Gametic Selection, Sex Ratio Bias, and Transitions Between Sex Determination Systems

Michael F Scott*¹ and Matthew M Osmond*², and Sarah P Otto²

^{*} These authors contributed equally to this work

 $^{^1}$ Department of Botany, University of British Columbia, #3529 - 6270 University Boulevard, Vancouver, BC, Canada V6T 1Z4

² Department of Zoology, University of British Columbia, #4200 - 6270 University Boulevard, Vancouver, BC, Canada V6T 1Z4 email: mfscott@biodiversity.ubc.ca, mmosmond@zoology.ubc.ca Contributions:

Abstract

Sex determination systems are remarkably dynamic; many studied taxa display transitions of sex-determining genes between chromosomes or the evolution of new sex-determining systems. Here, we utilize population genetic models to study the spread of novel sexdetermining systems where we also include haploid gametic selection, e.g., pollen or sperm competition. Haploid selected loci experience a form of sex-specific selection (because gametic competition occurs predominantly among haploids produced by males) and can also cause sex ratios at birth to become biased (because sex ratios are determined by the fertilization success of X- versus Y-bearing pollen/sperm). We find that the evolution of sex determination systems where mothers determine sex at birth (e.g., environmental sex determination where sex is determined at birth) is influenced by classic Fisherian sex ratio selection. (Maybe not true???) However, notably, we find that the spread of new genetic sex determination systems is not affected by sex ratio biases that are caused by gametic selection because sex ratios become biased after parental provisioning has occurred (even if pollen/sperm competition occurs within the mother). In addition, we find that linkage of an ancestral sex chromosome to a locus under haploid selection can favour transitions between male and female heterogamety (e.g., XY to ZW), which is not the case for any forms of diploid sex specific selection (e.g., sexually antagonistic selection). During these transitions, new sex-determining alleles spread despite breaking up favourable associations that build up between ancestral sex-determining loci and selected loci, reducing population mean fitness. Furthermore, a period of selection among haploids can favour the stable maintenance of polymorphic sex determination systems. Thus, our models offer several new insights to be explored as information about sex determination in non-model taxa accumulates.

Introduction

Animals and angiosperms exhibit extremely diverse sex determination systems, as reviewed by Bull (1983), Charlesworth and Mank (2010), Beukeboom and Perrin (2014), Bachtrog et al. (2014). Among species with genetic sex determination of diploid sexes, some taxa have heterogametic males (XY) and homogametic females (XX), including mammals and most dioecious plants (Ming et al. 2011); whereas other taxa have homogametic males (ZZ) and heterogametic females (ZW), including Lepidoptera and birds. Within several taxa, the chromosome that harbours the master sex-determining region changes. For example, transitions of the master sex-determining gene between chromosomes or the evolution of new master sex-determining genes have occurred in Salmonids (Li et al. 2011, Yano et al. 2012), Diptera (Vicoso and Bachtrog 2015), and Oryzias (Myosho et al. 2012). In addition, many gonochoric/dioecious clades with genetic sex determination exhibit transitions between male (XY) and female (ZW) heterogamety, including lizards (Ezaz et al. 2009), eight of 26 teleost fish families (Mank et al. 2006), true fruit flies (Tephritids, Vicoso and Bachtrog 2015), amphibians (Hillis and Green 1990), the angiosperm genus Silene (Slancarova et al. 2013), Coleoptera and Hemiptera (Beukeboom and Perrin 2014, plate 2). Indeed, in some cases, both male and female heterogametic sex determination systems can be found the same species, as exhibited by some cichlid species (Ser et al. 2010) and Rana rugosa (Ogata et al. 2007). In addition, multiple transitions have occurred between genetic and environmental sex determination systems, e.g., in reptiles and fishes (Conover and Heins 1987, Mank et al. 2006, Pokorná and Kratochvíl 2009, Ezaz et al. 2009, Pen et al. 2010, Holleley et al. 2015).

Predominant theories in which new sex determination systems are favoured by selection involve fitness differences between sexes (e.g., sexually antagonistic selection) or sex ratio selection. van Doorn and Kirkpatrick (2007; 2010) show that new sex determination loci can be favoured if they arise in close linkage with a locus that experiences sexual antagonism. For example, linkage allows favourable associations to build up between a male-beneficial allele and a neo-Y chromosome. Such associations can favour a new master sex-determining gene on a new chromosome (van Doorn and Kirkpatrick 2007) and can also favour a transition between male and female heterogamety (e.g., a ZW to XY transition, van Doorn and Kirkpatrick 2010). However, any sexually-antagonistic loci that are linked to the ancestral sex-determination locus will develop similar, favourable associations and select against the spread of a new sex-determination system.

It has been suggested that sex ratio selection could be a particularly important force driving transitions between sex-determining systems (Beukeboom and Perrin 2014, Chapter 7). For example, flexible sex determination systems may be favoured in order to exploit local environmental conditions that are optimal for males or females, which creates locally biased sex ratios (Charnov and Bull 1977, Werren and Taylor 1984, Pen et al. 2010). In addition, feminizing mutations may invade when female biased sex ratios are favoured due to selection among demes (Wilson and Colwell 1981, Vuilleumier et al. 2007). In other situations, sex ratio selection may favour transitions in order to restore equal sex ratios. For example, Kozielska et al. (2010) consider systems in which the ancestral sex chromosomes experience meiotic drive (e.g., where driving X or Y chromosomes are inherited disproportionately often), which causes sex ratios to become biased (Hamilton 1967). They find that new, unlinked sex-determining loci (masculinizing or feminizing mutations, i.e., neo-Y or neo-W loci) can then spread, restoring an even sex ratio.

Here, we use mathematical models to find the conditions under which new sex determination systems are favoured by selection where we include a period of selection among haploid gametes/gametophytes. Selection among haploid genotypes is thought to occur primarily among pollen/sperm, which can compete whenever there are more pollen/sperm than required for fertilization (Mulcahy et al. 1996, Joseph and Kirkpatrick 2004). Haploid selection

may be particularly common in plants, in which 60-70% of all genes are expressed in the male gametophyte and these genes exhibit stronger signatures of selection than random genes (Borg et al. 2009, Arunkumar et al. 2013, Gossmann et al. 2014). In addition, artificial selection pressures applied to male gametophytes cause the frequency of resistant alleles to increase (e.g., Hormaza and Herrero 1996, Ravikumar et al. 2003, Hedhly et al. 2004, Clarke et al. 2004). A smaller (but non-negligible) proportion of genes are thought to be expressed and selected in animal sperm, although precise estimates are uncertain (Zheng et al. 2001, Joseph and Kirkpatrick 2004, Vibranovski et al. 2010).

There are various ways in which a period of haploid selection could influence transitions between sex determination systems. Firstly, if we assume that haploid selection at any particular locus predominantly occurs in one sex (e.g., pollen/sperm competition), then such loci experience a form of sex-specific selection. In this respect, we might expect that haploid selection might affect transitions between sex determination systems in a similar manner to sex-specific diploid selection (as explored by van Doorn and Kirkpatrick 2007; 2010). That is, new masculizing mutations (neo-Y chromosomes) could be favoured via linkage associations with alleles that are beneficial in pollen/sperm. However, sex ratios can also become biased if there is linkage between the sex-determining region and a locus that harbours genetic variation in haploid fitness. For example, differences in fitness between X- and Y-bearing pollen tubes can cause the sex ratio among seeds to become biased when there is pollen competition (Lloyd 1974, Conn and Blum 1981, Stehlik and Barrett 2005; 2006, Field et al. 2012; 2013). It is not immediately clear how the spread of new sex determination systems would be influenced by the combination of sex ratio biases and favourable associations between haploid selected loci and sex-determining regions.

Surprisingly, our models show that haploid selection influences the evolution of new sex determination systems in a way that is distinct from both diploid sex-specific selection and sex ratio selection. We find that new genetic sex determination systems are not affected by any sex ratio biases caused by associations between sex-determining regions and haploid selected loci. In addition, we find that associations that build up between an ancestral sex-determining locus and a haploid-selected locus can favour transitions between male and female heterogamety (e.g., a neo-Z/neo-W locus spreads in an ancestrally XY system), despite the fact that these ancestral associations were built up by selection. This does not occur in models that do not include haploid selection.

NOTE RE: DRIVE. I expect drive (that occurs specifically in one sex, e.g., during spermatogenesis) to behave almost exactly like haploid selection. That is, I think that a XY-linked driver that is maintained by selection (e.g., because it causes sterility when homozygous, which is common in known drive systems) will only favour invasion of a more tightly linked neo-Y (worsening sex ratio biases) and could favour invasion of a neo-W. This may run counter to generic expectations from new sex chromosome systems evolving to balance the sex ratio. So, do you think it would significantly enhance the paper to model drive explicitly or just discuss it as being similar???

FOR RESULTS?

FROM PREVIOUS PAPER: The maintenance of polymorphism at loci that experience sex specific selection in both haploid and diploid phases was considered by Immler et al. Immler et al. (2012), demonstrating that polymorphisms can be maintained by sexually antagonistic selection or overdominance as well as by conflicting selection pressures in haploids and diploids (haploid-diploid conflict or ploidally antagonistic selection) or a combination of these selective regimes.

Model

We consider a three locus model. Locus \mathbf{X} is the ancestral sex-determining region (SDR), with alleles X and Y (or Z and W). Locus \mathbf{A} is a region under selection, with alleles A and a. And locus \mathbf{M} is a novel sex-determining region, with alleles M and m. With genotype MM at locus \mathbf{M} , the sex of a zygote is determined by the genotype at locus \mathbf{X} (XX become females and XY males, or ZW become females and ZZ males). With at least one m allele at locus \mathbf{M} , a zygote develops as a female with probability k and as a male with probability 1-k. With k=0, the novel sex determiner is a masculinizer (e.g., a neo-Y chromosome) and with k=1 the novel sex determiner is a feminizer (e.g., a neo-W chromosome). With intermediate k, locus \mathbf{M} is interpreted as an offspring controlled environmental sex-determination, where mothers with at least one m allele produce daughters with probability k.

The life-cycle begins with competition between haploid gametes/gametophytes from each sex, where selection depends on the sex of the diploid they came from and their allele at the $\bf A$ locus. Gametes/gametophytes from males then pair randomly with gametes/gametophytes from females. The resulting zygotes develop as males or females, depending on their genotypes at the $\bf X$ and $\bf M$ loci (or the $\bf M$ genotype of their mother in the case of maternal control). Diploids compete with others of the same sex, where selection depends on the sex of the individual and its genotype at the $\bf A$ locus. This is followed by meiosis with recombination. Recombination occurs between loci $\bf X$ and $\bf A$ with probability r, between loci $\bf A$ and $\bf M$ with probability R, and between loci $\bf X$ and $\bf M$ with probability χ . Any order of the loci can be modelled with appropriate choices of r, R, and χ . We track the frequency of haploid genotypes produced by each sex from one generation to the next (recursion equations in Sup. Mat.).

With allele M fixed at the M locus, sex is determined by locus X and an

equilibrium is reached at locus \mathbf{A} . We then examine the ability of a rare, novel sex-determiner, m, to increase in frequency from this equilibrium. Numerical simulations are used to examine when an invading mutation goes to fixation.

Results

Resident equilibrium and stability

With allele M fixed, we follow the dynamics of the frequency of A in gametes/gametophytes from the homogametic sex (e.g., in eggs from an XX female, p_X^f) and the frequencies of A in the two types of gametes/gametophytes from the heterogametic sex (e.g., in sperm from an XY male that are X-bearing, p_X^m , and Y-bearing, p_Y^m). Assuming selection is weak relative to recombination, the differences in the frequencies of A in each type of gamete/gametophyte are small, and we can solve for the mean frequency of A across all types (p_A) as well as the difference in the frequencies of A between two of the three types, to first order in selection. Linear stability analysis can then be used to determine the stability of this equilibrium. With no haploid selection our results reduce to those of van Doorn and Kirkpatrick (2007). However, with haploid selection a stable polymorphism at locus A no longer requires sexually antagonistic selection, as it can also be achieved by ploidally-antagonistic selection.

Sex chromosome turnover

The spread of a rare mutant m at the M locus in such a population is determined by the leading eigenvalue, λ , of the system described by the equations for the next generation frequency of haploid genotypes with the mutation. Below we present the results for invasion into an XY system assuming no competition among gametes/gametophytes from females and linear arrangement XAM. Equivalent results for invasions into a ZW system

can be derived by consistently switching the roles of males and females (van Doorn and Kirkpatrick 2010).

Neo-Y and neo-W

A rare, dominant neo-Y (k = 0) or neo-W (k = 1) is always expected to invade an ancestral XY system when the average growth rate of the mutant haplotypes (Am and am) is positive, $(g_A + g_a)/2 > 0$. When the growth rates of the mutant haplotypes without recombination (R = 0) are negative, $g_i^* < 0 \,\forall i \in \{A, a\}$, where $g_i < g_i^*$, the new sex-determining allele does not invade.

Otherwise, the new sex-determining allele increases in frequency on one **A** background and declines on the other, and invasion requires

$$R\left[\frac{p_X^f w_a^m}{g_A^*} + \frac{(1 - p_X^f) w_A^m}{g_a^*}\right] w_{Aa}^m < \nu^m, \tag{1}$$

for the neo-Y, and

$$R\left[\frac{(1-\bar{p}^{m})w_{a}^{m}}{g_{A}^{*}} + \frac{\bar{p}^{m}w_{A}^{m}}{g_{a}^{*}}\right]w_{Aa}^{f} < \nu^{f}$$
(2)

for the neo-W. Here $\bar{p}^m = (p_X^m + p_Y^m)/2$ is the average frequency of A in gametophytes produced by males, w_j^i is the relative viability fitness of sex $i \in \{m, f\}$ depending on their haploid or diploid genotype at the \mathbf{A} locus (with $j \in \{A, a, AA, Aa, aa\}$) and $\nu^i = p_X^f p_{\mathbf{X}^i}^m w_a^m w_{AA}^i + p_X^f (1 - p_{\mathbf{X}^i}^m) w_a^m w_{Aa}^i + (1 - p_X^f) p_{\mathbf{X}^i}^m w_a^m w_{Aa}^i + (1 - p_X^f) (1 - p_{\mathbf{X}^i}^m) w_a^m w_{aa}^i$, with $\mathbf{X}^m = Y$ and $\mathbf{X}^f = X$, is the mean relative fitness of resident individuals of sex i.

The new sex-determining allele is therefore expected to invade for any recombination rate, R, when the net flow of double recombinants is from the less fit to the more fit A background (making the terms inside the square brackets in Equations 1 and 2 negative). When the net flow of double recombinants is from the more fit to the less fit haplotype, the new sex-determining

allele can still invade when the rate of recombination between it and the selected locus, R, is small enough.

Assuming weak selection, we can solve for the invasion fitness of the new sex-determining allele, m, into the ancestral XY system explicitly, giving

$$\lambda_{m,XY} \approx 1 + V_A S_A C_m,\tag{3}$$

where $V_A = p_A(1-p_A)$ is the variance at the **A** locus, $S_A = s^f(t^m)^2/\left[rR\left(s^f + s^m\right)^2\right]$ (INTERPRETATION?), s^f and s^m are the respective selection coefficients for A in diploid females and males, t^m is the selection coefficient for A in gametes/gametophytes from males, and we've assumed equal dominance coefficients in the two sexes.

For the neo-Y we have $C_Y = (r - R)s^f$. The neo-Y can therefore invade whenever it is in tighter linkage with the selected locus than the ancestral sex-determining locus, r > R, provided locus **A** is polymorphic $(V_A > 0)$ and there is selection among both females $(s^f \neq 0)$ and gametes/gametophytes from males $(t^m \neq 0)$. This reduces to the weak linkage results of van Doorn and Kirkpatrick (2007) when we do not assume equal dominance coefficients in the two sexes, there is no haploid selection, and there is free recombination between locus **A** and one of the sex-determining regions.

For the neo-W we have $C_W = [(2r(1-R)-R)s^f + (1-2r)Rs^m]/2$. In this case, even when the novel sex-determining locus is in looser linkage with the selected locus than the ancestral sex-determining locus is, r < R, a neo-W can invade. For example, with R = 1/2 the neo-W invades if there is any linkage between the ancestral sex-determining and selected loci (r < 1/2), there is selection among gametes/gametophytes in males $(t^m \neq 0)$, and there is selection for or against A in both males and females $(s^m s^f > 0)$ that is stronger in males than in females $(|s^m| > |s^f|)$. This reduces to the weak linkage results of (van Doorn and Kirkpatrick 2010, Equation 3) when we do not assume equal dominance coefficients in the two sexes and there is no haploid selection.

Offspring-controlled neo-ESD

The growth rate of a rare, dominant offspring-controlled neo-ESD region that produces males or females with equal probability (k = 1/2) is

$$\lambda_{ESD,XY} \approx 1 + \frac{1}{2} \frac{(\lambda_{Y,XY} - 1) + (\lambda_{W,XY} - 1)}{2} \Big|_{R=1/2}$$
 (4)

Thus with k=1/2 the neo-ESD gets half of the advantages of a neo-W and half that of a neo-Y, but only has an effect one half of the time (the other half of the time it produces the same sex as the ancestral system would have). Recombination between the selected locus and the novel sex-determining locus, R, doesn't enter into the k=1/2 results because sex is essentially randomized each generation, preventing associations from building up between allele A and sex.

Maternally-controlled neo-ESD

One might think that when the sex of zygotes is under the control of mothers, there would be strong selection to balance the sex ratio among zygotes. However, we find that, as with offspring control, assuming weak selection the invasion fitness of a sex-determiner that is maternally controlled can be written

$$\lambda_{k,XY} \approx 1 + V_A S_A C_k,\tag{5}$$

where C_k is a term that depends on k. Of particular interest is k = 1/2 (i.e., when the mother perfectly balances the sex ratio of her offspring). When both recombination rates are small we have $C_{1/2} \approx R(s^m - s^f)/8 = \lim_{r\to 0} C_W/4$. This implies that, at least under tight linkage, the invasion of maternal ESD is independent of R (because $S_A \propto R^{-1}$) and can invade whenever a neo-W can.

Discussion

² Brief results summary.

DRAFT (improve): In Úbeda et al. (2015), the new sex determining locus spreads because it arises in linkage with a locus that experiences drive. They assume that drive occurs predominantly in one sex, e.g., during spermatogenesis or a 'killer' sperm. A driving allele is maintained at an intermediate frequency by selection, e.g., because it causes male sterility when homozygous (because all male sperm are killed). Y chromosomes that arise in linkage with the driving allele spread because they allow drive to occur more often, thus genetic sex determination with a sex ratio bias evolves. Thus Úbeda et al. (2015) also find that genetic sex determiners can invade, despite causing sex ratios to become biased. Finally, they show that autosomal 'restorers' that negate the effects of meiotic drive can invade and restore an equal sex ratio. OTHER RESTORERS. Indeed, alleles that negate the effect of sex-linked meiotic drivers and restore equal sex ratios have been identified (Stalker 1961, Smith 1975). A similar process occurs with cytoplasmic male sterility alleles (that cause biased sex ratios) and nuclear 'restorer' genotypes (Frank 1989). When sex ratio bias occurs due to haploid selection, a natural class of sex ratio 'restorers' exist because haploid selection often occurs in a context that is determined by the diploid parents. For example, the intensity of pollen competition can be manipulated by altering style length (Travers and Shea 2001, Lankinen and Skogsmyr 2001, Ruane 2009), delaying stigma receptivity (Galen et al. 1986, Lankinen and Madjidian 2011) and/or delaying pollen tube growth in the pistil (Herrero 2003). Where the X and Y have fitness differences, Hough et al. (2013) and Otto et al. (2015) demonstrated that mothers should generally evolve to balance sex ratios by reducing the intensity of haploid competition.

FROM THESIS: However, reducing competition among haploids also reduces the potential for harmful deleterious mutations to be purged. When deleterious mutations are included, the optimal intensity of haploid selection

can reflect a balance between maximizing offspring fitness and equalizing sex ratios.

As part of a collaborative project (Otto et al. 2015), I considered the evolution of the haploid 'selective arena' in cases where the X chromosome harbours a polymorphism that affects haploid fitness. Mothers again primarily evolve to restore equal sex ratios. However, modifying haploid selection also affects the X-linked genotypes that are inherited by offspring. Specifically, increasing the intensity of haploid selection increases the proportion of daughters (all progeny of X-bearing sperm/pollen are female) that inherit the allele with high haploid fitness. If this allele has high fitness in daughters, mothers can be selected to increase the intensity of haploid selection; otherwise, decreased selection among haploids is favoured. Thus, because altering haploid selection intensity affects the alleles that are inherited by daughters, mothers can favour slightly biased sex ratios. In addition, I found that stronger sex ratio biases can be favoured by paternal manipulations of the haploid 'selective arena' because fathers are strongly selected to maximize their own siring success (above selection to equalize the sex ratio).

Discuss patterns that might be looked for:

In broadcast spawning animal species (e.g., corals, many fish) and species where sperm typically requires greater longevity, haploid selection may be stronger because transcripts shared during spermatogenesis may become depleted (Immler et al. 2014). also, mating systems (e.g., fewer alleles are available during haploid competition in monogamous species), selfing rates, and estimates of pollen limitation could be used as indicators of the intensity of haploid selection

- We have results where polygenic sex determination is sometimes stable, may be worth mentioning:
- "Polygenic sex determination has been reported in many plants (e.g. Shannon & Holsinger 2007), fishes (Vandeputte et al. 2007; Ser et al. 2010;
 Liew et al. 2012), crustaceans (e.g. Battaglia 1958; Battaglia & Malesani

- 1959; Voordouw & Anholt 2002), bivalves (Haley 1977; Saavedra et al. 1997),
 gastropods (Yusa 2007a,b), and polychaetes (Bacci 1965, 1978; Premoli et al. 1996)." From Vuilleumier et al. 2007: "Polymorphism for sex-determining genes within or among populations has been reported in many species including houseflies, midges, woodlice, platyfish, cichlid fish, and frogs (Gordon, 1944; Kallman, 1970; Thomp-son, 1971; Macdonald, 1978; Bull, 1983; Rigaud et al., 1997; Caubet et al., 2000; Lande et al., 2001; Ogataet al., 2003; Lee et al., 2004; Mank et al., 2006)."
- (Check with Jim Bull that it's ok before including this speculation:) Finally, Hamilton (1967) pointed out that biased sex ratios can affect population size because the number of offspring in each generation is typically
 determined by the number of females. Population density can, in turn, affect the intensity of pollen/sperm competition in future generations because
 fewer males are available to donate pollen/sperm in a particular area. Thus,
 a feedback could occur between population densities and haploid selection,
 which has not yet been investigated.

References

- Arunkumar, R., E. B. Josephs, R. J. Williamson, and S. I. Wright. 2013. Pollen-specific, but not sperm-specific, genes show stronger purifying selection and higher rates of positive selection than sporophytic genes in Capsella grandiflora. Molecular biology and evolution 30:2475–2486.
- Bachtrog, D., J. E. Mank, C. L. Peichel, M. Kirkpatrick, S. P. Otto, T.-L. Ashman, M. W. Hahn, J. Kitano, I. Mayrose, R. Ming, N. Perrin, L. Ross,
 N. Valenzuela, J. C. Vamosi, and Tree of Sex Consortium. 2014. Sex determination: why so many ways of doing it? PLoS Biol 12:e1001899.
- Beukeboom, L. W., and N. Perrin. 2014. The evolution of sex determination. Oxford University Press, Oxford, UK.

- Borg, M., L. Brownfield, and D. Twell. 2009. Male gametophyte development: a molecular perspective. Journal of Experimental Botany 60:1465–1478.
- Bull, J. J. 1983. Evolution of sex determining mechanisms. The Benjamin Cummings Publishing Company.
- Charlesworth, D., and J. E. Mank. 2010. The birds and the bees and the flowers and the trees: lessons from genetic mapping of sex determination in plants and animals. Genetics 186:9–31.
- Charnov, E. L., and J. Bull. 1977. When is sex environmentally determined?
 Nature 266:828–830.
- Clarke, H. J., T. N. Khan, and K. H. M. Siddique. 2004. Pollen selection for chilling tolerance at hybridisation leads to improved chickpea cultivars. Euphytica 139:65–74.
- Conn, J. S., and U. Blum. 1981. Sex ratio of Rumex hastatulus: the effect of environmental factors and certation. Evolution 35:1108–1116.
- Conover, D. O., and S. W. Heins. 1987. Adaptive variation in environmental and genetic sex determination in a fish. Nature 326:496–498.
- Ezaz, T., S. D. Sarre, and D. O'Meally. 2009. Sex chromosome evolution in lizards: independent origins and rapid transitions. Cytogenetic and
 Genome Research 127:249–260.
- Field, D. L., M. Pickup, and S. C. H. Barrett. 2012. The influence of pollination intensity on fertilization success, progeny sex ratio, and fitness in a wind-pollinated, dioecious plant. International Journal of Plant Sciences 173:184–191.
- ———. 2013. Comparative analyses of sex-ratio variation in dioecious flowering plants. Evolution 67:661–672.

- Frank, S. A. 1989. The Evolutionary Dynamics of Cytoplasmic Male Sterility.

 American Naturalist 133:345–376.
- Galen, C., J. A. Shykoff, and R. C. Plowright. 1986. Consequences of stigma receptivity schedules for sexual selection in flowering plants. American Naturalist pages 462–476.
- Gossmann, T. I., M. W. Schmid, U. Grossniklaus, and K. J. Schmid. 2014. Selection-driven evolution of sex-biased genes Is consistent with sexual selection in *Arabidopsis thaliana*. Molecular biology and evolution 31:574–583.
- Hamilton, W. D. 1967. Extraordinary sex ratios. Science 156:477–488.
- Hedhly, A., J. I. Hormaza, and M. Herrero. 2004. Effect of temperature on pollen tube kinetics and dynamics in sweet cherry, *Prunus avium* (Rosaceae). American journal of botany 91:558–564.
- Herrero, M. 2003. Male and female synchrony and the regulation of mating in flowering plants. Philosophical Transactions of the Royal Society B:
 Biological Sciences 358:1019–1024.
- Hillis, D. M., and D. M. Green. 1990. Evolutionary changes of heterogametic sex in the phylogenetic history of amphibians. Journal of Evolutionary Biology 3:49–64.
- Holleley, C. E., D. O'Meally, S. D. Sarre, J. A. Marshall Graves, T. Ezaz,
 K. Matsubara, B. Azad, X. Zhang, and A. Georges. 2015. Sex reversal
 triggers the rapid transition from genetic to temperature-dependent sex.
 Nature 523:79–82.
- Hormaza, J. I., and M. Herrero. 1996. Male gametophytic selection as a plant breeding tool. Scientia horticulturae 65:321–333.

- Hough, J., S. Immler, S. Barrett, and S. P. Otto. 2013. Evolutionarily stable sex ratios and mutation load. Evolution 7:1915–1925.
- Immler, S., G. Arnqvist, and S. P. Otto. 2012. Ploidally antagonistic selection maintains stable genetic polymorphism. Evolution 66:55–65.
- Immler, S., C. Hotzy, G. Alavioon, E. Petersson, and G. Arnqvist. 2014.
 Sperm variation within a single ejaculate affects offspring development in
 Atlantic salmon. Biology letters 10:20131040.
- Joseph, S., and M. Kirkpatrick. 2004. Haploid selection in animals. Trends in Ecology & Evolution 19:592–597.
- Kozielska, M., F. J. Weissing, L. W. Beukeboom, and I. Pen. 2010. Segregation distortion and the evolution of sex-determining mechanisms. Heredity 104:100–112.
- Lankinen, A., and J. A. Madjidian. 2011. Enhancing pollen competition by delaying stigma receptivity: Pollen deposition schedules affect siring ability, paternal diversity, and seed production in *Collinsia heterophylla* (Plantaginaceae). American journal of botany 98:1191–1200.
- Lankinen, A., and I. Skogsmyr. 2001. Evolution of pistil length as a choice mechanism for pollen quality. Oikos 92:81–90.
- Li, J., R. B. Phillips, A. S. Harwood, B. F. Koop, and W. S. Davidson. 2011.
 Identification of the Sex Chromosomes of Brown Trout (Salmo trutta)
 and Their Comparison with the Corresponding Chromosomes in Atlantic Salmon (Salmo salar) and Rainbow Trout (Oncorhynchus mykiss). Cytogenetic and Genome Research 133:25–33.
- Lloyd, D. G. 1974. Female-predominant sex ratios in angiosperms, vol. 32.

 Heredity.

- Mank, J. E., D. E. L. Promislow, and J. C. Avise. 2006. Evolution of alternative sexdetermining mechanisms in teleost fishes. Biological Journal of the Linnean Society 87:83–93.
- Ming, R., A. Bendahmane, and S. S. Renner. 2011. Sex chromosomes in land plants. dx.doi.org 62:485–514.
- Mulcahy, D. L., M. Sari-Gorla, and G. B. Mulcahy. 1996. Pollen selection past, present and future. Sexual Plant Reproduction 9:353-356.
- Myosho, T., H. Otake, H. Masuyama, M. Matsuda, Y. Kuroki, A. Fujiyama,
 K. Naruse, S. Hamaguchi, and M. Sakaizumi. 2012. Tracing the Emergence
 of a Novel Sex-Determining Gene in Medaka, Oryzias luzonensis. Genetics
 191:163–170.
- Ogata, M., Y. Hasegawa, H. Ohtani, M. Mineyama, and I. Miura. 2007. The ZZ/ZW sex-determining mechanism originated twice and independently during evolution of the frog, Rana rugosa. Heredity 100:92–99.
- Otto, S. P., M. F. Scott, and S. Immler. 2015. Evolution of haploid selection in predominantly diploid organisms. Proceedings of the National . . .
- Pen, I., T. Uller, B. Feldmeyer, A. Harts, G. M. While, and E. Wapstra. 2010. Climate-driven population divergence in sex-determining systems. Nature 468:436–438.
- Pokorná, M., and L. Kratochvíl. 2009. Phylogeny of sexdetermining mechanisms in squamate reptiles: are sex chromosomes an evolutionary trap?
 Zoological Journal of the ... 156:168–183.
- Ravikumar, R. L., B. S. Patil, and P. M. Salimath. 2003. Drought tolerance in sorghum by pollen selection using osmotic stress. Euphytica 133:371–376.
- Ruane, L. G. 2009. Post-pollination processes and non-random mating among compatible mates. Evolutionary Ecology Research 11:1031–1051.

- Ser, J. R., R. B. Roberts, and T. D. Kocher. 2010. Multiple interacting loci control sex determination in lake Malawi cichlid fish. Evolution 64:486–501.
- Slancarova, V., J. Zdanska, B. Janousek, M. Talianova, C. Zschach, J. Zluvova, J. Siroky, V. Kovacova, H. Blavet, J. Danihelka, B. Oxelman, A. Widmer, and B. Vyskot. 2013. Evolution of sex determination systems with heterogametic males and females in *Silene*. Evolution 67:3669–3677.
- Smith, D. A. S. 1975. All-female broods in the polymorphic butterfly Danaus chrysippus L. and their ecological significance. Heredity 34:363–371.
- Stalker, H. D. 1961. The Genetic Systems Modifying Meiotic Drive in Drosophila Paramelanica. Genetics.
- Stehlik, I., and S. Barrett. 2005. Mechanisms governing sex-ratio variation in dioecious *Rumex nivalis*. Evolution 59:814–825.
- Stehlik, I., and S. C. H. Barrett. 2006. Pollination intensity influences sex ratios in dioecious Rumex nivalis, a wind-pollinated plant. Evolution 60:1207–1214.
- Travers, S. E., and K. Shea. 2001. Selection on pollen competitive ability in relation to stochastic factors influencing pollen deposition. Evolutionary Ecology Research 3:729–745.
- Ubeda, F., M. M. Patten, and G. Wild. 2015. On the origin of sex chromosomes from meiotic drive. Proceedings of the Royal Society B: Biological
 Sciences 282:20141932.
- van Doorn, G. S., and M. Kirkpatrick. 2007. Turnover of sex chromosomes induced by sexual conflict. Nature 449:909–912.
- ——. 2010. Transitions Between Male and Female Heterogamety Caused by Sex-Antagonistic Selection. Genetics 186:629–645.

- Vibranovski, M. D., D. S. Chalopin, H. F. Lopes, M. Long, and T. L. Karr. 2010. Direct evidence for postmeiotic transcription during *Drosophila melanogaster* spermatogenesis. Genetics 186:431–433.
- Vicoso, B., and D. Bachtrog. 2015. Numerous transitions of sex chromosomes in Diptera. PLoS Biol 13:e1002078.
- Vuillleumier, S., R. Lande, J. J. M. van Alphen, and O. Seehausen. 2007.
 Invasion and fixation of sex-reversal genes. Journal of Evolutionary Biology
 20:913–920.
- Werren, J. H., and P. D. Taylor. 1984. The effects of population recruitment on sex ratio selection. The American Naturalist 124:143–148.
- Wilson, D. S., and R. K. Colwell. 1981. Evolution of sex ratio in structured demes. Evolution 35:882–897.
- Yano, A., B. Nicol, E. Jouanno, E. Quillet, A. Fostier, R. Guyomard, and Y. Guiguen. 2012. The sexually dimorphic on the Y-chromosome gene (sdY) is a conserved male-specific Y-chromosome sequence in many salmonids. Evolutionary Applications 6:486–496.
- Zheng, Y., X. Deng, and P. A. Martin-DeLeon. 2001. Lack of sharing of Spam1 (Ph-20) among mouse spermatids and transmission ratio distortion. Biology of Reproduction 64:1730–1738.