

# class9- Alpha fold

AUTHOR

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## skip section 1

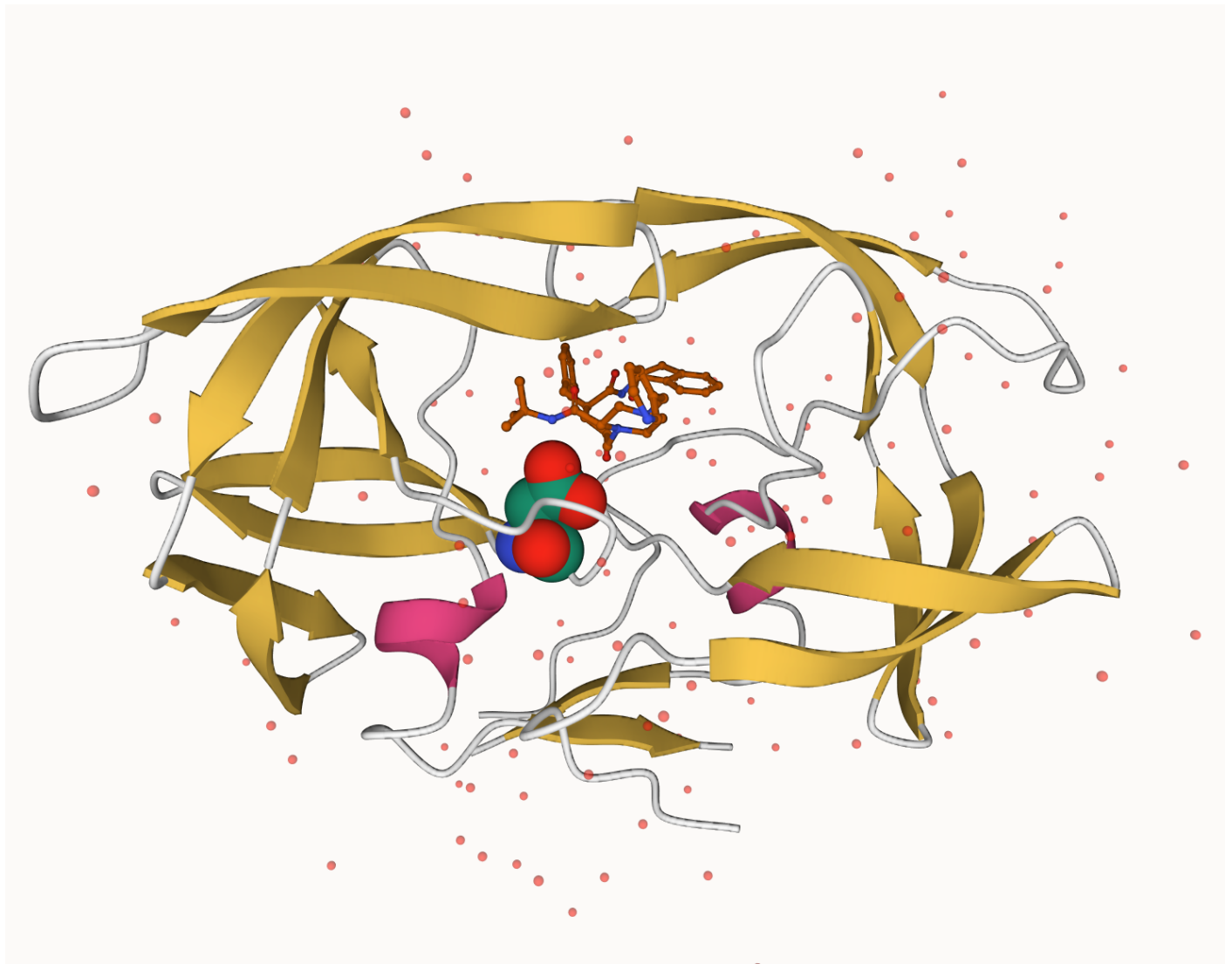
### The important role of water

Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure? Given that hydrogen is the smallest atom, it is not easy to see here.

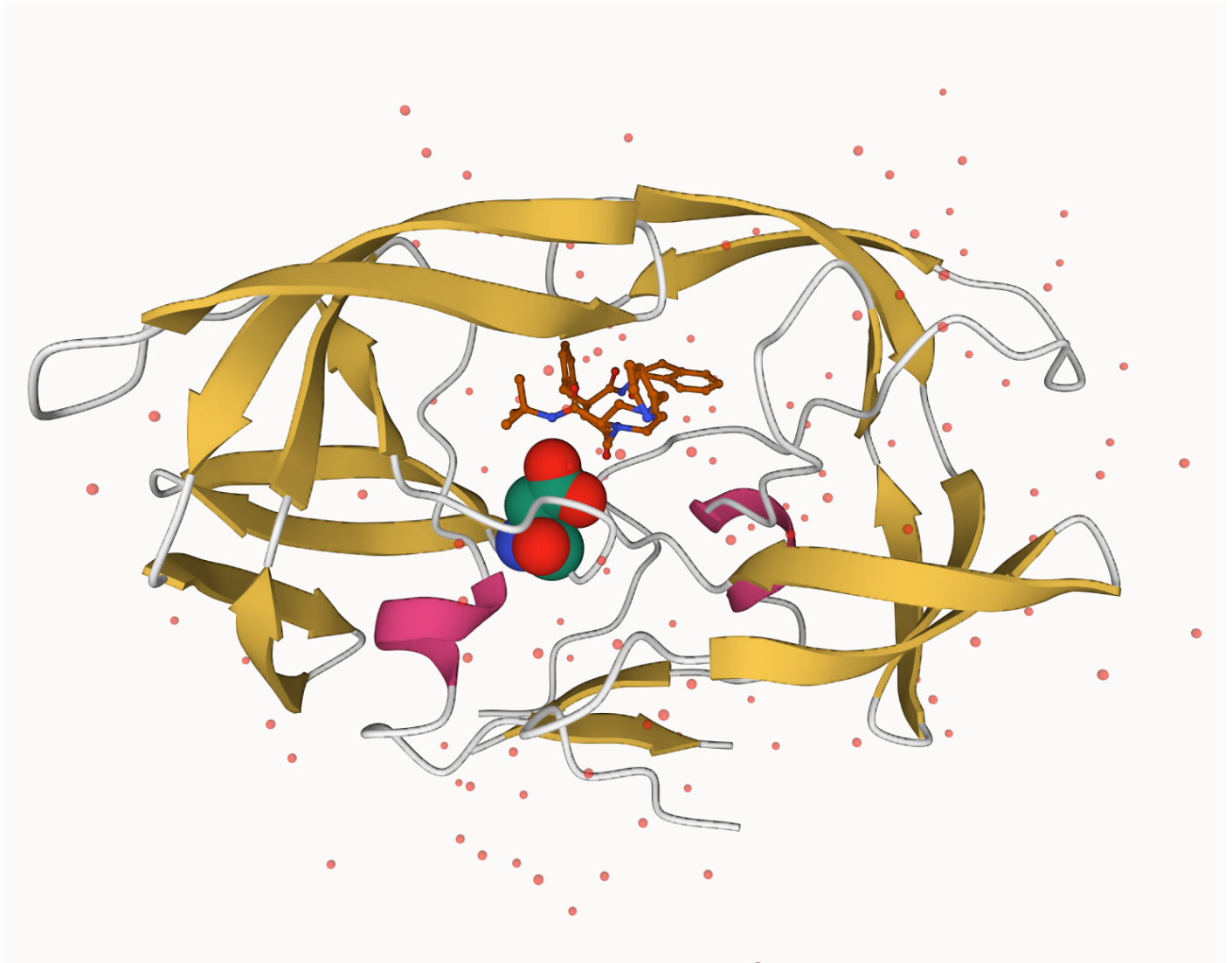
Q5: There is a critical "conserved" water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have

308

Q6: Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain and the critical water (we recommend "Ball & Stick" for these side-chains). Add this figure to your Quarto document.



## #Viewing PDB structures with Molstart



**3. Introduction to Bio3D in R** Bio3D is an R package for structural bioinformatics. Features include the ability to read, write and analyze biomolecular structure, sequence and dynamic trajectory data.

```
library(bio3d)
```

Reading PDB file data into R

```
pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
pdb
```

```
Call: read.pdb(file = "1hsg")
```

Total Models#: 1

Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)  
 Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 172 (residues: 128)  
 Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]

Protein sequence:

PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD  
 QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE  
 ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP  
 VNIIGRNLLTQIGCTLNF

+ attr: atom, xyz, seqres, helix, sheet,  
 calpha, remark, call

Q7: How many amino acid residues are there in this pdb object? There are 128 amino acid residues Q8:  
 Name one of the two non-protein residues? MK1 Q9: How many protein chains are in this structure? 2

```
attributes(pdb)
```

\$names

```
[1] "atom" "xyz" "seqres" "helix" "sheet" "calpha" "remark" "call"
```

\$class

```
[1] "pdb" "sse"
```

```
head(pdb$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40

	segid	elesy	charge
1	<NA>	N	<NA>
2	<NA>	C	<NA>
3	<NA>	C	<NA>
4	<NA>	O	<NA>
5	<NA>	C	<NA>
6	<NA>	C	<NA>

## Predicting functional motions of a single structure

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file  
PDB has ALT records, taking A only, rm.alt=TRUE

```
adk
```

```
Call: read.pdb(file = "6s36")
```

```
Total Models#: 1
```

```
Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)
```

```
Protein Atoms#: 1654 (residues/Calpha atoms#: 214)
```

```
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

```
Non-protein/nucleic Atoms#: 244 (residues: 244)
```

```
Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]
```

```
Protein sequence:
```

```
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLRAAVKSGSELGKQAKDIMDAGKLV  
DELVIALVKERIAQEDCRNGFLLDGFRTIPQADAMKEAGINVDYVLEFDVPDELIVDKI  
VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTRKDDQEETVRKRLVEYHQMTAPLIG  
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
```

```
+ attr: atom, xyz, seqres, helix, sheet,  
       calpha, remark, call
```

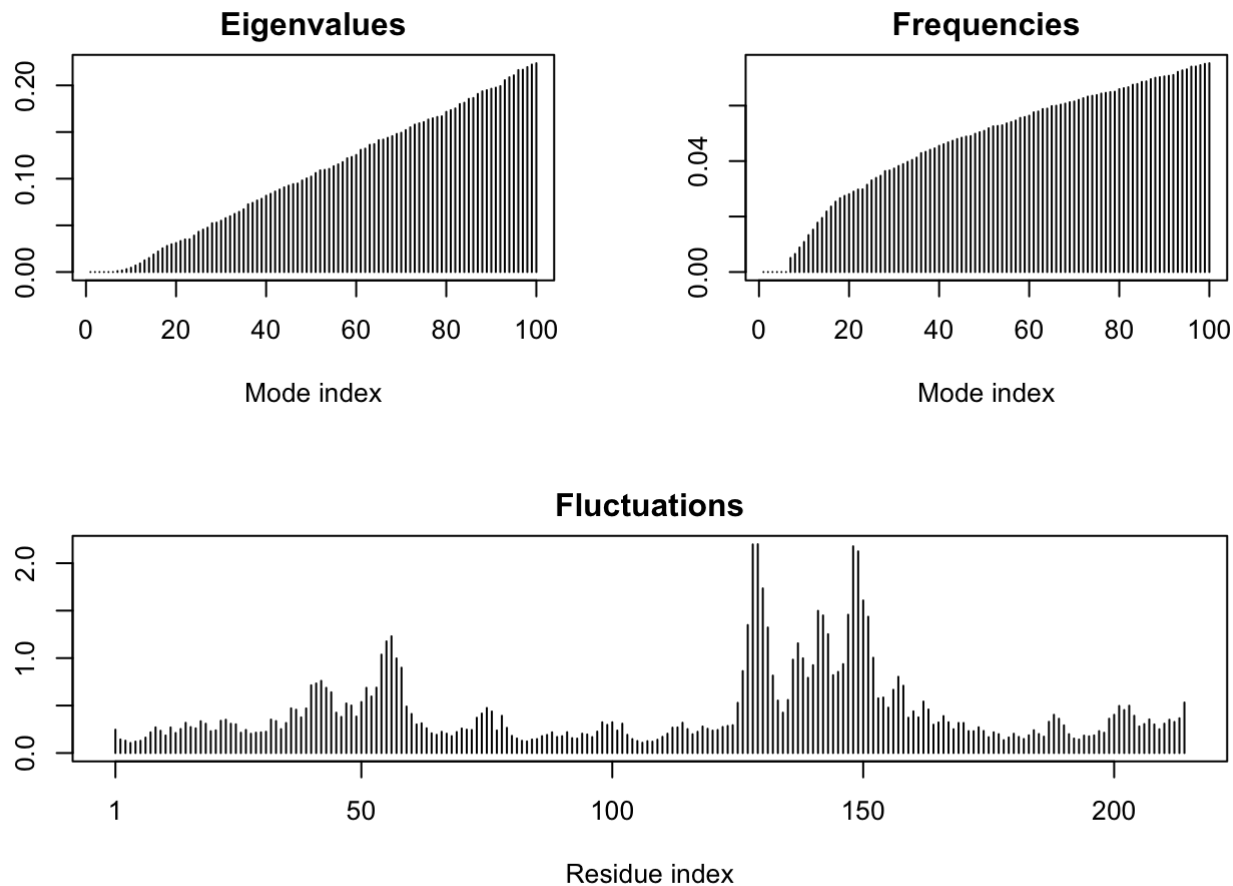
Normal mode analysis (NMA) it is a bioinformatics method for predicting functional motions. It will show us the parts of the protein that are "flexible" (ie. most dynamic). Perform flexibility prediction

```
m <- nma(adk)
```

```
Building Hessian... Done in 0.084 seconds.
```

```
Diagonalizing Hessian... Done in 0.249 seconds.
```

```
plot(m)
```



Make a movie of this moving by creating a trajectory

```
mktrj(m, file = 'adk_nma.pdb')
```

**\*\* 4. Comparative structure analysis of all Adenylate Kinase (ADK) structures** The goal of this section is to perform principal component analysis (PCA) on the complete collection of Adenylate kinase structures in the protein data-bank (PDB) **\*\* ADK → PDB database → homologous PDB get.seq() → blast.pdb() → get.pdb()**

```
# Install packages in the R console NOT your Rmd/Quarto file

#install.packages("bio3d")
#install.packages("devtools")
#install.packages("BiocManager")

#BiocManager::install("msa")
devtools::install_bitbucket("Grantlab/bio3d-view")
```

Skipping install of 'bio3d.view' from a bitbucket remote, the SHA1 (dd153987) has not changed since last install.

Use `force = TRUE` to force installation

Q10. Which of the packages above is found only on BioConductor and not CRAN? msa

Q11. Which of the above packages is not found on BioConductor or CRAN? bio3d.view

Q12. True or False? Functions from the devtools package can be used to install packages from GitHub and BitBucket? TRUE

First we get the sequence of ADK and use this to search the PDB database.

```
library(bio3d)
aa <- get.seq("1ake_a")
```

Warning in get.seq("1ake\_a"): Removing existing file: seqs.fasta

Fetching... Please wait. Done.

```
aa
```

```

      1      .      .      .      .      .      .      60
pdb|1AKE|A  MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLRAAVKSGSELGKQAKDIMDAGKLV
      1      .      .      .      .      .      .      60

      61      .      .      .      .      .      .      120
pdb|1AKE|A  DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVDRI
      61      .      .      .      .      .      .      120

     121      .      .      .      .      .      .      180
pdb|1AKE|A  VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG
     121      .      .      .      .      .      .      180

     181      .      .      .      214
pdb|1AKE|A  YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
     181      .      .      .      214
```

Call:

```
read.fasta(file = outfile)
```

Class:

```
fasta
```

Alignment dimensions:

```
1 sequence rows; 214 position columns (214 non-gap, 0 gap)
```

```
+ attr: id, ali, call
```

Q13. How many amino acids are in this sequence, i.e. how long is this sequence?

214 amino acids

```
hits <- NULL
hits$pdb.id <- c('1AKE_A', '6S36_A', '6RZE_A', '3HPR_A', '1E4V_A', '5EJE_A', '1E4Y_A', '3X2S_A',
```

```
pdb.annotate(hits$ pdb.id)
```

	structureId	chainId	macromoleculeType	chainLength	experimentalTechnique
1AKE_A	1AKE	A	Protein	214	X-ray
6S36_A	6S36	A	Protein	214	X-ray
6RZE_A	6RZE	A	Protein	214	X-ray
3HPR_A	3HPR	A	Protein	214	X-ray
1E4V_A	1E4V	A	Protein	214	X-ray
5EJE_A	5EJE	A	Protein	214	X-ray
1E4Y_A	1E4Y	A	Protein	214	X-ray
3X2S_A	3X2S	A	Protein	214	X-ray
6HAP_A	6HAP	A	Protein	214	X-ray
6HAM_A	6HAM	A	Protein	214	X-ray
4K46_A	4K46	A	Protein	214	X-ray
3GMT_A	3GMT	A	Protein	230	X-ray
4PZL_A	4PZL	A	Protein	242	X-ray

	resolution	scopDomain	pfam
1AKE_A	2.00	Adenylate kinase	Adenylate kinase, active site lid (ADK_lid)
6S36_A	1.60	<NA>	Adenylate kinase, active site lid (ADK_lid)
6RZE_A	1.69	<NA>	Adenylate kinase, active site lid (ADK_lid)
3HPR_A	2.00	<NA>	Adenylate kinase, active site lid (ADK_lid)
1E4V_A	1.85	Adenylate kinase	Adenylate kinase, active site lid (ADK_lid)
5EJE_A	1.90	<NA>	Adenylate kinase, active site lid (ADK_lid)
1E4Y_A	1.85	Adenylate kinase	Adenylate kinase, active site lid (ADK_lid)
3X2S_A	2.80	<NA>	Adenylate kinase, active site lid (ADK_lid)
6HAP_A	2.70	<NA>	Adenylate kinase, active site lid (ADK_lid)
6HAM_A	2.55	<NA>	Adenylate kinase, active site lid (ADK_lid)
4K46_A	2.01	<NA>	Adenylate kinase, active site lid (ADK_lid)
3GMT_A	2.10	<NA>	Adenylate kinase, active site lid (ADK_lid)
4PZL_A	2.10	<NA>	Adenylate kinase, active site lid (ADK_lid)

	ligandId
1AKE_A	AP5
6S36_A	CL (3),NA,MG (2)
6RZE_A	NA (3),CL (2)
3HPR_A	AP5
1E4V_A	AP5
5EJE_A	AP5,C0
1E4Y_A	AP5
3X2S_A	JPY (2),AP5,MG
6HAP_A	AP5
6HAM_A	AP5
4K46_A	ADP,AMP,P04
3GMT_A	S04 (2)
4PZL_A	CA,FMT,GOL

	ligandName
1AKE_A	BIS(ADENOSINE)-5'-PENTAPHOSPHATE
6S36_A	CHLORIDE ION (3),SODIUM ION,MAGNESIUM ION (2)
6RZE_A	SODIUM ION (3),CHLORIDE ION (2)
3HPR_A	BIS(ADENOSINE)-5'-PENTAPHOSPHATE
1E4V_A	BIS(ADENOSINE)-5'-PENTAPHOSPHATE

```

1E4V_A BIS(ADENOSINE)-5'-PENTAPHOSPHATE
5EJE_A BIS(ADENOSINE)-5'-PENTAPHOSPHATE,COBALT (II) ION
1E4Y_A BIS(ADENOSINE)-5'-PENTAPHOSPHATE
3X2S_A N-(pyren-1-ylmethyl)acetamide (2),BIS(ADENOSINE)-5'-PENTAPHOSPHATE,MAGNESIUM ION
6HAP_A BIS(ADENOSINE)-5'-PENTAPHOSPHATE
6HAM_A BIS(ADENOSINE)-5'-PENTAPHOSPHATE
4K46_A ADENOSINE-5'-DIPHOSPHATE,ADENOSINE MONOPHOSPHATE,PHOSPHATE ION
3GMT_A SULFATE ION (2)
4PZL_A CALCIUM ION,FORMIC ACID,GLYCEROL

```

## source

```

1AKE_A Escherichia coli
6S36_A Escherichia coli
6RZE_A Escherichia coli
3HPR_A Escherichia coli K-12
1E4V_A Escherichia coli
5EJE_A Escherichia coli 0139:H28 str. E24377A
1E4Y_A Escherichia coli
3X2S_A Escherichia coli str. K-12 substr. MDS42
6HAP_A Escherichia coli 0139:H28 str. E24377A
6HAM_A Escherichia coli K-12
4K46_A Photobacterium profundum
3GMT_A Burkholderia pseudomallei 1710b
4PZL_A Francisella tularensis subsp. tularensis SCHU S4

```

## structureTitle

1AKE\_A STRUCTURE OF THE COMPLEX BETWEEN ADENYLATE KINASE FROM ESCHERICHIA COLI AND THE INHIBITOR AP5A REFINED AT 1.9 ANGSTROMS RESOLUTION: A MODEL FOR A CATALYTIC TRANSITION STATE

6S36\_A

Crystal structure of E. coli Adenylate kinase R119K mutant

6RZE\_A

Crystal structure of E. coli Adenylate kinase R119A mutant

3HPR\_A

Crystal structure of V148G adenylate kinase from E. coli, in complex with Ap5A

1E4V\_A

Mutant G10V of adenylate kinase from E. coli, modified in the Gly-loop

5EJE\_A

Crystal structure of E. coli Adenylate kinase G56C/T163C double mutant in complex with Ap5a

1E4Y\_A

Mutant P9L of adenylate kinase from E. coli, modified in the Gly-loop

3X2S\_A

Crystal structure of pyrene-conjugated adenylate kinase

6HAP\_A

Adenylate kinase

6HAM\_A

Adenylate kinase

4K46\_A

Crystal Structure of Adenylate Kinase from Photobacterium profundum

3GMT\_A

Crystal structure of adenylate kinase from burkholderia pseudomallei



4PZL\_A

crystal structure of adenylate kinase from *Francisella tularensis* subsp. *tularensis* SCHU S4

		citation	rObserved	rFree
1AKE_A		Muller, C.W., et al. J Mol Biol (1992)	0.19600	NA
6S36_A		Rogne, P., et al. Biochemistry (2019)	0.16320	0.23560
6RZE_A		Rogne, P., et al. Biochemistry (2019)	0.18650	0.23500
3HPR_A	Schrank, T.P., et al. Proc Natl Acad Sci U S A (2009)		0.21000	0.24320
1E4V_A		Muller, C.W., et al. Proteins (1993)	0.19600	NA
5EJE_A	Kovermann, M., et al. Proc Natl Acad Sci U S A (2017)		0.18890	0.23580
1E4Y_A		Muller, C.W., et al. Proteins (1993)	0.17800	NA
3X2S_A		Fujii, A., et al. Bioconjug Chem (2015)	0.20700	0.25600
6HAP_A		Kantaev, R., et al. J Phys Chem B (2018)	0.22630	0.27760
6HAM_A		Kantaev, R., et al. J Phys Chem B (2018)	0.20511	0.24325
4K46_A		Cho, Y.-J., et al. To be published	0.17000	0.22290
3GMT_A	Buchko, G.W., et al. Biochem Biophys Res Commun (2010)		0.23800	0.29500
4PZL_A		Tan, K., et al. To be published	0.19360	0.23680

	rWork	spaceGroup
1AKE_A	0.19600	P 21 2 21
6S36_A	0.15940	C 1 2 1
6RZE_A	0.18190	C 1 2 1
3HPR_A	0.20620	P 21 21 2
1E4V_A	0.19600	P 21 2 21
5EJE_A	0.18630	P 21 2 21
1E4Y_A	0.17800	P 1 21 1
3X2S_A	0.20700	P 21 21 21
6HAP_A	0.22370	I 2 2 2
6HAM_A	0.20311	P 43
4K46_A	0.16730	P 21 21 21
3GMT_A	0.23500	P 1 21 1
4PZL_A	0.19130	P 32

#Download related files

```
files <- get.pdb(hits$ pdb, path = "pdbs", split=T, gzip = T)
```

Warning in get.pdb(hits\$ pdb, path = "pdbs", split = T, gzip = T): pdbs/  
1AKE.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$ pdb, path = "pdbs", split = T, gzip = T): pdbs/  
6S36.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$ pdb, path = "pdbs", split = T, gzip = T): pdbs/  
6RZE.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$ pdb, path = "pdbs", split = T, gzip = T): pdbs/  
3HPR.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$ pdb, path = "pdbs", split = T, gzip = T): pdbs/  
1E4V.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb, path = "pdbs", split = T, gzip = T): pdbs/  
5EJE.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb, path = "pdbs", split = T, gzip = T): pdbs/  
1E4Y.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb, path = "pdbs", split = T, gzip = T): pdbs/  
3X2S.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb, path = "pdbs", split = T, gzip = T): pdbs/  
6HAP.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb, path = "pdbs", split = T, gzip = T): pdbs/  
6HAM.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb, path = "pdbs", split = T, gzip = T): pdbs/  
4K46.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb, path = "pdbs", split = T, gzip = T): pdbs/  
3GMT.pdb.gz exists. Skipping download

Warning in get.pdb(hits\$pdb, path = "pdbs", split = T, gzip = T): pdbs/  
4PZL.pdb.gz exists. Skipping download



```
|-----| 320
|
|=====| 100%
```

## Align and superpose structures

```
# Align related PDBs
pdb<- pdbaln(files, fit = TRUE, exefile="msa")
```

Reading PDB files:

```
pdb<-pdbsplit_chain(1AKE_A.pdb)
pdb<-pdbsplit_chain(6S36_A.pdb)
pdb<-pdbsplit_chain(6RZE_A.pdb)
pdb<-pdbsplit_chain(3HPR_A.pdb)
pdb<-pdbsplit_chain(1E4V_A.pdb)
pdb<-pdbsplit_chain(5EJE_A.pdb)
pdb<-pdbsplit_chain(1E4Y_A.pdb)
pdb<-pdbsplit_chain(3X2S_A.pdb)
pdb<-pdbsplit_chain(6HAP_A.pdb)
pdb<-pdbsplit_chain(6HAM_A.pdb)
pdb<-pdbsplit_chain(4K46_A.pdb)
pdb<-pdbsplit_chain(3GMT_A.pdb)
pdb<-pdbsplit_chain(4PZL_A.pdb)
PDB has ALT records, taking A only, rm.alt=TRUE
. PDB has ALT records, taking A only, rm.alt=TRUE
. PDB has ALT records, taking A only, rm.alt=TRUE
. PDB has ALT records, taking A only, rm.alt=TRUE
.. PDB has ALT records, taking A only, rm.alt=TRUE
.... PDB has ALT records, taking A only, rm.alt=TRUE
. PDB has ALT records, taking A only, rm.alt=TRUE
...
```

## Extracting sequences

```
pdb<-seq(1 name: pdbs/split_chain/1AKE_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb<-seq(2 name: pdbs/split_chain/6S36_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb<-seq(3 name: pdbs/split_chain/6RZE_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb<-seq(4 name: pdbs/split_chain/3HPR_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb<-seq(5 name: pdbs/split_chain/1E4V_A.pdb
pdb<-seq(6 name: pdbs/split_chain/5EJE_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb<-seq(7 name: pdbs/split_chain/1E4Y_A.pdb
pdb<-seq(8 name: pdbs/split_chain/3X2S_A.pdb
pdb<-seq(9 name: pdbs/split_chain/6HAP_A.pdb
pdb<-seq(10 name: pdbs/split_chain/6HAM_A.pdb
PDB has ALT records, taking A only, rm.alt=TRUE
pdb<-seq(11 name: pdbs/split chain/4K46 A.pdb
```

PDB has ALT records, taking A only, rm.alt=TRUE

pdb/seq: 12 name: pdb/split\_chain/3GMT\_A.pdb

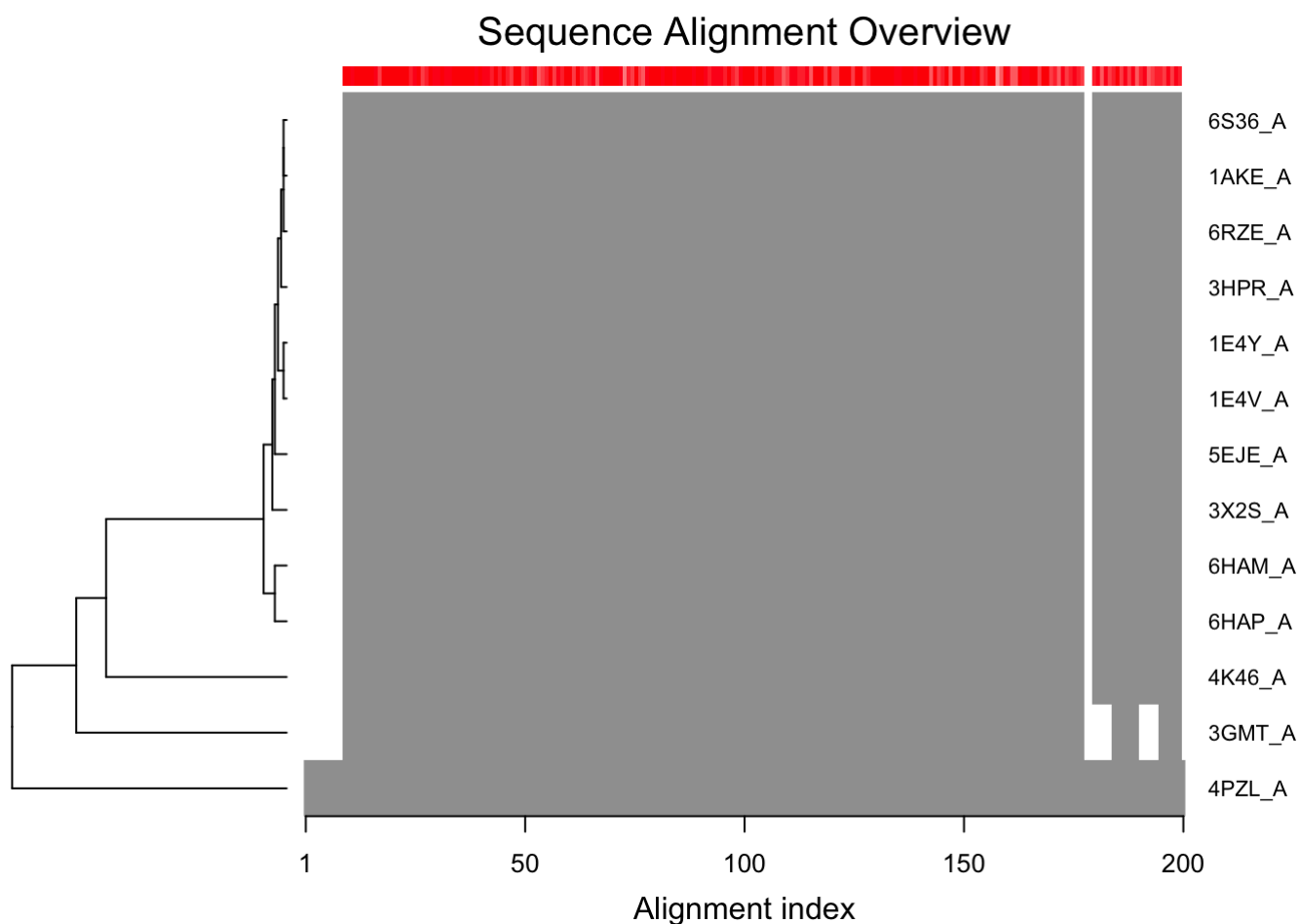
pdb/seq: 13 name: pdb/split\_chain/4PZL\_A.pdb

```
# Vector containing PDB codes for figure axis
```

```
ids <- basename.pdb(pdb$id)
```

```
# Draw schematic alignment
```

```
plot(pdb, labels=ids)
```



Annotate collected PDB structures

```
anno <- pdb.annotate(ids)
```

```
unique(anno$source)
```

- [1] "Escherichia coli"
- [2] "Escherichia coli K-12"
- [3] "Escherichia coli 0139:H28 str. E24377A"
- [4] "Escherichia coli str. K-12 substr. MDS42"
- [5] "Photobacterium profundum"
- [6] "Burkholderia pseudomallei 1710b"
- [7] "Francisella tularensis subsp. tularensis SCHU S4"

anno

	structureId	chainId	macromoleculeType	chainLength	experimentalTechnique
1AKE_A	1AKE	A	Protein	214	X-ray
6S36_A	6S36	A	Protein	214	X-ray
6RZE_A	6RZE	A	Protein	214	X-ray
3HPR_A	3HPR	A	Protein	214	X-ray
1E4V_A	1E4V	A	Protein	214	X-ray
5EJE_A	5EJE	A	Protein	214	X-ray
1E4Y_A	1E4Y	A	Protein	214	X-ray
3X2S_A	3X2S	A	Protein	214	X-ray
6HAP_A	6HAP	A	Protein	214	X-ray
6HAM_A	6HAM	A	Protein	214	X-ray
4K46_A	4K46	A	Protein	214	X-ray
3GMT_A	3GMT	A	Protein	230	X-ray
4PZL_A	4PZL	A	Protein	242	X-ray

	resolution	scopDomain	pfam
1AKE_A	2.00	Adenylate kinase	Adenylate kinase, active site lid (ADK_lid)
6S36_A	1.60	<NA>	Adenylate kinase, active site lid (ADK_lid)
6RZE_A	1.69	<NA>	Adenylate kinase, active site lid (ADK_lid)
3HPR_A	2.00	<NA>	Adenylate kinase, active site lid (ADK_lid)
1E4V_A	1.85	Adenylate kinase	Adenylate kinase, active site lid (ADK_lid)
5EJE_A	1.90	<NA>	Adenylate kinase, active site lid (ADK_lid)
1E4Y_A	1.85	Adenylate kinase	Adenylate kinase, active site lid (ADK_lid)
3X2S_A	2.80	<NA>	Adenylate kinase, active site lid (ADK_lid)
6HAP_A	2.70	<NA>	Adenylate kinase, active site lid (ADK_lid)
6HAM_A	2.55	<NA>	Adenylate kinase, active site lid (ADK_lid)
4K46_A	2.01	<NA>	Adenylate kinase, active site lid (ADK_lid)
3GMT_A	2.10	<NA>	Adenylate kinase, active site lid (ADK_lid)
4PZL_A	2.10	<NA>	Adenylate kinase, active site lid (ADK_lid)

	ligandId
1AKE_A	AP5
6S36_A	CL (3),NA,MG (2)
6RZE_A	NA (3),CL (2)
3HPR_A	AP5
1E4V_A	AP5
5EJE_A	AP5,C0
1E4Y_A	AP5
3X2S_A	JPY (2),AP5,MG
6HAP_A	AP5
6HAM_A	AP5
4K46_A	ADP,AMP,P04
3GMT_A	S04 (2)
4PZL_A	CA,FMT,GOL

	ligandName
1AKE_A	BIS(ADENOSINE)-5'-PENTAPHOSPHATE
6S36_A	CHLORIDE ION (3),SODIUM ION,MAGNESIUM ION (2)
6RZE_A	SODIUM ION (3),CHLORIDE ION (2)
3HPR_A	BIS(ADENOSINE)-5'-PENTAPHOSPHATE
1E4V_A	BIS(ADENOSINE)-5'-PENTAPHOSPHATE

```

1E4V_A BIS(ADENOSINE)-5'-PENTAPHOSPHATE
5EJE_A BIS(ADENOSINE)-5'-PENTAPHOSPHATE,COBALT (II) ION
1E4Y_A BIS(ADENOSINE)-5'-PENTAPHOSPHATE
3X2S_A N-(pyren-1-ylmethyl)acetamide (2),BIS(ADENOSINE)-5'-PENTAPHOSPHATE,MAGNESIUM ION
6HAP_A BIS(ADENOSINE)-5'-PENTAPHOSPHATE
6HAM_A BIS(ADENOSINE)-5'-PENTAPHOSPHATE
4K46_A ADENOSINE-5'-DIPHOSPHATE,ADENOSINE MONOPHOSPHATE,PHOSPHATE ION
3GMT_A SULFATE ION (2)
4PZL_A CALCIUM ION,FORMIC ACID,GLYCEROL

```

## source

```

1AKE_A Escherichia coli
6S36_A Escherichia coli
6RZE_A Escherichia coli
3HPR_A Escherichia coli K-12
1E4V_A Escherichia coli
5EJE_A Escherichia coli 0139:H28 str. E24377A
1E4Y_A Escherichia coli
3X2S_A Escherichia coli str. K-12 substr. MDS42
6HAP_A Escherichia coli 0139:H28 str. E24377A
6HAM_A Escherichia coli K-12
4K46_A Photobacterium profundum
3GMT_A Burkholderia pseudomallei 1710b
4PZL_A Francisella tularensis subsp. tularensis SCHU S4

```

## structureTitle

```

1AKE_A STRUCTURE OF THE COMPLEX BETWEEN ADENYLATE KINASE FROM ESCHERICHIA COLI AND THE
INHIBITOR AP5A REFINED AT 1.9 ANGSTROMS RESOLUTION: A MODEL FOR A CATALYTIC TRANSITION
STATE
6S36_A
Crystal structure of E. coli Adenylate kinase R119K mutant
6RZE_A
Crystal structure of E. coli Adenylate kinase R119A mutant
3HPR_A
Crystal structure of V148G adenylate kinase from E. coli, in complex with Ap5A
1E4V_A
Mutant G10V of adenylate kinase from E. coli, modified in the Gly-loop
5EJE_A
Crystal structure of E. coli Adenylate kinase G56C/T163C double mutant in complex with
Ap5a
1E4Y_A
Mutant P9L of adenylate kinase from E. coli, modified in the Gly-loop
3X2S_A
Crystal structure of pyrene-conjugated adenylate kinase
6HAP_A
Adenylate kinase
6HAM_A
Adenylate kinase
4K46_A
Crystal Structure of Adenylate Kinase from Photobacterium profundum
3GMT_A
Crystal structure of adenylate kinase from burkholderia pseudomallei

```

4PZL\_A

The

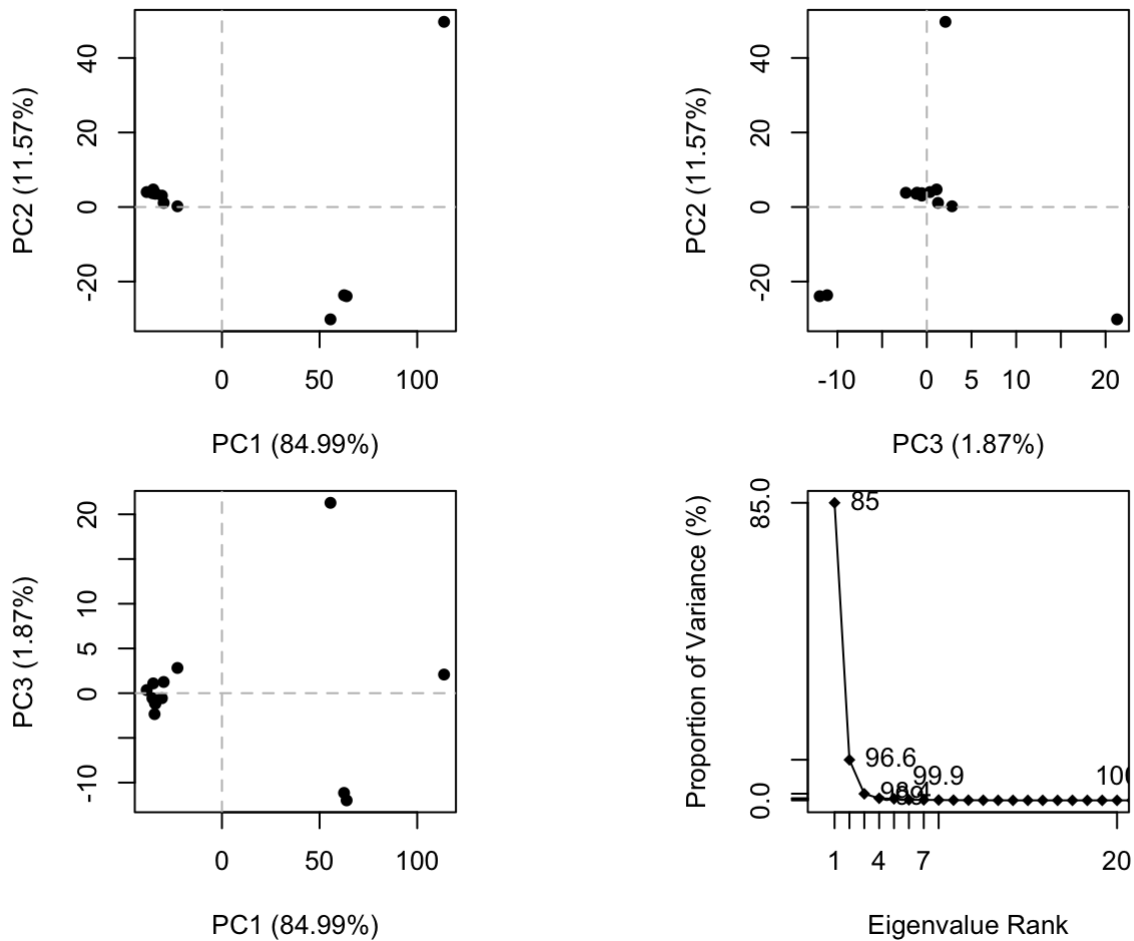
crystal structure of adenylate kinase from *Francisella tularensis* subsp. *tularensis* SCHU S4

		citation	rObserved	rFree
1AKE_A		Muller, C.W., et al. J Mol Biol (1992)	0.19600	NA
6S36_A		Rogne, P., et al. Biochemistry (2019)	0.16320	0.23560
6RZE_A		Rogne, P., et al. Biochemistry (2019)	0.18650	0.23500
3HPR_A	Schrank, T.P., et al. Proc Natl Acad Sci U S A (2009)		0.21000	0.24320
1E4V_A		Muller, C.W., et al. Proteins (1993)	0.19600	NA
5EJE_A	Kovermann, M., et al. Proc Natl Acad Sci U S A (2017)		0.18890	0.23580
1E4Y_A		Muller, C.W., et al. Proteins (1993)	0.17800	NA
3X2S_A		Fujii, A., et al. Bioconjug Chem (2015)	0.20700	0.25600
6HAP_A		Kantaev, R., et al. J Phys Chem B (2018)	0.22630	0.27760
6HAM_A		Kantaev, R., et al. J Phys Chem B (2018)	0.20511	0.24325
4K46_A		Cho, Y.-J., et al. To be published	0.17000	0.22290
3GMT_A	Buchko, G.W., et al. Biochem Biophys Res Commun (2010)		0.23800	0.29500
4PZL_A		Tan, K., et al. To be published	0.19360	0.23680

	rWork	spaceGroup
1AKE_A	0.19600	P 21 2 21
6S36_A	0.15940	C 1 2 1
6RZE_A	0.18190	C 1 2 1
3HPR_A	0.20620	P 21 21 2
1E4V_A	0.19600	P 21 2 21
5EJE_A	0.18630	P 21 2 21
1E4Y_A	0.17800	P 1 21 1
3X2S_A	0.20700	P 21 21 21
6HAP_A	0.22370	I 2 2 2
6HAM_A	0.20311	P 43
4K46_A	0.16730	P 21 21 21
3GMT_A	0.23500	P 1 21 1
4PZL_A	0.19130	P 32

Principal component analysis

```
# Perform PCA
pc.xray <- pca(pdbx)
plot(pc.xray)
```



```
# Calculate RMSD
rd <- rmsd(pdb)
```

Warning in rmsd(pdb): No indices provided, using the 204 non NA positions

```
# Structure-based clustering
hc.rd <- hclust(dist(rd))
grps.rd <- cutree(hc.rd, k=3)

plot(pc.xray, 1:2, col="grey50", bg=grps.rd, pch=21, cex=1)
```



