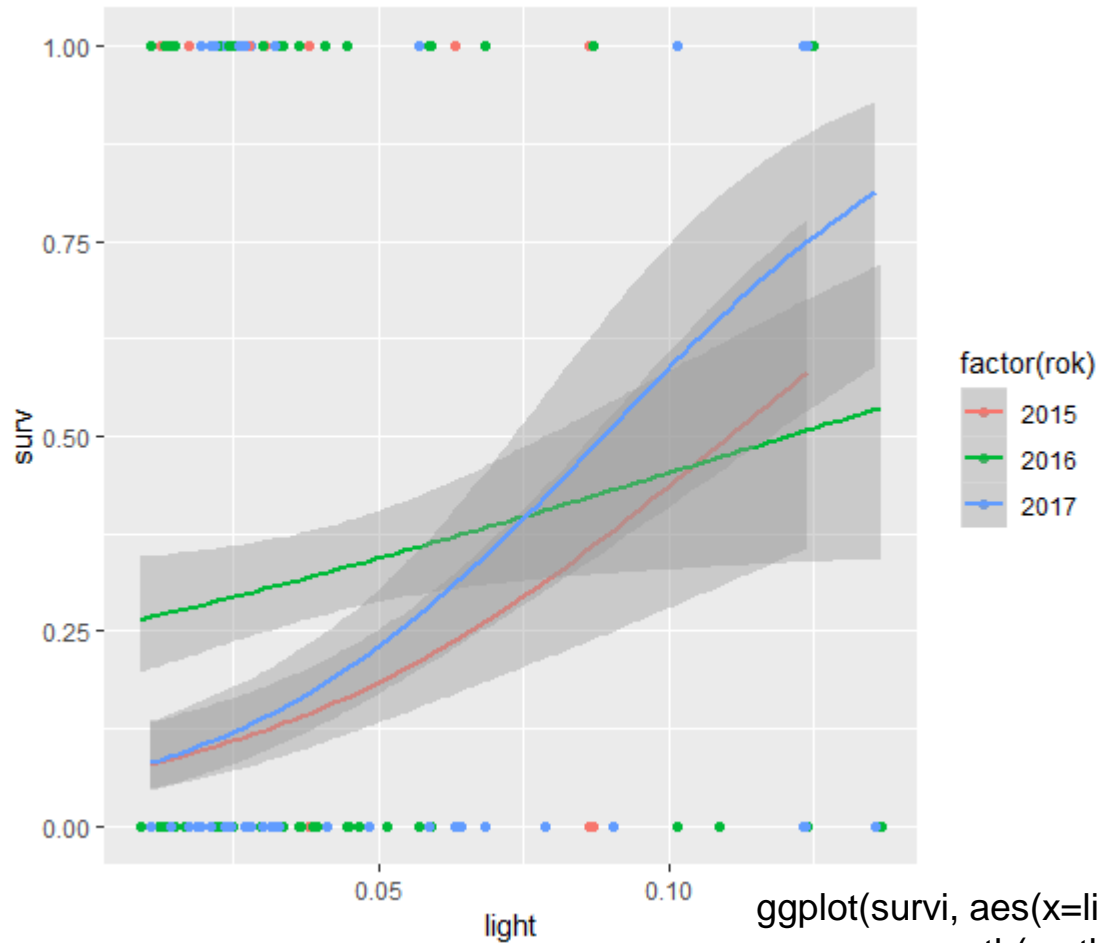
Przykład - przeżywalność siewek dębu czerwonego

Trzy lata, poletka w ramach bloków - efekty losowe



```
ggplot(survi, aes(x=light, y=surv, col=factor(blok)))+geom_point()  
+geom_smooth(method='glm',method.args=list(family='binomial'))
```

```
ggplot(survi, aes(x=light, y=surv))+geom_point()  
+geom_smooth(method='glm',method.args=list(family='binomial'))
```



Co się działo w tych latach?  
rok oznaczenia tasiemką,  
przeżywalność oceniona po roku

2015 - pierwsze tasiemkowanie  
2016 - dobra pomoc w terenie  
2017 - susza w 2018

```
ggplot(survi, aes(x=light, y=surv, col=factor(rok)))+geom_point()+  
+geom_smooth(method='glm',method.args=list(family='binomial'))
```

```
Call:
glm(formula = surv ~ light, family = binomial(link = "logit"),
  data = survi)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-1.4627	-0.6787	-0.6265	-0.5605	1.9640

Coefficients:

	Estimate	Std. Error	z value	Pr(> z )
(Intercept)	-1.978	0.159	-12.444	< 2e-16 ***
light	19.233	2.859	6.727	1.73e-11 ***

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 750.10 on 681 degrees of freedom  
Residual deviance: 704.07 on 680 degrees of freedom  
AIC: **708.07**

Number of Fisher Scoring iterations: 4

Generalized linear mixed model fit by maximum likelihood (Laplace Approximation) [glmerMod]

Family: binomial ( logit )  
Formula: surv ~ light + (1 | plot:blok) + (1 | rok)  
Data: survi  
AIC BIC logLik deviance df.resid  
**678.8** 696.9 -335.4 670.8 678

Scaled residuals:

Min	1Q	Median	3Q	Max
-2.0560	-0.4657	-0.3609	-0.2568	2.9111

Random effects:  
Groups Name Variance Std.Dev.  
plot:blok (Intercept) 0.89750 0.9474  
rok (Intercept) 0.09808 0.3132  
Number of obs: 682, groups: plot:blok, 127; rok, 3  
Fixed effects:

	Estimate	Std. Error	z value	Pr(> z )
(Intercept)	-2.243788	0.001140	-1968	<2e-16 ***
light	19.003078	0.001139	16677	<2e-16 ***

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

Correlation of Fixed Effects: **r.squaredGLMM(mod)**  
(Intr) R2m R2c  
light 0.000 theoretical 0.06727710 0.2839641  
convergence code: 0 delta 0.04502403 0.1900380

```
mod2<-glmer(surv~light+(light|plot:blok)+(1|rok),family=binomial(link='logit'),survi)
mod3<-glmer(surv~light+(light|plot:blok)+(light|rok),family=binomial(link='logit'),survi)
mod4<-glmer(surv~light+(1|plot:blok)+(light|rok),family=binomial(link='logit'),survi)
```

```
AIC(mod.lm, mod,mod2,mod3,mod4)
```

	df	AIC
mod.lm	2	708.0652
mod	4	678.8019
mod2	6	682.7923
mod3	8	681.2299
mod4	6	677.2307

Wniosek - random slop zależny od bloku i plotu - nie za bardzo, intercept - tak,  
random slop zależny od roku - tak



Generalized linear mixed model fit by maximum likelihood  
(Laplace Approximation) ['glmerMod']  
Family: binomial ( logit )  
Formula: surv ~ light + (1 | plot:blok) + (light | rok)  
Data: survi

AIC	BIC	logLik	deviance	df.resid
677.2	704.4	-332.6	665.2	676

Scaled residuals:

Min	1Q	Median	3Q	Max
-1.6123	-0.4963	-0.3524	-0.2451	3.0508

r.squaredGLMM(mod4)

	R2m	R2c
theoretical	0.07901921	0.2946918
delta	0.06523120	0.2432712

Random effects:

Groups	Name	Variance	Std.Dev.	Corr
plot:blok	(Intercept)	0.8019	0.8955	
rok	(Intercept)	0.4788	0.6919	
	light	58.6229	7.6566	-1.00

Number of obs: 682, groups: plot:blok, 127; rok, 3

Fixed effects:

	Estimate	Std. Error	z value	Pr(> z )
(Intercept)	-2.3249	0.4699	-4.948	7.51e-07 ***
light	20.7508	6.2316	3.330	0.000869 ***

---  
Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

Correlation of Fixed Effects:

(Intr)
light -0.891

convergence code: 0  
boundary (singular) fit: see ?isSingular

# Inny przykład – rozszerzenie modelu zgryzania

```
> mod.br<-glmmTMB(br~c+(1|set), ziformula = ~c+(1|set), data = prop,  
family=beta_family(), na.action = na.fail)
```

```
> summary(mod.br)
```

Family: beta ( logit )

Formula: br ~ c + (1 | set)

Zero inflation: ~c + (1 | set)

Data: prop

AIC	BIC	logLik	deviance	df.resid
-49.3	-37.4	31.6	-63.3	33

Random effects:

Conditional model:

Groups Name	Variance	Std.Dev.
set (Intercept)	0.02431	0.1559

Number of obs: 40, groups: set, 10

Zero-inflation model:

Groups Name	Variance	Std.Dev.
-------------	----------	----------

set (Intercept)	1.913e-08	0.0001383
-----------------	-----------	-----------

Number of obs: 40, groups: set, 10

Dispersion parameter for beta family (): 32.2

Conditional model:

	Estimate	Std. Error	z value	Pr(> z )
(Intercept)	-1.456134	0.386194	-3.770	0.000163 ***
c	0.015184	0.006423	2.364	0.018071 *

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

Zero-inflation model:

	Estimate	Std. Error	z value	Pr(> z )
(Intercept)	-8.40446	3.71108	-2.265	0.0235 *
c	0.08317	0.05067	1.642	0.1007

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

# GLMM Zero-Inflated Poisson

hab – siedlisko (las/pole)

part – część ciała

osobnik – ID sarny

```
mod.total<-glmmTMB(all~hab*part+(1|osobnik),ziformula =  
~hab*part,family = poisson,kl2)  
Family: poisson ( log )  
Formula:      all ~ hab * part + (1 | osobnik)  
Zero inflation: ~hab * part  
Data: kl2
```

AIC	BIC	logLik	deviance	df.resid
1813.4	1897.2	-885.7	1771.4	379

Random effects:

Conditional model:

Groups	Name	Variance	Std.Dev.
--------	------	----------	----------

osobnik	(Intercept)	1.769	1.33
---------	-------------	-------	------

Number of obs: 400, groups: osobnik, 80

Conditional model:

	Estimate	Std. Error	z value	Pr(> z )
(Intercept)	-1.0045	0.3973	-2.528	0.01147 *
habforest	1.0377	0.5141	2.018	0.04356 *
partfront leg	1.1313	0.3202	3.534	0.00041 ***

parthead	0.5646	0.3344	1.689	0.09131 .
parhind leg	0.5445	0.3348	1.626	0.10389
partneck	1.4791	0.3165	4.674	2.95e-06 ***
habforest:partfront leg	0.5420	0.4102	1.321	0.18640
habforest:parthead	0.9245	0.4230	2.186	0.02885 *
habforest:parhind leg	0.7593	0.4241	1.790	0.07339 .
habforest:partneck	0.5047	0.4063	1.242	0.21412
---				
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1				

Zero-inflation model:

	Estimate	Std. Error	z value	Pr(> z )
(Intercept)	0.89642	0.59288	1.512	0.13054
habforest	-0.47429	0.75932	-0.625	0.53222
partfront leg	-16.33248	549.24286	-0.030	0.97628
parthead	-4.66956	5.47047	-0.854	0.39333
parhind leg	-2.84490	1.06626	-2.668	0.00763 **
partneck	-14.16956	444.83585	-0.032	0.97459
habforest:partfront leg	-2.41148	2253.92771	-0.001	0.99915
habforest:parthead	2.61471	5.51341	0.474	0.63533
habforest:parhind leg	0.04727	1.35836	0.035	0.97224
habforest:partneck	10.40834	444.83724	0.023	0.98133

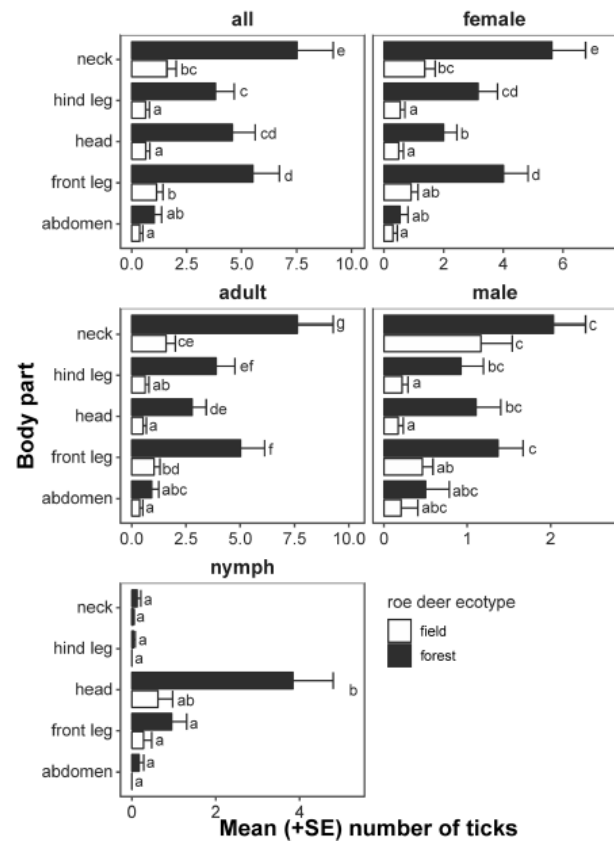
# Jak wyciągnąć wartości? cld i emmeans

```
cld(emmeans(mod.total, ~hab*part,type='response'))
```

hab	part	rate	SE	df	lower.CL	upper.CL	.group
field	abdomen	0.366	0.146	379	0.168	0.80	1
field	hind leg	0.631	0.179	379	0.361	1.10	1
field	head	0.644	0.184	379	0.368	1.13	1
forest	abdomen	1.034	0.340	379	0.542	1.97	12
field	front leg	1.135	0.298	379	0.678	1.90	2
field	neck	1.607	0.414	379	0.968	2.67	23
forest	hind leg	3.807	0.859	379	2.443	5.93	3
forest	head	4.582	1.029	379	2.947	7.13	34
forest	front leg	5.510	1.215	379	3.572	8.50	4
forest	neck	7.516	1.645	379	4.888	11.56	5

```
library(multcomp)
library(emmeans)
```

.group – test Tukeya



# Problemy z glmmTMB

Są – trzeba się na to nastawić że nie zawsze model wychodzi

Najczęściej – za dużo zmiennych w modelu – brak konwergencji – trzeba zmniejszyć liczbę zmiennych i efektów losowych

Nieraz pomaga zamiana zakresu jednostek (np. 5%=0,05) lub skalowanie predyktorów  
*MuMIn::dredge()* liczy się zwykle kilkadziesiąt razy dłużej

czasem problem z optymalizacją kodu – warto dodać w funkcji glmmTMB argument  
*control=glmmTMBControl(optimizer=optim,optArgs=list(method="BFGS"))*

więcej

<https://cran.r-project.org/web/packages/glmmTMB/vignettes/troubleshooting.html>

# Zagęszczenie siewek buka w PNGS – przykład

- $n=32$  poletka, każde 100 m<sup>2</sup>
- zależne przestrzennie – wzdłuż trzech rzek (random intercept riv)
- zaczynamy od modelu globalnego

```
mod.fsy1.glob<-glmmTMB(`Fagus sylvatica seedlings`~ekto+DIFN+ph+PC1  
+deadAB+(1|riv),data=od.df,family=poisson(),na.action=na.fail)#,  
control=glmmTMBControl(optimizer=optim,optArgs=list(method="BFGS")))
```

Warning message:

```
In (function (start, objective, gradient = NULL, hessian = NULL, :  
  NA/NaN function evaluation
```

# warning mnie niepokoi, wrzucam control

- `mod.fsy1.glob<-glmmTMB(`Fagus  
sylvatica  
seedlings`~ekto+DIFN+ph+PC1+de  
adAB+(1|riv),data=od.df,family=pois  
son(),na.action=na.fail,  
control=glmmTMBControl(optimizer  
=optim,optArgs=list(method="BFGS"  
)))`

- DHARMA:

`testZeroInflation(simulateResiduals(mod.fsy1.glob))`

DHARMA zero-inflation test via comparison to  
expected zeros with

simulation under H0 = fitted model

data: simulationOutput

ratioObsSim = 11.218, p-value < 2.2e-16

alternative hypothesis: two.sided

`> testDispersion(simulateResiduals(mod.fsy1.glob))`

DHARMA nonparametric dispersion test via sd  
of residuals fitted  
vs. simulated

data: simulationOutput

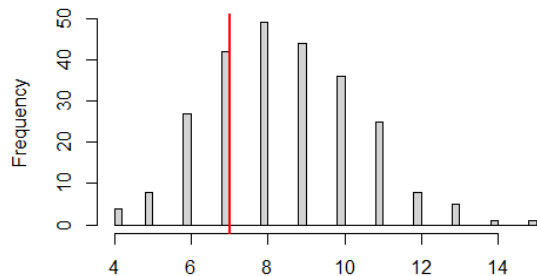
dispersion = 36.199, p-value < 2.2e-16

alternative hypothesis: two.sided

# Problem jest raczej z rozkładem niż z zerami

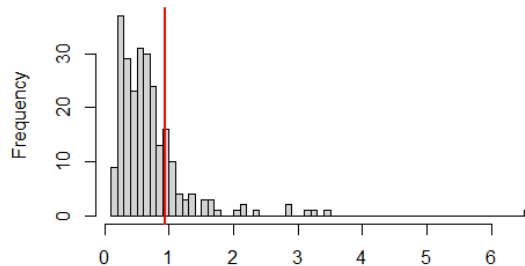
```
mod.fsy1.glob<-glmmTMB(`Fagus sylvatica  
seedlings`~ekto+DIFN+ph+PC1+deadAB  
+(1|riv),data=od.df,family=nbinom1(),na.action=na.fail,  
control=glmmTMBControl(optimizer=optim,optArgs=list(method="BFGS")))
```

DHARMA zero-inflation test via comparison to  
expected zeros with simulation under  $H_0$  = fitted  
model



Simulated values, red line = fitted model. p-value (two.sided) = 0.648

DHARMA nonparametric dispersion test via sd of  
residuals fitted vs. simulated



Simulated values, red line = fitted model. p-value (two.sided) = 0.392



# VIF

```
car::vif(mod.fsy1.glob)
```

```
Error in cov2cor(v) : 'V' is not a square numeric matrix
```

```
In addition: Warning message:
```

```
In vif.default(mod.fsy1.glob) : No intercept: vifs may not be sensible.
```

```
#Trzeba inaczej –sprawdzić vify na glmer lub glm:
```

```
car::vif(glmer(`Fagus sylvatica
```

```
seedlings` ~ekto+DIFN+ph+PC1+deadAB+(1|riv),data=od.df, family='poisson'))
```

ekto	DIFN	ph	PC1	deadAB
2.104512	2.995334	1.721692	1.271933	1.179923

#selekcja *mod.fsy1.dr* <- dredge(*mod.fsy1.glob*)

	cond((Int))	disp((Int))	cond(deadAB)	cond(DIFN)	cond(ekto)	cond(PC1)	cond(ph)	df	logLik	AICc	delta	weight
29	-7.910384	+	NA	NA	-0.3632284	-0.20213164	2.696815	6	-106.4601	228.4203	0.0000000	NA
17	-9.551967	+	NA	NA	NA	NA	2.891595	4	-109.6077	228.7539	0.3335784	NA
27	-7.365725	+	NA	-0.3621034	NA	-0.18528601	2.552524	6	-106.8939	229.2878	0.8674838	NA
21	-6.766921	+	NA	NA	-0.2183832	NA	2.364828	5	-108.6916	229.7832	1.3629522	NA
31	-6.729590	+	NA	-0.1854773	-0.2647744	-0.21090792	2.459733	7	-106.1833	231.2361	2.8158257	NA
3	3.917263	+	NA	-0.4346936	NA	NA	NA	4	-110.9277	231.3939	2.9735975	NA
30	-7.985040	+	0.0009138896	NA	-0.3838958	-0.20329323	2.710139	7	-106.3247	231.5190	3.0986756	NA
18	-9.569007	+	0.0001250795	NA	NA	NA	2.893913	5	-109.6056	231.6113	3.1909908	NA
11	4.182706	+	NA	-0.5721939	NA	-0.16090004	NA	5	-109.7486	231.8971	3.4768187	NA
13	4.222116	+	NA	NA	-0.5290239	-0.18556971	NA	5	-109.8506	232.1012	3.6808992	NA
5	3.877793	+	NA	NA	-0.3620902	NA	NA	4	-111.4513	232.4410	4.0207200	NA
28	-7.377845	+	0.0001473191	-0.3625592	NA	-0.18404854	2.553505	7	-106.8906	232.6507	4.2304411	NA
22	-6.747674	+	0.0007817547	NA	-0.2326178	NA	2.357123	6	-108.6234	232.7467	4.3264260	NA
20	-6.020576	+	0.0004486092	-0.2484428	NA	NA	2.188312	6	-108.6790	232.8580	4.4376678	NA

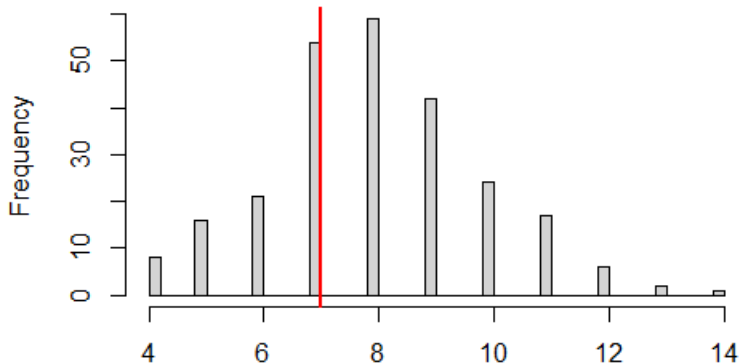
Showing 1 to 14 of 32 entries

Console: E:\Nauka\Projekty badawcze\pnps\pnps2021\Banal/

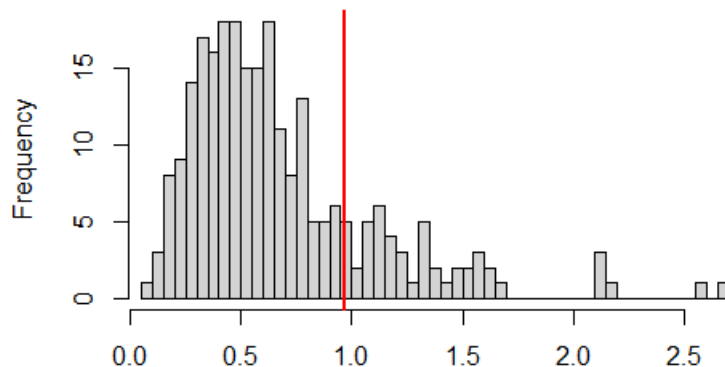
# model finalny

```
mod.fsy1.fin<-glmmTMB(`Fagus sylvatica  
seedlings`~ekto+PC1+ph+(1|riv),data=od.df,family=nbinom1(),na.action=na.fail,  
control=glmmTMBControl(optimizer=optim,optArgs=list(method="BFGS")))
```

**DHARMA zero-inflation test via comparison to  
expected zeros with simulation under H0 = fitted  
model**



**DHARMA nonparametric dispersion test via sd of  
residuals fitted vs. simulated**



# summary(mod.fsy1.fin)

Family: nbinom1 ( log )

Formula: `Fagus sylvatica seedlings` ~ ekto +  
PC1 + ph + (1 | riv)

Data: od.df

AIC	BIC	logLik	deviance	df.resid
224.9	233.5	-106.5	212.9	25

Random effects:

Conditional model:

Groups Name	Variance	Std.Dev.
riv (Intercept)	3.427e-05	0.005854

Number of obs: 31, groups: riv, 3

Dispersion parameter for nbinom1 family (): 35.6

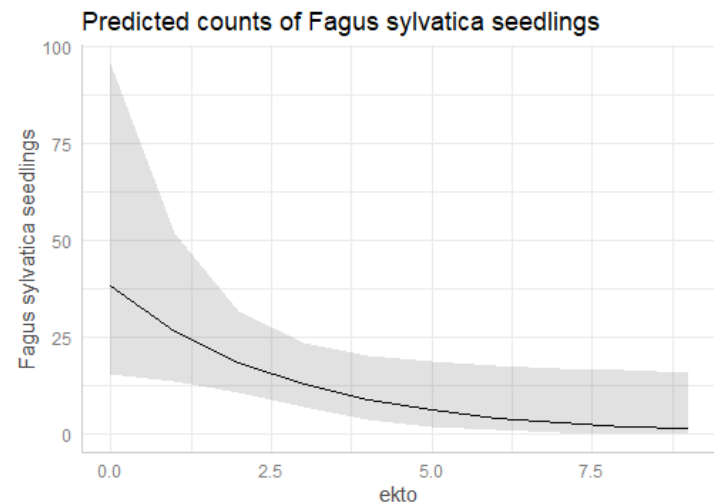
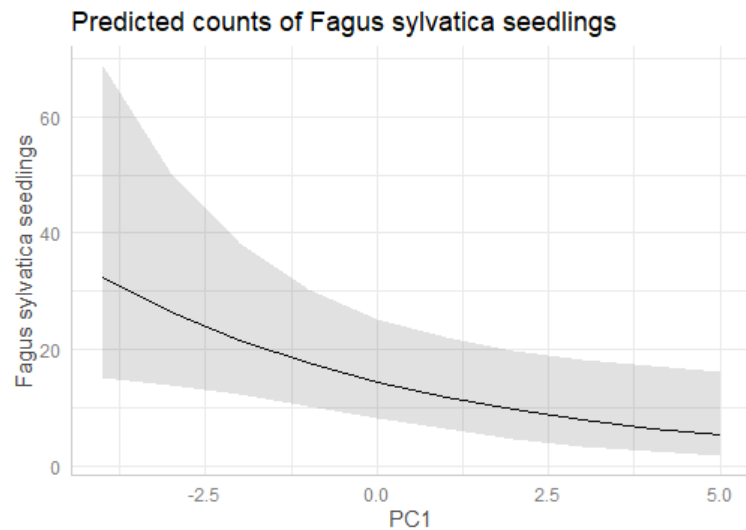
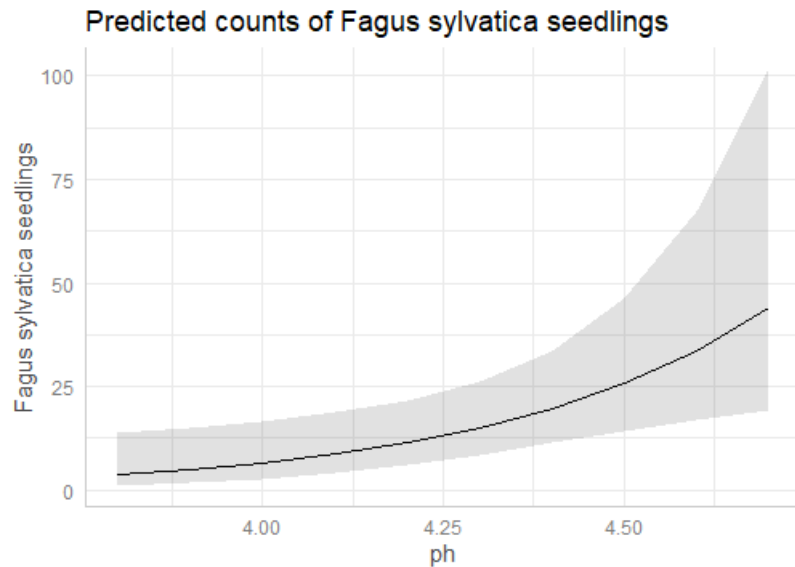
Conditional model:

	Estimate	Std. Error	z value	Pr(> z )
(Intercept)	-7.91038	4.61592	-1.714	0.08658 .
ekto	-0.36323	0.17454	-2.081	0.03743 *
PC1	-0.20213	0.08532	-2.369	0.01783 *
ph	2.69681	1.01302	2.662	0.00776 **

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

```
plot(ggpredict(mod.fsy1.fin))
```



# Uogólnione modele addytywne - GAM

Generalized Additive Models

Po co? nieparametryczne - nie zawsze rozkłady odpowiadają rzeczywistym danym i założeniom modeli

Co zamiast funkcji liniowych? Funkcje sklepane (splines)

Zamiast składników liniowych - składniki nieliniowe

Wada: słabsza możliwość wykorzystania, trudniejsze interpretacyjnie

# Funkcje sklejane (splines)

dla  $x \in (-\infty, -10)$   $y=12x+2$

dla  $x \in [-10, 2)$   $y=17^x$

dla  $x \in [2, 7)$   $y=2-x^3$

dla  $x \in [7, \infty)$   $y=2x^2-3.122$

mamy cztery funkcje, sklejone w jedną (wygładzoną)

Mamy trzy punkty przegięcia - w których funkcja zmienia bieg

# Przykład zastosowania

wzorce rozwoju korzeni drobnych

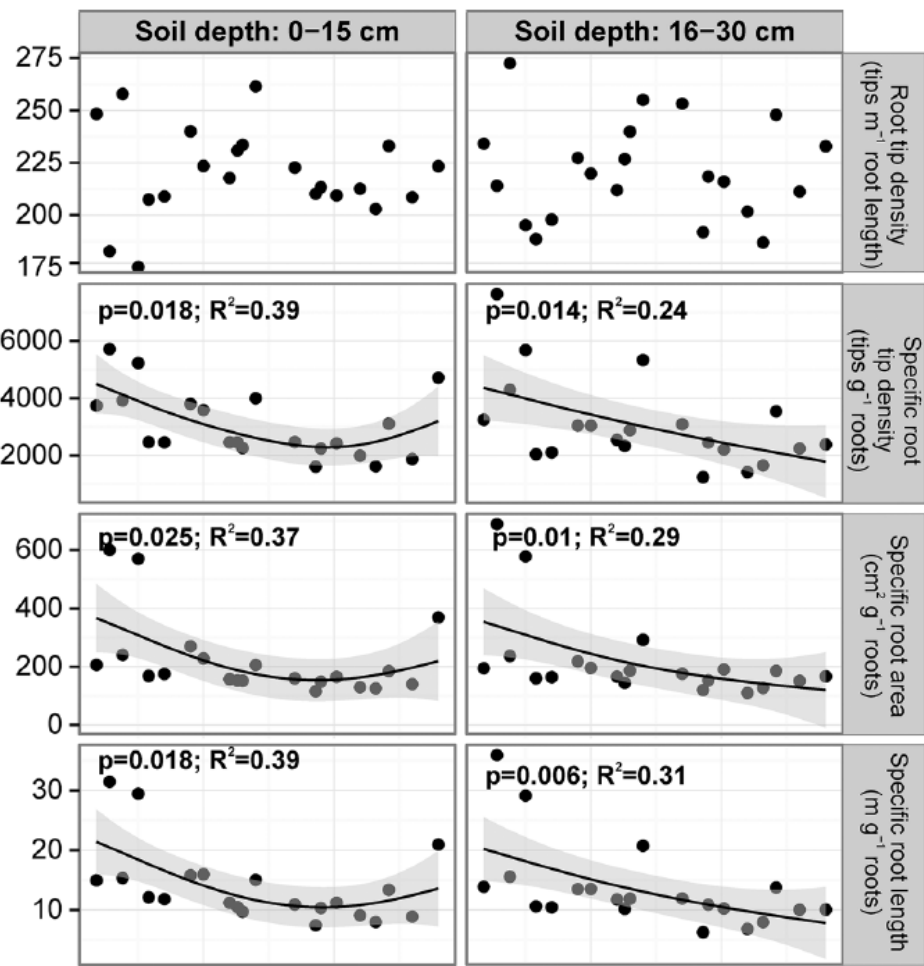
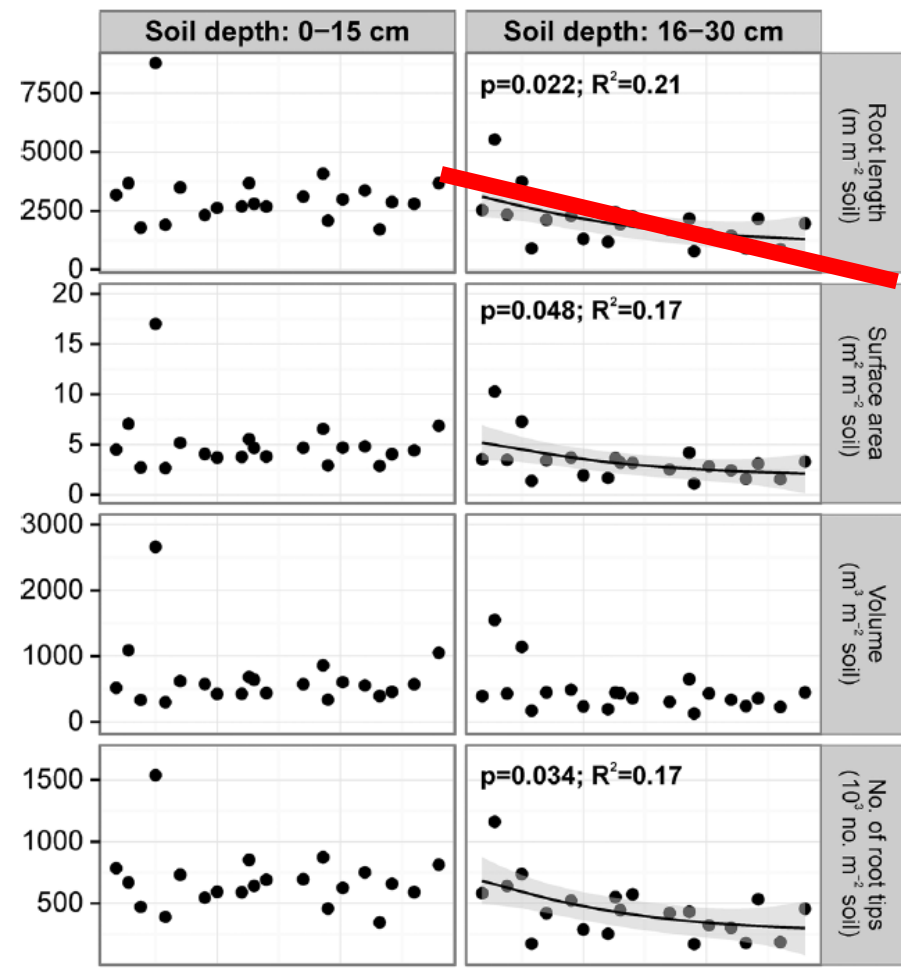


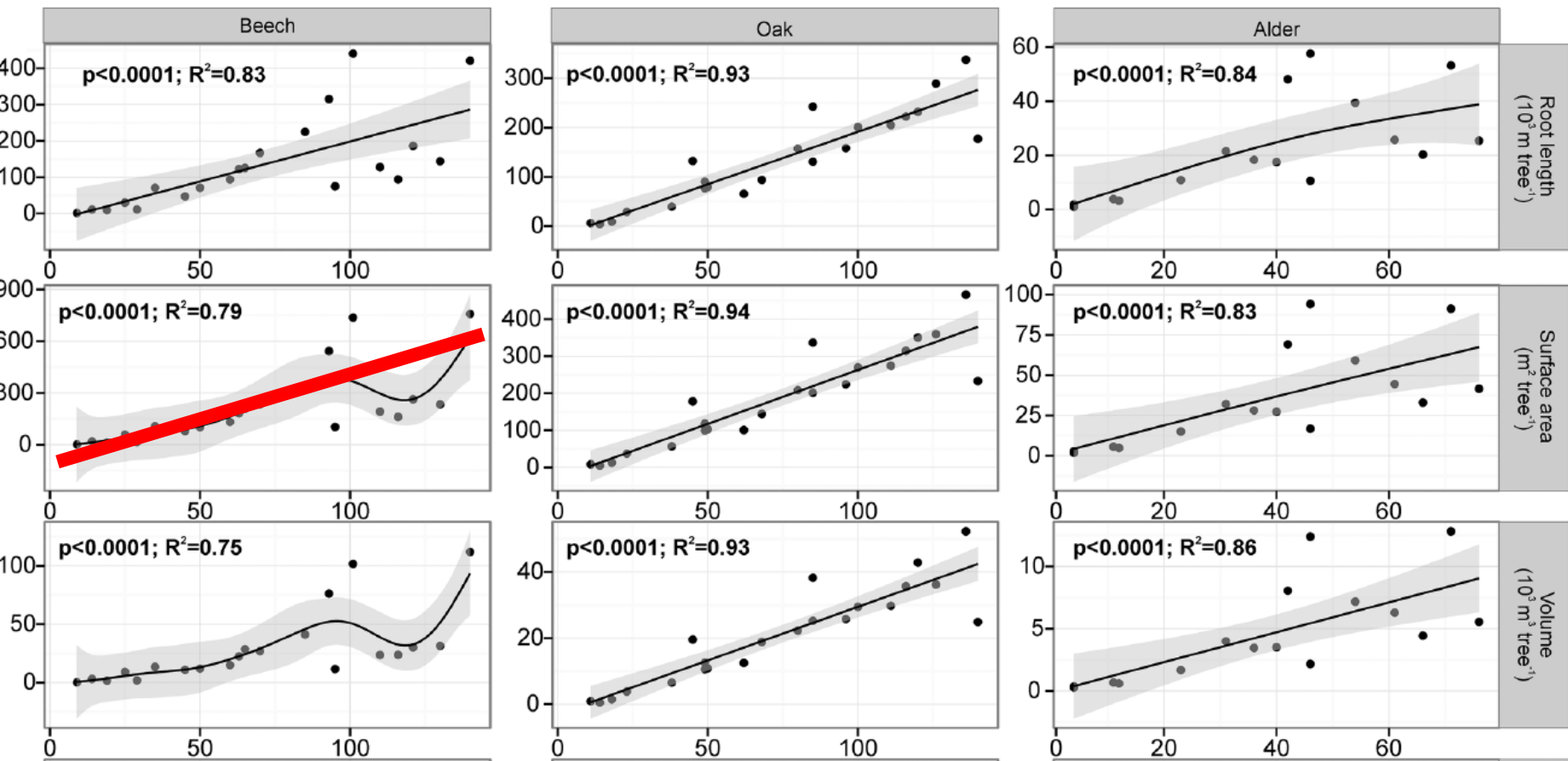
RESEARCH ARTICLE

## Tree Age Effects on Fine Root Biomass and Morphology over Chronosequences of *Fagus sylvatica*, *Quercus robur* and *Alnus glutinosa* Stands

Andrzej M. Jagodzinski<sup>1,2\*</sup>, Jędrzej Ziółkowski<sup>2</sup>, Aleksandra Warnkowska<sup>2</sup>, Hubert Prais<sup>2</sup>







```

> library(mgcv)
> model1<-gam(AB~s(V),data=sosny)
> summary(model1)

```

```

Family: gaussian
Link function: identity

```

```

Formula:
AB ~ s(V)

```

```

Parametric coefficients:

```

	Estimate	Std. Error	t value	Pr(> t )
(Intercept)	21.5752	0.3845	56.11	<2e-16 ***

```

---
signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

```

```

Approximate significance of smooth terms:

```

	edf	Ref.df	F	p-value
s(V)	3.701	4.546	460.4	<2e-16 ***

```

---
signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

```

```

R-sq.(adj) =  0.965   Deviance explained = 96.7%
GCV = 12.125   Scale est. = 11.385       n = 77

```

```

>

```

w modelu zamiast V  
mamy s(V), czyli  
funkcję sklejaną

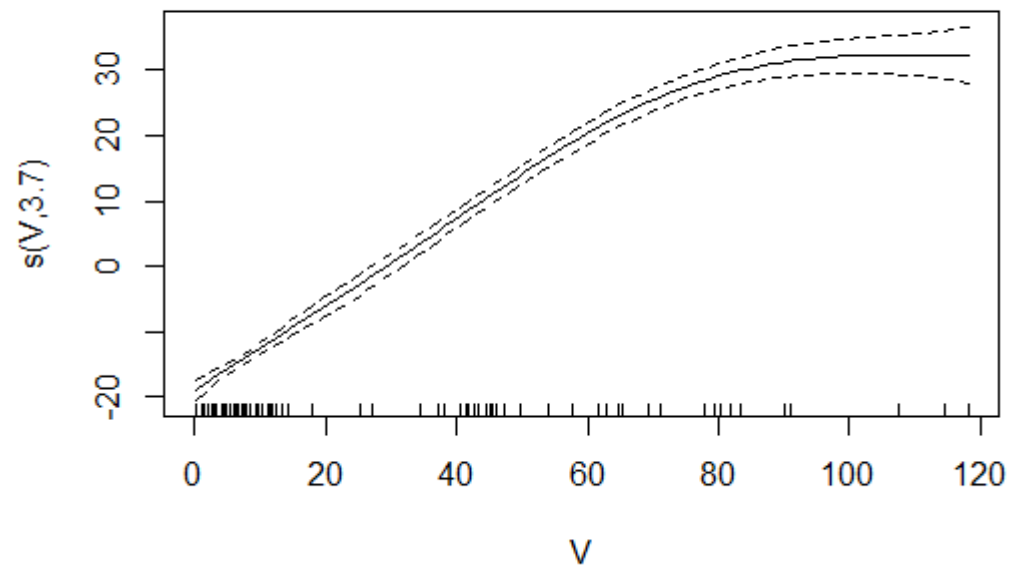
jest to wielomian  
stopnia 3,7  
(patrzymy na edf)

edf - estimated degrees  
of freedom

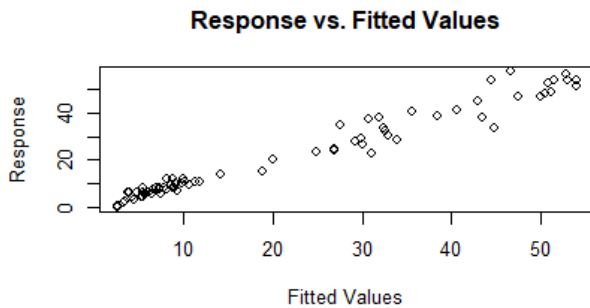
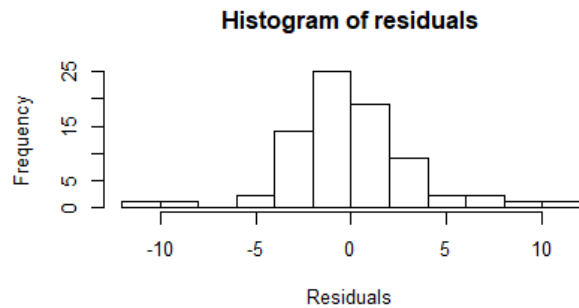
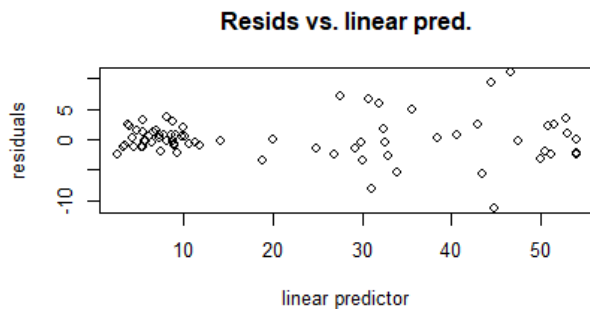
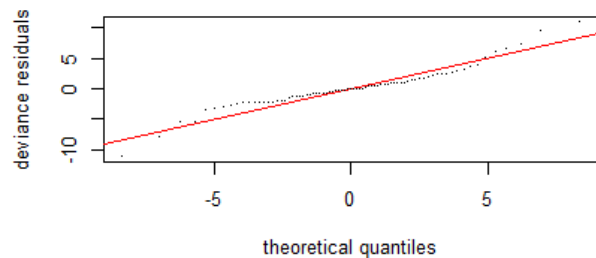
Ref.df - reference  
degrees of freedom

F - statystyka testowa

plot(model1)



```
par(mfrow=c(2,2))  
gam.check(model1)
```



```
> AIC(model1)
```

```
[1] 412.3526
```

```
> AIC(model0)
```

```
[1] 666.9126
```

# poprzednie modele

```
> AIC(lm(AB~1,data=sosny)) #model zerowy
```

```
[1] 666.9126
```

```
> AIC(lm(AB~V,data=sosny)) #model liniowy
```

```
[1] 467.6902
```

```
> AIC(lm(AB~poly(V,2),data=sosny)) #model kwadratowy
```

```
[1] 422.4431
```

```
> AIC(nls(AB~a*V^b,data=sosny,start=list(a=1,b=-2))) #model potęgowy
```

```
[1] 441.9817
```

```
> AIC(gam(AB~s(V),data=sosny)) #GAM
```

```
[1] 412.3526
```

```
> plot(modelbr)
> modelbr<-gam(BR~s(Age),data=sosny)
> summary(modelbr)
```

Family: gaussian  
Link function: identity

Formula:  
BR ~ s(Age)

Parametric coefficients:

	Estimate	Std. Error	t value	Pr(> t )
(Intercept)	5.4899	0.3988	13.77	<2e-16 ***

---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

Approximate significance of smooth terms:

	edf	Ref.df	F	p-value
s(Age)	7.751	8.566	9.005	3.45e-09 ***

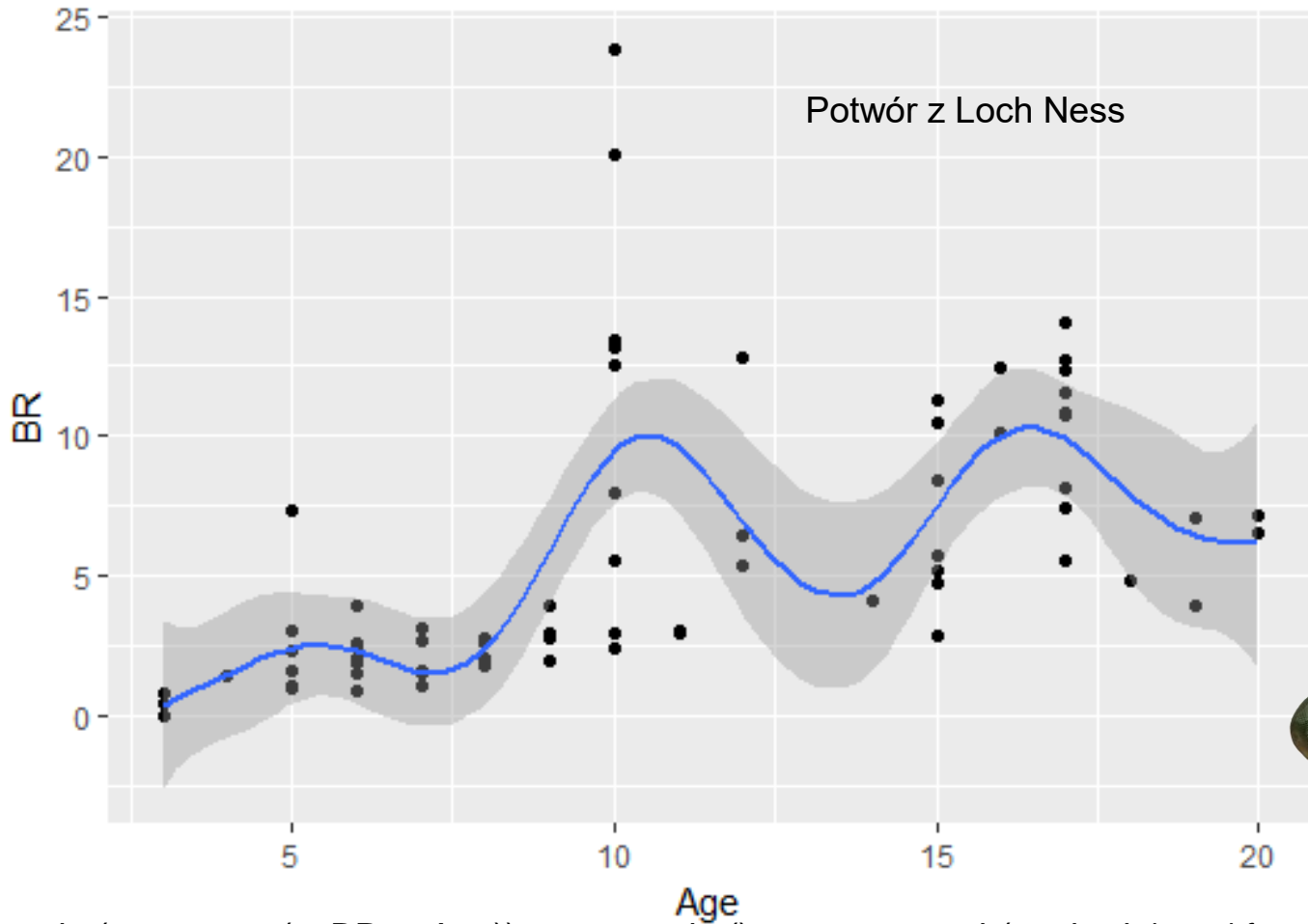
---

Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

R-sq.(adj) = 0.486    Deviance explained = 53.9%  
GCV = 13.814    Scale est. = 12.244    n = 77

```
> |
```





albo Nagini



<https://harrypotter.fandom.com/pl/wiki/Nagini>

```
ggplot(sosny, aes(y=BR,x=Age))+geom_point()+geom_smooth(method='gam',formula=y~s(x))
```

# With **four** parameters I can fit an elephant, and with **five** I can make him wiggle his trunk.

## Drawing an elephant with four complex parameters

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We define four complex numbers representing the parameters needed to specify an elephantine shape. The real and imaginary parts of these complex numbers are the coefficients of a Fourier coordinate expansion, a powerful tool for reducing the data required to define shapes. © 2010 American Association of Physics Teachers  
[DOI: 10.1119/1.3254017]

A turning point in Freeman Dyson's life occurred during a meeting in the Spring of 1953 when Enrico Fermi criticized the complexity of Dyson's model by quoting Johnny von Neumann: "With four parameters I can fit an elephant, and with five I can make him wiggle his trunk." Since then it has become a well-known saying among physicists, but nobody has successfully implemented it.

To parametrize an elephant, we note that its perimeter can be described as a set of points  $(x(t), y(t))$ , where  $t$  is a parameter that can be interpreted as the elapsed time while going along the path of the contour. If the speed is uniform,  $t$  becomes the arc length. We expand  $x$  and  $y$  separately<sup>2</sup> as a Fourier series

$$x(t) = \sum_{k=0}^{\infty} (A_k^x \cos(kt) + B_k^x \sin(kt)), \quad (1)$$

$$y(t) = \sum_{k=0}^{\infty} (A_k^y \cos(kt) + B_k^y \sin(kt)), \quad (2)$$

where  $A_k^x$ ,  $B_k^x$ ,  $A_k^y$ , and  $B_k^y$  are the expansion coefficients. The lower indices  $k$  apply to the  $k$ th term in the expansion, and the upper indices denote the  $x$  or  $y$  expansion, respectively.

Using this expansion of the  $x$  and  $y$  coordinates, we can analyze shapes by tracing the boundary and calculating the coefficients in the expansions (using standard methods from Fourier analysis). By truncating the expansion, the shape is smoothed. Truncation leads to a huge reduction in the information necessary to express a certain shape compared to a pixelated image, for example. Székely *et al.*<sup>3</sup> used this approach to segment magnetic resonance imaging data. A similar approach was used to analyze the shapes of red blood cells,<sup>4</sup> with a spherical harmonics expansion serving as a 3D generalization of the Fourier coordinate expansion.

The coefficients represent the best fit to the given shape in the following sense. The  $k=0$  component corresponds to the center of mass of the perimeter. The  $k=1$  component corresponds to the best fit ellipse. The higher order components

trace out elliptical corrections analogous to Ptolemy's epicycles.<sup>5</sup> Visualization of the corresponding ellipses can be found at Ref. 6.

We now use this tool to fit an elephant with four parameters. Wei<sup>7</sup> tried this task in 1975 using a least-squares Fourier sine series but required about 30 terms. By analyzing the picture in Fig. 1(a) and eliminating components with amplitudes less than 10% of the maximum amplitude, we obtained an approximate spectrum. The remaining amplitudes were

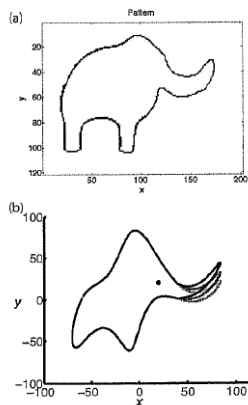


Fig. 1. (a) Outline of an elephant. (b) Three snapshots of the wagging trunk.

~John vonNeumann

# Czy jest to biologicznie uzasadnione?

Co się dzieje w wieku 10 lat?

Czy coś takiego przejdzie?

na obronie doktoratu - zależy od audytorium

w dobrym czasopiśmie - nie bardzo

im większy stopień wielomianu/głębokość splinu tym lepsze dopasowanie

na czym nam zależy?

**overfitting**

# Overfitting i ekstrapolacja

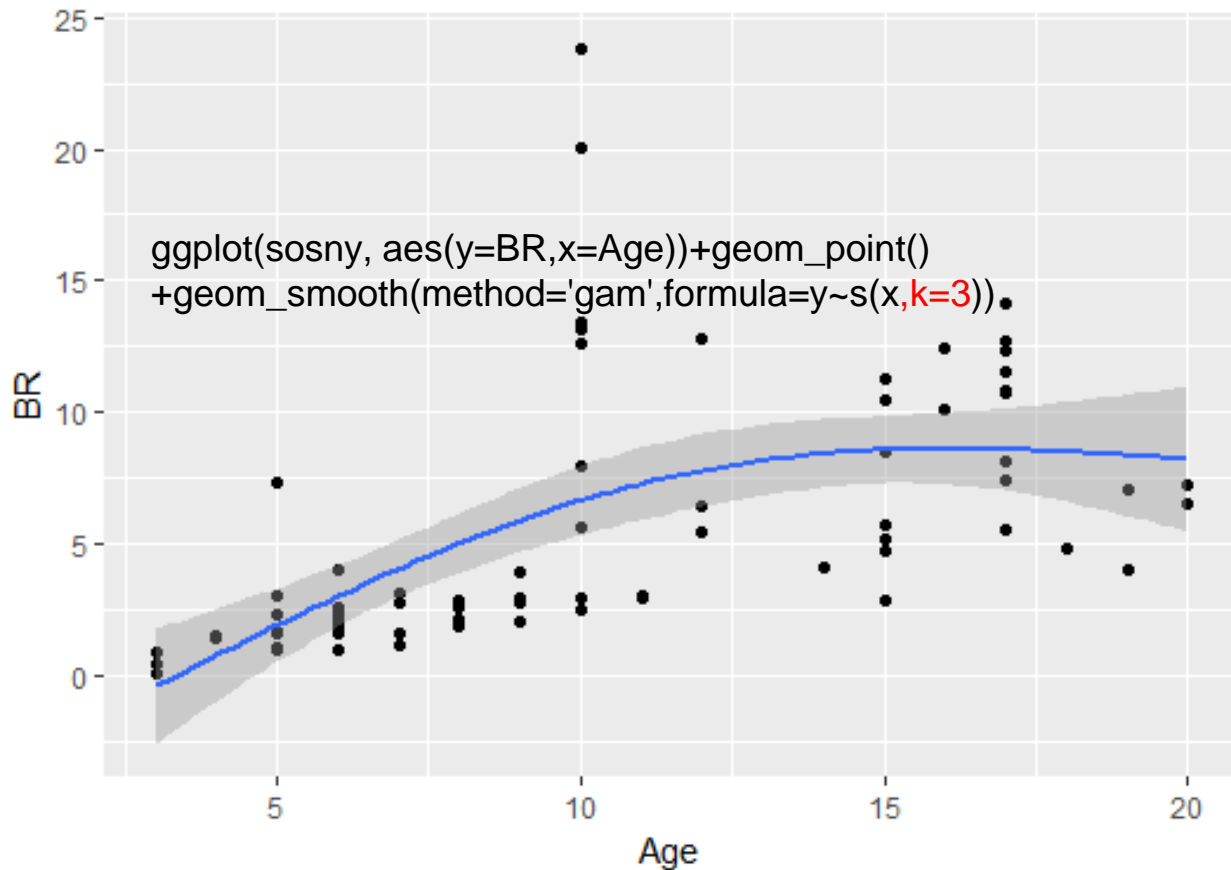
overfitting - model dobrze działa na zbiorze treningowym, ale źle na testowym  
brak możliwości ekstrapolacji

Po co robimy modele? Aby coś uogólnić, wyciągnąć trend

Oddzielić ziarno od plew - wiedzę biologiczną od elementów lokalnych i szumu

Stąd założenia o niezależności prób, wielkość próby, powtórzenia ...

# rozwiązanie - ograniczyć spline



```
', formula=y~s(x,k=3))  
> modelbr<-gam(BR~s(Age,k=3), data=sosny)  
> summary(modelbr)
```

Family: gaussian  
Link function: identity

Formula:  
BR ~ s(Age, k = 3)

Parametric coefficients:

	Estimate	Std. Error	t value	Pr(> t )
(Intercept)	5.4899	0.4488	12.23	<2e-16 ***

---  
Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

Approximate significance of smooth terms:

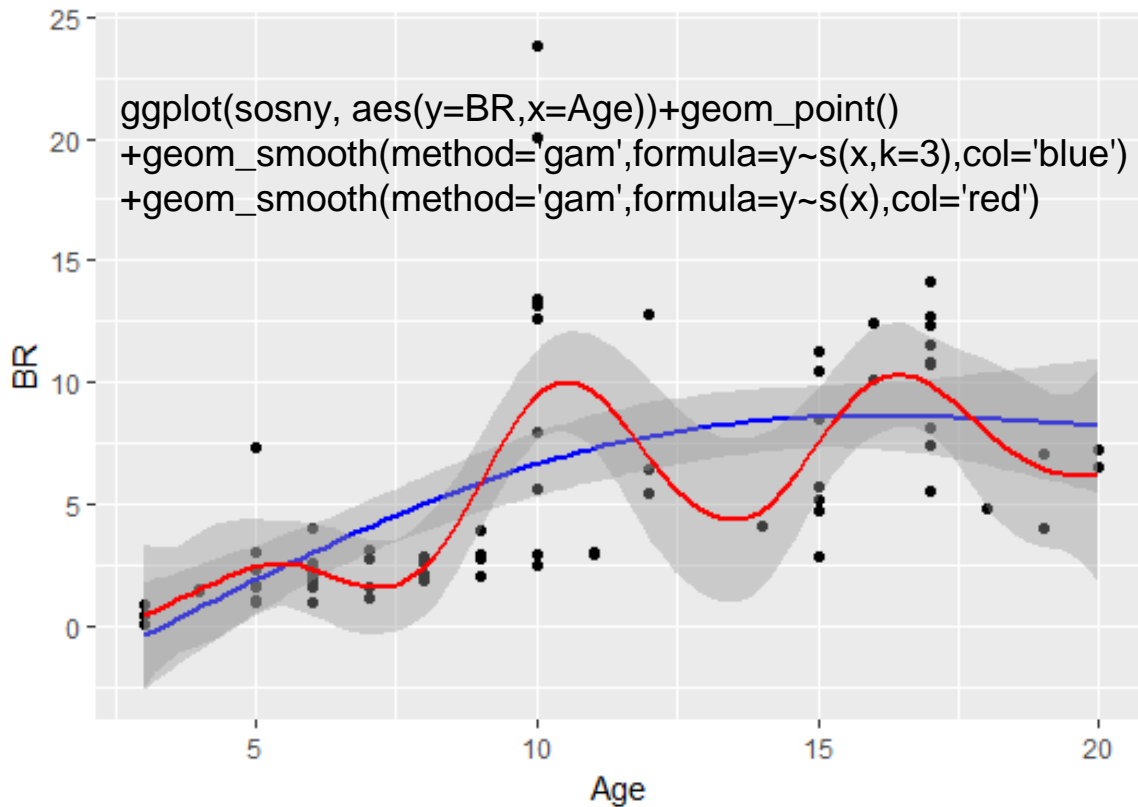
	edf	Ref.df	F	p-value
s(Age)	1.87	1.983	22.44	6.6e-08 ***

---  
Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

R-sq.(adj) = 0.349    Deviance explained = 36.5%  
GCV = 16.112    Scale est. = 15.512    n = 77

```
> |
```

# Tracimy trochę R2 ale model jest bardziej sensowny



... all models are approximations. Essentially, **all models are wrong, but some are useful**. However, the approximate nature of the model must always be borne in mind....

~George E. P. Box





BIAŁOWIESKA SZKOŁA STATYSTYKI