

# Haploid Selection, Sex Ratio Bias, and Transitions Between sex-determination systems

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Contributions:

## Abstract

2 Sex-determination systems are remarkably dynamic; many taxa display  
4 shifts in the location of sex-determining loci or the evolution of entirely  
6 new sex-determining systems. Predominant theories for why we observe  
8 such transitions generally conclude that novel sex-determining systems are  
10 favoured by selection if they equalise the sex ratio or increase linkage with  
12 a sexually-antagonistic locus. We use population genetic models to extend  
14 these theories in two ways: (1) We explicitly consider how selection on very  
16 tightly sex-linked loci influences the spread of novel sex-determiners. We  
18 find that tightly sex-linked genetic variation can favour the spread of new  
20 sex-determination systems in which the heterogametic sex changes (XY to  
22 ZW or ZW to XY) and the new sex-determining region is less closely linked  
24 (or unlinked) to the sex-linked locus under selection; a result that is not found  
26 with loose sex-linkage. (2) We also consider selection upon haploid geno-  
28 types either during gametic competition (e.g., pollen/sperm competition) or  
meiosis (i.e., non-Mendelian segregation); selective processes that typically  
occur in one sex or the other. With haploid selection, we again find that  
transitions between male and female heterogamety can occur even if the new  
sex-determining region is less closely linked to the locus under selection.  
Haploid selection in the heterogametic sex can also cause sex ratio biases,  
which may increase or decrease with the spread of new sex chromosomes.  
Thus, transitions between sex-determination systems cannot be simply pre-  
dicted by selection to equalise the sex-ratio. Overall, our models reveal  
that transitions between sex-determination systems, particularly transitions  
where the heterogametic sex changes, can be driven by loci in previously un-  
expected genomic locations that experience selection during diploid and/or  
haploid phases. These results might be reflected in the lability with which  
sex-determination systems evolve.

## Introduction

30 Animals and angiosperms exhibit extremely diverse sex-determination systems  
31 (reviewed in Bull 1983, Charlesworth and Mank 2010, Beukeboom and Perrin  
32 2014, Bachtrog et al. 2014). Among species with genetic sex determination of  
33 diploid sexes, some taxa have heterogametic males (XY) and homogametic fe-  
34 males (XX), including mammals and most dioecious plants (Ming et al. 2011);  
35 whereas other taxa have homogametic males (ZZ) and heterogametic females (ZW),  
36 including Lepidoptera and birds. Within several taxa, the chromosome that har-  
37 bours the master sex-determining region changes. For example, transitions of the  
38 master sex-determining gene between chromosomes or the evolution of new mas-  
39 ter sex-determining genes have occurred in Salmonids (Li et al. 2011, Yano et al.  
40 2012), Diptera (Vicoso and Bachtrog 2015), and *Oryzias* (Myosho et al. 2012). In  
41 addition, many gonochoric clades with genetic sex determination exhibit transi-  
42 tions between male (XY) and female (ZW) heterogamety, including snakes ([Gamble et al. 2017, Current Biology](#)), lizards (Ezaz et al. 2009), eight of 26 teleost  
43 fish families (Mank et al. 2006), true fruit flies (Tephritids, Vicoso and Bachtrog  
44 2015), amphibians (Hillis and Green 1990), the angiosperm genus *Silene* (Slan-  
45 carova et al. 2013), [the angiosperm family Salicaceae \(Pucholt et al. 2015, 2017\)](#),  
46 and Coleoptera and Hemiptera (Beukeboom and Perrin 2014, plate 2). Indeed, in  
47 some cases, both male and female heterogametic sex-determination systems can  
48 be found in the same species, as exhibited by some cichlid species (Ser et al. 2010)  
49 and *Rana rugosa* (Ogata et al. 2007) ([Miura 2007](#)). In addition, multiple transitions  
50 have occurred between genetic and environmental sex-determination systems, e.g.,  
51 in reptiles and fishes (Conover and Heins 1987, Mank et al. 2006, Pokorná and  
52 Kratochvíl 2009, Ezaz et al. 2009, Pen et al. 2010, Holleley et al. 2015).

53 Predominant theories accounting for the spread of new sex-determination sys-  
54 tems by selection involve fitness differences between sexes (e.g., sexually antag-  
55 onistic selection) or sex-ratio selection. van Doorn and Kirkpatrick (2007; 2010)  
56 show that new sex-determining loci can be favoured if they arise in closer link-  
57 age with a locus that experiences sexual antagonism. Tighter linkage allows a

stronger favourable association to build up between a male-beneficial allele, and  
60 a neo-Y chromosome, for example. Such associations can favour a new master  
sex-determining gene on a new chromosome (van Doorn and Kirkpatrick 2007)  
62 and can also favour a transition between male and female heterogamety (e.g., a  
ZW to XY transition, van Doorn and Kirkpatrick 2010). However, any sexually-  
64 antagonistic loci that are more closely linked to the ancestral sex-determination  
locus will develop similar, favourable associations and hinder the spread of a new  
66 sex-determination system.

The sex ratio is directly affected by the sex-determination system, and it has  
68 therefore been suggested that sex-ratio selection is a dominant force in the evolution  
of sex determination (e.g., Bull 1983, p 66-67; Beukeboom and Perrin 2014,  
70 Chapter 7). ‘Fisherian’ sex-ratio selection favours a 1:1 zygotic sex ratio when as-  
suming that males and females are equally costly to produce (Fisher 1930, Charnov  
72 1982). This follows from the fact that, for an autosomal locus, half of the ge-  
netic material is inherited from a male and half from a female (West 2009). Thus,  
74 if the population sex ratio is biased towards one sex, the average per-individual  
contribution of genetic material to the next generation from the opposite sex is  
76 greater. Therefore, a mutant that increases investment in the rarer sex will spread  
via the higher per-individual contributions made by that sex. In the case of sex-  
78 chromosome evolution, Kozielska et al. (2010) consider systems in which the an-  
cestral sex chromosomes experience meiotic drive (e.g., where driving X or Y  
80 chromosomes are inherited disproportionately often), which causes sex ratios to  
become biased (Hamilton 1967). They find that new, unlinked sex-determining  
82 loci (masculinizing or feminizing mutations, i.e., neo-Y or neo-W loci) can then  
spread, which restore an even sex ratio.

84 Here we use mathematical models to find the conditions under which new  
sex-determination systems spread when individuals experience selection at both  
86 diploid and haploid stages. Even in animal and plant species that have much  
larger and more conspicuous diploid phases than haploid phases, many loci ex-  
88 perience significant haploid selection through gamete competition and/or meiotic

drive (Mulcahy et al. 1996, Joseph and Kirkpatrick 2004). We use the term ‘meiotic drive’ to refer to the biased (non-Mendelian) segregation of genotypes during gamete production (from one parent) and the term ‘gametic competition’ to refer to selection upon haploid genotypes within a gamete/gametophyte pool (potentially from multiple parents); the term ‘haploid selection’ encompasses both processes.

94     Genetic mapping experiments, which are typically designed to minimize selection in diploids, have revealed segregation distortion in various species, including  
96     mice, Drosophila, Rice, Maize, Wheat, Barley, Cotton... In some of these cases,  
98     biased segregation has been attributed to meiotic drive and/or gametic selection  
   (Leppala et al. 2013, Didion et al. 2015, 2016 Xu et al 2013 (rice), Fishman...).

Meiotic drive generally occurs either during the production of male or female  
100    gametes only (Úbeda and Haig 2005, Lindholm et al. 2016). Because there are  
102    typically many more pollen/sperm than required for fertilization, gametic competi-  
104    tion is also typically sex specific, occurring primarily among male gametes. Gametic  
106    competition may be particularly common in plants, in which 60-70% of all  
108    genes are expressed in the male gametophyte and these genes exhibit stronger sig-  
110    natures of selection than random genes (Borg et al. 2009, Arunkumar et al. 2013,  
112    Gossmann et al. 2014). In addition, artificial selection pressures applied to male  
114    gametophytes are known to cause a response to selection (e.g., Hormaza and Her-  
116    rero 1996, Ravikumar et al. 2003, Hedhly et al. 2004, Clarke et al. 2004). A smaller  
118    proportion of genes are thought to be expressed and selected during competition in  
   animal sperm, although precise estimates are uncertain (Zheng et al. 2001, Joseph  
   and Kirkpatrick 2004, Vibranovski et al. 2010). Recent studies have demonstrated  
   that sperm competition can alter haploid allele frequencies and increase offspring  
   fitness (Immler et al. 2014) (Alavioon et al. 2017).

114    There are various ways in which a period of haploid selection could influence  
116    transitions between sex-determination systems. If we assume that haploid selec-  
118    tion at any particular locus predominantly occurs in one sex (e.g., meiotic drive  
   during spermatogenesis), then such loci experience a form of sex-specific selec-  
   tion. In this respect, we might expect that haploid selection would affect transitions

between sex-determination systems in a similar manner to sex-specific diploid selection (as explored by van Doorn and Kirkpatrick 2007; 2010). That is, new masculinizing mutations (neo-Y chromosomes) could be favoured via associations with alleles that are beneficial in the male haploid stage. On the other hand, sex ratios can also become biased by linkage between the sex-determining region and a locus that harbours genetic variation in haploid fitness. For example, there are several known cases of sex-ratio bias caused by sex-linked meiotic drive alleles (Burt and Trivers 2006, Chapter 3) or selection among X- and Y-bearing pollen (Lloyd 1974, Conn and Blum 1981, Stehlik and Barrett 2005; 2006, Field et al. 2012; 2013). It is not immediately clear how the spread of new sex-determination systems would be influenced by the combination of sex-ratio biases and associations between haploid selected loci and sex-determining regions.

We find that sex-ratio biases caused by haploid selection can exert Fisherian sex-ratio selection upon novel sex-determiners but that their spread is also determined by selection on genetically-associated alleles. Consequently, it is possible for selection on linked alleles to drive turnover between sex-determining systems despite causing transitory or even permanent increases in sex-ratio bias. In addition to considering haploid selection, another novel development in our model is that we consider loci that are under diploid and/or haploid selection and also in very tight linkage with the ancestral sex-determining region. Even in the absence of haploid selection, we show that transitions between male and female heterogamy can then evolve despite the fact that the neo-sex-determining locus is less closely linked to a locus under selection and therefore disrupts favourable ancestral associations between sex and the alleles selected in that sex.

## Model

144 Change all  $\alpha^\delta$  to  $(1 + \alpha_\Delta^\delta)$ .

146 switch between  $\chi$  and  $\rho$  in all places because  $\chi$  is used for double recombination events.

Change  $\zeta$  to represent zygotic sex ratio of males, consistent with  $q$  and figures.

148 We consider transitions between ancestral and novel sex-determining systems  
149 using a three-locus model, each locus having two alleles. Locus **X** is the ancestral  
150 sex-determining region, with alleles  $X$  and  $Y$  (or  $Z$  and  $W$ ). Locus **A** is a locus  
151 under selection, with alleles  $A$  and  $a$ . Locus **M** is a novel sex-determining region,  
152 at which the null allele ( $M$ ) is initially fixed in the population such that sex of  
153 zygotes is determined by the genotype at the ancestral sex-determining region, **X**;  
154  $XX$  genotypes become females and  $XY$  become males (or  $ZW$  become females  
155 and  $ZZ$  become males). To evaluate the evolution of new sex-determination sys-  
156 tems, we consider the invasion, fixation, maintenance, and/or loss of novel sex-  
157 determining alleles ( $m$ ) at the **M** locus. We assume that the **M** locus is epistatically  
158 dominant over the **X** locus such that zygotes with at least one  $m$  allele develop as  
159 females with probability  $k$  and as males with probability  $1 - k$ , regardless of the  
160 **X** locus genotype. With  $k = 0$ , the  $m$  allele is a masculinizer (i.e., a neo-Y) and  
161 with  $k = 1$  the  $m$  allele is a feminizer (i.e., a neo-W). With intermediate  $k$ , we can  
162 interpret  $m$  as an environmental sex determination (ESD) allele, such that zygotes  
163 develop as females in a proportion ( $k$ ) of the environments they experience.

164 In each generation, we census the genotype frequencies in male and female  
165 gametes/gametophytes (hereafter gametes) before gametic competition. A full de-  
166 scription of our model, including recursion equations, is given in the Appendix.  
First, competition occurs among male gametes (sperm/pollen competition) and  
167 among female gametes (egg/ovule competition) separately. Selection during ga-  
168 metic competition depends on the **A** locus genotype, relative fitnesses are given  
169 by  $w_A^\varphi$  and  $w_a^\varphi$  ( $\varphi \in \{\text{♀}, \text{♂}\}$ ; see table 1). We assume that all gametes compete for  
170 fertilization during gametic competition, which assumes a polygamous mating sys-  
171 tem. Gametic competition in monogamous mating systems is, however, equivalent  
172 to meiotic drive in our model (described below), as both only alter the frequency  
173 of gametes produced by heterozygotes. After gametic competition, random mating  
174 occurs between male and female gametes. The resulting zygotes develop as males  
175 or females, depending on their genotypes at the **X** and **M** loci. Diploid males and

females then experience selection, with relative fitnesses  $w_{AA}^\varphi$ ,  $w_{Aa}^\varphi$ , and  $w_{aa}^\varphi$ . The  
 178 next generation of gametes is produced by meiosis, during which recombination  
 and sex-specific meiotic drive can occur. Recombination (i.e., an odd number of  
 180 cross-overs) occurs between loci **X** and **A** with probability  $r$ , between loci **A** and  
**M** with probability  $R$ , and between loci **X** and **M** with probability  $\chi$ . Any linear  
 182 order of the loci can be modelled with appropriate choices of  $r$ ,  $R$ , and  $\chi$  (see Ta-  
 ble S.1). Individuals that are heterozygous at the **A** locus may experience meiotic  
 184 drive; a gamete produced by  $Aa$  heterozygotes of sex  $\varphi$  bear allele  $A$  with probab-  
 ity  $\alpha^\varphi$ . Thus, the **A** locus can experience sex-specific gametic competition, diploid  
 186 selection, and/or meiotic drive.

Table 1: Relative fitness of different genotypes in sex  $\varphi \in \{\text{♀}, \text{♂}\}$

Genotype	Relative fitness during gametic competition
A	$w_A^\varphi = 1 + t^\varphi$
a	$w_a^\varphi = 1$
Genotype	Relative fitness during diploid selection
AA	$w_{AA}^\varphi = 1 + s^\varphi$
Aa	$w_{Aa}^\varphi = 1 + h^\varphi s^\varphi$
aa	$w_{aa}^\varphi = 1$
Genotype	Transmission during meiosis in $Aa$ heterozygotes
A	$\alpha^\varphi = 1/2 + \alpha_\Delta^\varphi / 2$
a	$1 - \alpha^\varphi = 1/2 - \alpha_\Delta^\varphi / 2$

## Results

188 The model outlined above describes both ancestrally- $XY$  and ancestrally- $ZW$   
 sex-determination systems if we relabel the two sexes as being ancestrally ‘het-  
 190 erogametic’ or ancestrally ‘homogametic’. Without loss of generality, we primar-  
 ily refer to the ancestrally heterogametic sex as male and the ancestrally homoga-

<sup>192</sup> metic sex as female. That is, we describe an ancestral XY sex-determination system but our model is equally applicable to an ancestral ZW sex-determination system (relabelling the ancestrally-heterogametic sex as female and the ancestrally-homogametic sex as male).

<sup>196</sup> **Generic invasion by a neo-Y or neo-W**

The evolution of a new sex-determination system requires that a rare mutant allele at the novel sex-determining locus,  $m$ , increases in frequency when rare. The spread of a rare mutant  $m$  at the **M** locus is determined by the leading eigenvalue,  $\lambda$ , of the system of eight equations describing the frequency of eggs and sperm carrying the  $m$  allele in the next generation (equations S.1). This system simplifies substantially in a number of cases of interest. Dominant neo-Y (when  $k = 0$ ) or neo-W alleles (when  $k = 1$ ) are only found in male diploids (neo-Y) or female diploids (neo-W) such that their growth rate ultimately depends only on the change in frequency of  $m$ -bearing gametes produced by males or by females, respectively. Furthermore, if the  $m$  allele is fully epistatically dominant over the ancestral sex-determining system, phenotypes are not affected by the genotype at the ancestral sex-determining region (**X** locus). Thus, the invasion of rare dominant neo-Y or neo-W alleles is determined by the largest eigenvalue that solves a quadratic characteristic polynomial, **Mention the possibility that the other roots yield the leading eigenvalue somewhere.**  $\lambda^2 + b\lambda + c = 0$ . Here,  $b = -(\lambda_{mA} + \lambda_{ma}) + (\rho_{mA} + \rho_{ma})$  and  $c = (\lambda_{mA} - \rho_{mA})(\lambda_{ma} - \rho_{ma}) - \rho_{mA}\rho_{ma}$ , where  $\lambda_{mi}$  is the multiplicative growth rate of mutant haplotypes on background  $i \in \{A, a\}$ , without accounting for loss due to recombination, and  $\rho_{mi}$  is the rate at which mutant haplotypes on background  $i \in \{A, a\}$  recombine onto the other **A** locus background in heterozygotes (see Table 2). The  $\lambda_{mi}$  and  $\rho_{mi}$ , and thus the spread of the mutant  $m$  allele, depend on the frequency of alleles at the **A** and **X** loci in the ancestral population. In the ancestral population, it is convenient to follow the frequency of the  $A$  allele among female gametes (eggs),  $p_X^\varnothing$ , and among X-bearing,  $p_X^\delta$ , and among Y-bearing,  $p_Y^\delta$ , male gametes (sperm/pollen). We also track the fraction of male gametes that are

Y-bearing,  $q$ , which may deviate from 1/2 due to meiotic drive in males.

222

Table 2: Parameters determining invasion of mutant neo-Y and neo-W alleles into an ancestrally XY system

neo-Y ( $k = 0$ )
$\lambda_{mA} = \{2(1 - \zeta)\}^{-1} [p_X^{\varphi} w_A^{\varphi} w_A^{\delta} w_{AA}^{\delta} + 2(1 - p_X^{\varphi}) w_a^{\varphi} w_A^{\delta} w_{Aa}^{\delta} \alpha^{\delta}] / (\bar{w}_H^{\varphi} \bar{w}_H^{\delta} \bar{w}^{\delta})$
$\lambda_{ma} = \{2(1 - \zeta)\}^{-1} [(1 - p_X^{\varphi}) w_a^{\varphi} w_a^{\delta} w_{aa}^{\delta} + 2p_X^{\varphi} w_A^{\varphi} w_a^{\delta} w_{Aa}^{\delta} (1 - \alpha^{\delta})] / (\bar{w}_H^{\varphi} \bar{w}_H^{\delta} \bar{w}^{\delta})$
$\rho_{mA} = R \{2(1 - \zeta)\}^{-1} [2(1 - p_X^{\varphi}) w_a^{\varphi} w_A^{\delta} w_{Aa}^{\delta} \alpha^{\delta}] / (\bar{w}_H^{\varphi} \bar{w}_H^{\delta} \bar{w}^{\delta})$
$\rho_{ma} = R \{2(1 - \zeta)\}^{-1} [2p_X^{\varphi} w_A^{\varphi} w_a^{\delta} w_{Aa}^{\delta} (1 - \alpha^{\delta})] / (\bar{w}_H^{\varphi} \bar{w}_H^{\delta} \bar{w}^{\delta})$
neo-W ( $k = 1$ )
$\lambda_{mA} = (2\zeta)^{-1} [\bar{p}^{\delta} w_A^{\varphi} w_A^{\delta} w_{AA}^{\varphi} + 2(1 - \bar{p}^{\delta}) w_a^{\delta} w_A^{\varphi} w_{Aa}^{\varphi} \alpha^{\varphi}] / (\bar{w}_H^{\varphi} \bar{w}_H^{\delta} \bar{w}^{\varphi})$
$\lambda_{ma} = (2\zeta)^{-1} [(1 - \bar{p}^{\delta}) w_a^{\delta} w_a^{\varphi} w_{aa}^{\varphi} + 2\bar{p}^{\delta} w_A^{\delta} w_a^{\varphi} w_{Aa}^{\varphi} (1 - \alpha^{\varphi})] / (\bar{w}_H^{\varphi} \bar{w}_H^{\delta} \bar{w}^{\varphi})$
$\rho_{mA} = R (2\zeta)^{-1} [2(1 - \bar{p}^{\delta}) w_a^{\delta} w_A^{\varphi} w_{Aa}^{\varphi} \alpha^{\varphi}] / (\bar{w}_H^{\varphi} \bar{w}_H^{\delta} \bar{w}^{\varphi})$
$\rho_{ma} = R (2\zeta)^{-1} [2\bar{p}^{\delta} w_A^{\delta} w_a^{\varphi} w_{Aa}^{\varphi} (1 - \alpha^{\varphi})] / (\bar{w}_H^{\varphi} \bar{w}_H^{\delta} \bar{w}^{\varphi})$

$\bar{p}^{\delta} = (1 - q)p_X^{\delta} + qp_Y^{\delta}$  is the average frequency of the  $A$  allele among X- and Y-bearing male gametes.

$\zeta$  is the zygotic sex ratio (fraction female)

$\bar{w}^{\delta}$  is the mean fitness of diploids of sex  $\delta$ , see Table S.2

$\bar{w}_H^{\delta}$  is the mean fitness of haploids from sex  $\delta$ , see Table S.2

224 We are particularly concerned with the conditions under which a rare neo-sex-determining allele increases in frequency, which occurs when the largest eigenvalue,  $\lambda$ , is greater than one. If the average change in frequency of the two haplotypes that carry the  $m$  allele ( $Am$  and  $am$ ) is positive, invasion will always occur, 226 i.e., if  $(\lambda_{mA} + \lambda_{ma})/2 > 1$  then  $\lambda > 1$ . If neither haplotype increases in frequency 228 ( $\lambda_{mA}, \lambda_{ma} < 1$ ), the  $m$  allele will not invade. Otherwise, the new sex-determining 230 allele increases in frequency on one A background and declines on the other, and invasion requires

$$\rho_{ma} (\lambda_{mA} - 1) + \rho_{mA} (\lambda_{ma} - 1) > 0. \quad (1)$$

232 For example, if we assume that only the  $mA$  haplotype has a positive growth rate

( $\lambda_{ma} < 1 < \lambda_{mA}$ ), the second term on the left-hand side of (1) is negative and  
234 invasion requires that the growth rate of *mA* haplotypes and the rate at which they  
are produced by recombination is sufficiently large relative to that of *ma* haplo-  
236 types. In other words, invasion requires that the average growth rate of the two  
haplotypes, weighted by the rates they are created by recombination, is positive.

238 Table 2 illustrates a number of key points about the invasion of neo-Y and  
neo-W mutations. First, Fisherian sex-ratio selection will favour the spread of a  
240 neo-Y if the ancestral zygotic sex ratio is biased towards females,  $\zeta > 1/2$  (i.e.,  
 $\zeta > 1/2$  causes the first factor of the  $\lambda_{mi}$  to be greater than one for a neo-Y and  
242 vice versa for a neo-W). However, the spread of a neo-Y (neo-W) also depends on  
the male (female) fitness of associated alleles (terms in square brackets). Second,  
244 invasion by a neo-Y (neo-W) allele does not directly depend on the fitness of female  
(male) diploids (for a given set of equilibrium allele frequencies). This is because  
246 a dominant neo-Y (neo-W) is always found in males (females), and therefore the  
frequency of the neo-Y (neo-W) allele,  $m$ , only changes in males (females). Finally,  
248 invasions by a neo-Y and a neo-W are qualitatively different. This is because a  
gamete with the ancestral- or neo-Y always pairs with a female gamete containing  
250 an X, and both develop into males. By contrast, a gamete with a neo-W can pair  
with an X or Y male gamete, developing into a female, while female gametes  
252 without the neo-W can become female (when paired with X) or male (when paired  
with Y). Consequently, the types of females produced differ in the frequency of *A*  
254 alleles they obtain from mating.

In order to explicitly determine the conditions under which a rare neo-sex-  
256 determining allele spreads, we must calculate the equilibrium frequency of the *A*  
allele in the ancestral population (i.e.,  $\hat{p}_X^{\varnothing}$ ,  $\hat{p}_X^{\delta}$ , and  $\hat{p}_Y^{\delta}$ ). Since only the A locus expe-  
258 riences selection directly, any deterministic evolution requires that there is a poly-  
morphism at the A locus. Polymorphisms can be maintained by mutation-selection  
260 balance or transiently present during the spread of beneficial alleles. However,  
polymorphisms maintained by selection can maintain alleles at higher allele fre-  
262 quencies for longer periods. Here, we focus of polymorphisms maintained by se-

lection, where the  $A$  allele reaches a stable intermediate equilibrium frequency  
264 under the ancestral sex-determination system before the neo-sex-determining allele ( $m$ ) arises. We can analytically calculate the allele frequency of the  $A$  allele  
266 using two alternative simplifying assumptions: (1) the  $A$  locus is within (or tightly linked to) the non-recombining region around the ancestral SDR ( $r \approx 0$ ) or (2) selection is weak relative to recombination ( $s^{\delta}, t^{\delta}, \alpha_{\Delta}^{\delta}$  of order  $\epsilon \ll 1$ ).  
268

Change to  $\hat{p}$  throughout as we assume that allele frequencies change slowly  
270 such that lambda is unaffected

### Tight linkage with the ancestral sex-determining region

272 The ancestral equilibrium allele frequencies and their stability conditions are given  
in the appendix. When there is complete linkage between the ancestral sex-determining  
274 region and the  $A$  locus ( $r = 0$ ), either the  $A$  allele or the  $a$  allele must be fixed on  
the Y. Because the labelling of alleles is arbitrary, we will assume that the  $a$  locus  
276 is fixed on the Y ( $p_Y^{\delta} = 0$ ), without loss of generality. If there are two alleles main-  
tained at the  $A$  locus, the X can either be fixed for the  $A$  allele ( $\hat{p}_X^{\delta} = \hat{p}_X^{\delta} = 1$ ) or  
278 polymorphic ( $0 < \hat{p}_X^{\delta}, \hat{p}_X^{\delta} < 1$ ).

280 A neo-Y will never invade an ancestral XY system that already has tight linkage  
with the locus under selection ( $r = 0$ , for details see supplementary *Mathematica*  
file). A neo-Y haplotype with the same allele as the ancestral Y is neutral ( $\lambda_{ma} = 1$ )  
282 and does not change in frequency. The other neo-Y haplotype will not spread  
( $\lambda_{mA} < 1$ ) given that the initial equilibrium is stable. Therefore, a neo-Y mutation  
284 cannot spread ( $\lambda \leq 1$ ) in an ancestral XY system that is at equilibrium with all se-  
lected loci within the non-recombining region around the SDR. In essence, through  
286 tight linkage with the  $A$  locus, the ancestral Y becomes strongly specialized on the  
allele that has the highest fitness across male haploid and diploid phases. Given  
288 that the ancestral Y is at this equilibrium, it is not possible for a neo-Y to create  
males that have higher fitness than the ancestral Y.

290 Neo-W alleles, on the other hand, can invade an ancestral XY system under  
some conditions (the full invasion conditions are given in the appendix; equations

292 S.6 and S.7). That is, selection on loci within the non-recombining region of the  
 293 SDR can favour the invasion of a less closely linked neo-W, see Figure 1. This re-  
 294 sult is unexpected given the results of van Doorn and Kirkpatrick (2010), who did  
 295 not explicitly calculate equilibrium allele frequencies under tight linkage and gen-  
 296 erally concluded that heterogametic transitions occur when neo-sex-determining  
 297 alleles are in tighter linkage with loci under sex-specific diploid selection. To de-  
 298 velop an understanding (intuition) for how this happens, we focus on cases where  
 299 there is no haploid selection and discuss the effects of haploid selection in the  
 300 appendix.

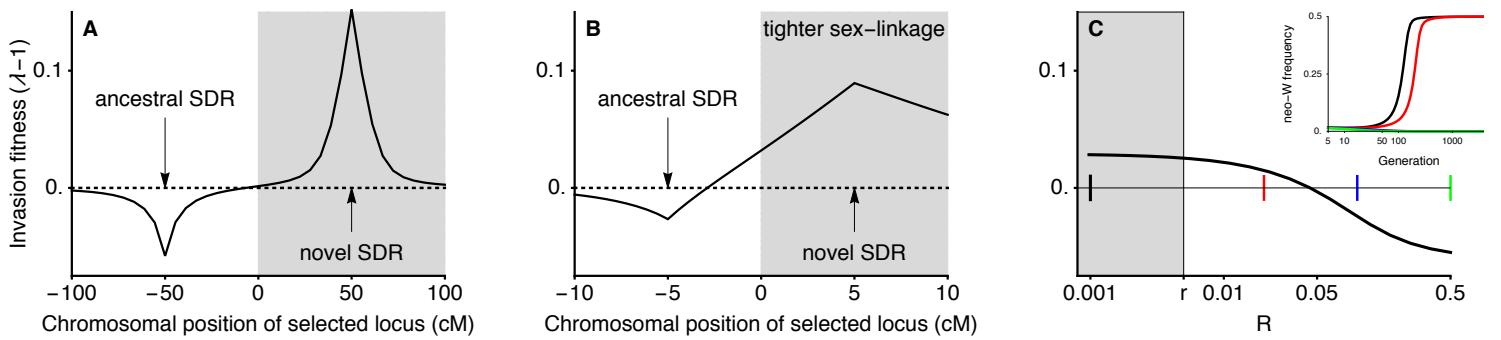


Figure 1: Transitions between XY and ZW systems can occur even when the neo-SDR is more loosely linked to a locus under sexually-antagonistic selection (here, without haploid selection  $t^\delta = \alpha_\Delta^\delta = 0$ ). In panel A, linkage is loose enough relative to selection that the weak selection analytical results hold, and a neo-W can only invade when it is more tightly linked with the selected locus ( $R < r$ ; shaded region). In panel B, linkage is tight enough relative to selection that the weak selection analytical results do not hold, and a neo-W can only invade even when it is less tightly linked with the selected locus ( $r < R$ ; unshaded region). In panel C we vary the recombination rate between the neo-W and the selected locus ( $R$ ) for a fixed recombination rate between the ancestral-SDR and the selected locus ( $r = 0.005$ ). Coloured markers show recombination rates for which the temporal dynamics of invasion are plotted in the inset, demonstrating that neo-W alleles can fix (reach frequency 0.5 among female gametes) if they are more (black) or less (red) closely linked to a locus experiencing sexually-antagonistic selection. A very loosely linked neo-W does not spread in this case (blue and green lines overlap and go to 0). Indeed, we show that neo-W invasion fitness is negative when  $R = 1/2$  and  $s^q s^\delta < 0$ ,  $0 < h^\delta < 1$  in the supplementary *Mathematica* file. Fitness parameters are shown by an asterisk in Figure 2:  $w_{AA}^q = 1.05$ ,  $w_{aa}^q = 1.2$ ,  $w_{aa}^\delta = w_{AA}^\delta = 0.85$ ,  $w_{Aa}^\delta = 1$ .

If we categorise the  $a$  allele as being ancestrally ‘male-beneficial’ via the fact  
 302 that it is fixed on the Y, then  $\lambda_{mA} > 1$  indicates that the neo-W spreads when found  
 with the ancestrally ‘female-beneficial’ allele. Broadly, this is possible because

304 the ancestral X chromosome is not able to perfectly specialise on the ‘female-  
305 beneficial’ allele due to the fact that X’s are sometimes found in males. For ex-  
306 ample, when the *a* allele is favoured in males, a polymorphism of *A* and *a* alleles  
307 can be maintained on the X despite directional selection in favour of the *A* al-  
308 lele in females ( $s^F > 0$ ,  $0 < h^F < 1$ ). Figure 2A indicates that  $\lambda_{mA}$  tends to be  
309 larger than one with sexually-antagonistic selection where the *a* allele is strongly  
310 favoured in males ( $w_{aa}$  much larger than  $w_{Aa}$ ). In this case the *a* allele is at high  
311 frequency among XX females is high due to selection upon the X in males. By  
312 contrast, W-*A* haplotypes will only create females with high fitness (*AA* or *Aa*  
313 genotypes) and can therefore have higher fitness than ancestral females. When  
314 only one neo-W haplotype can has a positive growth rate (see Figure 2), a neo-W  
315 can invade as long as equation (1) is satisfied, which may require that the recom-  
316 bination rate, *R*, is small enough. Nevertheless, because we assume here that *r* is  
317 small, these results indicate that a more loosely linked sex-determining region can  
318 spread. Therefore, tightly sex-linked loci that experience sexually-antagonistic se-  
319 lection can drive heterogametic transitions in which the neo-SDR is less closely  
320 linked to the locus under selection (Figure 1).

Given that the *a* allele can be considered ancestrally ‘male-beneficial’ because  
321 it is fixed on the Y, it is surprising that neo-W-*a* haplotypes can sometimes be  
322 favoured by selection in females ( $\lambda_{ma} > 1$ ). Again, this occurs because ancestral  
323 X’s also experience selection in males, in which they will always be paired with  
324 a Y-*a*. Hence, if there is overdominance in males, X-*A* Y-*a* males have high fit-  
325 ness and the *A* allele is favoured by selection on the X in males. Therefore, the  
326 X can be polymorphic or even fixed for the *A* allele despite favouring the *a* allele  
327 during selection in females (e.g., see outlined region in Figure 2B and Lloyd and  
328 Webb 1977, Otto 2014). In such cases, neo-W-*a* haplotypes can spread because  
329 they create more *Aa* and *aa* females when pairing with an X from males and be-  
330 cause they bring Y-*a* haplotypes into females, in which case females are always *aa*.  
331 As discussed in the appendix, this scenario where neo-W’s associated with *a* are  
332 favoured can also occur with haploid selection, even without overdominance (e.g.,

334 when  $a$  is female-beneficial and favoured by haploid selection in male gametes).

In some cases, both W-A and W- $a$  haplotypes can spread, e.g., when  $AA$  individuals have low fitness in females yet the  $A$  is polymorphic or fixed on the X due to overdominance in males (Figure 2B and 2C). Both neo-W-A and neo-W- $a$  haplotypes then produce fewer unfit  $AA$  females. This is true for the neo-W-A haplotype because it can pair with a  $Y - a$  haplotype and still be female. Whenever both haplotypes have positive growth rates, invasion by a neo-W is expected regardless of its linkage with the selected locus (i.e., even unlinked neo-W alleles can invade, see Figures S.1 and S.2 for examples).

What can we discuss about haploid selection here. Perhaps the fact that overdominance is not required for  $\lambda_{Ma} > 1$  when there is haploid selection? We also don't yet discuss the fact that polymorphic equilibria (mixed systems) can be stable.

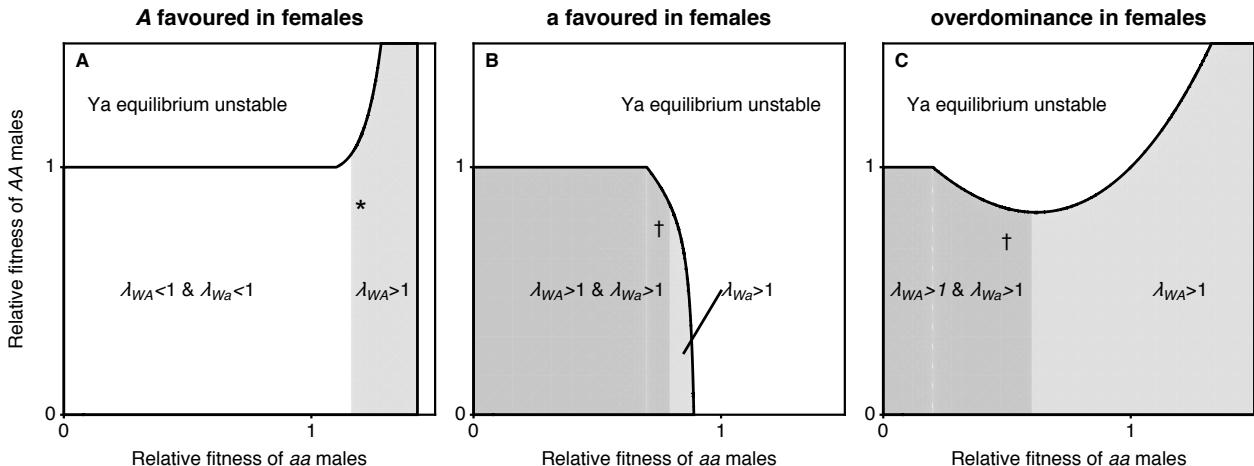


Figure 2: When the ancestral-XY locus is tightly linked to a locus under selection ( $r = 0$ ), one or both neo-W haplotypes can spread. We vary the fitness of male homozygotes relative to heterozygotes ( $w_{Aa}^\delta = 1$ ) and only consider stable equilibria at which both  $A$  locus alleles are maintained and the  $a$  allele is initially fixed on the Y, region outlined. Here, selection in females can favour the  $A$  allele (panel A,  $w_{aa}^\delta = 0.85$ ,  $w_{AA}^\delta = 1.05$ ), favour the  $a$  allele (panel B,  $w_{aa}^\delta = 1.05$ ,  $w_{AA}^\delta = 0.85$ ), or be overdominant (panel C,  $w_{aa}^\delta = w_{AA}^\delta = 0.6$ ). If  $\lambda_{wA}$  or  $\lambda_{wa}$  is greater than one, then a rare neo-W can spread for, at least, some values of  $R > r$ . For the parameter values marked with an asterisk, example invasion dynamics are shown in Figure 1C. Where both  $\lambda_{wA}$  and  $\lambda_{wa}$  are greater than one, a neo-W will spread when rare, regardless of linkage with the selected locus (for any  $R$ ). Figure S.1 shows two examples using the parameters marked with a dagger. Here, there is no haploid selection  $t^\delta = \alpha_\Delta^\delta = 0$ .

## Loose linkage with the ancestral sex-determining region

348 Assuming that selection is weak relative to all recombination rates ( $r$ ,  $R$  and  $\chi$ ),  
 we denote the leading eigenvalues describing the invasion of a neo-Y ( $k = 0$ ) and  
 350 a neo-W ( $k = 1$ ) into an ancestrally XY system by  $\lambda_{Y',XY}$  and  $\lambda_{W',XY}$ , respectively.  
 To leading order in selection, these are:

$$\lambda_{Y',XY} = 1 + V_A S_A^2 \frac{(r - R)}{rR} + O(\epsilon^3) \quad (2)$$

352 and

$$\lambda_{W',XY} = \lambda_{Y',XY} + (2\alpha_\Delta^\delta - 2\alpha_\Delta^? + t^\delta - t^?) (\hat{p}_Y^\delta - \hat{p}_X^\delta) / 2 + O(\epsilon^3) \quad (3)$$

where  $V_A = \bar{p}(1 - \bar{p})$  is the variance in the frequency of  $A$  and  $S_A = (D^\delta + \alpha_\Delta^\delta + t^\delta) - (D^? + \alpha_\Delta^? + t^?)$  describes sex differences in selection for the  $A$  versus  $a$  across diploid selection, meiosis, and gametic competition. The diploid selection term,  
 354  $D^\delta = (\bar{p}s^\delta + (1 - \bar{p})h^\delta s^\delta) - (\bar{p}h^\delta s^\delta + (1 - \bar{p}))$ , is the difference in fitness between  $A$   
 356 and  $a$  alleles in diploids of sex  $\delta \in \{\text{♀}, \text{♂}\}$ , where  $\bar{p}$  is the leading-order probability  
 358 of mating with an  $A$ -bearing gamete from the opposite sex (see Appendix). The  
 difference in  $A$ -allele-frequency among Y-bearing sperm versus X-bearing sperm  
 360 is given by  $\hat{p}_Y^\delta - \hat{p}_X^\delta = V_A (D^\delta - D^? + \alpha_\Delta^\delta - \alpha_\Delta^? + t^\delta - t^?) (1 - 2r) / 2r$ .

The neo-sex-determining allele,  $m$ , will spread if  $\lambda_{m,XY} > 1$ . Equation (2)  
 362 demonstrates that under weak selection a neo-Y will invade an XY system if and  
 only if it is more closely linked to the selected locus than the ancestral sex-determining  
 364 region (i.e., if  $R < r$ ; note that  $V_A S_A^2$  is strictly positive as long as  $A$  is polymor-  
 phic). This echoes our tight linkage results above where a neo-Y could never invade  
 366 if  $r \approx 0$  and is consistent with the results of van Doorn and Kirkpatrick (2007),  
 who considered diploid selection only and also found that homogametic transitions  
 368 (XY to XY or ZW to ZW) can only occur when the neo-sex-determining locus is  
 more closely linked to a locus under sexually-antagonistic selection.

370 With weak selection and no haploid selection ( $t^\delta = \alpha_\Delta^\delta = 0$ ), the spread of

a neo-W is equivalent to the spread of a neo-Y ( $\lambda_{W',XY} = \lambda_{Y',XY}$ ), such that heterogametic transitions (XY to ZW or ZW to XY) can also occur only if the neo-sex-determining region is more closely linked to a locus under selection ( $R < r$ ),  
372 as found by van Doorn and Kirkpatrick (2010). With haploid selection, however,  
374 the additional term in equation (3) can be positive, which can allow, for example,  
376 neo-W invasion ( $\lambda_{W',XY} > 1$ ) even when the neo-sex-determining region is less  
closely linked to the selected locus ( $R > r$ ).

378 Equation (3) shows that, with weak selection, neo-W alleles can invade an XY  
system for a large number of selective regimes. To clarify the parameter space  
380 under which  $\lambda_{W',XY} > 1$ , we consider several special cases. Firstly, if the **A** locus  
is unlinked to the ancestral sex-determining region ( $r = 1/2$ ), a more closely linked  
382 neo-W ( $R < 1/2$ ) can always invade because there is then no association between  
 $A$  alleles and sex chromosomes in males,  $(\hat{p}_Y^\delta - \hat{p}_X^\delta) = 0$ , see equation (S.5). The  
384 second term in equation (3) then disappears and invasion depends only on the sign  
of  $(r - R)$ . Indeed, invasion typically occurs when the neo-W is more closely linked  
386 to the selected locus than the ancestral sex-determining region, under a variety of  
selective regimes (Figure 3). Secondly, we can simplify the discussion of cases  
388 where invasion occurs despite looser sex-linkage,  $R > r$ , by focussing on the  
special case where  $R = 1/2$  and  $r < 1/2$  (e.g., the selected locus is on the ancestral  
390 sex chromosome and the novel sex-determining locus arises on an autosome). In  
table 3 we give the conditions where invasion occurs when we further assume  
392 that haploid selection only occurs in one sex (e.g., during male meiosis only) and  
dominance coefficients are equal in the two sexes,  $h^q = h^\delta$ . When there is no  
394 gametic competition and meiotic drive is in one sex only, an unlinked neo-W can  
invade as long as the same allele is favoured during diploid selection in males and  
396 females ( $s^q s^\delta > 0$ , see Figure 3B and Figure 4B). When there is no meiotic drive  
and gametic competition occurs in one sex only, an unlinked neo-W can invade as  
398 long as the same allele is favoured in male and female diploid selection and there  
are sex differences in selection of one type (e.g.,  $s^q(s^\delta - s^q) > 0$ , see Figure 3C).  
400 These special cases indicate that neo-W invasion can occur for a relatively large

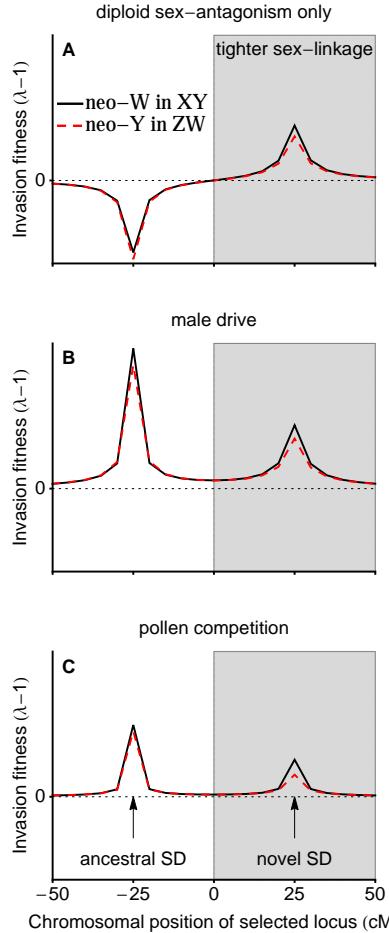


Figure 3: A neo-W can invade an XY system under a large number of selective regimes. In panel A, there is no haploid selection ( $t^\delta = \alpha_\Delta^\delta = 0$ ) and selection in diploids is sexually antagonistic ( $s^\delta = -s^q = 1/10$ ,  $h^\delta = 1 - h^q = 3/10$ ), in which case the neo-sex-determining allele can only invade if it is more closely linked to the selected locus ( $R < r$ , gray region; but see Figure 1B for the case of very tight linkage). In panel B, male drive ( $\alpha_\Delta^\delta = -1/20$ ,  $t^\delta = \alpha_\Delta^\delta = 0$ ) opposes selection in diploids (no sex-differences:  $s^\delta = 1/10$ ,  $h^\delta = 7/10$ ), in which case the neo-sex-determining allele can invade regardless of linkage. In panel C, gametic competition in males ( $t^\delta = -1/10$ ,  $t^q = \alpha_\Delta^q = 0$ ) opposes selection in diploids (sex-differences:  $s^\delta = 1/20$ ,  $s^q = 3/20$ ,  $h^\delta = 7/10$ ), in which case the neo-sex-determining allele can once again invade regardless of linkage. We use Haldane's map function (Equation 3 in Haldane 1919) to convert from map distance (centiMorgans, cM) to the probability of recombination (an odd number of cross-over events). **Check the mismatch between red and black lines here: probably because of adding or subtracting from 1.**

fraction of the parameter space, even if the neo-W uncouples the sex-determining  
 402 locus from a locus under selection.

Previous research suggests that when the ancestral sex-determining locus is

Table 3: Invasion conditions for unlinked neo-W ( $R = 1/2$ ,  $r < 1/2$ ) into ancestral XY with one form of haploid selection

Scenario	Assumptions	neo-W spreads ( $\lambda_{W',XY} > 1$ ) if
male drive only	$h^\delta = h^\varnothing, t^\varnothing = t^\delta = \alpha_\Delta^\varnothing = 0$	$s^\varnothing s^\delta > 0$
female drive only	$h^\delta = h^\varnothing, t^\varnothing = t^\delta = \alpha_\Delta^\delta = 0$	$s^\varnothing s^\delta > 0$
sperm competition only	$h^\delta = h^\varnothing, t^\varnothing = \alpha_\Delta^\varnothing = \alpha_\Delta^\delta = 0$	$s^\varnothing(s^\delta - s^\varnothing) > 0$
egg competition only	$h^\delta = h^\varnothing, t^\delta = \alpha_\Delta^\varnothing = \alpha_\Delta^\delta = 0$	$s^\delta(s^\varnothing - s^\delta) > 0$

404 linked to a locus that experiences haploid selection (e.g., meiotic drive), a new,  
405 unlinked sex-determining locus invades in order to restore equal sex ratios (Koziel-  
406 ska et al. 2010). Consider, for example, the case where the A locus is linked to the  
407 ancestral-SDR ( $r < 1/2$ ) and experiences meiotic drive in males only (e.g., dur-  
408 ing spermatogenesis but not during oogenesis,  $\alpha_\Delta^\delta \neq 0, \alpha_\Delta^\varnothing = 0$ ), without gametic  
409 competition ( $t^\varnothing = t^\delta = 0$ ). In this case, the zygotic sex ratio can be initially biased  
410 only if the ancestral sex-determining system is XY (Figure 4B). We might there-  
411 fore expect a difference in the potential for XY to ZW and ZW to XY transitions.  
412 However, to leading order with selection weak relative to recombination, we find  
413 that sex ratio selection favours the spread of a neo-W (through the first terms in  
414 table 2) by an amount that is equal in magnitude to the fitness effects of alleles  
415 associated with new sex-determining alleles (second terms in table 2). Thus, in-  
416 vasion by a neo-W into an XY system and invasion by a neo-Y into a ZW system  
417 occur under the same conditions ( $\lambda_{Y',XY} = \lambda_{W',ZW}$  and  $\lambda_{Y',ZW} = \lambda_{W',XY}$ , at least  
418 to order  $\epsilon^2$ ). For example, in Figure 4B neo-W alleles invade an ancestrally XY  
419 system where females are initially rare because the ancestral-Y is associated with  
420 a male meiotic drive allele. However, Figure 4A shows that a neo-Y can invade  
421 an ancestrally ZW system under the same conditions. In fact, where  $R < 1/2$  the  
422 neo-Y becomes associated with the male meiotic drive allele such that the zygotic  
423 sex ratio evolves to become biased towards males.

424 The green curves in Figure 4 demonstrate a case where transitions between  
425 male and female heterogamety occur even though the new sex-determining re-  
426 gion is unlinked to a locus that experiences haploid and diploid selection. We

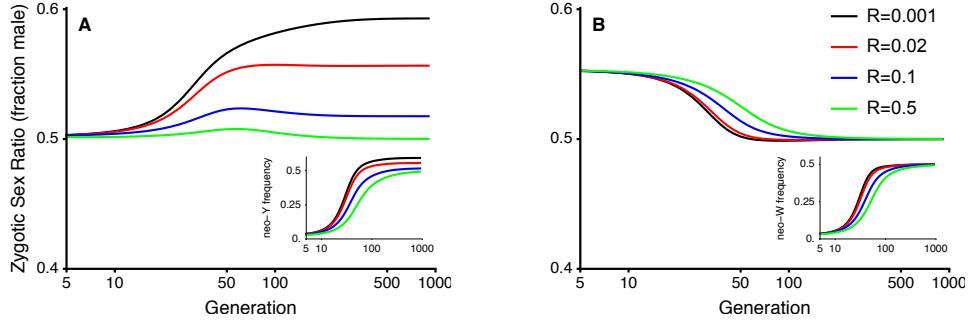


Figure 4: Fisherian sex-ratio selection alone is not a good predictor of turnover between sex-determining systems. In this figure, selection is pliodally antagonistic with haploid selection favouring the  $a$  allele during male meiosis ( $s^q = s^\delta = 0.2$ ,  $h^q = h^\delta = 0.7$ ,  $t^q = t^\delta = \alpha_\Delta^\delta = 0$ ). In panel A, male drive in an ancestral ZW system has no affect on the zygotic sex ratio, yet a neo-Y can invade and replace the ancestral sex-determination system (inset shows neo-Y frequency among male gametes, the ancestral W also goes to fixation during this transition). When  $R < 1/2$ , the neo-Y becomes associated with the allele favoured by drive, causing the zygotic sex ratio to become biased, hence the frequency of neo-Y among male gametes can be higher than 0.5 (inset). In panel B, male drive in an ancestral XY system causes a male bias, allowing a neo-W to invade and replace the ancestral sex-determination system (inset shows neo-W frequency among female gametes, the ancestral Y also goes to fixation), which balances the zygotic sex ratio. Parameters:  $s^q = s^\delta = 0.2$ ,  $h^q = h^\delta = 0.7$ ,  $t^q = t^\delta = \alpha_\Delta^\delta = 0$ ,  $\alpha_\Delta^\delta = -0.2$ ,  $r = 0.02$ .

use these green curves to discuss why heterogametic transitions can occur when  
 428  $R = 1/2$  and  $r < 1/2$ , as in Table 3. In Figure 4B, an unlinked neo-W can spread  
 because the zygotic sex ratio is ancestrally male biased. However, in Figure 4A, an  
 430 unlinked neo-Y spreads despite the fact that the ancestral zygotic sex ratio is even.  
 In this case, the the male meiotic drive allele,  $a$ , is initially more common among  
 432 ancestral-Z-bearing eggs than ancestral-W-bearing eggs because the Z is found in  
 males more often than the W and  $r < 1/2$  (equation S.5). Polymorphism at the A  
 434 locus is maintained by counter-selection against the  $a$  allele in diploids and there-  
 fore ancestral-ZZ males have generally low diploid fitness. A freely recombining  
 436 neo-Y ( $R = 1/2$ ) is not directly favoured or disfavoured by male meiotic drive  
 because it is equally likely to be segregate with the  $A$  or  $a$  allele when found in  
 438 a heterozygote. The neo-Y spreads because it produces males with high diploid  
 fitness through matings with ancestral-W-bearing female gametes, which are more  
 440 likely to carry the  $A$  allele. Thus, a key factor in explaining why heterogametic  
 transitions can occur when  $R > r$  is that that the neo-SDR determines sex in the  
 442 diploid phase but recombination occurs before any subsequent haploid selection.

## Environmental sex determination

<sup>444</sup> We next consider the case where the new sex-determining mutation,  $m$ , causes sex to be determined probabilistically or by heterogeneous environmental conditions  
<sup>446</sup> (environmental sex determination, ESD), with individuals carrying allele  $m$  developing as females with probability  $k$ . Here, we do not assume that the environmental  
<sup>448</sup> conditions that determine sex also differentially affect the fitness of males versus females. Such correlations can favour environmental sex-determination systems  
<sup>450</sup> that allow each sex to be produced in the environment in which it has highest fitness; in the absence of these correlations, previous theory would predict that ESD  
<sup>452</sup> is favoured when it produces more equal sex ratios than the ancestral system (see reviews by Charnov 1982, Bull 1983, West 2009).

<sup>454</sup> The characteristic polynomial determining the eigenvalues (equations S.1) does not factor for ESD mutants as it does for  $k = 0$  or  $k = 1$ . We therefore focus  
<sup>456</sup> on weak selection here. Assuming weak selection, the spread of the new sex-determining region is given by

$$\begin{aligned}\lambda_{ESD',XY} = & 1 + (1 - 2k)^2 V_A S_A^2 \frac{r - R}{rR} \\ & + \frac{k(\hat{p}_Y^\delta - \hat{p}_X^\delta)}{2} (k(2\alpha_\Delta^\delta - 2\alpha_\Delta^\varnothing + t^\delta - t^\varnothing) - 4(1 - k)S_A) + O(\epsilon^3),\end{aligned}\tag{4}$$

<sup>458</sup> which reduces to  $\lambda_{Y',XY}$  when  $k = 0$  and  $\lambda_{W',XY}$  when  $k = 1$ .

Of particular interest are ESD mutations that cause half of their carriers to  
<sup>460</sup> develop as females and half as males ( $k = 1/2$ , creating equal sex ratios), the spread of which is given by

$$\lambda_{ESD',XY} = 1 + \frac{1}{2} \frac{(\lambda_{Y',XY|R=1/2} - 1) + (\lambda_{W',XY|R=1/2} - 1)}{2} + O(\epsilon^3),\tag{5}$$

<sup>462</sup> where  $\lambda_{Y',XY|R=1/2}$  and  $\lambda_{W',XY|R=1/2}$  represent  $\lambda_{Y',XY}$  and  $\lambda_{W',XY}$  when evaluated at  $R = 1/2$  (equations 2 and 3). That is, recombination between the selected locus

and the novel sex-determining locus,  $R$ , doesn't enter into the  $k = 1/2$  results. This is because sex is essentially randomized each generation, preventing associations from building up between allele  $A$  and sex. Equation (5) shows that the neo-ESD gets half of the fitness of a feminizing mutation (neo- $W$ ) and half of the fitness of a masculinizing mutation (neo- $Y$ ), but only has an effect one half of the time (the other half of the time it produces the same sex as the ancestral system would have, to leading order). As discussed above,  $\lambda_{Y',XY|R=1/2}$  is necessarily less than one, but  $\lambda_{W',XY|R=1/2}$  can be greater than one if there is haploid selection. That is, when there is haploid selection, ESD mutations can invade an ancestrally-XY system because they generate females that are either rare or have high fitness, in the same manner as a neo- $W$ .

Significantly, equation (5) is the same whether ESD is invading an ancestrally XY or ZW system (because  $\lambda_{Y',XY} = \lambda_{W',ZW}$  and  $\lambda_{W',XY} = \lambda_{Y',ZW}$ ). Thus, because the sex ratio is only biased by male haploid selection when the ancestral sex-determination system is XY, Fisherian sex-ratio selection alone does not explain the invasion of ESD under weak selection. Specifically, with male haploid selection, the neo-ESD is equally likely to invade when it equalizes the zygotic sex ratio (through  $\lambda_{W',XY}$ ) and when it doesn't (through  $\lambda_{Y',ZW}$ ). In addition, we note that ESD may not invade, even if the sex ratio is initially biased (e.g., with drive in males only,  $r < 1/2$ ,  $h^{\varphi} = h^{\delta}$ , and  $s^{\varphi}s^{\delta} < 0$ , such that  $\lambda_{W',XY} < 1$ , see Table 3).

## Discussion

Two predominant theories explaining the remarkably high frequency of transitions between sex-determination systems are sexually-antagonistic selection and sex-ratio selection (reviewed in Blaser et al. 2012) ([van Doorn, 2014, sexual development](#)). The former predicts that neo-sex-determining alleles can invade when they arise in closer linkage with a sexually-antagonistic locus (van Doorn and Kirkpatrick 2007; 2010). The latter predicts that new sex-determining systems are generally favoured if they result in more equal sex-ratios than the ancestral sys-

492 tem. Firstly, we show that selection (including sexually-antagonistic selection) on  
493 loci within or near the non-recombining region of the ancestral sex-determining  
494 region can favour heterogametic transitions (XY to ZW or ZW to XY) to new  
495 sex-determining systems that have looser linkage (e.g., see Figure 1). Secondly,  
496 assuming that selection is weak relative to recombination, we show that new sex-  
497 determining alleles are typically favoured if they are more closely linked to a locus  
498 under haploid selection, which is the only condition favouring homogametic trans-  
499 missions (XY to XY or ZW to ZW). In addition, with haploid selection and weak  
500 selection, heterogametic transitions (XY to ZW or ZW to XY) can occur even when  
501 the new sex-determining region is less closely linked to the locus under selection  
502 (e.g., see Figure 4).

503 Sex-ratio biases caused by haploid selection can facilitate heterogametic transi-  
504 tions between sex-determining systems. For instance, alleles favoured by haploid  
505 selection in males often become associated with the Y, which leads to a male-  
506 biased zygotic sex-ratio. This male bias increases the potential for a neo-W to  
507 invade (Table 2), which can equalize the sex-ratio (e.g., see Figure 4B, for related  
508 examples see Kozielska et al. 2010, Úbeda et al. 2015). However, sex-ratio se-  
509 lection can be overwhelmed by additional selective effects (e.g., when a linked  
510 allele is beneficial for male diploids but detrimental for female diploids; Table 3),  
511 preventing the neo-W from invading. Indeed, transitions between sex-determining  
512 systems can even lead to stronger sex-ratio biases. For example, where a neo-Y  
513 invades and is linked with a locus that experiences haploid selection in male ga-  
514 metes, the sex ratio evolves to become biased (e.g., see Figure 4A and step 1 in  
Úbeda et al. 2015). Furthermore, with weak selection, we find that there is no  
516 difference in conditions allowing XY to ZW and ZW to XY transitions, indicating  
517 that sex chromosome transitions are not predominantly predicted by their effect on  
518 the sex-ratio (i.e., the sex-ratio bias created by male haploid selection facilitates  
519 the spread of a neo-W into an XY system the same way that male haploid selection  
520 drives the spread of a neo-Y into a ZW system with a 1:1 sex ratio). Thus, haploid  
selection can favour heterogametic transitions both via sex-ratio selection and via

522 fitness effects of alleles that are associated with the neo-sex-determining allele,  
523 and these selection pressures are predicted to often be of roughly equal magnitude  
524 (unless linkage is tight).

We have shown that the spread of new sex determination systems can be driven  
526 by loci experiencing haploid selection. Because haploid selection can cause transi-  
527 tions that increase or decrease sex-linkage, haploid selection may lead to less  
528 stability, and greater potential for cycling, in sex-determination systems (e.g., the  
529 final state of the red line in Figure 4A is the starting state in Figure 4B). In par-  
530 ticular, if haploid selection is strong but selective differences between male and  
531 female diploids are weak, we find that heterogametic transitions (XY to ZW or  
532 vice versa) are favoured more strongly than homogametic transitions (e.g., with  
533  $|D^\delta - D^q| << |\alpha_\Delta^\delta - \alpha_\Delta^q + t^\delta - t^q|$  we have  $\lambda_{W',XY} > \lambda_{Y',XY}$ ; equations 3 and S.5).  
534 Turnovers driven by haploid selection may help to explain the relative rarity of  
535 heteromorphic sex chromosomes in plants, which are thought to experience more  
536 selection during their multicellular haploid stage. For example, among relatively  
537 few dioecious clades in which multiple species have well characterized sex chro-  
538 mosomes (Ming et al. 2011), heterogametic transitions have been inferred in *Silene*  
539 subsection *Otites* (Slancarova et al. 2013) and in *Salicaceae* (Pucholt et al. 2015,  
540 2017). Furthermore, assuming that transitions from dioecy to hermaphroditism  
541 (equal parental investment in male and female gametes) are favoured in a simi-  
542 lar manner to the ESD examined here (equal probability of zygotes developing as  
543 males or females), our results suggest that competition during the haploid stage  
544 could drive transitions between dioecy and hermaphroditism, which are frequent  
545 in plants (Käfer et al., 2017, Sabath et al., 2017).

546 In support of their role in sex chromosome turnover, genes expected to be un-  
547 der sexually-antagonistic selection (e.g., those causing bright male colouration)  
548 have been found on recently derived sex chromosomes (Lindholm and Breden  
549 2002, Tripathi et al. 2009, Ser et al. 2010). Our results show that, if loci ex-  
550periencing overdominance and/or sexually-antagonistic selection can be identi-  
551 fied in close linkage with the ancestral sex-determining locus (rather than only

552 the novel sex-determining locus), then they could also be implicated in driving  
553 heterogametic transitions between sex-determination systems. As noted by van  
554 Doorn and Kirkpatrick (2010), it would be prudent to compare closely related  
555 clades in order to determine whether observed polymorphisms predate a transi-  
556 tion in sex-determination or arose afterwards. In addition, we show haploid se-  
557 lection on loci around either the ancestral- or the novel-sex-determining regions  
558 could have had a role in driving sex chromosome turnover. A recent transcript-  
559 ome analysis in *Rumex*, suggests a role for gametic competition in the evolution  
560 of sex-determination systems, showing that Y-linked genes are have higher expres-  
561 sion in haploid pollen than autosomal genes ([check this is accurate](#)). Interestingly,  
562 haploid-expression is also more common on the autosome that is orthologous to the  
563 sex chromosomes in closely related species suggesting that new sex chromosomes  
564 may have been favoured through their association with haploid selected alleles on  
565 these chromosomes ([Sandler et al., 2017, Personal Communication](#)).

566 We assume that sex-determining alleles do not experience direct selection ex-  
567 cept via their associations with sex and selected alleles. However, in some cases,  
568 there may be significant degeneration around the sex-limited allele (Y or W) in the  
569 ancestral sex-determining region because recessive deleterious mutations and/or  
570 deletions accumulate around the Y or W sex-determining regions (Rice 1996,  
571 Charlesworth and Charlesworth 2000, Bachtrog 2006, Marais et al. 2008). During  
572 heterogametic transitions (XY to ZW or ZW to XY), but not homogametic transi-  
573 tions (XY to XY or ZW to ZW), any recessive deleterious alleles linked to the Y or  
574 W are revealed to selection in YY or WW individuals (Bachtrog et al. 2014). This  
575 phenomenon was studied by van Doorn and Kirkpatrick (2010), who found that  
576 degeneration can prevent fixation of a neo-W or a neo-Y allele, leading to a mixed  
577 sex-determination system where the ancestral and new sex-determining loci are  
578 both segregating. However, they noted that very rare recombination events around  
579 the ancestral sex-determining region can allow these heterogametic transitions to  
580 complete. Degeneration around the Y or W could explain why heterogametic transi-  
581 tions are not observed to be much more common than homogametic transitions

despite the fact that our models demonstrate that they are favoured under a wider range of conditions. For example, Vicoso and Bachtrog (2015) found a dozen sex chromosome configurations among Dipteran species but only one transition between male and female heterogamety.

Another simplification that we made is that meiotic drive involves only a single locus with two alleles. However, many meiotic drive systems involve an interaction with another locus at which alleles may ‘suppress’ the action of meiotic drive (Burt and Trivers 2006, Lindholm et al. 2016) Taylor,1999. Thus, the dynamics of meiotic drive alleles can be heavily dependent on the interaction between two loci and the recombination rate between them, which in turn can be affected by sex-linkage if there is reduced recombination between sex chromosomes (Hurst and Pomiankowski 1991). Furthermore, in some cases, a driving allele may act by killing any gametes that carry a ‘target’ allele at another locus, in which case there can be fertility effects which can affect the equilibrium frequency of a meiotic drive allele (Holman et al. 2015). In polygamous mating systems, the intensity of pollen/sperm competition can depend on the density of males available to donate pollen/sperm, which can itself depend on the sex ratio (Taylor and Jaenike 2002). In terms of our model, this implies that the strength of gametic competition ( $t^\delta$ ) may both determine and be determined by the sex ratio. How the evolution of new sex-determining mechanisms could be influenced by two-locus meiotic drive and/or by ecological feedbacks under different mating systems remains to be studied.

We have shown that tight sex-linkage and haploid selection can drive previously unexpected transitions between sex-determination systems. In particular, both can select for neo-sex-determining loci that are more loosely linked. In addition, haploid selection alone can cause transitions analogous to those caused by purely sexually-antagonistic selection, eliminating the need for differences in selection between male and female diploids. Perhaps counterintuitively, transitions involving haploid selection can be driven by sex-ratio selection or cause sex-ratio biases to evolve. We therefore argue that haploid selection should be considered

612 as a pivotal factor in the evolution of sex-determination systems. Overall, our re-  
613 sults suggest several new scenarios under which new sex-determination systems  
614 are favoured, which could help to explain why the evolution of sex-determination  
systems is so dynamic.

616 Discuss polymorphic mating systems somewhere? Say that haploid selection  
makes this particularly likely (I think there are examples with gametic competition  
618 and weak selection, whereas the vD&K, 2010 results suggest that it's not possible  
with weak selection and diploid selection alone)? This might be best as a section  
620 added to the appendix. When giving an example of polymorphic, make sure it's  
not just that variation was lost at the A locus. The following examples copied and  
622 pasted from from Vuilleumier et al. 2007 and vD&K, 2010, might be added to this  
section.

624 “Polygenic sex determination has been reported in many plants (e.g. Shannon  
& Holsinger 2007), fishes (Vandeputte et al. 2007; Ser et al. 2010; Liew et al.  
626 2012), crustaceans (e.g. Battaglia 1958; Battaglia & Malesani 1959; Voordouw  
& Anholt 2002), bivalves (Haley 1977; Saavedra et al. 1997), gastropods (Yusa  
628 2007a,b), and polychaetes (Bacci 1965, 1978; Premoli et al. 1996).” From Vuilleu-  
mier et al. 2007: “Polymorphism for sex-determining genes within or among pop-  
630ulations has been reported in many species including houseflies, midges, woodlice,  
platyfish, cichlid fish, and frogs (Gordon, 1944; Kallman, 1970; Thompson, 1971;  
632 Macdonald, 1978; Bull, 1983; Rigaud et al., 1997; Caubet et al., 2000; Lande et  
al., 2001; Ogata et al., 2003; Lee et al., 2004; Mank et al., 2006).” Also check  
634 Kallman (1984) -from vD&K, 2010.

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# Appendix

## 822 Recursion Equations

In each generation we census the genotype frequencies in male and female ga-  
824 metes/gametophytes (hereafter, gametes) between meiosis (and any meiotic drive)  
and gametic competition. At this stage we denote the frequencies of X- and Y-  
826 bearing gametes from males and females  $x_i^\delta$  and  $y_i^\delta$ , where  $\delta \in \{\delta, \Omega\}$  specifies  
the sex of the diploid that the gamete came from.  $i \in \{1, 2, 3, 4\}$  specifies the  
828 genotype at the selected locus **A** and at the novel sex-determining locus **M** where  
 $1 = MA$ ,  $2 = Ma$ ,  $3 = mA$ , and  $4 = ma$ . The gamete frequencies from each sex  
830 sum to one,  $\sum_i x_i^\delta + y_i^\delta = 1$ .

Competition then occurs among gametes of the same sex (e.g., among eggs  
832 and among sperm separately) according to the genotype at the **A** locus ( $w_1^\delta = w_3^\delta = w_A^\delta$ ,  $w_2^\delta = w_4^\delta = w_a^\delta$ , see Table 1). The genotype frequencies after gametic  
834 competition are  $x_i^{\delta,s} = w_i x_i^\delta / \bar{w}_H^\delta$  and  $y_i^{\delta,s} = w_i y_i^\delta / \bar{w}_H^\delta$ , where  $\bar{w}_H^\delta = \sum_i w_i x_i^\delta +$   
 $w_i y_i^\delta$  is the mean fitness of male ( $\delta = \delta$ ) or female ( $\delta = \Omega$ ) gametes.

836 Random mating then occurs between gametes to produce diploid zygotes. The  
frequencies of  $XX$  zygotes are then denoted as  $xx_{ij}$ ,  $XY$  zygotes as  $xy_{ij}$ , and  $YY$   
838 zygotes as  $yy_{ij}$ , where **A** and **M** locus genotypes are given by  $i, j \in \{1, 2, 3, 4\}$ , as  
above. In  $XY$  zygotes, the haplotype inherited from an X-bearing gamete is given  
840 by  $i$  and the haplotype from a Y-bearing gamete is given by  $j$ . In  $XX$  and  $YY$   
zygotes, individuals with diploid genotype  $ij$  are equivalent to those with diploid  
842 genotype  $ji$ ; for simplicity, we use  $xx_{ij}$  and  $yy_{ij}$  with  $i \neq j$  to denote the average of  
these frequencies,  $xx_{ij} = (x_i^{\Omega,s} x_j^{\delta,s} + x_j^{\Omega,s} x_i^{\delta,s})/2$  and  $yy_{ij} = (y_i^{\Omega,s} y_j^{\delta,s} + y_j^{\Omega,s} y_i^{\delta,s})/2$ .

844 Denoting the **M** locus genotype by  $b \in \{MM, Mm, mm\}$  and the **X** locus  
genotype by  $c \in \{XX, XY, YY\}$ , zygotes develop as females with probability  
846  $k_{bc}$ . Therefore, the frequencies of  $XX$  females are given by  $xx_{ij}^\Omega = k_{bc} xx_{ij}$ ,  $XY$   
females are given by  $xy_{ij}^\Omega = k_{bc} xy_{ij}$ , and  $YY$  females are given by  $yy_{ij}^\Omega = k_{bc} yy_{ij}$ .  
848 Similarly,  $XX$  male frequencies are  $xx_{ij}^\delta = (1 - k_{bc}) xx_{ij}$ ,  $XY$  male frequencies  
are  $xy_{ij}^\delta = (1 - k_{bc}) xy_{ij}$ , and  $YY$  males frequencies are  $yy_{ij}^\delta = (1 - k_{bc}) yy_{ij}$ . This

850 notation allows both the ancestral and novel sex-determining regions to determine  
 851 zygotic sex according to an  $XY$  system, a  $ZW$  system, or an environmental sex-  
 852 determining system. In addition, we can consider any epistatic dominance rela-  
 853 tionship between the two sex-determining loci. Here, we assume that the ancestral  
 854 sex-determining system (**X** locus) is  $XY$  ( $k_{MMXX} = 1$  and  $k_{MMXY} = k_{MMYY} = 0$ )  
 855 or  $ZW$  ( $k_{MMZZ} = 0$  and  $k_{MMZW} = k_{MMWW} = 1$ ) and epistically recessive to  
 856 a dominant novel sex-determining locus, **M** ( $k_{Mmc} = k_{mmc} = k$ ).

Selection among diploids then occurs according to the diploid genotype at the  
 857 **A** locus,  $l \in \{AA, Aa, aa\}$ , for an individual of type  $ij$  (see Table 1). The diploid  
 858 frequencies after selection in sex  $\hat{\varphi}$  are given by  $xx_{ij}^{\hat{\varphi},s} = w_l^{\hat{\varphi}} xx_{ij}/\bar{w}^{\hat{\varphi}}$ ,  $xy_{ij}^{\hat{\varphi},s} =$   
 859  $w_l^{\hat{\varphi}} xy_{ij}/\bar{w}^{\hat{\varphi}}$ , and  $yy_{ij}^{\hat{\varphi},s} = w_l^{\hat{\varphi}} yy_{ij}/\bar{w}^{\hat{\varphi}}$ , where  $\bar{w}^{\hat{\varphi}} = \sum_{i=1}^4 \sum_{j=1}^4 w_l^{\hat{\varphi}} xx_{ij} + w_l^{\hat{\varphi}} xy_{ij} +$   
 860  $w_l^{\hat{\varphi}} yy_{ij}$  is the mean fitness of individuals of sex  $\hat{\varphi}$ .

861 Finally, these diploids undergo meiosis to produce the next generation of ga-  
 862 metes. Recombination and sex-specific meiotic drive occur during meiosis. Here,  
 863 we allow any relative locations for the SDR, **A**, and **M** loci by using three param-  
 864 eters to describe the recombination rates between them.  $R$  is the recombination  
 865 rate between the **A** locus and the **M** locus,  $\chi$  is the recombination rate between the  
 866 **M** locus and the **X** locus, and  $r$  is the recombination rate between the **A** locus and  
 867 the **X** locus. Table S.1 shows the value of  $\chi$  in the absence of cross-over inter-  
 868 ference for each possible ordering of the loci. During meiosis in sex  $\hat{\varphi}$ , meiotic drive  
 869 occurs such that, in  $Aa$  heterozygotes, a fraction  $\alpha^{\hat{\varphi}}$  of gametes produced carry the  
 870  $A$  allele and  $(1 - \alpha^{\hat{\varphi}})$  carry the  $a$  allele.

Table S.1:  $\chi$  substitutions for different loci orders (assuming no interference)

Order of loci	
SDR-A-M	$\chi = R(1 - r) + r(1 - R)$
SDR-M-A	$\chi = (r - R)/(1 - 2R)$
A-SDR-M	$\chi = (R - r)/(1 - 2r)$

871 Among gametes from sex  $\hat{\varphi}$ , the frequencies of haplotypes (before gametic

competition) in the next generation are given by

$$\begin{aligned}
x_1^{\phi'} = & xx_{11}^{\phi,s} + xx_{13}^{\phi,s}/2 + (xx_{12}^{\phi,s} + xx_{14}^{\phi,s})\alpha^{\phi} \\
& - R(xx_{14}^{\phi,s} - xx_{23}^{\phi,s})\alpha^{\phi} \\
& + (xy_{11}^{\phi,s} + xy_{13}^{\phi,s})/2 + (xy_{12}^{\phi,s} + xy_{14}^{\phi,s})\alpha^{\phi} \\
& - r(xy_{12}^{\phi,s} - xy_{21}^{\phi,s})\alpha^{\phi} - \chi(xy_{13}^{\phi,s} - xy_{31}^{\phi,s})/2 \\
& + \left\{ -(R+r+\chi)xy_{14}^{\phi,s} + (r+\chi-R)xy_{41}^{\phi,s} \right. \\
& \left. + (R+r-\chi)xy_{23}^{\phi,s} + (R+\chi-r)xy_{32}^{\phi,s} \right\} \alpha^{\phi}/2
\end{aligned} \tag{S.1a}$$

$$\begin{aligned}
x_2^{\phi'} = & xx_{22}^{\phi,s} + xx_{24}^{\phi,s}/2 + (xx_{12}^{\phi,s} + xx_{23}^{\phi,s})\alpha^{\phi} \\
& - R(xx_{23}^{\phi,s} - xx_{14}^{\phi,s})\alpha^{\phi} \\
& (xy_{22}^{\phi,s} + xy_{24}^{\phi,s})/2 + (xy_{21}^{\phi,s} + xy_{23}^{\phi,s})(1 - \alpha^{\phi}) \\
& - r(xy_{21}^{\phi,s} - xy_{12}^{\phi,s})(1 - \alpha^{\phi}) - \chi(xy_{24}^{\phi,s} - xy_{42}^{\phi,s})/2 \\
& + \left\{ -(R+r+\chi)xy_{23}^{\phi,s} + (r+\chi-R)xy_{32}^{\phi,s} \right. \\
& \left. + (R+r-\chi)xy_{14}^{\phi,s} + (R+\chi-r)xy_{41}^{\phi,s} \right\} (1 - \alpha^{\phi})/2
\end{aligned} \tag{S.1b}$$

$$\begin{aligned}
x_3^{\phi'} = & xx_{33}^{\phi,s} + xx_{13}^{\phi,s}/2 + (xx_{23}^{\phi,s} + xx_{34}^{\phi,s})\alpha^{\phi} \\
& - R(xx_{23}^{\phi,s} - xx_{14}^{\phi,s})\alpha^{\phi} \\
& (xy_{33}^{\phi,s} + xy_{31}^{\phi,s})/2 + (xy_{32}^{\phi,s} + xy_{34}^{\phi,s})\alpha^{\phi} \\
& - r(xy_{34}^{\phi,s} - xy_{43}^{\phi,s})\alpha^{\phi} - \chi(xy_{31}^{\phi,s} - xy_{13}^{\phi,s})/2 \\
& + \left\{ -(R+r+\chi)xy_{32}^{\phi,s} + (r+\chi-R)xy_{23}^{\phi,s} \right. \\
& \left. + (R+r-\chi)xy_{41}^{\phi,s} + (R+\chi-r)xy_{14}^{\phi,s} \right\} \alpha^{\phi}/2
\end{aligned} \tag{S.1c}$$

$$\begin{aligned}
x_4^{\phi'} = & xx_{44}^{\phi,s} + xx_{34}^{\phi,s}/2 + (xx_{14}^{\phi,s} + xx_{24}^{\phi,s})\alpha^\phi \\
& - R(xx_{14}^{\phi,s} - xx_{23}^{\phi,s})\alpha^\phi \\
& (xy_{44}^{\phi,s} + xy_{42}^{\phi,s})/2 + (xy_{41}^{\phi,s} + xy_{43}^{\phi,s})(1 - \alpha^\phi) \\
& - r(xy_{43}^{\phi,s} - xy_{34}^{\phi,s})(1 - \alpha^\phi) - \chi(xy_{42}^{\phi,s} - xy_{24}^{\phi,s})/2 \\
& + \left\{ -(R + r + \chi)xy_{41}^{\phi,s} + (r + \chi - R)xy_{14}^{\phi,s} \right. \\
& \left. + (R + r - \chi)xy_{32}^{\phi,s} + (R + \chi - r)xy_{23}^{\phi,s} \right\} (1 - \alpha^\phi)/2
\end{aligned} \tag{S.1d}$$

$$\begin{aligned}
y_1^{\phi'} = & yy_{11}^{\phi,s} + yy_{13}^{\phi,s}/2 + (yy_{12}^{\phi,s} + yy_{14}^{\phi,s})\alpha^\phi \\
& - R(yy_{14}^{\phi,s} - yy_{23}^{\phi,s})\alpha^\phi \\
& (xy_{11}^{\phi,s} + xy_{31}^{\phi,s})/2 + (xy_{21}^{\phi,s} + xy_{41}^{\phi,s})\alpha^\phi \\
& - r(xy_{21}^{\phi,s} - xy_{12}^{\phi,s})\alpha^\phi - \chi(xy_{31}^{\phi,s} - xy_{13}^{\phi,s})/2 \\
& + \left\{ -(R + r + \chi)xy_{41}^{\phi,s} + (r + \chi - R)xy_{14}^{\phi,s} \right. \\
& \left. + (R + r - \chi)xy_{32}^{\phi,s} + (R + \chi - r)xy_{23}^{\phi,s} \right\} \alpha^\phi/2
\end{aligned} \tag{S.1e}$$

$$\begin{aligned}
y_2^{\phi'} = & yy_{22}^{\phi,s} + yy_{24}^{\phi,s}/2 + (yy_{12}^{\phi,s} + yy_{23}^{\phi,s})\alpha^\phi \\
& - R(yy_{23}^{\phi,s} - yy_{14}^{\phi,s})\alpha^\phi \\
& (xy_{22}^{\phi,s} + xy_{42}^{\phi,s})/2 + (xy_{12}^{\phi,s} + xy_{32}^{\phi,s})(1 - \alpha^\phi) \\
& - r(xy_{12}^{\phi,s} - xy_{21}^{\phi,s})(1 - \alpha^\phi) - \chi(xy_{42}^{\phi,s} - xy_{24}^{\phi,s})/2 \\
& + \left\{ -(R + r + \chi)xy_{32}^{\phi,s} + (r + \chi - R)xy_{23}^{\phi,s} \right. \\
& \left. + (R + r - \chi)xy_{41}^{\phi,s} + (R + \chi - r)xy_{14}^{\phi,s} \right\} (1 - \alpha^\phi)/2
\end{aligned} \tag{S.1f}$$

$$\begin{aligned}
y_3^{\phi'} = & yy_{33}^{\phi,s} + yy_{13}^{\phi,s}/2 + (yy_{23}^{\phi,s} + yy_{34}^{\phi,s})\alpha^\phi \\
& - R(yy_{23}^{\phi,s} - yy_{14}^{\phi,s})\alpha^\phi \\
& (xy_{33}^{\phi,s} + xy_{13}^{\phi,s})/2 + (xy_{23}^{\phi,s} + xy_{43}^{\phi,s})\alpha^\phi \\
& - r(xy_{43}^{\phi,s} - xy_{34}^{\phi,s})\alpha^\phi - \chi(xy_{13}^{\phi,s} - xy_{31}^{\phi,s})/2 \\
& + \left\{ -(R + r + \chi)xy_{23}^{\phi,s} + (r + \chi - R)xy_{32}^{\phi,s} \right. \\
& \left. + (R + r - \chi)xy_{14}^{\phi,s} + (R + \chi - r)xy_{41}^{\phi,s} \right\} \alpha^\phi/2
\end{aligned} \tag{S.1g}$$

$$\begin{aligned}
y_4^{\delta'} = & yy_{44}^{\delta,s} + yy_{34}^{\delta,s}/2 + (yy_{14}^{\delta,s} + yy_{24}^{\delta,s})\alpha^{\delta} \\
& - R(yy_{14}^{\delta,s} - yy_{23}^{\delta,s})\alpha^{\delta} \\
& (xy_{44}^{\delta,s} + xy_{24}^{\delta,s})/2 + (xy_{14}^{\delta,s} + xy_{34}^{\delta,s})(1 - \alpha^{\delta}) \\
& - r(xy_{34}^{\delta,s} - xy_{43}^{\delta,s})(1 - \alpha^{\delta}) - \chi(xy_{24}^{\delta,s} - xy_{42}^{\delta,s})/2 \\
& + \left\{ -(R + r + \chi)xy_{14}^{\delta,s} + (r + \chi - R)xy_{41}^{\delta,s} \right. \\
& \left. + (R + r - \chi)xy_{23}^{\delta,s} + (R + \chi - r)xy_{32}^{\delta,s} \right\}(1 - \alpha^{\delta})/2
\end{aligned} \tag{S.1h}$$

874

The full system is therefore described by 16 recurrence equations (three diallelic loci in two sexes,  $2^3 \times 2 = 16$ ). However, not all diploid types are produced under certain sex-determination systems. For example, with the *M* allele fixed and an ancestral *XY* sex determination, there are *XX* males, *XY* females, or *YY* females ( $x_3^{\delta} = x_4^{\delta} = y_4^{\delta} = y_3^{\delta} = y_i^{\delta} = 0$ ). In this case, the system only involves six recursion equations, which we assume below to calculate the equilibria.

## Resident equilibrium and stability

In the resident population (allele *M* fixed), we follow the frequency of *A* in X-bearing female gametes,  $p_X^{\varphi}$ , and X-bearing male gametes,  $p_X^{\delta}$ , and Y-bearing male gametes,  $p_Y^{\delta}$ . We also track the total frequency of Y among male gametes,  $q$ , which may deviate from 1/2 due to meiotic drive in males. These four variables determine the frequencies of the six resident gamete types:  $x_1^{\varphi} = p_X^{\varphi}$ ,  $x_2^{\varphi} = 1 - p_X^{\varphi}$ ,  $x_1^{\delta} = (1 - q)p_X^{\delta}$ ,  $x_2^{\delta} = (1 - q)(1 - p_X^{\delta})$ ,  $y_1^{\delta} = qp_Y^{\delta}$ , and  $y_2^{\delta} = q(1 - p_Y^{\delta})$ . Mean fitnesses in the resident population are given in table S.2.

Various forms of selection can maintain a polymorphism at the **A** locus, including sexually antagonistic selection, overdominance, conflicts between diploid selection and selection upon haploid genotypes (ploidally antagonistic selection, Immler et al. 2012), or a combination of these selective regimes.

In particular special cases, e.g., no sex-differences in selection or meiotic drive ( $s^{\delta} = s^{\varphi}$ ,  $h^{\delta} = h^{\varphi}$ , and  $\alpha^{\delta} = \alpha^{\varphi} = 1/2$ ), the equilibrium allele frequency and sta-

Table S.2: mean fitnesses in the resident population ( $M$  fixed, XY sex determination)

Sex & Life Cycle Stage	Mean Fitness
female gametes ( $\bar{w}_H^q$ )	$p_X^q w_A^q + (1 - p_X^q) w_a^q$
male gametes ( $\bar{w}_H^\delta$ )	$\bar{p}^\delta w_A^\delta + (1 - \bar{p}^\delta) w_a^\delta$
females ( $\bar{w}^q$ )	$\{p_X^q w_A^q p_X^\delta w_A^\delta w_{Aa}^q + (1 - p_X^q) w_a^q p_X^\delta w_A^\delta w_{Aa}^q + p_X^q w_A^q (1 - p_X^\delta) w_a^\delta w_{Aa}^q + (1 - p_X^q) w_a^q (1 - p_X^\delta) w_a^\delta w_{aa}^q\} / \{\bar{w}_H^q \bar{w}_H^\delta \zeta\}$
males ( $\bar{w}^\delta$ )	$\{p_X^q w_A^q p_Y^\delta w_A^\delta w_{Aa}^\delta + (1 - p_X^q) w_a^q p_Y^\delta w_A^\delta w_{Aa}^\delta + p_X^q w_A^q (1 - p_Y^\delta) w_a^\delta w_{Aa}^\delta + (1 - p_X^q) w_a^q (1 - p_Y^\delta) w_a^\delta w_{aa}^\delta\} / \{\bar{w}_H^q \bar{w}_H^\delta (1 - \zeta)\}$
zygotic sex ratio $\zeta$	$\{(1 - q)(p_X^\delta w_A^\delta + (1 - p_X^\delta) w_a^\delta)\} / \bar{w}_H^\delta$

ability can be calculated analytically without assuming anything about the relative  
 896 strengths of selection and recombination. However, here, we focus on two regimes  
 (tight linkage and weak selection) in order to make fewer assumptions about fit-  
 898 nesses.

### Recombination weak relative to selection (tight linkage between A and X)

900 We first calculate the equilibrium frequency of the Y and A alleles in the ances-  
 901 tral population when the recombination rate between the X and A loci is small  
 902 ( $r$  of order  $\epsilon$ ). Selection at the A locus will not affect evolution at the novel sex-  
 903 determining locus, M, if one allele is fixed on all backgrounds. We therefore focus  
 904 on the five equilibria that maintain both A and a alleles, four of which are given to  
 leading order by:

$$(A) \quad \hat{p}_Y^\delta = 0, \quad \hat{q} = \frac{1}{2} - \frac{(\alpha^\delta - 1/2)w_{Aa}^\delta \phi}{w_{Aa}^\delta \phi + w_{aa}^\delta \psi}, \quad (\text{S.2a})$$

$$\hat{p}_X^\delta = \frac{w_a^\delta \phi}{w_a^\delta \phi + w_A^\delta \psi}, \quad \hat{p}_X^\delta = \frac{2\alpha^\delta w_{Aa}^\delta \phi}{2\alpha^\delta w_{Aa}^\delta \phi + w_{AA}^\delta \psi}$$

$$(A') \quad \hat{p}_Y^\delta = 1, \quad \hat{q} = \frac{1}{2} + \frac{(\alpha^\delta - 1/2)w_{Aa}^\delta \phi'}{w_{Aa}^\delta \phi' + w_{AA}^\delta \psi'}, \quad (\text{S.2b})$$

$$\hat{p}_X^\delta = 1 - \frac{w_A^\delta \phi'}{w_A^\delta \phi' + w_a^\delta \psi'}, \quad \hat{p}_X^\delta = 1 - \frac{2(1 - \alpha^\delta)w_{Aa}^\delta \phi'}{2(1 - \alpha^\delta)w_{Aa}^\delta \phi' + w_{aa}^\delta \psi'}$$

$$(B) \quad \hat{p}_Y^\delta = 0, \quad \hat{p}_X^\delta = 1, \quad \hat{p}_X^\delta = 1, \quad \hat{q} = 1 - \alpha^\delta \quad (\text{S.2c})$$

$$(B') \quad \hat{p}_Y^\delta = 1, \quad \hat{p}_X^\delta = 0, \quad \hat{p}_X^\delta = 0, \quad \hat{q} = \alpha^\delta \quad (\text{S.2d})$$

$$\begin{aligned} \phi &= \alpha^\delta w_A^\delta w_{Aa}^\delta (w_a^\delta w_{aa}^\delta + 2\alpha^\delta w_A^\delta w_{Aa}^\delta) - w_a^\delta w_A^\delta w_{aa}^\delta w_{aa}^\delta \\ \psi &= (1 - \alpha^\delta) w_a^\delta w_{Aa}^\delta (w_a^\delta w_{aa}^\delta + 2\alpha^\delta w_A^\delta w_{Aa}^\delta) - 2\alpha^\delta w_A^\delta w_A^\delta w_{Aa}^\delta w_{AA}^\delta \\ \phi' &= (1 - \alpha^\delta) w_a^\delta w_{Aa}^\delta (w_A^\delta w_{AA}^\delta + 2(1 - \alpha^\delta) w_a^\delta w_{Aa}^\delta) - w_A^\delta w_A^\delta w_{AA}^\delta w_{AA}^\delta \\ \psi' &= \alpha^\delta w_A^\delta w_{Aa}^\delta (w_A^\delta w_{AA}^\delta + 2(1 - \alpha^\delta) w_a^\delta w_{Aa}^\delta) - 2(1 - \alpha^\delta) w_A^\delta w_A^\delta w_{Aa}^\delta w_{aa}^\delta \end{aligned}$$

906 A fifth equilibrium (*C*) also exists where *A* is present at an intermediate frequency  
 907 on the Y chromosome ( $0 < \hat{p}_Y^\delta < 1$ ). However, equilibrium (*C*) is never locally  
 908 stable when  $r \approx 0$  and is therefore not considered further. Thus, the Y can either  
 910 be fixed for the *a* allele (equilibria *A* and *B*) or the *A* allele (equilibria *A'* and  
 911 *B'*). The X chromosome can then either be polymorphic (equilibria *A* and *A'*) or  
 912 fixed for the alternative allele (equilibria *B* and *B'*). Since equilibria (*A*) and  
 913 (*B*) are equivalent to equilibria (*A'*) and (*B'*) with the labelling of *A* and *a* alleles  
 914 interchanged, we discuss only equilibria (*A*) and (*B*), in which the Y is fixed for the  
 915 *a* allele. If there is no haploid selection ( $\alpha^\delta = 1/2, w_A^\delta = w_a^\delta = 1$ ), these equilibria  
 916 are equivalent to those found by Lloyd and Webb (1977) and Otto (2014).

917 We next calculate when (*A*) and (*B*) are locally stable for  $r = 0$ . According  
 918 to the ‘small parameter theory’ (Karlin and McGregor 1972*a;b*), these stability

properties are unaffected by small amounts of recombination between the SDR and A locus, although equilibrium frequencies may be slightly altered. For the  $a$  allele to be stably fixed on the Y is stable if  $\bar{w}_{Y_a}^\delta > \bar{w}_{YA}^\delta$  where  $\bar{w}_{Y_a}^\delta = w_a^\delta(2p_X^\varphi(1 - \alpha^\delta)w_A^\varphi w_{Aa}^\delta + (1 - p_X^\varphi)w_a^\varphi w_{aa}^\delta)$  and  $\bar{w}_{YA}^\delta = w_A^\delta(p_X^\varphi w_A^\varphi w_{AA}^\delta + 2(1 - p_X^\varphi)\alpha^\delta w_a^\varphi w_{Aa}^\delta)$ . That is,  $Ya$  haplotypes must have higher fitness than  $YA$  haplotypes. Substituting in  $p_X^\varphi = \hat{p}_X^\varphi$  from above, fixation of the  $a$  allele on the Y requires that  $\gamma_i > 0$  where  $\gamma_{(A)} = w_a^\delta(2(1 - \alpha^\delta)w_{Aa}^\delta\phi + w_{aa}^\delta\psi) - w_A^\delta(w_{AA}^\delta\phi + 2\alpha^\delta w_{Aa}^\delta\psi)$  for equilibrium (A) and  $\gamma_{(B)} = 2(1 - \alpha^\delta)w_a^\delta w_{Aa}^\delta - w_A^\delta w_{AA}^\delta$  for equilibrium (B). Stability of a polymorphism on the X chromosome (equilibrium A) further requires that  $\phi > 0$  and  $\psi > 0$ . Fixation of the  $a$  allele on the X (equilibrium B) can be stable only if equilibrium (A) is not and requires  $\psi < 0$  and  $w_A^\varphi w_{AA}^\varphi > (1 - \alpha^\varphi)w_a^\varphi w_{Aa}^\varphi$ .

check last condition and the stability condition below are correct

### 930 Selection weak relative to recombination (weak selection)

Here, we assume that selection and meiotic drive are weak relative to recombination ( $s^\varphi, t^\varphi, \alpha_\Delta^\varphi$  of order  $\epsilon$ ). The maintenance of a polymorphism at the A locus then requires that

$$0 < -((1 - h^\varphi)s^\varphi + (1 - h^\delta)s^\delta + t^\varphi + t^\delta + \alpha_\Delta^\varphi + \alpha_\Delta^\delta) \quad (\text{S.3})$$

and     $0 < (h^\varphi s^\varphi + h^\delta s^\delta + t^\varphi + t^\delta + \alpha_\Delta^\varphi + \alpha_\Delta^\delta).$

which indicates that a polymorphism can be maintained by various selective regimes.

Given that a polymorphism is maintained at the A locus by weak selection, the frequencies of A in each type of gamete are the same ( $\hat{p}_X^\varphi = \hat{p}_X^\delta = \hat{p}_Y^\delta = \bar{p}$ ) and given, to leading order, by

$$\bar{p} = \frac{h^\varphi s^\varphi + h^\delta s^\delta + t^\varphi + t^\delta + \alpha_\Delta^\varphi + \alpha_\Delta^\delta}{(2h^\varphi - 1)s^\varphi + (2h^\delta - 1)s^\delta} + O(\epsilon). \quad (\text{S.4})$$

Differences in frequency between gamete types are of  $O(\epsilon)$ :

$$\begin{aligned}
\hat{p}_X^\delta - \hat{p}_X^q &= V_A(D^\delta - D^q + \alpha_\Delta^\delta - \alpha_\Delta^q) + O(\epsilon^2) \\
\hat{p}_Y^\delta - \hat{p}_X^q &= V_A(D^\delta - D^q + \alpha_\Delta^\delta - \alpha_\Delta^q + (1-2r)(t^\delta - t^q))/2r + O(\epsilon^2) \quad (\text{S.5}) \\
\hat{p}_Y^\delta - \hat{p}_X^\delta &= V_A(D^\delta - D^q + \alpha_\Delta^\delta - \alpha_\Delta^q + t^\delta - t^q)(1-2r)/2r + O(\epsilon^2)
\end{aligned}$$

where  $V_A = \bar{p}(1-\bar{p})$  is the variance in the frequency of  $A$  and  $D^\delta = (\bar{p}s^\delta + (1-\bar{p})h^\delta s^\delta) - (\bar{p}h^\delta s^\delta + (1-\bar{p}))$  corresponds to the difference in fitness between  $A$  and  $a$  alleles in diploids of sex  $\delta \in \{\text{♀}, \text{♂}\}$  ( $\bar{p}$  is the leading-order probability of mating with an  $A$ -bearing gamete from the opposite sex). The frequency of  $Y$  among male gametes depends upon the difference in the frequency of the  $A$  allele between  $X$ - and  $Y$ -bearing male gametes and the strength of meiotic drive in favour of the  $A$  allele in males,  $q = 1/2 + \alpha_\Delta^\delta(\hat{p}_Y^\delta - \hat{p}_X^\delta)/2 + O(\epsilon^3)$ . Without gametic competition or drive ( $\alpha_\Delta^\delta = t^\delta = 0$ ) our results reduce to those of van Doorn and Kirkpatrick (2007).

## 948 Invasion conditions

Cover the other parts of the characteristic polynomial here.

950 A rare neo-Y or neo-W will spread from a given ancestral equilibrium when  
the leading eigenvalue,  $\lambda$ , of the Jacobian matrix derived from the eight mutant  
952 recursion equations (given by S.1c,d,g,h), evaluated at the ancestral equilibrium, is  
greater than one. However, because a neo-Y (neo-W) is always in males (females)  
954 and is epistemically dominant to the ancestral sex-determining locus, we need only  
two recursion equations (e.g., tracking the change in the frequency of neo-Y- $A$  and  
956 neo-Y- $a$  gametes from males) and thus the leading eigenvalue is the largest solution  
to a quadratic characteristic polynomial  $\lambda^2 + b\lambda + c = 0$  as described in the text  
958 (Table 2).

For tight linkage between the ancestral sex-determining locus and the selected  
960 locus we can calculate each of these terms exactly, while for weak selection we  
take a Taylor series of the leading eigenvalue. With weak selection, the leading

<sup>962</sup> eigenvalue,  $\lambda$ , for any  $k$ , is given up to order  $\epsilon^2$  by equation (4).

### Tight linkage between A and X (recombination weak relative to selection)

<sup>964</sup> Here, we explore the conditions under which a neo-W invades an XY system as-  
 suming that the A locus is initially in tight linkage with the ancestral sex-determining  
<sup>966</sup> region ( $r \approx 0$ ). We disregard neo-Y mutations, which never spread given that the  
 ancestral population is at a stable equilibrium (see supplementary *Mathematica*  
<sup>968</sup> notebook for proof).

<sup>970</sup> Starting with the simpler equilibrium ( $B$ ), the terms of the characteristic poly-  
 nomial are

$$\lambda_{mA} = (2\alpha^\delta w_A^\delta)^{-1} \frac{w_A^\varphi}{w_A^\delta} \frac{[w_A^\delta \alpha^\delta w_{AA}^\varphi + w_a^\delta (1 - \alpha^\delta) w_{Aa}^\varphi 2\alpha^\varphi]}{w_{AA}^\varphi} \quad (\text{S.6a})$$

$$\lambda_{ma} = (2\alpha^\delta w_A^\delta)^{-1} \frac{w_a^\varphi}{w_A^\delta} \frac{[w_A^\delta \alpha^\delta w_{Aa}^\varphi 2(1 - \alpha^\varphi) + w_a^\delta (1 - \alpha^\delta) w_{aa}^\varphi]}{w_{AA}^\varphi} \quad (\text{S.6b})$$

$$\rho_{mA} = (2\alpha^\delta w_A^\delta)^{-1} \frac{w_A^\varphi}{w_A^\delta} \frac{R w_a^\delta (1 - \alpha^\delta) w_{Aa}^\varphi \alpha^\varphi}{w_{AA}^\varphi} \quad (\text{S.6c})$$

$$\rho_{ma} = (2\alpha^\delta w_A^\delta)^{-1} \frac{w_a^\varphi}{w_A^\delta} \frac{R w_A^\delta \alpha^\delta w_{Aa}^\varphi (1 - \alpha^\varphi)}{w_{AA}^\varphi} \quad (\text{S.6d})$$

Haploid selection impacts the spread of neo-W haplotypes in three ways. Firstly,  
<sup>972</sup> the zygotic sex ratio becomes male biased,  $\zeta < 1/2$ , when the  $a$  allele (which is  
 fixed on the Y) is favoured during competition among male gametes or by meiotic  
<sup>974</sup> drive in males. Specifically, at equilibrium ( $B$ ), the sex ratio is  $\zeta = \alpha^\delta w_A^\delta / \bar{w}_H^\delta$   
 where  $\bar{w}_H^\delta = [(1 - \alpha^\delta) w_a^\delta + \alpha^\delta w_A^\delta]$  has been canceled in equations (S.6) to leave the  
<sup>976</sup> term  $(2\alpha^\delta w_A^\delta)^{-1}$ . Male biased sex ratios facilitate the spread of a neo-W because  
 neo-W alleles cause the zygotes that carry them to develop as the rarer, female,  
<sup>978</sup> sex.

Secondly, haploid selection in females selects on neo-W haplotypes directly. At

980 equilibrium (*B*), the fitness of female gametes under the ancestral sex-determining  
 981 system is  $w_A^{\varphi}$  such that the relative fitnesses of neo-W-*A* and neo-W-*a* haplotypes  
 982 during female gametic competition are  $w_A^{\varphi}/w_A^{\varphi}$  and  $w_a^{\varphi}/w_A^{\varphi}$ , see terms in equation  
 983 (S.6). Meiotic drive in females will also change the proportion of gametes that  
 984 carry the *A* versus *a* alleles, which will be produced by heterozygous females in  
 985 proportions  $(1 + \alpha_{\Delta}^{\varphi})$  and  $(1 - \alpha_{\Delta}^{\varphi})$ , respectively. These terms are only associated  
 986 with heterozygous females, i.e., they are found alongside  $w_{Aa}^{\varphi}$ .

Thirdly, haploid selection in males affects the diploid genotypes of females by  
 988 altering the allele frequencies in the male gametes that female gametes pair with.  
 At equilibrium (*B*), neo-W female gametes will mate with X-*A* male gametes with  
 990 probability  $\alpha^{\delta} w_A^{\delta}/\bar{w}_H^{\delta}$  and Y-*a* male gametes with probability  $(1 - \alpha^{\delta})w_a^{\delta}/\bar{w}_H^{\delta}$ ,  
 where the  $\bar{w}_H^{\delta}$  terms have been canceled in (S.6). Thus, for example, neo-W-*A*  
 992 haplotypes are found in *AA* female diploids with probability  $\alpha^{\delta} w_A^{\delta}/\bar{w}_H^{\delta}$  (first term  
 in square brackets in equation S.6a) and in *Aa* female diploids with probability  
 994  $(1 - \alpha^{\delta})w_a^{\delta}/\bar{w}_H^{\delta}$  (see equation S.6c and second term in square brackets in equation  
 S.6a).

996 The other terms in equations (S.6) are more easily interpreted if we assume  
 that there is no haploid selection in either sex, in which case  $\lambda_{mA} > 1$  when  $w_{Aa}^{\varphi} >$   
 998  $w_{AA}^{\varphi}$  and  $\lambda_{ma} > 1$  when  $(w_{Aa}^{\varphi} + w_{aa}^{\varphi})/2 > w_{AA}^{\varphi}$ . These conditions cannot be met  
 under purely sexually-antagonistic selection, where *a* is directionally favoured in  
 1000 males ( $w_{AA}^{\delta} > w_{Aa}^{\delta} > w_{aa}^{\delta}$ ) and *A* is directionally favoured in females ( $w_{AA}^{\varphi} >$   
 $w_{Aa}^{\varphi} > w_{aa}^{\varphi}$ ). Essentially, the X is already as specialized as possible for the female  
 1002 beneficial allele (XA is fixed), and the neo-W often makes daughters with the Y-*a*  
 haplotype, increasing the flow of *a* alleles into females, which reduces the fitness  
 1004 of those females.

If selection doesn't uniformly favour *A* in females, however, neo-W-*A* haplo-  
 1006 types and/or neo-W-*a* haplotypes can spread ( $\lambda_{mA} > 1$  and/or  $\lambda_{ma} > 1$ ) at this  
 equilibrium. A neo-W can spread alongside the *A* allele ( $\lambda_{mA} > 1$ ), despite the  
 1008 fact that a neo-W brings Ya haplotypes into females, when  $w_{Aa}^{\varphi} > w_{AA}^{\varphi}$ . In this  
 case the *a* allele is favoured by selection in females despite *A* being fixed on the

1010 X. For this equilibrium to be stable, X-A must be sufficiently favoured in males  
 1011 to keep the frequency of XA at one (specifically, from the stability conditions, we  
 1012 must have  $w_{Aa}^\delta / ((w_{aa}^\delta + w_{Aa}^\delta)/2) > w_{Aa}^\varphi / w_{AA}^\varphi$ ).

Under this same condition,  $w_{Aa}^\varphi > w_{AA}^\varphi$ , the neo-W can also spread alongside  
 1014 the  $a$  allele ( $\lambda_{ma} > 1$ ) if there is sufficiently strong underdominance in females  
 1015 ( $w_{aa}^\varphi > w_{Aa}^\varphi$ ), such that  $(w_{Aa}^\varphi + w_{aa}^\varphi)/2 > w_{AA}^\varphi$ . In this case,  $a$  is not favored in  
 1016 females near the equilibrium where females are AA (comparing  $Aa$  to  $AA$  geno-  
 1017 types) and yet the neo-W can spread with  $a$  because it produces female  $aa$  individ-  
 1018 uals by capturing Y- $a$  haplotypes.

Similar equations can be derived for equilibrium (A) by substituting the equi-  
 1020 librium allele frequencies into Table 2.

$$\lambda_{mA} = \frac{a}{b} \left[ w_{AA}^\varphi w_{Aa}^\delta w_A^\delta \alpha^\delta \phi + 2w_{Aa}^\varphi \alpha^\varphi w_a^\delta \frac{c}{d} \right] / w_a^\varphi \quad (\text{S.7a})$$

$$\lambda_{ma} = \frac{a}{b} \left[ 2w_{Aa}^\varphi (1 - \alpha^\varphi) w_{Aa}^\delta w_A^\delta \alpha^\delta \phi + w_{aa}^\varphi w_a^\delta \frac{c}{d} \right] / w_A^\varphi \quad (\text{S.7b})$$

$$\rho_{mA} = \frac{a}{b} R \left[ 2w_{Aa}^\varphi \alpha^\varphi w_a^\delta \frac{c}{d} \right] / w_a^\varphi \quad (\text{S.7c})$$

$$\rho_{ma} = \frac{a}{b} R \left[ 2w_{Aa}^\varphi (1 - \alpha^\varphi) w_{Aa}^\delta w_A^\delta \alpha^\delta \phi \right] / w_A^\varphi \quad (\text{S.7d})$$

where

$$a = w_a^\varphi \phi + w_A^\varphi \psi \quad (\text{S.8a})$$

$$b = w_{AA}^\varphi \phi (2w_{Aa}^\delta w_A^\delta \alpha^\delta \phi) + w_{Aa}^\varphi \psi (2w_{Aa}^\delta w_A^\delta \alpha^\delta \phi + w_{AA}^\delta w_a^\delta \psi) + w_{aa}^\varphi \psi (w_{AA}^\delta w_a^\delta \psi) \quad (\text{S.8b})$$

$$c = 2(w_{Aa}^\delta \phi)^2 (1 - \alpha^\delta) \alpha_\delta + w_{Aa}^\delta \phi (w_{AA}^\delta \psi + w_{aa}^\delta \psi \alpha^\delta) + w_{aa}^\delta \psi w_{AA}^\delta \psi \quad (\text{S.8c})$$

$$d = 2w_{Aa}^\delta \alpha^\delta \phi + w_{aa}^\delta \psi \quad (\text{S.8d})$$

1022 As with equilibrium (B), haploid selection again modifies invasion fitnesses  
 by altering the sex-ratio and the diploid genotypes of females and directly select-

1024 ing upon female gametes. The only difference is that resident XX females are no  
 1025 longer always homozygote  $AA$  and males are no longer always heterozygote  $Aa$ .  
 1026 Thus the effect of haploid selection in males is reduced, as is the difference in fit-  
 1027 ness between neo-W haplotypes and resident X haplotypes, as both can be on any  
 1028 diploid or haploid background.

The other terms are easier to interpret in the absence of haploid selection. For  
 1030 instance, without haploid selection, the neo-W- $A$  haplotype spreads ( $\lambda_{mA} > 1$ ) if  
 and only if

$$2(w_{Aa}^{\varphi} - w_{aa}^{\varphi})w_{AA}^{\delta}\psi^2 > (w_{AA}^{\varphi} - w_{Aa}^{\varphi})w_{Aa}^{\delta}\phi(\phi - \psi) \quad (\text{S.9})$$

1032 where  $\phi - \psi = w_{AA}^{\varphi}w_{Aa}^{\delta} - w_{aa}^{\varphi}w_{aa}^{\delta}$  and both  $\phi$  and  $\psi$  are positive when equilibrium  
 (A) is stable. In contrast to equilibrium (B), a neo-W haplotype can spread under  
 1034 purely sexually-antagonistic selection ( $w_{AA}^{\delta} < w_{Aa}^{\delta} < w_{aa}^{\delta}$  and  $w_{AA}^{\varphi} > w_{Aa}^{\varphi} > w_{aa}^{\varphi}$ ).  
 The neo-W- $A$  can spread as long as it becomes associated with females that bear  
 1036 more  $A$  alleles than observed at equilibrium (A).

Without haploid selection, the neo-W- $a$  haplotype spreads ( $\lambda_{ma} > 1$ ) if and  
 1038 only if

$$(w_{aa}^{\varphi} + w_{Aa}^{\varphi} - 2w_{AA}^{\varphi})w_{Aa}^{\delta}\phi^2 + (w_{aa}^{\varphi} - w_{Aa}^{\varphi})(w_{Aa}^{\delta} + 2w_{AA}^{\delta})\phi\psi > 0 \quad (\text{S.10})$$

This condition cannot be met with purely sexually antagonistic selection (as both  
 1040 terms on the left-hand side would then be negative), but it can be met under other  
 circumstances. For example, with overdominance in males there is selection for  
 1042 increased  $A$  frequencies on X chromosomes in males, which are always paired  
 with Y- $a$  haplotypes. Directional selection for  $a$  in females can then maintain a  
 1044 polymorphism at the A locus on the X. This scenario selects for a modifier that  
 increases recombination between the sex chromosomes (e.g., blue region of Figure  
 1046 2d in Otto 2014) and facilitates the spread of neo-W- $a$  haplotypes, which create  
 more females bearing more  $a$  alleles than the ancestral X chromosome does.

## 1048 Supplementary Figures

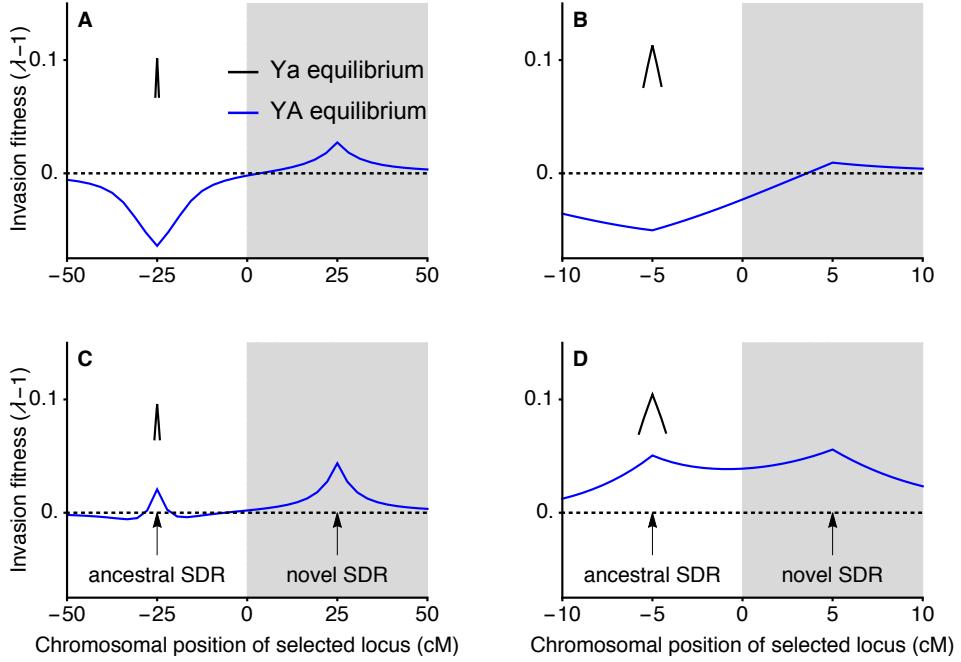


Figure S.1: Neo-W alleles can spread when loci under diploid selection are tightly linked to the ancestral sex determining locus ( $r \approx 0$ ). In panels A and B, the  $a$  allele is favoured in females ( $w_{aa}^{\varphi} = 1.05$ ,  $w_{Aa}^{\varphi} = 1$ ,  $w_{AA}^{\varphi} = 0.85$ ) and selection in males is overdominant ( $w_{aa}^{\delta} = w_{AA}^{\delta} = 0.75$ ). In panels C and D, selection in males and females is overdominant ( $w_{aa}^{\varphi} = w_{AA}^{\varphi} = 0.6$ ,  $w_{aa}^{\delta} = 0.5$ ,  $w_{AA}^{\delta} = 0.7$ ,  $w_{Aa}^{\delta} = 1$ ). These parameters are marked by a dagger in Figure 2, which shows that neo-W invasion is expected for any  $R$  when the  $a$  allele is nearly fixed on the Y (black lines). Equilibria where the  $A$  allele is more common among Y-bearing male gametes can also be stable for these parameters (blue lines). The weak selection approximation holds when all recombination rates are large relative to selection (around 0 in panels A and C), in which case neo-W alleles should spread if they are more tightly linked to the selected locus (positive invasion fitness in the grey region). However, when linkage is tight (panels C and D and when the selected locus is near the SDR), this prediction breaks down. Here, there is no haploid selection  $t^{\varphi} = \alpha_{\Delta}^{\varphi} = 0$ .

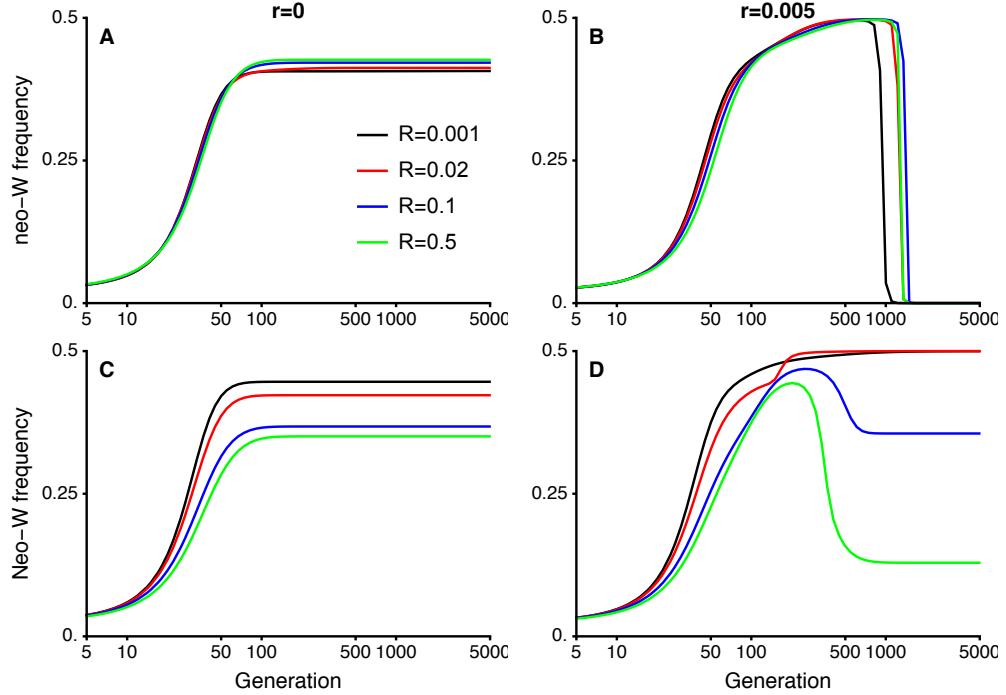


Figure S.2: Following invasion by a neo-W allele, there can be a complete transition to a new sex-determination system, maintenance of polymorphism at both ancestral-XY and neo-ZW sex determining regions, or loss of the new sex-determining allele. Here we plot the frequency of the neo-W allele among female gametes; as the neo-W reaches frequency 0.5, polymorphism at the ancestral XY locus is lost with Y becoming fixed such that sex is determined only by the ZW allele carried by a female gamete. Panels A, C and D show cases where a steady state is reached with the neo-W at a frequency below 0.5, in which case ancestral-X and Y alleles also both segregate. In all cases, we assume that the  $a$  allele is initially more common than the  $A$  allele on the Y ( $Y_a$  is fixed when  $r = 0$ ). When  $r > 0$  (panels B and D),  $YA$  haplotypes created by recombination can become more common than  $Ya$  haplotypes as the neo-W spreads. In B, this leads to loss of the neo-W and the system goes to an equilibrium with  $Xa$  and  $YA$  haplotypes fixed (A'), such that all females have the high fitness genotype  $aa$  and all males  $Aa$ . For the parameters in B, neo-W alleles have negative invasion fitness when the  $YA$  haplotype is ancestrally more common than  $Ya$  (see blue line in Figure S.2A and S.2B). In contrast, the neo-W is not lost in panel D (see blue line in Figure S.2C and S.2D). Fitness parameters are the same as in Figure S.2, the  $a$  allele is favoured in females ( $w_{aa}^q = 1.05$ ,  $w_{Aa}^\delta = 1$ ,  $w_{AA}^\delta = 0.85$ ) and there is overdominant selection in males ( $w_{aa}^\delta = w_{AA}^\delta = 0.75$ ) in panels A and B. In panels C and D, selection in males and females is overdominant ( $w_{aa}^q = w_{AA}^q = 0.6$ ,  $w_{aa}^\delta = 0.5$ ,  $w_{AA}^\delta = 0.7$ ,  $w_{Aa}^\delta = 1$ ). These parameters are marked by a dagger in Figure 2. Here, there is no haploid selection  $t^\delta = \alpha_\Delta^\delta = 0$ .

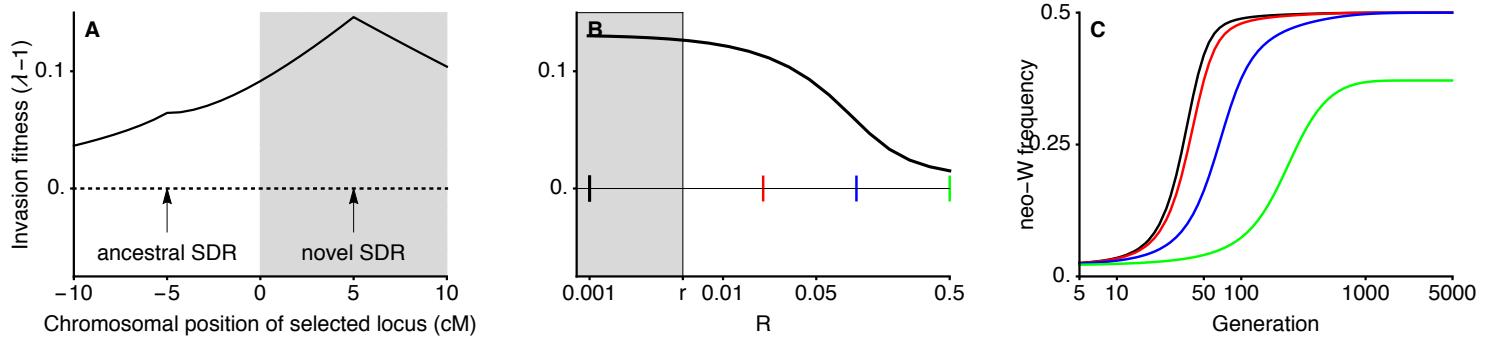


Figure S.3: When there is haploid selection and sexually-antagonistic selection, a neo-W may invade for any  $R$ . Check that we mention Sally's result that invasion cannot occur with sexually-antagonistic selection and  $R = 1/2$  (currently only in legend for figure 1) Add asterisk to Figure S.5B to show parameters used in this plot.  $w_{aa}^{\varphi} = 1.05$ ,  $w_{Aa}^{\delta} = 1$ ,  $w_{AA}^{\varphi} = 0.85$ ,  $w_{aa}^{\delta} = 0.85$ ,  $w_{AA}^{\varphi} = 1.05$ ,  $\alpha_{\Delta}^{\delta} = -0.08$ .

Add Sally's figure showing lambda for small  $r$  near equil A versus near equil B. Add references to this figure to appendix where we discuss whether lambdas can be greater than 1 with sexually antagonistic selection.

Perhaps it would also be useful to add an 8 panel figure that features ploidally antagonistic selection. For each type of haploid selection (gametic competition/meiotic drive in males/females), give a regionplot where  $h^{\delta} = h^{\varphi}$ , e.g.,  $h^{\delta} = h^{\varphi} = 0.75$  (or perhaps the value of  $h$  we use in the regionplots we have, in which  $w_{aa} = 0.85$ ,  $w_{Aa} = 1$ ,  $w_{AA} = 1.05$ ). Matt made a figure like this before but both Ya and YA equilibria were plotted and there was no outline showing where the Ya equilibrium is stable (as in Figure 2). In Matt's plot the axes were  $s^{\delta}$  and  $\alpha_{\Delta}^{\delta}$ . Add an asterisk to each region plot and show invasion in another panel, using those parameters and various  $R$  (e.g., in the style of S.2). In an email, Sally has an example of ploidally-antagonistic selection where the neo-W fixes and  $R = 1/2$ . This would cover that case and more.

We could also give versions of Figure 2 where there is also haploid selection of various types. Haplod selection can favour  $A$  or  $a$ , so this would involve 4x 6-panel figures. Started looking at this in Figure S.4 and Figure S.5, add female

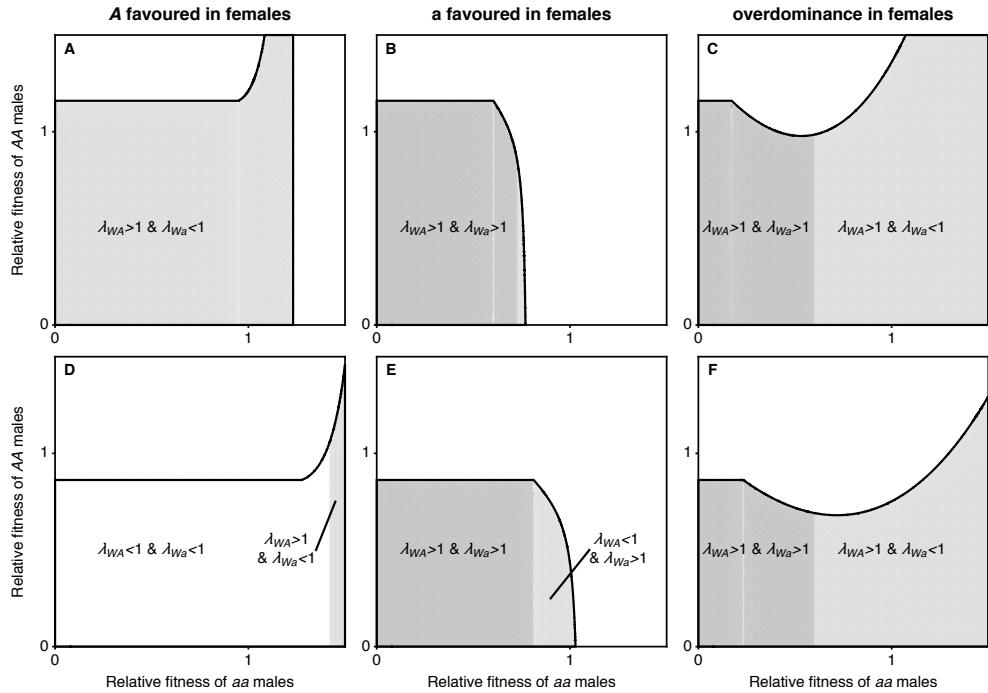


Figure S.4: ABC,  $w_a^\delta = 1.16$ ,  $w_A^\delta = 1$ . DEF,  $w_a^\delta = 1$ ,  $w_A^\delta = 1.16$ . I thought that re-running this with  $w_a^\delta = 1.16$ ,  $w_A^\delta = 0.84$  and  $w_a^\delta = 0.84$ ,  $w_A^\delta = 1.16$  will mean that it matches exactly with the meiotic drive example. Checking for panel A, the result was qualitatively similar (e.g., region where both  $\lambda$ 's are greater than one in panel A, as you might expect from equation (S.6), however the region where a polymorphism is stable is also altered so they don't exactly match.

1066 haploid selection. Try to integrate into the discussion of haploid selection? e.g.,  
 1067 male haploid selection ones generally show effect of sex ratio, increasing both  
 1068 lambdas when female biased (top rows).

1070 Perhaps, for one set of parameters, we should plot the dynamics of all the dif-  
 1071 ferent alleles. E.g., we could use the same parameters used in 4. The main purpose  
 1072 would be to show what happens to the ancestral SDR during turnover. We could  
 1073 also show an example where XY and ZW sex determining systems are both poly-  
 1074 morphic and stable (e.g., using one of the curves in Figure S.2 and the green curve  
 1075 in Figure S.3). I think there are also examples with looser sex linkage and pollen  
 1076 competition that lead to a mixed sex-determination system. We should probably  
 have a short section in the appendix discussing this.

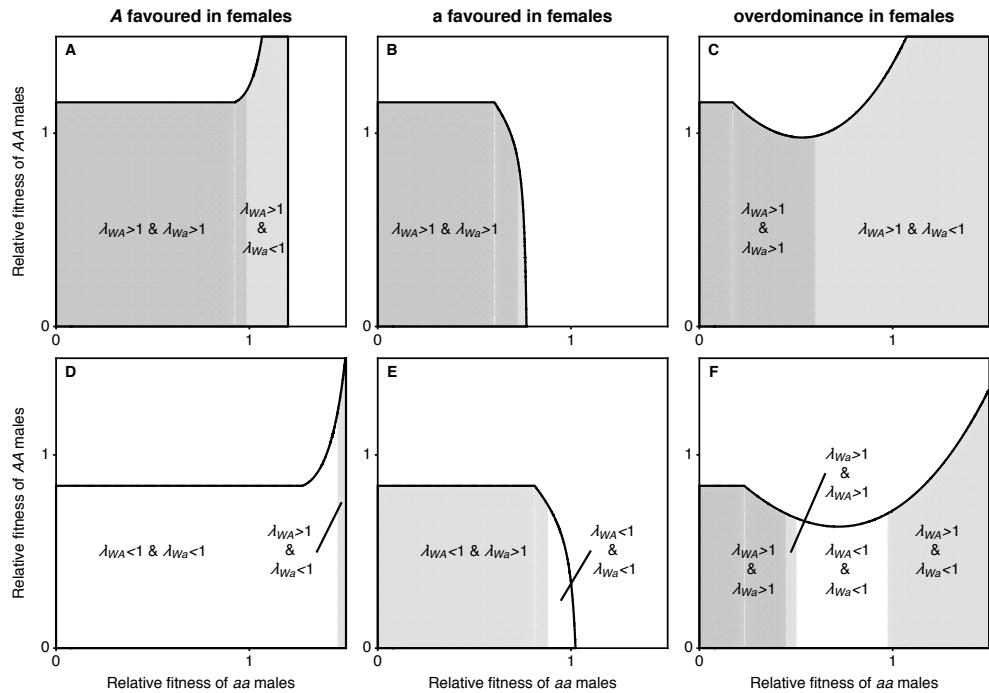


Figure S.5: ABC,  $\alpha_{\Delta}^{\delta} = -0.08$  DEF,  $\alpha_{\Delta}^{\delta} = 0.08$ . Panel F mislabelled, should have  $\lambda_{Wa} > 1$  &  $\lambda_{WA} < 1$  as the upper label that has the line.

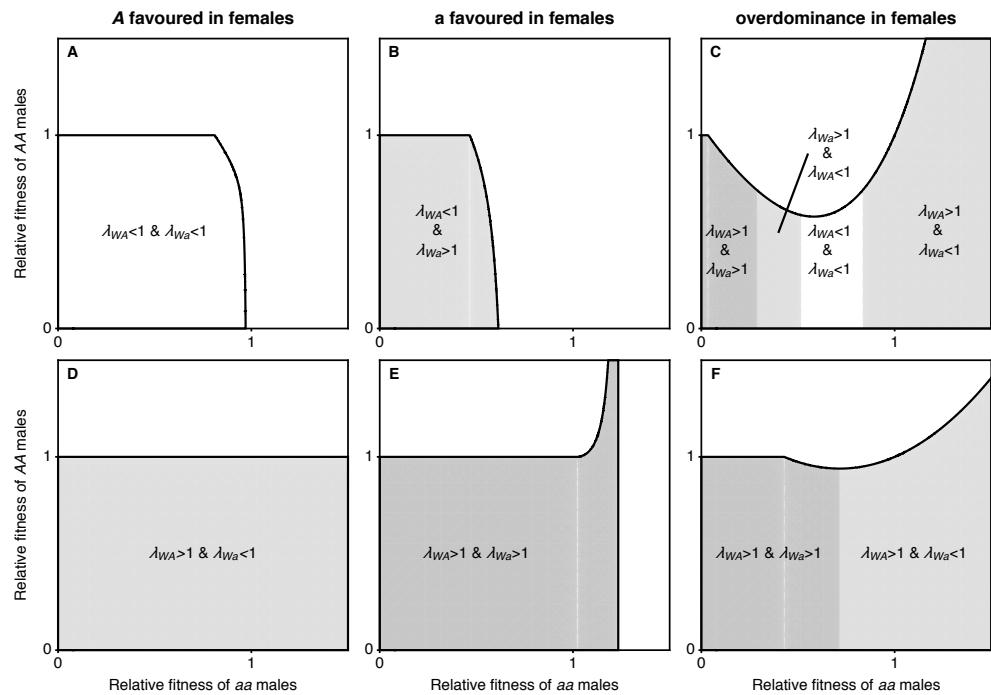


Figure S.6: ABC,  $\alpha_{\Delta}^{\varnothing} = -0.08$  DEF,  $\alpha_{\Delta}^{\varnothing} = 0.08$ .  $\lambda$ s are increased for the haplotype that is favoured by female haploid selection, the stability conditions are also affected.