Standardized network reconstruction of CHO cell metabolism

Kieran Smallbone

Manchester Centre for Integrative Systems Biology 131 Princess Street, Manchester M1 7DN, UK kieran.smallbone@manchester.ac.uk

Abstract

We have created a genome-scale network reconstruction of chinese hamster ovary (CHO) cell metabolism. Existing reconstructions were improved in terms of annotation standards, to facilitate their subsequent use in dynamic modelling. The resultant network is available from ChoNet (http://cho.sf.net/).

ChoNet

The structure of metabolic networks can be determined by a reconstruction approach, using data from genome annotation, metabolic databases and chemical databases [1]. We built upon an existing reconstruction of the metabolic network of CHO cells that was based on genomic and literature data (Selvarasu *et al.* [2]). This model contains 1065 genes, 1545 metabolic reactions, and 1218 unique metabolites. Use of *in silico* modelling allows characterisation internal metabolic behaviour during growth and non-growth phases [2].

Selvarasu et al. suffers from the use of non-standard names and is not annotated with methods that are machine-readable. The model was thus updated according to existing community-driven annotation standards [3]. The reconstruction is described and made available in Systems Biology Markup Language (SBML) (http://sbml.org/, [4]), an established community XML format for the mark-up of biochemical models that is understood by a large number of software applications. The network is available from ChoNet (http://cho.sf.net/). As supplied, the model has an optimal growth rate of 0.0257 flux units.

Annotation

The highly-annotated network is primarily assembled and provided as an SBML file. Specific model entities, such as species or reactions, are annotated using ontological terms. These annotations, encoded using the resource description framework (RDF) [5], provide the facility to assign definitive terms to individual components, allowing software to identify such components unambiguously and thus link model components to existing data resources [6]. Minimum Information Requested in the Annotation of Models (MIRIAM, [7]) –compliant annotations have been used to identify components unambiguously by associating them with

one or more terms from publicly available databases registered in MIRIAM resources [8]. Thus this network is entirely traceable and is presented in a computational framework.

Six different databases are used to annotate entities in the network (see Table 1). The Systems Biology Ontology (SBO) [9] is also used to semantically discriminate between entity types. Five different SBO terms are used to annotate entities in the network (see Table 2).

example	identifier	database
ChoNet	10029	taxonomy
ChoNet	22252269	pubmed
cytosol	GO:0005737	obo.go
N-methylhistamine	CHEBI:29009	chebi
1-oxidoreductase	1.1.99.1	ec-code
1-oxidoreductase	218865	ncbigene

Table 1: MIRIAM annotations used in the model.

example	SBO term	interpretation
cytosol	290	compartment
N-methylhistamine	247	metabolite
N-methylhistamine	176	biochemical reaction
AATRA20	185	transport reaction
biomass objective function	397	modelling reaction

Table 2: SBO terms used in the model.

Use

We maintain the distinction between the CHO cell GEnome scale Network REconstruction (GENRE) [10] and its derived GEnome scale Model (GEM) [11]. This is important to differentiate between the established biochemical knowledge included in a GENRE and the modelling assumptions required for analysis or simulation with a GEM. A GENRE serves as a structured knowledge base of established biochemical facts, while a GEM is a model which supplements the established biochemical information with additional (potentially hypothetical) information to enable computational simulation and analysis [12]. Reactions added to the GEM include the biomass objective function – a sink representing cellular growth – and hypothetical transporters.

Three versions of the network are made available:

- <organism>_<version>.xml, a GEM for use in flux analyses, provided in Flux Balance Constraints (FBC) format [13]
- <organism>_<version>_cobra.xml, the same GEM network, provided in Cobra format [14]

• <organism>_<version>_recon.xml, a GENRE containing only reactions for which there is experimental evidence

EcoliNet and YeastNet

EcoliNet (http://ecoli.sf.net/) and YeastNet (http://yeast.sf.net/) are annotated metabolic network of Escherichia coli and Saccharomyces cerevisiae S288c, respectively, that are periodically updated by a team of collaborators from various research groups. The three networks are structured identically to facilitate comparative studies.

Acknowledgements This work is deliverable 4.3 of the EU FP7 (KBBE) grant 289434 "BioPreDyn: New Bioinformatics Methods and Tools for Data-Driven Predictive Dynamic Modelling in Biotechnological Applications".

References

- [1] Palsson BØ, Thiele I: A protocol for generating a high-quality genome-scale metabolic reconstruction. *Nature Protoc* 2010, 5:91–121. doi:10.1038/nprot.2009.203
- [2] Selvarasu S, Ho YS, Chong WP, Wong NS, Yusufi FN, Lee YY, Yap MG, Lee DY: Combined in silico modeling and metabolomics analysis to characterize fed-batch CHO cell culture. *Biotechnol Bioeng* 2012, **109**:1415–1429. doi:10.1002/bit.24445
- [3] Herrgård MJ, Swainston N, Dobson P, Dunn WB, Arga KY, Arvas M, Blüthgen N, Borger S, Costenoble R, Heinemann M, Hucka M, Le Novére N, Li P, Liebermeister W, Mo M, Oliveira AP, Petranovic D, Pettifer S, Simeonidis E, Smallbone K, Spasić I, Weichart D, Brent R, Broomhead DS, Westerhoff HV, Kırdar B, Penttilä M, Klipp E, Palsson BØ, Sauer U, Oliver SG, Mendes P, Nielsen J, Kell DB: A consensus yeast metabolic network obtained from a community approach to systems biology. Nature Biotechnol 2008, 26:1155–1160. doi:10.1038/nbt1492
- [4] Hucka M, Finney A, Sauro H, Bolouri H, Doyle J, Kitano H, Arkin A, Bornstein B, Bray D, Cornish-Bowden A, Cuellar A, Dronov S, Gilles E, Ginkel M, Gor V, Goryanin I, Hedley W, Hodgman T, Hofmeyr J, Hunter P, Juty N, Kasberger J, Kremling A, Kummer U, Le Novère N, Loew L, Lucio D, Mendes P, Minch E, Mjolsness E, Nakayama Y, Nelson M, Nielsen P, Sakurada T, Schaff J, Shapiro B, Shimizu T, Spence H, Stelling J, Takahashi K, Tomita M, Wagner J, Wang J: The systems biology markup language (SBML): a medium for representation and exchange of biochemical network models. Bioinformatics 2003, 19:524–531. doi:10.1093/bioinformatics/btg015
- [5] Wang XS, Gorlitsky R, Almeida JS: From XML to RDF: how semantic web technologies will change the design of 'omic' standards. *Nature Biotechnol* 2005, 23:1099–1103. doi:10.1038/nbt1139

- [6] Kell DB, Mendes P: The markup is the model: reasoning about systems biology models in the Semantic Web era. J Theor Biol 2008, 252:538–543. doi:10.1016/j.jtbi.2007.10.023
- [7] Le Novére N, Finney A, Hucka M, Bhalla US, Campagne F, Collado-Vides J, Crampin EJ, Halstead M, Klipp E, Mendes P, Nielsen P, Sauro H, Shapiro B, Snoep JL, Spence HD, Wanner BL: Minimum information requested in the annotation of biochemical models (MIRIAM). Nature Biotechnol 2005, 23:1509–1515. doi:10.1038/nbt1156
- [8] Laibe C, Le Novére N: MIRIAM resources: tools to generate and resolve robust cross-references in Systems Biology. BMC Syst Biol 2008, 252:538–543. doi:10.1186/1752-0509-1-58
- [9] Courtot M., Juty N., Knüpfer C., Waltemath D., Zhukova A., Drger A., Dumontier M., Finney A., Golebiewski M., Hastings J., Hoops S., Keating S., Kell D.B., Kerrien S., Lawson J., Lister A., Lu J., Machne R., Mendes P., Pocock M., Rodriguez N., Villeger A., Wilkinson D.J., Wimalaratne S., Laibe C., Hucka M., Le Novére N.: Controlled vocabularies and semantics in systems biology. Mol Syst Biol 2011, 7:-543. doi:10.1038/msb.2011.77
- [10] Price ND, Reed JL, Palsson BØ: Genome-scale models of microbial cells: evaluating the consequences of constraints. Nat Rev Microbiol 2004, 2:886–897. doi:10.1038/nrmicro1023
- [11] Feist AM, Herrgård MJ, Thiele I, Reed JL, Palsson BØ: Reconstruction of biochemical networks in microorganisms. Nat Rev Microbiol 2008, 7:129–143. doi:10.1038/nrmicro1949
- [12] Heavner BD, Smallbone K, Barker B, Mendes P, Walker LP: Yeast 5 an expanded reconstruction of the Saccharomyces cerevisiae metabolic network. *BMC Syst Biol* 2012, **6**:55. doi:10.1186/1752-0509-6-55
- [13] Olivier BG, Bergmann FT: Flux Balance Constraints, Version 1 Release 1. Available from COMBINE. 2013.
- [14] Schellenberger J, Que R, Fleming RM, Thiele I, Orth JD, Feist AM, Zielinski DC, Bordbar A, Lewis NE, Rahmanian S, Kang J, Hyduke DR, Palsson BØ: Quantitative prediction of cellular metabolism with constraint-based models: the COBRA Toolbox v2.0. Nat Protoc 2011, 6:1290–1307. doi:10.1038/nprot.2011.308.4