Similarity Searches on Sequence Databases

Lorenza Bordoli Swiss Institute of Bioinformatics EMBnet Course, Zürich, October 2004

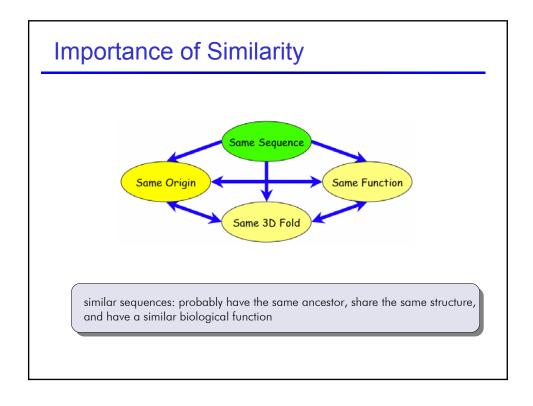


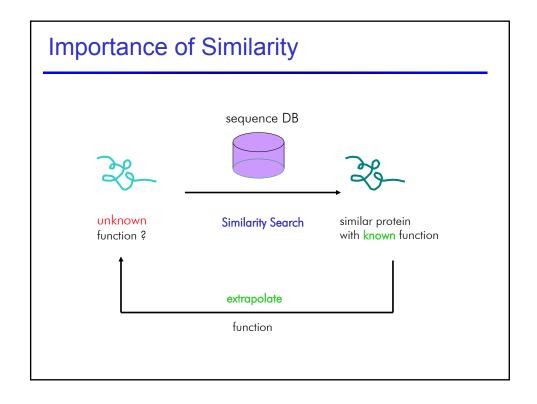
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Outline

- · Importance of Similarity
- · Heuristic Sequence Alignment:
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 - FASTA algorithm
 - BLAST algorithm
- Assessing the significance of sequence alignment
 - Raw score, normalized (bits) score, P-value, E-Value
- BLAST:
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 - Choosing the right Parameters
- Other members of the BLAST family





Importance of Similarity

Rule-of-thumb:

If your sequences are more than 100 amino acids long (or 100 nucleotides long) you can considered them as homologues if 25% of the aa are identical (70% of nucleotide for DNA). Below this value you enter the twilight zone.

Twilight zone = protein sequence similarity between \sim 0-20% identity: is not statistically significant, i.e. could have arisen by chance.

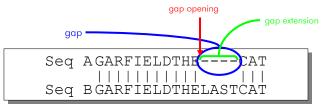


Beware:

- E-value (Expectation value)
- Length of the segments similar between the two sequences
- The number of insertions/deletions

Insertions and deletions

Gap penalties



- Opening a gap penalizes an alignment score
- Each extension of a gap penalizes the alignment's score
- The gap opening penalty is in general higher than the gap extension penalties (simulating evolutionary behavior)
- The raw score of a gapped alignment is the sum of all amino acid substitutions from which we subtract the gap opening and extension penalties.

Alignment

Alignement types:

• Global Alignment between the complete sequence A and the

complete sequence B

• Local Alignment between a sub-sequence of A and a sub-

sequence of B

Computer implementation (Algorithms):

Dynamic programing (exact algorithm)

Global Needleman-WunschLocal Smith-Waterman

Heuristic Sequence Alignment

- With the Dynamic Programming algorithm, one obtain an alignment in a time that is proportional to the product of the lengths of the two sequences being compared. Therefore when searching a whole database the computation time grows linearly with the size of the database. With current databases calculating a full Dynamic Programming alignment for each sequence of the database is too slow (unless implemented in a specialized parallel hardware).
- The number of searches that are presently performed on whole genomes creates a need for faster procedures.
- ⇒ Two methods that are least 50-100 times faster than dynamic programming were developed: FASTA and BLAST

Heuristic Sequence Alignment: Principle

- Dynamic Programming: computational method that provide in mathematical sense the best alignment between two sequences, given a scoring system.
- Heuristic Methods (e.g. BLAST, FASTA) they prune the search space by using fast approximate methods to select the sequences of the database that are likely to be similar to the query and to locate the similarity region inside them
 - =>Restricting the alignment process:
 - Only to the selected sequences
 - Only to some portions of the sequences (search as small a fraction as possible of the cells in the dynamic programming matrix)

Heuristic Sequence Alignment: Principle

- These methods are heuristic; i.e., an empirical method of computer programming in which rules of thumb are used to find solutions.
- They almost always works to find related sequences in a database search but does not have the underlying guarantee of an optimal solution like the dynamic programming algorithm (But good ones often do).
- Advantage: This methods that are least 50-100 times faster than dynamic programming therefore better suited to search databases.

FASTA & BLAST: story

1985 : FASTP (D. Lipman and W. Pearson)

Global gapped alignments

1988: FASTA (W. Pearson and D. Lipman)

Local gapped alignments

1990 : BLAST1

(S. Altschul, W. Gish, W. Miller, E. Myers, and D. Lipman)

Local ungapped alignments

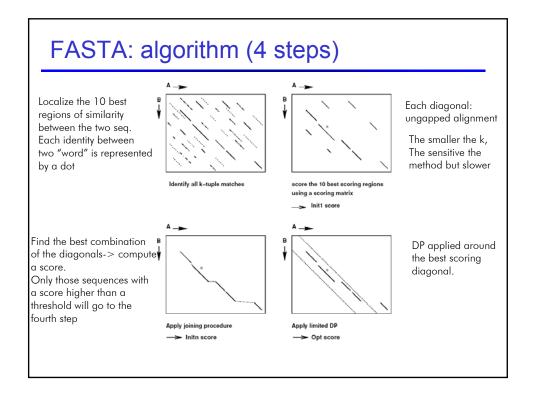
Gapped BLASTs:

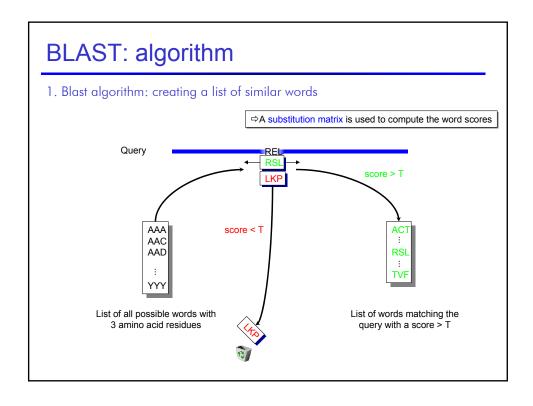
1996: WU-BLAST2 (W. Gish)

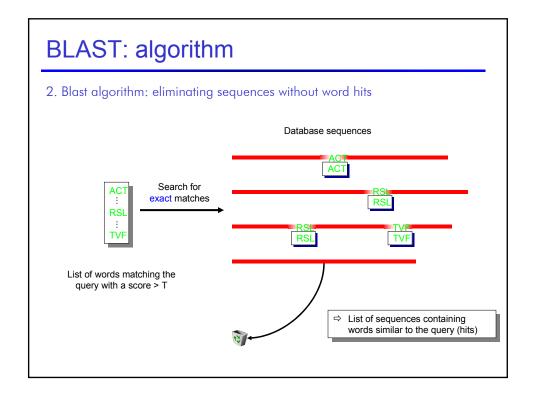
1997: NCBI-BLAST2 (and PSI-BLAST)

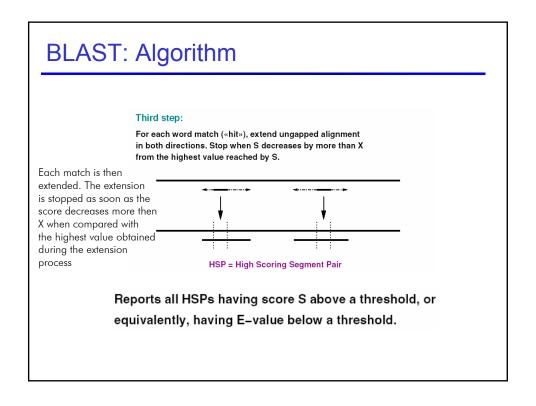
(S. Altschul, T. Madden, A. Schaffer, J. Zhang, Z. Zhang,

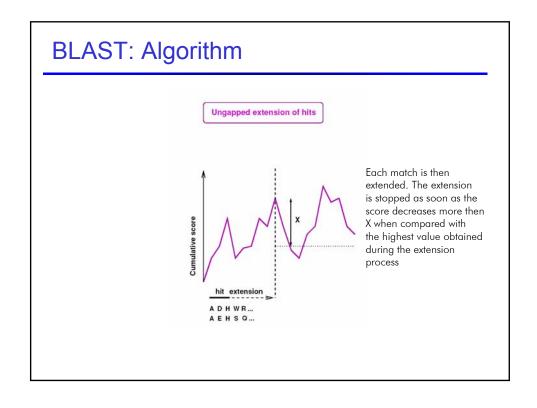
W. Miller and D. Lipman)

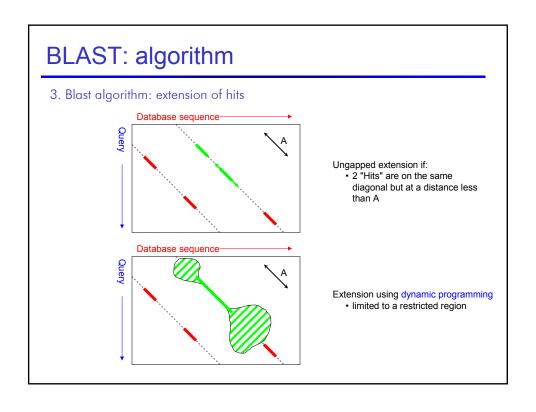


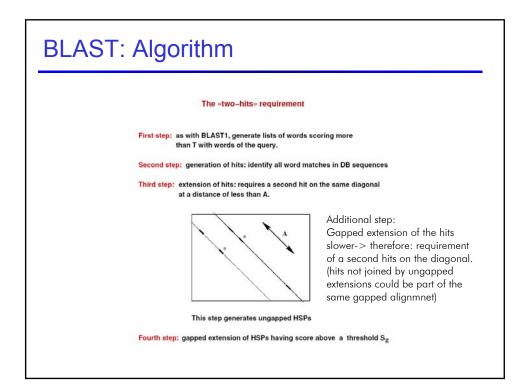












Assessing the significance of sequence alignment

- Scoring System:
 - 1. Scoring (Substitution) matrix (or match mismatch for DNA): In proteins some mismatches are more acceptable than others. Substitution matrices give a score for each substitution of one amino-acid by another (e.g. PAM, BLOSUM)
 - 2. Gap Penalties: simulate as closely as possible the evolutionary mechanisms involved in gap occurrence. Gap opening penalty: Counted each time a gap is opened in an alignment and Gap extension penalty: Counted for each extension of a gap in an alignment.
- Based on a given scoring system: you can calculate the raw score of the alianment
 - Raw score= sum of the amino acid substitution scores and gap penalties

Assessing the significance of sequence alignment



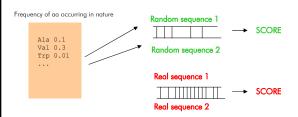
Caveats:

- 1. We need a normalised (bit) score to compare different alignments, based on different scoring systems, e.g. different substitution matrices.
- 2. A method to asses the statistical significance of the alignment is needed (is an alignment biological relevant?): E-value

Assessing the significance of sequence alignment

- How?
 - ⇒ Evaluate the probability that a score between random or unrelated sequences will reach the score found between two real sequences of interest:

If that probability is very low, the alignment score between the real sequences is significant.



If SCORE > SCORE => the alignment between the real sequences is significant

Assessing the significance of sequence alignment

Statistics derived from the scores:



- P-value
 - ⇒ Probability that an alignment with this score occurs by chance in a database of this size
 - ⇒ The closer the P-value is towards 0, the better the alignment



- E-value
 - ⇒ Number of matches with this score one can expect to find by chance in a database of size N
 - ⇒ The closer the e-value is towards 0, the better the alignment
- Relationship between E-value and P-value:
 - \Rightarrow In a database containing N sequences $E = P \times N$

BLAST

Basic Local Alignment Search Tool

A Blast for each query Different programs are available according to the type of query Program Query Database protein protein blastp blastn nucleotide nucleotide blastx nucleotide protein protein tblastn nucleotide protein protein tblastx nucleotide nucleotide protein protein

BLASTing protein sequences

blastp = Compares a protein sequence with a protein database

If you want to find something about the function of your protein, use **blastp** to compare your protein with other proteins contained in the databases; identify common regions between proteins, or collect related proteins (phylogenetic analysis;

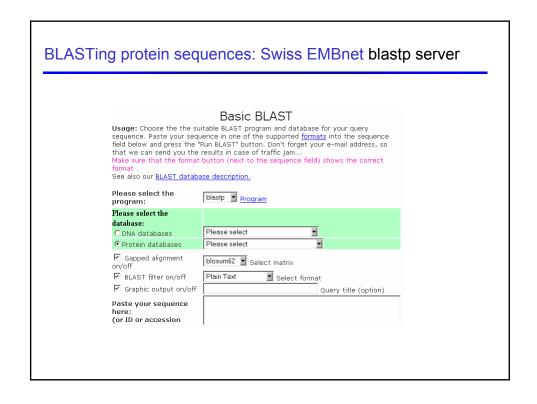
tblastn = Compares a protein sequence with a nucleotide database

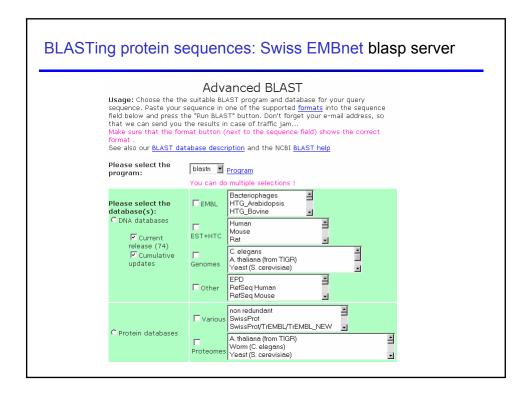
If you want to discover new genes encoding proteins (from multiple organisms), use **tblastn** to compare your protein with DNA sequences translated into their six possible reading frames; map a protein to genomic DNA;

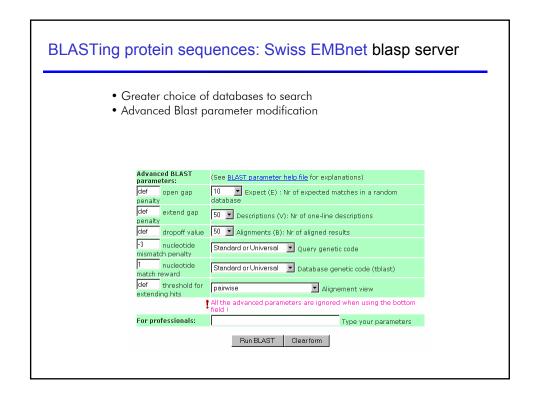
BLASTing protein sequences

Three of the most popular **blastp** online services:

- NCBI (National Center for Biotechnology Information) server: http://www.ncbi.nlm.nih.gov/BLAST
- ExPASy server: http://www.expasy.org/tools/blast/
- Swiss EMBnet server (European Molecular Biology network): http://www.ch.embnet.org/software/bBLAST.html (basic) http://www.ch.embnet.org/software/aBLAST.html (advanced)





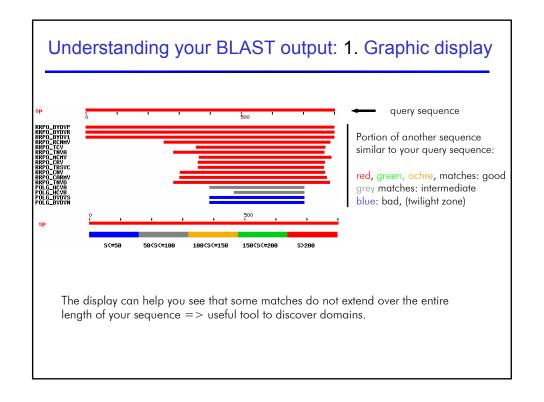


Understanding your BLAST output

- 1. Graphic display:
 - shows you where your query is similar to other sequences
- 2. Hit list:

the name of sequences similar to your query, ranked by similarity

- 3. The alignment:
 - every alignment between your query and the reported hits
- 4. The parameters:
 - a list of the various parameters used for the search



Understanding your BLAST output: 2. Hit list

```
Sequences producing significant alignments:

sp|F09505|RRPO BYDVP Putative RNA-directed RNA polymerase (EC 2... 1652 0.0 sp|F29045|RRPO BYDVR Putative RNA-directed RNA polymerase (EC 2... 1635 0.0 sp|F29044|RRPO BYDVI Putative RNA-directed RNA polymerase (EC 2... 1635 0.0 sp|F22956|RRPO RCMMV Putative RNA-directed RNA polymerase (EC 2... 1635 0.0 sp|F22956|RRPO RCMMV Putative RNA-directed RNA polymerase (EC 2... 367 e-101 sp|F17460|RRPO TCV Probable RNA-directed RNA polymerase (EC 2.7. 286 1e-76 sp|F22958|RRPO TNVA RNA-directed RNA polymerase (EC 2.7.7.48) [C... 280 1e-74

Sequence ac number and name Description Bit score E-value
```

- Sequence ac number and name: Hyperlink to the database entry: useful annotations
- Description: better to check the full annotation
- Bit score (normalized score): A measure of the similarity between the two sequences:
 the higher the better (matches below 50 bits are very unreliable)
- E-value: The lower the E-value, the better. Sequences identical to the query have an E-value of 0. Matches above 0.001 are often close to the twilight zone. As a rule-of-thumb an E-value above 10-4 (0.0001) is not necessarily interesting. If you want to be certain of the homology, your E-value must be lower than 10-4

Understanding your BLAST output: 3. Alignment



A good alignment should not contain too many gaps and should have a few patches of high similarity, rather than isolated identical residues spread here and there

Understanding your BLAST output: 4. Parameters

Database: swiss_nr
Posted date: Jan 12, 2002 5:06 AH
Number of letters in database: 103,264

Database: swiss_varsplic_nr
Posted date: Jan 12, 2002 5:07 AM
Number of sequences in database: 103,264

Database: swiss_varsplic_nr
Posted date: Jan 12, 2002 5:07 AM
Number of letters in database: 2,521,853
Number of sequences in database: 3785

Lambda K H
0.318 0.137 0.425

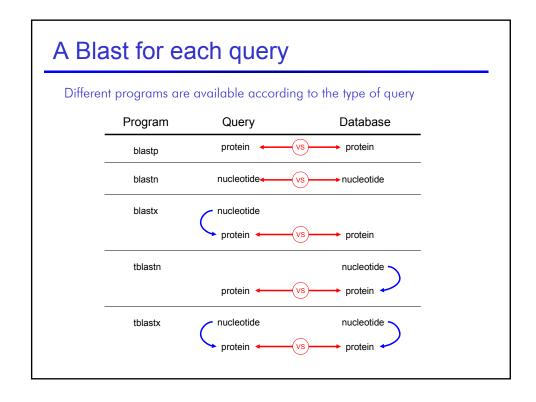
Gapped
Lambda K H
0.267 0.0410 0.140

Matrix: BLOSUM62

Gap Penalties: Existence: 11, Extension: 1
Number of Hits to DB: 79,326,108
Number of sequences: 107049
Number of sequences: 107049
Number of sequences better than 10.0: 152
Number of HSP's better than 10.0 vithout gapping: 72
Number of HSP's better than 10.0 vithout gapping: 72
Number of HSP's better than 10.0 vithout gapping: 72
Number of HSP's that attempted gapping in prelim test: 80
Number of HSP's successfully gapped in prelim test: 7745
Number of HSP's gapped (non-prelim): 314
length of database: 40,578,901
effective length of database: 28,054,168
effective search space: 23565501120
effective search space: 23565501120

Search details (at the bottom of the results)

- Size of the database searched
- Scoring system parameters
- Details about the number of hits found



BLASTing DNA sequences

- BLASTing DNA requires operations similar to BLASTing proteins BUT does not always work so well.
- It is faster and more accurate to BLAST proteins (blastp) rather than nucleotides. If you know the reading frame in your sequence, you're better off translating the sequence and BLASTing with a protein sequence.
- Otherwise:

Different BLAST Programs Available for DNA Sequences						
Program Query Database Usage						
blastn	DNA	DNA	Very similar DNA sequences			
tblastx	T DNA	TDNA	Protein discovery and ESTs			
blastx	T DNA	Protein	Analysis of the query DNA sequence			

T= translated

BLASTing DNA sequences

blastn = Compares a DNA sequence with a DNA database;

Mapping oligonucleotides, cDNAs, and PCR products to a genome; annotating genomic DNA; screening repetitive elements; cross-species sequence exploration;

tblastx = Compares a DNA translated into protein with a DNA database translated into protein;

Cross-species gene prediction at the genome or transcript level (ESTs); searching for genes not yet in protein databases;

blastx = Compares a DNA translated into protein with a protein sequence database;

Finding protein-coding genes in genomic cDNA; determining if a cDNA corresponds to a known protein;

BLASTing DNA sequences: choosing the right BLAST

Question	Answer		
Am I interested in non-coding DNA?	Yes: Use blastn . Never forget that blastn is only for closely related DNA sequences (more than 70% identical)		
Do I want to discover new proteins?	Yes: Use tblastx.		
Do I want to discover proteins encoded in my query DNA sequence?	Yes: Use blastx .		
Am I unsure of the quality of my DNA?	Yes: Use blastx if you suspect your DNA sequence is the coding for a protein but it may contain sequencing errors.		

- Pick the right database: choose the database that's compatible with the BLAST program you want to use (in general!)
- Restrict your search: Database searches on DNA are slower. When possible, restrict your search to the subset of the database that you're interested in (e.g. only the Drosophila genome)
- Shop around: Find the BLAST server containing the database that you're interested in
- Use filtering: Genomic sequences are full of repetitions: use some filtering

BLASTting DNA: BLASTN output

- DNA double-stranded molecule => genes may occur on either strand
- plus strand (the query sequence), minus strand (reverse complement)
- If the similarity between query and subject is on the same strand: plus/plus
- If the minus strand of the query sequence is similar to a database sequence: plus/minus with the subject sequence in reverse coordinates (flipped)

BLASTting DNA: BLASTX output

- Query sequence: translated in the 3 reading frames, on both plus and minus strand: +1,+2,+3 (plus strand) and -1, -2, -3 (minus strand)
- Matches on the plus strand: +1,+2,+3
- · Matches on the minus strand: query coordinates are inverted

```
Score = 790 bits (2040), Expect = 0.0
Identities = 520/1381 (37%), Positives = 745/1381 (53%), Gaps = 36/1381 (2%)
Frame = +3
Query: 156 SEMNVNMKYQLPNFTAETPIQNVVLHKHH--IYLGAVNYIYVLNDKDLQKVAEYKTGPVL 329
S +N ++ Y +P F A PIQN+V + + +Y+ + N I +N + L+KV E +TGPV
Sbjct: 31 SPVNFSVVYTMPFFQAGGPIQNIVNNSFYQEVYVASQNVIEAVN-QSLEKVWELRTGPV- 88

Score = 64.5 bits (169), Expect = 1.7e-258
Identities = 30/34 (88%), Positives = 34/34 (100%), Gaps = 3/34 (2%)
Frame = -1
Query: 1071 SEMNVNMKYQLPNFTAETPIQNVVLHKHH--IYLGAVNYIYVLNDKDLQKVAEYKTGPVL 970
S +N ++ Y +P F A PIQN+V + + +Y+ + N I +N + L+KV E +TGPV
Sbjct: 722 SPVNFSVVYTMPFFQAGGPIQNIVNNSFYQEVYVASQNVIEAVN-QSLEKVWELRTGPV- 755
```

BLASTting DNA: TBLASTN output

 Alignments similar to BLASTX, except that the database and query are exchanged (e.g. on minus strand the database sequence has flipped coordinates)

BLASTting DNA: TBLASTX output

- Both query and database have strand and frame
- Alignments may have any combination of frames

```
Score = 790 bits (2040), Expect = 0.0

Identities = 520/1381 (37%), Positives = 745/1381 (53%), Gaps = 36/1381 (2%)

Frame = +3/+3

Query: 156 SEMNVNMKYQLPNFTAETPIQNVVLHKHH--IYLGAVNYIYVLNDKDLQKVAEYKTGPVL 329

S +N ++ Y +P F A PIQN+V + + +Y+ + N I +N + L+KV E +TGPV

Sbjct: 31 SPVNFSVVYTMPFFQAGGPIQNIVNNSFYQEVYVASQNVIEAVN-QSLEKVWELRTGPV- 88

Score = 64.5 bits (169), Expect = 1.7e-258

Identities = 30/34 (88%), Positives = 34/34 (100%), Gaps = 3/34 (2%)

Frame = -1/+2

Query: 1071 SEMNVNMKYQLPNFTAETPIQNVVLHKHH--IYLGAVNYIYVLNDKDLQKVAEYKTGPVL 970

S +N ++ Y +P F A PIQN+V + + +Y+ + N I +N + L+KV E +TGPV

Sbjct: 722 SPVNFSVVYTMPFFQAGGPIQNIVNNSFYQEVYVASQNVIEAVN-QSLEKVWELRTGPV- 755
```

Choosing the right Parameters

• The default parameters that BLAST uses are quite optimal and well tested. However for the following reasons you might want to change them:

Some Reasons to Change BLAST Default Parameters					
Reason	Parameters to Change				
The sequence you're interested in contains many identical residues; it has a biased composition.	Sequence filter (automatic masking)				
BLAST doesn't report any results	Change the substitution matrix or the gap penalties.				
Your match has a borderline E-value	Change the substitution matrix or the gap penalties to check the match robustness.				
BLAST reports too many matches	Change the database you're searching OR filter the reported entries by keyword OR increase the number of reported matches OR increase Expect, the E-value threshold.				

Choosing the right Parameters: sequence masking

- When BLAST searches databases, it makes the assumption that the average composition of any sequence is the same as the average composition of the whole database.
- However this assumption doesn't hold all the time, some sequences have biased compositions, e.g. many proteins contain patches known as low-complexity regions: such as segments that contain many prolines or glutamic acid residues.
- If BLAST aligns two proline-rich domains, this alignment gets a very good E-value because of the high number of identical amino acids it contains. BUT there is a good chance that these two proline-rich domains are not related at all.
- In order to avoid this problem, sequence masking can be applied.



Choosing the right Parameters: DNA masking

- DNA sequences are full of sequences repeated many times: most of genomes contain many such repeats, especially the human genome (60% are repeats).
- If you want to avoid the interference of that many repeats, select the Human Repeats check box that appears in the blastn page of NCBI or the Xblast-repsim filter

Options for advanced blasting Limit by entrez or select from: (none) query Choose filter Low complexity Human repeats Mask for lookup table only Mask lower case • Or at the swiss EMBnet server (advanced BLAST):

Plain Text BLAST filter on/off ▼ Select format ✓ Xblast-repsim filter

Controlling the BLAST output

• If your query belongs to a large protein family, the BLAST output may give you troubles because the databases contain too many sequences nearly identical to yours => preventing you from seeing a homologous sequence less closely related but associated with experimental information; so how to proceed?

1) Choosing the right database

If BLAST reports too many hits, search for Swiss-Prot (100 times smaller) rather than NR; or search only one genome

2) Limit by Entrez query (NCBI)

For instance, if you want BLAST to report proteases only and to ignore proteases from the HIV virus, type "protease NOT hiv1[Organism]"

Expect

Change the cutoff for reporting hits, to force BLAST to report only good hits with a low cutoff

Changing the BLAST alignment parameters

- Among the parameters that you can change on the BLAST servers two important ones have to do with the way BLAST makes the alignments: the gap penalites (gap costs) and the substitution matrix (matrix) or match/mismatch parameters (DNA).
- Use a substitution matrix adapted to the expected divergence of the searched sequences (nevertheless most of the time BLOSUM62 works well):
 - BLOSUM 80: increase selectivity (exclude false positive, missing true positives) (closest to PAM120)
 - BLOSUM 45: increase sensitivity (more true matches, incluse false positives) (closest to PAM250)

Changing the BLAST alignment parameters

Most of the BLAST searches fall into one of two categories: 1. mapping and 2. exploring;

- Mapping: finding the position of one sequence within another (e.g. finding a gene within a genome) => you can expect the alignments to be nearly identical, and the coordinates are generally the focus of the results;
- Exploring: the goal is usually to find functionally related sequences => the
 alignment and alignment statistics (score, E-value, percent identity, ...) are often of
 greatest importance

Alignment parameters: BLASTN protocols

- 1. When sequences are expected to be nearly identical (mapping): $\pm 1/-3$ matchmismatch parameters:
 - Mapping oligos: filering (turned off): we want the entire oligo to match; -G 2 -E 1
 - Mapping nonspliced DNA to a genome: mask repeats; increase the word size (faster): -W 30; -G 1 –E 3;
 - Mapping cDNA/EST: mask repeats; reduce word size (-W 15) to see short exons;
 G 1 –E 3; low E-value to cut down false positives (-e 1e-20);
- 2. cross-species exploration (search for genes, regulatory elements, RNA genes): $\pm 1/-1$ match-mismatch parameters, $\pm 1/-1$ with the sensitivity:
 - Annotating Genomic DNA with ESTs (similar transcripts for genes no transcripts have been isolated yet): mask repeats; -G 1 –E 2; set low E-value to cut down false positives (-e 1e-20);

Alignment parameters: BLASTP protocols

Most BLASTP searches fall under the exploring category: try to learn about your query sequence by comparing it to other proteins:

- Standard search (default parameters): balances speed and sensitivity; not ideal for very distant proteomes;
- Fast insensitive search: when performing multiple searches (but not for sequences that have less than 50 percent identity); sequences are expected to be very similar: BLOSUM80, set low E-value (-e 1e-5); -G 9 –E 2;
- Slow, sensitive search: looking for distant relatives; set E higher (-e 100); BLOSUM45;

Changing the BLAST alignment parameters

• Guidelines from BLAST tutorial at NCBI (http://www.ncbi.nlm.nih.gov/Education/BLASTinfo/information3.html)

Step 3. Choose the appropriate search parameters or use default settings.

Choosing Parameters for Protein-Based BLAST Searches.

		Special Cases			
	Default	Short Query	Large Sequence Family	Ungapped BLAST	
Filter	on	off	on	on	
Scoring Matrix	BLOSUM62	PAM30 for 35 and under	BLOSUM62	BLOSUM62	
Word Size	3	3, or reduce to 2	3	3	
E value	10	1000 or more	10	10	
Gap costs	11,1	11,1	11,1	4	
Alignments	50	50	2000	50	

Alignment parameters: BLASTX protocols

BLASTX is generally used to find protein coding genes in genomic DNA or to identify proteins encoded by transcripts (exploring, but sometimes mapping):

- Gene finding in genomic DNA: mask repeats; BLOSUM62; higher E-value (-e 100) don't want to miss low-scoring genes;
- Annotating ESTs: what protein do they encode?; slightly less sensitive parameters than the default ones: set low E-value (-e 1e-10) to prevent misclassification;

Alignment parameters: TBLASTN protocols

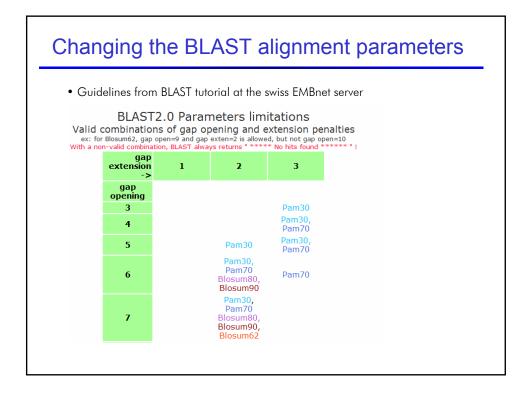
Similar to BLASTX but with TBLAST you map a protein to a genome or search EST databases for related protein not yet in the protein database:

- Mapping a protein to a genome: set a low E-value (-e 1e-5);
- Annotating ESTs: what protein do they encode?;

Alignment parameters: TBLASTX protocols

Coding sequences evolve slowly compared to the DNA: TBLASTX for gene-prediction for genomes that are appropriately diverged: not too much (human vs. E.coli) or not enough (human vs. chimpanzees)

- Finding undocumented genes in genomic DNA: mask repeats;
- Transcript of unknown function: first BLASTX and then (if no results) TBLASTX with ESTs databases;



Conclusions

Blast: the most used database search tool

- Fast and very reliable even for a heuristic algorithm
- Does not necessarily find the best alignment, but most of the time it finds the best matching sequences in the database
- Easy to use with default parameters
- Solid statistical framework for the evaluation of scores

but...

• The biologist's expertise is still essential to the analysis of the results!

Tips and tricks

- For coding sequences always search at the protein level
- Mask low complexity regions
- Use a substitution matrix adapted to the expected divergence of the searched sequences (nevertheless most of the time BLOSUM62 works well)
- If there are only matches to a limited region of your query, cut out that region and rerun the search with the remaining part of your query

BLAST Family

- Faster algorithm for genomic search:
 - MegaBLAST (NCBI): http://www.ncbi.nih.gov/BLAST/
 - and SSAHA (Ensembl): http://www.ensembl.org/

This program is optimized for aligning sequences that differ slightly as a result of sequencing or other similar "errors". (larger word size is used as default to speed up the search)

• PSI-BLAST and PHI-BLAST-> Thursday

Acknowledgments & References

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- Jean-Michel Claverie & Cedric Notredame, Bioinformatics for Dummies, Wiley Publishing