# Topological Machine Learning for Low Data Medical Imaging

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#### Abstract

Deep Learning (DL) has revolutionized medical image analysis by providing automated techniques to extract valuable insights from large datasets. However, challenges such as interpretability and reliance on extensive labeled data persist. Topological Data Analysis (TDA) has emerged as a complementary approach that captures underlying topological structures in data, potentially enhancing the performance of DL models.

In this paper, we present a comprehensive evaluation of TDA methods for computer-aided diagnosis from two perspectives. First, we examine the effectiveness of topological methods in data-limited settings by comparing the standalone performance of DL models, TDA approaches, and their fusion. Our results demonstrate that integrating topological features into DL models significantly improves performance when labeled data are scarce. Second, we assess the standalone performance of TDA methods in data-rich environments using the MedM-NIST collection, which includes over 600K images across 12 2D and 6 3D medical imaging datasets. Our experiments reveal that while TDA methods do not outperform DL models on 2D datasets, they achieve competitive results on 3D imaging tasks. These findings suggest that the fusion of TDA and DL methods can enhance the accuracy and robustness of computer-aided diagnosis, particularly in low-data or 3D imaging scenarios.

**Keywords:** Medical Image Analysis, Topological Data Analysis, Cubical Persistence, CNNs, Computer-aided Diagnosis, MedMNIST

Data and Code Availability Our datasets are publicly available at https://medmnist.com. We provide our code at https://github.com/Kausta/topo-net

Institutional Review Board (IRB) Our study does not require IRB approval.

#### 1. Introduction

Deep Learning (DL) has become a cornerstone in medical image analysis, providing efficient, effective, and automated techniques to extract valuable insights from vast amounts of medical image data (Chan et al., 2020). These models excel at learning from large datasets, identifying patterns in medical images that are often imperceptible to the human eye, and improving the accuracy of disease detection and diagnosis. The rapid progress in machine learning has significantly enhanced the identification, classification, and quantification of patterns in medical imaging, achieving high precision in computeraided diagnosis (Fujita, 2020). Despite these advances, challenges such as interpretability and the reliance on vast amounts of labeled data remain persistent. Therefore, complementary approaches that enhance computational feasibility, interpretability, and robustness are critical (Sarvamangala et al., 2022).

In the past decade, topological data analysis (TDA) has emerged as a promising approach that complements existing ML models in medical image analysis (Skaf et al., 2022). TDA focuses on extracting and analyzing the underlying topological structures in data, enabling a deeper understanding of complex relationships and patterns. By cap-

turing topological features, TDA provides an additional layer of information, enriching the overall analysis (Choe and Ramanna, 2022; Hensel et al., 2021). Inspired by its successful application in various domains, TDA has shown potential as a powerful feature extraction tool that can enhance the performance of state-of-the-art (SOTA) models in medical image analysis (Singh et al., 2023).

In this paper, we present a comprehensive analysis of topological machine learning methods for computer-aided diagnosis from two distinct perspectives. First, we examine the effectiveness of topological methods in data-limited settings by comparing the standalone performance of several deep learning models, topological approaches, and their fusion. Our results show that in low-data scenarios, incorporating topological features significantly enhances the performance of deep learning models. Second, we assess the standalone performance of topological methods in data-rich environments across various medical imaging domains. To conduct this evaluation, we utilize the MedMNIST collection, which consists of 12 sets of 2D and 6 sets of 3D medical imaging benchmark datasets, comprising over 600,000 images. Our experiments reveal that while topological methods do not outperform DL models on 2D datasets, they achieve competitive results on 3D imaging tasks.

#### Our contributions:

- We provide a comprehensive evaluation of topological methods in medical image analysis, comparing its performance with deep learning models in both data-limited and data-rich settings.
- Our experiments show that integrating topological features into DL models significantly improves performance in data-limited environments, highlighting the value of TDA in scenarios with scarce labeled data.
- We assess TDA's performance in data-rich environments using the MedMNIST collection, demonstrating competitive results on 3D imaging tasks.
- We offer insights into the fusion of TDA and DL methods, enhancing computer-aided diagnosis with improved accuracy and robustness.

### 2. Related Work

CNNs in Computer-Aided Diagnosis. Medical image analysis has witnessed a transformative

revolution with the advent of DL techniques. By harnessing various computer vision algorithms, DL has emerged as a key tool in extracting valuable insights from medical images. From accurate segmentation and classification to detecting subtle abnormalities, DL algorithms are empowering clinicians and researchers to unlock a wealth of information and make more informed decisions. In recent years, the dominance of traditional hand-crafted feature approaches has been overshadowed by the emergence of CNNs (Zhou et al., 2021; Halalli and Makandar, 2018). Numerous successful deep convolutional architectures have emerged for this purpose, including AlexNet (Krizhevsky et al., 2017), as well as the ResNet (He et al., 2016), Inception (Szegedy et al., 2017), MobileNet (Howard et al., 2017), EfficientNet (Tan and Le, 2019) and many more. Following the success of the attention mechanism and Transformer networks (Vaswani et al., 2017) in other domains, Vision Transformers (Dosovitskiy et al., 2021) achieved significant success in the domain of image analysis, leading to models like Swin Transformer (Liu et al., 2021, 2022). Despite their success, persistent challenges include the interpretability of results and the ongoing requirement for substantial volumes of labeled data, as highlighted in the study by Salehi et al. (2023). For a thorough review of deep learning methods in medical imaging, and computeraided diagnosis, see excellent reviews by Aggarwal et al. (2021); Shamshad et al. (2023); Abdou (2022).

Recent advances in medical image analysis in limited data settings have focused on mitigating the challenges of overfitting and enhancing generalization by leveraging techniques such as data augmentation (Chlap et al., 2021), transfer learning (Kora et al., 2022), and synthetic data generation using generative adversarial networks (GANs) (Chen et al., 2022). Few-shot learning has gained significant traction in this space, enabling models to learn from a small number of labeled examples by utilizing metalearning techniques or adapting pre-trained models to new tasks with minimal data (Singh et al., 2021; Feng et al., 2021). Self-supervised learning and contrastive learning approaches have also shown promise, leveraging large amounts of unlabeled data to extract meaningful representations, which are then finetuned for specific medical imaging tasks (Shurrab et al., 2022). These approaches, combined with careful model regularization, have proven effective in improving the robustness and accuracy of models in limited-data scenarios (Jiao et al., 2023).

TDA in Medical Image Analysis. Topological Data Analysis (TDA) is a novel approach for studying complex data by identifying both local and global structures at various scales, addressing challenges related to data dimensionality, differences in data collection methodologies, and varying scales (Coskunuzer and Akcora, 2024; Hensel et al., 2021). In the past decade, TDA has found widespread applications across multiple domains, including image analysis, neurology, cardiology, hepatology, gene-level and single-cell transcriptomics, drug discovery, evolution, and protein structural analysis (Skaf et al., 2022). By leveraging inherent topological features, and with its ability to capture hidden patterns of images, TDA opens up new opportunities for tasks such as image segmentation, object recognition, image registration, and image reconstruction (Choe and Ramanna, 2022). commonly used tool from TDA in image analysis is persistent homology (PH) has shown its effectiveness in several medical imaging domains, e.g., histopathological images (Yadav et al., 2023), MRI images (François et al., 2024), and CT scans (Somasundaram et al., 2021). Recently, TDA methods have been increasingly utilized to improve the performance of deep learning models across various medical imaging domains, as demonstrated in studies such as those by Peng et al. (2024); McGuire et al. (2023). A comprehensive review of TDA methods in medical imaging and biomedicine can be found in the excellent survey (Singh et al., 2023).

### 3. Methodology

Our methodology leverages topological techniques, specifically cubical persistence, to extract image embeddings. We then develop two ML models: *Topo-Med*, which evaluates the standalone performance of topological features, and *Topo-Net*, which combines topological vectors with deep learning methods.

#### 3.1. Cubical Persistence

Persistent homology (PH) is a powerful mathematical tool used in TDA to study the shape and structure of complex datasets. The main idea behind PH is to systematically assess the evolution of various hidden patterns in the data as we vary a scale parameter (Coskunuzer and Akcora, 2024). While PH is used as a very effective feature extraction method for various data types (e.g., point clouds, networks),

in this paper, we focus only on the image setting, in particular *cubical persistence*. While we describe PH construction in simple terms for non-experts, one can find more details in the textbook by Dey and Wang (2022). Basically, PH can be described as a 3-step process: Inducing a nested sequence of topological spaces from data *Filtration*, recording the evolution of topological changes in this sequence *Persistence Diagram*, and finally, converting persistence diagrams into vectors *Vectorization*.

Step 1 - Constructing Filtrations. We first describe the process for 2D images. Then, we explain how to adapt this construction to 3D images. For a given 2D image  $\mathcal{X}$  (say  $r \times s$  resolution), the filtration step is to create a nested sequence of binary images (aka cubical complexes) (Kaji et al., 2020). To create such a sequence, one can use grayscale (or other color channels) values  $\gamma_{ij} \in [0, 255]$  of each pixel  $\Delta_{ij} \subset \mathcal{X}$ . In particular, for a sequence of grayscale values, called thresholds,  $(0 \le t_1 < t_2 <$  $\cdots < t_N \le 255$ ), one obtains a nested sequence of binary images  $\mathcal{X}_1 \subset \mathcal{X}_2 \subset \cdots \subset \mathcal{X}_N$  such that  $\mathcal{X}_n = \{\Delta_{ij} \subset \mathcal{X} \mid \gamma_{ij} \leq t_n\}$  (See Figure 1). In other words, we start with an empty  $r \times s$  image and start activating (coloring black) pixels when their grayscale value reaches the given threshold. This is called *sub*level filtration for  $\mathcal{X}$  with respect to a given function (grayscale color values in this case). Similarly, when  $\mathcal{X}$  is a 3D image (say  $r \times s \times t$  resolution), we consider  $\mathcal{X}$  as a 3D cubical complex of size  $r \times s \times t$ . Again, to create our nested sequence of 3D binary images, we use grayscale values  $\gamma_{ijk} \in [0, 255]$  of each voxel  $\Delta_{ijk} \subset \mathcal{X}$ . Then, for a sequence of grayscale values  $(0 \le t_1 < t_2 < \cdots < t_N \le 255)$ , one obtains 3D binary images  $\mathcal{X}_1 \subset \mathcal{X}_2 \subset \cdots \subset \mathcal{X}_N$  where  $\mathcal{X}_n = \{\Delta_{ijk} \subset \mathcal{X} \mid \gamma_{ijk} \leq t_n\}$  as before.

Step 2 - Persistence Diagrams. PH keeps track of the evolution of topological features in the filtration sequence, and records as persistence diagram. In particular, if a topological feature  $\sigma$  first appears in  $\mathcal{X}_m$  and disappears in  $\mathcal{X}_n$  with  $1 \leq m < n \leq N$ , we call the corresponding thresholds  $t_m$  as birth time  $b_{\sigma}$  and  $t_n$  the death time  $d_{\sigma}$  of the topological feature  $\sigma$ ,

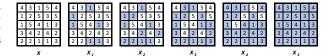


Figure 1: For the  $5 \times 5$  image  $\mathcal{X}$  with the given pixel values, **the sublevel filtration** is the sequence of binary images  $\mathcal{X}_1 \subset \mathcal{X}_2 \subset \mathcal{X}_3 \subset \mathcal{X}_4 \subset \mathcal{X}_5$ .

i.e.  $b_{\sigma} = t_m$  and  $d_{\sigma} = t_n$ . Then, PD is the collection of all such 2-tuples  $\operatorname{PD}_k(\mathcal{X}) = \{(b_{\sigma}, d_{\sigma})\}$  where k represent the dimension of the topological features. The difference  $d_{\sigma} - b_{\sigma}$  is called lifespan of  $\sigma$ . More formally,  $k^{th}$  persistence diagram can be defined as  $\operatorname{PD}_k(\mathcal{X}) = \{(b_{\sigma}, d_{\sigma}) \mid \sigma \in H_k(\mathcal{X}_i) \text{ for} b_{\sigma} \leq i < d_{\sigma}\}$  where  $H_k(\mathcal{X}_i)$  is the  $k^{th}$  homology group of the cubical complex  $\mathcal{X}_i$ . Hence,  $\operatorname{PD}_k(\mathcal{X})$  is the collection of 2-tuples marking the birth and death times of k-dimensional holes  $\{\sigma\}$  (connected components, loops, and voids) in the filtration sequence  $\{\mathcal{X}_i\}$ . e.g., for Figure 1,  $\operatorname{PD}_0(\mathcal{X}) = \{(1,\infty), (1,2), (1,2), (1,3)\}$  marking the connected components, while  $\operatorname{PD}_1(\mathcal{X}) = \{(2,4), (3,5), (4,5)\}$  marking the holes in respective binary images in Figure 1.

Step 3 - Vectorizations. Persistence Diagrams (PDs) which are collections of 2-tuples, are not particularly practical for use with ML tools. Instead, a widely adopted approach is to convert the PD information into a vector or a function, a process known as "vectorization" (Ali et al., 2023). A commonly used function for this purpose is the Betti function which keeps track of the number of "alive" topological features at a given threshold. In particular, the Betti function is a step function with  $\beta_0(t_n)$  the count of connected components in the binary image  $\mathcal{X}_n$ , and  $\beta_1(t_n)$  the number of holes (loops) in  $\mathcal{X}_n$ . In ML applications, Betti functions are usually taken as a vector  $\overrightarrow{\beta_k}$  of size N with entries  $\beta_k(t_n)$  for  $1 \leq n \leq N$ . i.e.,  $\overrightarrow{\beta_k}(\mathcal{X}) = [\beta_k(t_1) \dots \beta_k(t_N)]$ . e.g., for Figure 1, we have  $\overrightarrow{\beta_0}(\mathcal{X}) = [4\ 2\ 1\ 1\ 1]$  which are the count of connected components in binary image  $\{\mathcal{X}_i\}$  while  $\overline{\beta_1'}(\mathcal{X}) = [0 \ 1 \ 2 \ 2 \ 0]$  are the counts of holes in  $\{\mathcal{X}_i\}$ . i.e.,  $\beta_0(1) = 4$  is the count of components in  $\mathcal{X}_1$  and  $\beta_1(3) = 2$  is the count of holes (loops) in  $\mathcal{X}_3$ .

There are various other ways to convert PDs into a vector, e.g., persistence images, persistence land-scapes, and kernel methods (Ali et al., 2023).

**Topological Vectors.** For a given 2D (or 3D) image  $\mathcal{X}$ , say at  $r \times s$  (or  $r \times s \times t$ ) size, to produce its topological feature vectors, we first obtain a sublevel filtration, a sequence of binary images,  $\mathcal{X}_1 \subset \mathcal{X}_2 \cdots \subset \mathcal{X}_N$  by using the grayscale values of each pixel. If  $\mathcal{X}$  is a color image, one can replace grayscale values with RGB color channels. Here, N is the number of thresholds equally dividing [0, 255] color interval. In practice, N = 50 or 100 works well. The next step is to get the persistence diagram of this sublevel filtration. The final step is to choose a good

vectorization. In our experiments, we use Betti functions and Silhouette functions with different powers. If the image is 2D, only meaningful dimensions are k=0 and k=1. If the image is 3D, then the only meaningful dimensions are k=0,1,2. e.g., assuming N=100 and Betti functions as vectorization, for each image, we obtain  $\overrightarrow{\beta_0}$  and  $\overrightarrow{\beta_1}$  vectors where each is 100 dimensional. After concatenation, this results in 200-dimensional embeddings for 2D images, and 300-dimensional embeddings for 3D images.

#### 3.2. ML Models

Once we have obtained the topological feature vectors, the next step involves applying ML tools to analyze the effectiveness of these features. Essentially, each medical image is represented as a 200-dimensional vector (300-dimensional for 3D images). The classification task treats these embeddings as a point cloud and aims to identify distinct clusters corresponding to different classes. To evaluate the utility of these vector in computer-aided diagnosis, we employed two different ML models. In the first model, Topo-Net, we utilized topological vectors in existing deep learning models. We developed simpler and scalable second model, Topo-Med, to test the standalone performance of these topological vectors.

Topo-Net for Limited Data. In our first model, Topo-Net (Figure 2), we assess the impact of incorporating topological features into CNN architectures trained from scratch on datasets with limited samples. The motivation behind this approach lies in the complementary roles of topological features and convolutional layers: topological features capture the global structure of an image, providing embeddings

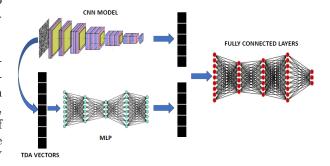


Figure 2: **Topo-Net Model.** In this model, we integrate our topological features of images obtained in the Topo-Med model with convolutional vectors generated by a CNN backbone, followed by processing through a fully connected layer.

that are less dependent on data quantity, while convolutional layers focus on localized patterns, where learning is highly influenced by the amount of available data. By merging these two perspectives, we hypothesize that the model will exhibit improved performance. Furthermore, we deliberately chose a simple architecture (Figure 2) to isolate and evaluate the contribution of topological vectors in enhancing DL models under limited-data settings.

For the 2D tasks, the Topo-Net model is crafted by selecting a base model from a range of state-of-the-art CNN models, including ResNet50 (He et al., 2016) as a baseline similar to the original MedMnist experiments (Yang et al., 2023a) and EfficientNet-B0 (Tan and Le, 2019) and Swin Transformer V2-Tiny (Liu et al., 2022) as more up-to-date alternatives. These were chosen for their robust feature extraction capabilities and performance in image classification tasks.

Topo-Net extends these models with an auxiliary feature network, taking Betti features as input. This Betti network uses a Multilayer Perceptron (MLP) to learn and extract features from the Betti numbers. Then, a classifier head consisting of an MLP is used to merge the CNN-based features and the auxiliary features from the Betti network and predict the output class. This allows the topological features to be incorporated with the convolutional features in a simple and network-agnostic way, allowing easy modification of other networks for limited-data scenarios where topological features can be obtained without additional training or data.

The 3D models are constructed similarly with an auxiliary Betti number MLP and a classifier MLP. These MLPs are used with a ResNet-18 (He et al., 2016) backbone where the 2D convolutions are converted into 2.5D, 3D, or ACS convolutions from the previous work ACSConv (Yang et al., 2021a), similar to the original MedMnist experiments. Similar to the 2D case, this allows the usage of topological features like Betti numbers without requiring significant network architecture changes, as the auxiliary networks can be added to any 3D CNN-based backbone.

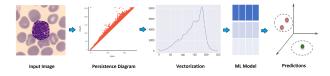


Figure 3: **Topo-Med model.** In our basic model, we simply extract topological feature vectors from images and apply standard ML models.

Topo-Med model. To evaluate the standalone performance of topological vectors across various medical domains, we introduce a simple ML model, Topo-Med (Figure 3). In this model, we use two types of classifiers with our topological vectors. The first is eXtreme Gradient Boosting (XGBoost), a treebased method widely recognized for its strong performance. The second is the Multilayer Perceptron (MLP), a feedforward neural network known for its flexibility and robustness. These models offer diverse strengths for classification tasks, particularly in highdimensional settings. MLP typically performs well with large and imbalanced datasets, while XGBoost consistently delivers reliable results across different scenarios. Further details on our ML procedures and hyperparameter tuning are provided in Section 4.

### 4. Experiments

#### 4.1. Experimental Setup

Datasets. In our experiments, we utilized the publicly available MedMNIST dataset collection, which is widely recognized in the medical imaging community (https://medmnist.com). MedMNIST comprises twelve pre-processed 2D datasets and six pre-processed 3D datasets sourced from various modalities, representing diverse classification tasks and varying data scales. The dataset includes predefined training and test splits and offers multiple resolutions, ranging from 28x28 to 224x224 for 2D images, and from 28x28x28 to 64x64x64 for 3D images (Yang et al., 2023b). Detailed information about these datasets is provided in Table 9 in Appendix.

**Topological Vectors.** For both our models, Topo-Net and Topo-Med, we first generated topological feature vectors. For each dataset, we applied sublevel filtration using 100 evenly spaced thresholds within the range [0, 255] for the grayscale channel. Betti vectorization was then applied to the resulting persistence diagrams. This process yielded 100-dimensional Betti vectors,  $\overrightarrow{\beta_i}(\mathcal{X})$ , for each dimension of an image  $\mathcal{X}$ . After concatenation, we obtained a 200-dimensional vector for each MedMNIST2D dataset and a 300-dimensional vector for each MedMNIST3D dataset.

**Topo-Net Hyperparameters.** We followed the hyperparameters from the original MedMNIST experiments as closely as possible (Yang et al., 2023b). We used a batch size of 128 for all 2D models with 224x224 images and a batch size of 16 for all 3D mod-

Table 1: **2D** Limited Data Performances. AUC results for vanilla-CNN and Topo-CNN models using ResNet-50, EfficientNetB0, and Swin Transformer backbones in limited data settings on the MedMNIST 2D datasets. The TDA columns display the standalone performance of topological vectors (Topo-Med). For each backbone, the top AUC **improvements** are highlighted in **blue**, and highest performance for each sample training size is <u>underlined</u>. Accuracy results are given in Table 5.

			D	ermaM!	NIST					Ti	ssueMN	IST					P	neuMN	IST		
#	TDA	Res	T-Res	Eff	T-Eff	Swin	T-Swin	TDA	Res	T-Res	Eff	T-Eff	Swin	T-Swin	TDA	Res	T-Res	Eff	T-Eff	Swin	T-Swin
100	0.8163	0.7068	0.6783	0.6775	0.6790	0.6149	0.6576	0.7273	0.6169	0.6316	0.5973	0.6336	0.5670	0.6303	0.7466	0.8968	0.9221	0.9092	0.8862	0.8509	0.8215
250	0.8217	0.7942	0.7829	0.7687	0.7520	0.6339	0.7432	0.7345	0.6356	0.6800	0.6253	0.6583	0.5851	0.6686	0.7532	0.8960	0.9296	0.9157	0.9178	0.6352	0.8690
500	0.8342	0.8181	0.8261	0.8073	0.7894	0.6737	0.7169	0.7421	0.6393	0.6947	0.6233	0.6731	0.5980	0.6841	0.8061	0.9213	0.9446	0.9340	0.9210	0.8351	0.8678
1000	0.8451	0.8625	0.8446	0.8454	0.8511	0.6719	0.7467	0.7322	0.6967	0.7177	0.6911	0.7033	0.5788	0.6994	0.8029	0.9443	0.9586	0.9359	0.9252	0.7868	0.8944
2000	0.8665	0.8784	0.8634	0.8502	0.8766	0.7900	0.7898	0.7289	0.7260	0.7440	0.7087	0.7328	0.5659	0.7118	0.8525	0.9532	0.9605	0.9178	0.9413	0.7369	0.8912
			В	loodMN	IST					Pa	thMNI	ST					0	CTMN	IST		
#	TDA	Res	B T-Res		IST T-Eff	Swin	T-Swin	TDA	Res				Swin	T-Swin	TDA	Res	O T-Res		IST T-Eff	Swin	T-Swin
			T-Res	Eff	T-Eff		T-Swin    0.9549			T-Res	Eff	T-Eff					T-Res	Eff	T-Eff		
100	0.9129	0.9403	T-Res	Eff 0.8474	T-Eff	0.8080		0.7660	0.8379	T-Res   0.8745	Eff 0.8270	T-Eff	0.7179	0.8818	0.5379	0.6956	T-Res	Eff 0.5079	T-Eff 0.6220	0.5149	0.5821
100 250	0.9129 0.9487	0.9403 0.9822	T-Res 0.9639	Eff 0.8474 0.9249	T-Eff 0.9527	0.8080 0.8262	0.9549	0.7660 0.8379 <u>0</u>	0.8379 0.9380	T-Res   0.8745   0.9310	Eff 0.8270 0.8943	T-Eff   0.9042   0.9147	0.7179	0.8818	0.5379 0.5691	$\frac{0.6956}{0.8178}$	T-Res 0.6909 0.8006	Eff 0.5079 <b>0.6283</b>	T-Eff 0.6220 0.7717	0.5149 0.5943	0.5821
100 250 500	0.9129 0.9487 0.9629	0.9403 0.9822 0.9865	T-Res 0.9639 0.9902	Eff 0.8474 0.9249 0.9621	T-Eff   0.9527   0.9823   0.9865	0.8080 0.8262	0.9549 0.9832 0.9863	0.7660 0.8379 <u>0</u>	0.8379 0.9380 0.9560	T-Res   0.8745   0.9310   0.9284	Eff 0.8270 0.8943	T-Eff   0.9042   0.9147   0.9390	0.7179 <b>0.5664</b>	0.8818 <b>0.9084</b>	0.5379 0.5691 0.6043	$\begin{array}{c} \underline{0.6956} \\ \underline{0.8178} \\ \underline{0.8734} \end{array}$	T-Res   0.6909   0.8006   0.8414	Eff 0.5079 <b>0.6283</b> 0.7536	T-Eff 0.6220 0.7717 0.8273	0.5149 0.5943 0.5789	0.5821 0.6979

Table 2: **3D Limited Data Performances.** AUC results for vanilla-CNN and Topo-CNN models using ResNet-18 + 2.5D, ResNet-18 + 3D, and ResNet-18 + ACS backbones in limited data settings on the MedMNIST 3D datasets.

			Adr	enalMN	IST3D					Ves	selMNI	ST3D		
#	TDA	R18-ACS	T-R18-ACS	R18-3D	T-R18-3D	R18-2.5D	T-R18-2.5D	TDA	R18-ACS	T-R18-ACS	R18-3D	T-R18-3D	R18-2.5D	T-R18-2.5D
50	0.8426	0.8358	0.7259	0.7970	0.8349	0.6734	0.7472	0.7733	0.7330	0.7236	0.7725	0.6890	0.6562	0.6861
100	0.8529	0.7987	0.8472	0.8396	0.8453	0.7352	0.7160	0.8388	0.6505	0.7290	0.7679	0.7546	0.7420	0.6629
200	0.8566	0.7999	0.8343	0.8540	0.8577	0.6697	0.7607	0.8801	0.6678	0.7866	0.7628	0.7910	0.7700	0.7860
250	0.8511	0.7863	0.7932	0.8570	0.8113	0.6962	0.7748	0.9194	0.8050	0.8175	0.8184	0.8065	0.7819	0.8029
500	0.8592	0.8958	0.8497	0.8636	0.8048	0.8079	0.8169	0.9171	0.8175	0.8458	0.9274	0.8542	0.8024	0.8584

els with 64x64x64 inputs. We used cross-entropy loss with Adam optimizer with an initial learning rate of 0.001, and we trained the models for 100 epochs, multiplying the learning rate by 0.1 after 50 epochs and 75 epochs. Key performance metrics are incorporated, including accuracy and AUC.

For the Topo-Net variants, an auxiliary three-layer Betti MLP with 256 hidden layer dimensions and a two-layer classifier MLP with 256 hidden layer dimension were used. In addition, a feature size of 128 for CNN and 128 for the auxiliary Betti MLP was chosen based on initial hyperparameter tests comparing different ratios of feature sizes for the CNN and the Betti MLP. For the CNNs, original normalization and activations were used, and ReLU activations were used for all MLPs. The data augmentations followed were the same as the MedMnist experiments (no augmentation for the 2D datasets, multiplying the training set by a random value in [0, 1] during training and multiplying the images by a fixed coefficient of 0.5 during evaluation for the 3D datasets).

**Topo-Med Hyperparameters.** We trained both XGBoost and MLP models on our datasets. The XGBoost model was set up for multi-class classification with *multi:softmax* and *binary:logistic* for bi-

nary classification as the objective and the number of classes specified by MedMNIST in each dataset. For the MLP model, the architecture included one input layer, three hidden layers with the first two having 256 units each and ReLU activation, and the third with 128 units and ReLU activation, and a dropout layer with a rate of 0.5. The output layer used a softmax activation function. The model was compiled with the categorical crossentropy loss function and the Adam optimizer with a learning rate of 0.001. We trained the MLP models for 100 epochs with a batch size of 32.

Computational Complexity & Runtime. PH calculation can be computationally intensive, especially in high-dimensional data (Otter et al., 2017). However, for 2D images, it is relatively efficient, with a time complexity of approximately  $\mathcal{O}(|\mathcal{P}|^{2.3})$ , where  $|\mathcal{P}|$  denotes the total number of pixels (Milosavljević et al., 2011). This means the computational effort grows almost quadratically with image size. In contrast, subsequent steps like vectorization and ML are comparatively lightweight.

We conducted all experiments on a highperformance computing system configured with one node, 20 tasks, and the 256i partition. The 256i par-

Table 3: Topo-Med vs. DL. Performance comparison of seven deep learning models vs. standalone performance of topological vectors on MedMNIST collection using full datasets. Detailed results for the seven baseline models are provided in Table 8, while additional performance metrics for Topo-Med are presented in Table 7 in the Appendix.

	Pat	thM	Che	$_{ m estM}$	Der	maM	OC	TM	Pne	euM	Reti	naM
Methods	AUC	ACC	AUC	ACC	AUC	ACC	AUC	ACC	AUC	ACC	$ \overline{\mathrm{AUC}} $	ACC
Best Baseline	0.989	0.909	0.778	0.948	0.920	0.754	0.963	0.771	0.991	0.946	0.750	0.531
Topo-Med	0.942	0.683	0.787	0.530	0.904	0.669	0.710	0.450	0.845	0.762	0.728	0.458
	Bre	astM	Bloc	odM	Tiss	ueM	Orga	nAM	Orga	nCM	Orga	nSM
Methods	AUC	ACC	AUC	ACC	AUC	ACC	AUC	ACC	AUC	ACC	$ \overline{\mathrm{AUC}} $	ACC
Best Baseline	0.919	0.861	0.998	0.966	0.941	0.703	0.997	0.935	0.994	0.920	0.974	0.813
Topo-Med	0.821	0.737	0.973	0.798	0.837	0.450	0.921	0.523	0.894	0.489	0.910	0.532
	Organ	M3D	Nodule	M3D	Fractur	eM3D	Adrena	alM3D	Vessel	M3D	Synaps	eM3D
Methods	AUC	ACC	AUC	ACC	AUC	ACC	AUC	ACC	AUC	ACC	AUC	ACC
Best Baseline	0.996	0.907	0.914	0.874	0.750	0.571	0.839	0.754	0.930	0.928	0.851	0.795
Topo-Med	0.837	0.554	0.808	0.736	0.653	0.480	0.864	0.769	0.920	0.887	0.730	0.730

tition is equipped with 20 Intel Ivy Bridge cores and 256 GB of memory. Generating both Betti-0 and Betti-1 vectors from the OCTMNIST dataset, which contains 109,309 images of size  $28 \times 28$ , took 50 minutes and 54 seconds. The remaining ML tasks were significantly faster, all completed in under a minute. For smaller datasets, the generation of topological vectors was much faster.

#### 4.2. Results and Discussion

**Limited Data Performance.** In Tables 1 and 2, we present our results across various limited data settings. For the 2D datasets, we used 100, 250, 500, 1000, and 2000 training images with the backbones ResNet50, EfficientNetB0, and Swin Transformers. For the 3D datasets, we used 50, 100, 200, 250, and 500 training images with the backbones ResNet-18 + 2.5D, ResNet-18 + 3D, and ResNet-18 + ACS. Additionally, in Figure 4, we provide a comprehensive summary of AUC performance across selected MedMNIST datasets, illustrating the impact of TDAenhanced models on CNN performance. This figure demonstrates how integrating topological data analysis (TDA) consistently improves model robustness and generalization, particularly in limited data scenarios. All models were evaluated on the original test set of the corresponding MedMNIST dataset, consisting of 2,005 images for DermaMNIST, 47,280 images for TissueMNIST, 624 images for PneuMNIST, 3,421 images for BloodMNIST, 7,180 images for PathM-NIST, 1,000 images for OCTMNIST, 298 images for AdrenalMNIST3D, and 382 images for VesselM-NIST3D.

Corresponding accuracy results are detailed in Tables 5 and 6, and additional performance metrics for the Topo-Med model are presented in Table 7 in the Appendix. For the 2D datasets, our results show that topological vectors consistently enhance deep learning models in limited data scenarios. In particular, the performance of Swin Transformers improves by as much as 30%. Similarly, the EfficientB0 model shows significant gains, with improvements of up to 15%. Although the synergy between ResNet50 and topological vectors is less substantial, we still observe improvements of up to 6%. When considering the best results for each limited data setting, topological vectors, either on their own or in combination with a model, achieve the highest performance in nearly all settings, with few exceptions in the PathMNIST and OCTMNIST datasets.

In 3D datasets, the performance of topological vectors is even more remarkable. In nearly all limited data scenarios, topological vectors outperform deep learning (DL) models when used alone. This difference is especially significant in the VesselMNIST3D dataset. Furthermore, the improvement that topological vectors provide to various backbone models is consistent, ranging from 3% to 12%.

These results support our hypothesis that in datalimited settings, topological vectors provide critical complementary information to deep learning models, showing great promise in addressing this challenge. Notably, even state-of-the-art models like Swin Transformers struggle with limited data, and our results suggest that integrating topological vectors has strong potential to mitigate this issue.

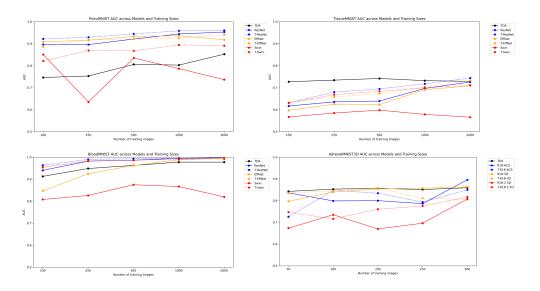


Figure 4: Summary of AUC performance across selected MedMNIST datasets.

Table 4: Effect of Resolution & ML Classifiers on TDA performance. Performance comparison of topological vectors on high (128×128) vs. low (28×28) resolution images with XGBoost and MLP Classifiers. Colored bottom rows give the difference between the best performances of low vs. high-resolution models. Blue numbers represent high res model is better, and red numbers represent low res model is better.

	Pat	hM	Che	stM	Deri	naM	OC	$\mathbf{TM}$	Pne	euM	Reti	naM	Brea	astM	Blo	odM	Tiss	ueM
Methods	AUC	ACC	$\overline{ ext{AUC}}$	ACC	$\overline{ ext{AUC}}$	ACC	$\overline{ ext{AUC}}$	ACC	AUC	ACC	AUC	ACC	$\overline{ ext{AUC}}$	ACC	AUC	ACC	$\overline{ ext{AUC}}$	ACC
Low + XGB	0.943	0.712	0.567	0.638	0.684	0.665	0.820	0.505	0.838	0.732	0.618	0.453	0.795	0.752	0.972	0.810	0.813	0.485
Low + MLP	0.942	0.683	0.787	0.530	0.904	0.669	0.710	0.450	0.845	0.762	0.728	0.458	0.821	0.737	0.973	0.798	0.837	0.450
High + XGB	0.962	0.746	0.623	0.635	0.796	0.685	0.852	0.475	0.888	0.789	0.614	0.433	0.768	0.821	0.997	0.944	0.829	0.511
High + MLP	0.949	0.689	0.779	0.632	0.921	0.678	0.673	0.433	0.894	0.832	0.671	0.395	0.854	0.794	0.996	0.936	0.862	0.489
High vs. Low	1.9%	3.4%	0.8%	0.3%	1.7%	1.6%	3.2%	3.0%	4.9 %	7.0 %	5.7%	2.5%	3.3%	6.9 %	2.4 %	13.4 %	2.5 %	2.6 %
	Orga	nAM	Orga	nCM	Orga	nSM	Organ	ıM3D	Nodul	eM3D	Fractu	reM3D	Adre	nalM3I	) Vess	elM3D	Synaps	seM3D
Methods	- 6								Nodul					nalM3I		elM3D		seM3D ACC
Methods Low + XGB	- 6													ACC	AUC	ACC		
	AUC	ACC	AUC	ACC	AUC	ACC	AUC	ACC	AUC	ACC	AUC	ACC	AUC	0.769	0.662	2 ACC 2 0.887	AUC	ACC
Low + XGB	AUC 0.924	<b>ACC</b> 0.565	AUC 0.900	<b>ACC</b> 0.543	AUC 0.894	<b>ACC</b> 0.546	AUC 0.933	<b>ACC</b> 0.585	AUC 0.768	<b>ACC</b> 0.823	AUC 0.625	<b>ACC</b> 0.463	AUC 0.749	0.769 0.769	0.662 0.920	2 <b>ACC</b> 2 0.887 0 0.887	AUC 0.783	<b>ACC</b> 0.810
Low + XGB Low + MLP	AUC 0.924 0.921	ACC 0.565 0.523	AUC 0.900 0.894	ACC 0.543 0.489	0.894 0.910	ACC 0.546 0.532	AUC 0.933 0.837	ACC 0.585 0.554	0.768 0.808	0.823 0.736	0.625 0.653	ACC 0.463 0.48	0.749 0.864	0.769 0.769	0.662 0.920 0.694	0.887 0.887 0.887	AUC 0.783 0.730	0.810 0.730

Topo-Med & Entire Dataset. In Tables 3 and 8, we present the standalone performance of topological vectors using the full dataset and compare it with SOTA benchmarks provided by MedMNIST (Yang et al., 2023b) for 28x28 (or 28x28x28) size images. For 2D datasets, the seven baselines include two pretrained CNNs (for two image sizes): ResNet-18 and ResNet-50 (He et al., 2016), and three AutoML models: auto-sklearn (Feurer et al., 2022), AutoKeras (Jin et al., 2019), and Google AutoML Vision (Bisong, 2019). For 3D datasets, eight baselines include ResNet-18 and ResNet-50 with 2.5D/3D/ACS convo-

lutions (Yang et al., 2021b), and the AutoML methods auto-sklearn and AutoKeras.

While we report only the top-performing baselines in Table 3, comprehensive performance metrics for all baselines can be found in Appendix B. Our results establish a new benchmark for the MedMNIST collection, highlighting that in data-rich scenarios, topological vectors offer competitive performance but generally do not outperform DL models across most medical imaging domains. With a few exceptions, the Topo-Med model achieves results within 1-10% of the best-performing DL models. This indicates that while topological vectors are a promising alter-

native in limited data settings, they often fall short in data-rich environments. However, effective integration of topological vectors with DL models holds significant potential for developing more robust and accurate models, a direction we plan to explore in our future work.

High vs. Low Resolution. We also analyzed how image resolution affects the performance of our topological model. As shown in Section 4.2, while higher resolution generally leads to improved performance, this is not always the case. Blue percentages indicate instances where the high-resolution model outperforms, while red percentages highlight where the low-resolution model does better. Overall, we interpret that while TDA methods offer comparable performance with greater computational efficiency on low-resolution images, higher resolution is preferable when performance is the priority, as it better captures topological patterns in medical imaging.

ML Classifiers. Comparing the performances of the ML part of our model, we observe that MLP is giving slightly better performance than XGBoost for 2D datasets. For 3D datasets, XGBoost is consistently better. The rough interpretation is that for small datasets, XGBoost would be a better choice as MLP needs a large training set for good performance.

**Limitation.** A key limitation of the TDA approach in medical imaging is its strong dependence on the specific domain. While it excels in certain diagnostic tasks, such as dermal, retinal, and blood-related imaging, it falls short when compared to deep learning (DL) models in other domains, like tissue or organ imaging (e.g., abdominal CT scans). When sufficient data is available, DL methods generally learn more effectively, adapting better to downstream tasks and delivering more accurate results. The reliance on topological feature vectors makes the TDA approach highly domain-specific, limiting its applicability across general medical imaging tasks. However, our experiments indicate that TDA vectors capture fundamentally different information from DL models and, when properly integrated, have the potential to enhance the performance of existing DL models.

## 5. Conclusion

In this study, we evaluated the effectiveness of topological features across various medical imaging domains. Our findings demonstrate that topological

methods hold significant promise in addressing the limitations of DL models in data-limited settings, a common challenge in medical imaging. Notably, in 3D settings, topological vectors showed remarkable standalone performance, outperforming all DL models, even when they were augmented with topological features. However, in data-rich environments, DL models generally exhibited superior performance compared to TDA approaches. Importantly, our results highlight that integrating topological feature vectors can enhance the accuracy and robustness of DL models. Future research will focus on harnessing this potential to develop specialized clinical decision support systems tailored to specific medical domains by combining TDA outputs with DL models, ultimately aiming for more advanced and reliable medical image analysis systems.

### Acknowledgements

This work was partially supported by the National Science Foundation under grants DMS-2202584, 2229417, and DMS-2220613 and by Simons Foundation under grant # 579977. The authors acknowledge the Texas Advanced Computing Center (TACC) at UT Austin for computational resources which contributed to the research results reported within this paper.

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## Appendix A. Further Performance Metrics for Limited Data

In Tables 5 and 6, we present the accuracy results of our models in limited data settings. We further present the additional performance metrics for Topo-Med model in these datasets for limited data settings in Table 7.

Table 5: **2D** Limited Data. Accuracy results for vanilla-CNN and Topo-CNN models using ResNet-50, EfficientNetB0, and Swin Transformer backbones in limited data settings on the MedMNIST 2D datasets. The TDA columns display the standalone performance of topological vectors using an MLP (Topo-Med). For each backbone, the top AUC **improvements** are highlighted in **blue**, and the highest performance within each row for each dataset is **underlined**.

			Bl	oodMN	IST					О	CTMN.	ST					Pa	athMN	IST		
#	TDA	Res	T-Res	Eff	T-Eff	Swin	T-Swin	TDA	Res	T-Res	Eff	T-Eff	Swin	T-Swin	TDA	Res	T-Res	Eff	T-Eff	Swin	T-Swin
100	0.7366	0.7653	0.7866	0.5077	0.7463	0.4291	0.7638	0.3000	0.4710	0.4420	0.2480	0.3720	0.2390	0.3250	0.4187	0.6029	0.6418	0.5387	0.7077	0.3050	0.6659
250	0.8158	0.8834	0.9062	0.6942	0.8375	0.4505	0.8690	0.3410	0.5630	0.5060	0.3270	0.4810	0.314	0.402	0.6383	0.6974	0.7113	0.6109	0.7072	0.1180	0.6760
500	0.8603	0.8942	0.9220	0.7662	0.8787	0.4744	0.8781	0.3830	0.6230	0.6310	0.4830	0.5200	0.2500	0.3980	0.5840	0.8194	0.7155	0.6469	0.7234	0.0825	0.6947
1000	0.8948	0.9395	0.9304	0.9035	0.9073	0.4616	0.8994	0.4040	0.7290	0.6990	0.5750	0.6200	0.2510	0.4050	0.6164	0.8187	0.7253	0.7227	0.6985	0.2056	0.6845
2000	0.9176	0.9594	0.9494	0.9532	0.9281	0.3975	0.9182	0.3980	0.7820	0.7760	0.6770	0.7820	0.2500	0.4120	0.6448	0.8191	0.7765	0.7926	0.7467	0.4721	0.6669
			De	rmaMN	NIST					Ti	ssueMN	IST					P	neuMN	IST		
#	TDA	Res	De T-Res	ermaMN	NIST T-Eff	Swin	T-Swin	TDA	Res			IST T-Eff	Swin	T-Swin	TDA	Res	T-Res		IST T-Eff	Swin	T-Swin
# 100				Eff	T-Eff			TDA	<u> </u>		Eff	T-Eff		T-Swin 0.4014	"		T-Res	Eff	T-Eff		
# 100 250		0.5840	T-Res	Eff	<b>T-Eff</b> 0.6195		0.5800	"	0.4007	T-Res	Eff	T-Eff 0.4074	0.3226		0.6955	0.7612	T-Res 0.7821	Eff 0.6282	T-Eff 0.7949	0.8045	0.7276
	0.6050 0.5786	0.5840	T-Res 0.6160	Eff	T-Eff 0.6195 0.6589	0.6459	0.5800 0.6613	0.3886	0.4007 0.4181	T-Res 0.394	Eff   0.3886   0.3789	T-Eff 0.4074 0.3931	0.3226 0.1487	0.4014 <b>0.4358</b>	0.6955 0.7532	0.7612 0.8157	T-Res 0.7821	Eff 0.6282 0.8013	T-Eff 0.7949 0.8237	0.8045	0.7276
250 500	$\begin{array}{c} 0.6050 \\ 0.5786 \\ 0.6030 \end{array}$	0.5840 0.6324 0.6459	T-Res 0.6160 0.6185	Eff 0.6643 0.6464	T-Eff 0.6195 0.6589	0.6459 <b>0.6668</b>	0.5800 0.6613 0.6524	0.3886 0.3912 0.4033	0.4007 0.4181 0.4336	T-Res 0.394 0.3896	Eff   0.3886   0.3789   0.4018	T-Eff 0.4074 0.3931 0.4505	0.3226 0.1487 0.3355	0.4014 0.4358 0.4568	0.6955 0.7532 0.8061	0.7612 0.8157 <b>0.8285</b>	T-Res 0.7821 <u>0.8574</u>	Eff 0.6282 0.8013 0.8189	T-Eff 0.7949 0.8237	$\begin{array}{ c c c }\hline 0.8045 \\ 0.3750 \\ \hline \end{array}$	0.7276 <b>0.8061</b>

Table 6: **3D Limited Data Performances.** Accuracy results for vanilla-CNN and Topo-CNN models using ResNet-18 + 2.5D, ResNet-18 + 3D, and ResNet-18 + ACS backbones in limited data settings on the MedMNIST 3D datasets.

			Ad	renalMN	NIST3D					Ve	sselMN	IST3D		
#	TDA	R18-ACS	T-R18-ACS	R18-3D	T-R18-3D	R18-2.5D	T-R18-2.5D	TDA	R18-ACS	T-R18-ACS	R18-3D	T-R18-3D	R18-2.5D	T-R18-2.5D
50	0.7282	0.7752	0.7685	0.8054	0.2819	0.5503	0.3926	0.7016	0.8874	0.8063	0.8848	0.8298	0.8874	0.8743
100	0.7483	0.8255	0.2617	0.7819	0.2416	0.7785	0.7886	0.8717	0.7880	0.8874	0.6545	0.8874	0.8874	0.8874
200	0.7685	0.5503	0.2919	0.8054	0.7550	0.6477	0.7685	0.8717	0.8874	$\underline{0.9031}$	0.8115	0.8796	0.8874	0.8874

Table 7: Standalone performance of topological vectors with different sample sizes on MedMNIST datasets.

		Tissue	MNIST (8)			OCTI	MNIST (4)			Blood	MNIST (8)	
Samples	Acc	AUC	Precision	Recall	Acc	AUC	Precision	Recall	Acc	AUC	Precision	Recall
100	0.3886	0.7273	0.4044	0.3746	0.3000	0.5379	0.2986	0.2980	0.7366	0.9129	0.7436	0.7308
250	0.3912	0.7345	0.4062	0.3776	0.3410	0.5691	0.3417	0.3410	0.8158	0.9487	0.8218	0.8118
500	0.4033	0.7421	0.4178	0.3887	0.3830	0.6043	0.3841	0.3810	0.8603	0.9629	0.8639	0.8591
1000	0.3977	0.7322	0.4073	0.3869	0.4040	0.6315	0.4124	0.4000	0.8948	0.9714	0.8961	0.8921
2000	0.4088	0.7289	0.4154	0.4018	0.3980	0.6381	0.4074	0.3940	0.9176	0.9778	0.9186	0.9170
		Bronet	MNIST (2)				FRITCH (a)			T (1.3		
		Dieast	WINIST (2)			Pneul	MNIST (2)			PathN	ANIST (9)	
Samples	Acc	AUC	Precision	Recall	Acc	AUC	Precision	Recall	Acc	AUC	ANIST (9) Precision	Recall
Samples 100	<b>Acc</b> 0.7115			Recall 0.7115	Acc   0.6955		. ,	Recall 0.6955	Acc   0.5187		( )	Recall 0.5174
•		AUC	Precision			AUC	Precision			AUC	Precision	
100	0.7115	<b>AUC</b> 0.7617	Precision 0.7115	0.7115	0.6955	<b>AUC</b> 0.7466	Precision 0.6955	0.6955	0.5187	<b>AUC</b> 0.7660	Precision 0.5194	0.5174
100 250	0.7115 0.7051	AUC 0.7617 0.7490	0.7115 0.7051	0.7115 0.7051	0.6955	AUC 0.7466 0.8283	0.6955 0.7532	0.6955 0.7532	0.5187 0.6383	AUC 0.7660 0.8379	0.5194 0.6411	0.5174 0.6376

## Appendix B. Topo-Med vs. SOTA Baselines on Entire Dataset

In Table 8, we present the performance metrics of seven baseline models on the MedMNIST datasets (using 28x28 and 28x28x28 sized images), compared with our Topo-Med model, which incorporates topological vectors and an MLP. This comparison evaluates the standalone performance of topological vectors against SOTA models when applied to the entire dataset (Table 9). Baseline performances are reported from Yang et al. (2023b).

Table 8: **Topo-Med vs. DL.** Performance comparison across various DL models and Topo-Med model on the MedMNIST2D (28x28) and MedMNIST3D (28x28x28) datasets. The best performance is given in bold, and the second best is underlined.

	Pat	hM	Che	stM	Deri	naM	OC	$\mathbf{TM}$	Pne	euM	Reti	naM
Methods	AUC	ACC	AUC	ACC	AUC	ACC	AUC	ACC	AUC	ACC	AUC	ACC
ResNet-18 (28)	0.983	0.907	0.768	0.947	0.917	0.735	0.943	0.743	0.944	0.854	0.717	0.524
ResNet-18 (224)	0.989	0.909	0.773	0.947	0.920	0.754	0.958	0.763	0.956	0.864	0.710	0.493
ResNet-50 (28)	0.990	0.911	0.769	0.947	0.913	0.735	0.952	0.762	0.948	0.854	0.726	0.528
ResNet-50 (224)	0.989	0.892	0.773	0.948	0.912	0.731	0.958	0.776	0.962	0.884	0.716	0.511
auto-sklearn	0.934	0.716	0.649	0.779	0.902	0.719	0.887	0.601	0.942	0.855	0.690	0.515
AutoKeras	0.959	0.834	0.742	0.937	0.915	0.749	0.955	0.763	0.947	0.878	0.719	0.503
Google AutoML	0.944	0.728	0.778	0.948	0.914	0.768	0.963	0.771	0.991	0.946	0.750	0.531
Topo-Med	0.942	0.683	0.787	0.530	0.904	0.669	0.710	0.450	0.845	0.762	0.728	0.458
	Brea	$_{ m stM}$	Bloc	odM	Tiss	ueM	Orga	nAM	Orga	nCM	Orga	nSM
Methods	AUC	ACC	AUC	ACC	AUC	ACC	AUC	ACC	AUC	ACC	AUC	ACC
ResNet-18 (28)	0.901	0.863	0.998	0.958	0.930	0.676	0.997	0.935	0.992	0.900	0.972	0.782
ResNet-18 (224)	0.891	0.833	0.998	0.963	0.933	0.681	0.998	0.951	0.994	0.920	0.974	0.778
ResNet-50 (28)	0.857	0.812	0.997	0.956	0.931	0.680	0.997	0.935	0.992	0.905	0.972	0.770
ResNet-50 (224)	0.866	0.842	0.997	0.950	0.932	0.680	0.998	0.947	0.993	0.911	0.975	0.785
auto-sklearn	0.836	0.803	0.984	0.878	0.828	0.532	0.963	0.762	0.976	0.829	0.945	0.672
AutoKeras	0.871	0.831	0.998	0.961	0.941	0.703	0.994	0.905	0.990	0.879	0.974	0.813
${\it Google AutoML}$	0.919	0.861	0.998	0.966	0.924	0.673	0.990	0.886	0.988	0.877	0.964	0.749
Topo-Med	0.821	0.737	0.973	0.798	0.837	0.450	0.921	0.523	0.894	0.489	0.910	0.532
	Orga	nM3D	Nodul	leM3D	Fractu	reM3D	Adren	alM3D	Vesse	lM3D	Synaps	seM3D
Methods	$\overline{ ext{AUC}}$	ACC	AUC	ACC	AUC	ACC	AUC	ACC	AUC	ACC	AUC	ACC
ResNet-18 $+$ 2.5D	0.977	0.788	0.838	0.835	0.587	0.451	0.718	0.772	0.748	0.846	0.634	0.696
ResNet-18 + 3D	0.996	0.907	0.863	0.844	0.712	0.508	0.827	0.721	0.874	0.877	0.820	0.745
ResNet-18 + ACS	0.994	0.900	0.873	0.847	0.714	0.497	0.839	0.754	0.930	0.928	0.705	0.722
ResNet-50 + 2.5D	0.974	0.769	0.835	0.848	0.552	0.397	0.732	0.763	0.751	0.877	0.669	0.735
ResNet-50 + 3D	0.994	0.883	0.875	0.847	0.725	0.494	0.828	0.745	0.907	0.918	0.851	0.795
ResNet-50 + ACS		0.889	0.886	0.841	0.750	0.517	0.828	0.758	0.912	0.858	0.719	0.709
auto-sklearn	0.977	0.814	0.914	0.874	0.628	0.453	0.828	0.802	0.910	0.915	0.631	0.730
AutoKeras	0.979	0.804	0.844	0.834	0.642	0.458	0.804	0.705	0.773	0.894	0.538	0.724
Topo-Med	0.837	0.554	0.808	0.736	0.653	0.480	0.864	0.769	0.920	0.887	0.730	0.730

## Appendix C. Dataset Details

In our experiments, we utilized the publicly available MedMNIST dataset, which covers a diverse array of medical imaging domains and formats, comprising over 600,000 images. MedMNIST is a standardized and extensive collection of biomedical image datasets, supporting a range of tasks and scales—from 100 to 100,000 samples. It provides uniform preprocessing, predefined train-validation-test splits, and image sizes varying from 28x28 to 224x224 for 2D, and from 28x28x28 to 64x64x64 for 3D data. Detailed information about the dataset is presented in Table 9. More information can be found at https://medmnist.com

Table 9: MedMNIST2D and MedMNIST3D Dataset Details.

MedMNIST2D	Data Modality	# Classes	# Samples	Train/Valid/Test
PathMNIST	Colon Pathology	9	107,180	89,996 / 10,004 / 7,180
ChestMNIST	Chest X-ray	14	112,120	78,468 / 11,219 / 22,433
DermaMNIST	Dermatoscope	7	10,015	7,007 / 1,003 / 2,005
OCTMNIST	Retinal OCT	4	109,309	97,477 / 10,832 / 1,000
PneuMNIST	Chest X-ray	2	5,856	4,708 / 524 / 624
RetinaMNIST	Fundus Camera	5	1,600	1,080 / 120 / 400
BreastMNIST	Breast Ultrasound	2	780	546 / 78 / 156
BloodMNIST	Blood Cell Microscope	8	17,092	11,959 / 1,712 / 3,421
TissueMNIST	Kidney Cortex Microscope	8	236,386	165,466 / 23,640 / 47,280
OrganAMNIST	Abdominal CT	11	58,850	34,581 / 6,491 / 17,778
OrganCMNIST	Abdominal CT	11	23,660	13,000 / 2,392 / 8,268
OrganSMNIST	Abdominal CT	11	25,221	$13{,}940 \ / \ 2{,}452 \ / \ 8{,}829$
OrganMNIST3D	Abdominal CT	11	1,743	972 / 161 / 610
NoduleMNIST3D	Chest CT	2	1,633	1,158 / 165 / 310
AdrenalMNIST3D	Abdominal CT	2	1,584	1,188 / 98 / 298
FractureMNIST3D	Chest CT	3	1,370	1,027 / 103 / 240
VesselMNIST3D	Brain MRA	2	1,909	1,335 / 192 / 382
${\bf Synapse MNIST3D}$	Electron Microscope	2	1,759	1,230 / 177 / 352