**Paper Outline**

**Intro**

Version 1

Every year, in the late summer or early fall, the mixotrophic cilliate, *Mesodinium sp.*, forms impressive non-toxic red water blooms in the Columbia River Estuary. Previously referred to in past publications on the Columbia River Estuary as *Mesodinium rubrum* (*= Myrionecta rubra*)(Lohmann, 1908; Jankowski, 1976), the closely related *Mesodinium sp*. that blooms in this estuary shares a similarly unique relationship with its cryptophyte prey. *M.* *rubrum* is among the marine microzooplankton that have been observed to harbor the plastids of their prey, allowing these predators to function as mixotrophs, capable of utilizing both phagotrophic and photosynthetic mechanisms to acquire carbon (Crawford, 1989).

It has been hypothesized that because of this predator-prey interaction, the dynamics of the *Mesodinium sp.* bloom have the potential to be influenced by it's cryptophyte prey, as well as a number of physical drivers. The Columbia River Estuary is highly influenced by both tidal forcing and the seasonality of freshwater discharge, which results in an extended summer saltwater intrusion (Chawla et al., 2008). This saltwater intrusion and subsequent temporary decrease in turbulence has recently been shown, along with coinciding neap tides, to correlate with the initiation of the yearly *Mesodinium sp.* bloom in the estuary (Herfort et al., 2011a). Much less is known about how the dynamics of the cryptophytes in the Columbia River Estuary might control the *Mesodinium sp.* bloom.

To better understand the role of this cryptophyte-*Mesodinium sp.* relationship within the Columbia River Estuary bloom, we... explain what we are doing here

Version 2

Every year, in the late summer or early fall, impressive non-toxic red water blooms appear in the Columbia River Estuary, consisting of both cryptophyte algae and the mixotrophic ciliate, *Mesodinium sp.* Insert info about naming etc here.

Reasons for the timing, patchiness, and persistence of the bloom are not yet fully understood. Past research on the estuary has focused largely on understanding the dynamics of the *Mesodinium sp.*, but it has been hypothesized that the cryptophytes may be important in controlling the bloom, as they are both a source of prey and photosynthetic ability to the ciliate. Chloroplasts originating from the cryptophyte, *Teleaulax amphioexa,* have been found inside the *Mesodinium sp.* in the Columbia River Estuary, conferring photosynthetic capabilities (Herfort et al., 2011b). Though free-living T. amphioexa are low in number during the red tide blooms (Herfort et al., 2011b), the presence and growth dynamics (?) of these cryptophytes have the potential to be key factors in the proliferation of their ciliate predator. In order to monitor abundance and growth rates of the cryptophytes in the Columbia River Estuary over the course of a red-water bloom, we have employed novel methods utilizing continuous flow cytometry and the application of a size-structured growth model...

**Methods**

Study Area

The Columbia River Estuary, located in between Washington and Oregon (fig), is a turbid and often highly stratified system. It is characterized by its dynamic physical processes and strong influence from diurnal and semi-diurnal tides (Jay 1984). The estuary also has a short residence time, with flushing in the range of 0.5-5 days (Neal, 1972).

Sample Collection

SeaFlow was stationed on a dock outside of Astoria, OR (fig) and set up to run continuous measurements of surface water for up to blah days each week for four weeks in September-October 2013. Discrete samples for *Mesodinium sp.* enumeration and for flow cytometry were taken at three depths. Additional samples were collected for analysis of nutrients.

Division Rate Lab Verification

For verification of the division rates found using the size-structured growth model, cultures of the cryptophyte, *Rhodomonas sp*. (CCMP 755), were grown in flasks at 13 °C in f/2 seawater media under a (16:8) light-dark cycle. The abundances of daily 1mL samples of Rhodomonas sp. were determined via cell counts using a Sedgewick-Rafter slide. Growth rates were calculated using blah equation.

A single replicate of the *Rhodomonas sp.* cultures was brought to a volume of 20L in a blah carboy with a concentration of 50 cells/mL and mixed using a magnetic carboy stirbar. SeaFlow was run on the culture under a (16:8) light-dark cycle for 4 days. During the first day running SeaFlow, 1mL triplicate samples of the *Rhodomonas sp.* culture were taken every 2 hours for 28 hours, preserved with 20L of 50% gluteraldehyde, flash frozen, and stored at -80 °C for cell cycle analysis.

*Mesodinium sp.* Enumeration

Nutrient Analysis

SeaFlow Data Analysis

**Results**

**Discussion**

notes

Though pigmented plastids and nuclei of cryptophyte origin were identified within *M. rubrum* cells early on (Barber et al., 1969; Taylor and Blackbourne, 1971), it was not until a cultured isolate of *M. rubrum* from the Antarctic became available that more directed laboratory studies on the ciliate-cryptophyte relationship were able to be done (Gustafson et al., 2000). In culture experiments where *M. rubrum* was fed with the cryptophyte, *Teleaulax acuta*, it was shown that the ciliate required this cryptophyte prey to sustain its growth rates and that the number of cryptophyte nuclei within the ciliate increased when fed (Gustafson et al., 2000). Further work on the Antarctic isolate demonstrated that the endogenous organelles originated from the same cryptophyte prey species added to cultures in experiments using a fluorescence *in situ* hybridization (FISH) probe (Johnson et al., 2007). In addition, the cryptophyte nuclei remained transcriptionally active, despite the dissolution of the cryptophyte membrane and other structures after being ingested. The sequestered chloroplasts inside *M. rubrum* were able to divide, with the cryptophyte nuclei aiding in their maintenance- without these nuclei, the chloroplasts could not replicate and decreased in number (Johnson et al., 2007).