
Environmental Laboratory Documentation

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This textbook is written and maintained in [Github](#) via [Sphinx](#). It uses and refers to AguaClara code and functions in `aide_design`. Listed below are the versions of the programs we use:

Table 1: These are the software versions used to compile this textbook

Software	version
Sphinx	1.7.5
<code>aide_design</code>	0.0.12
Anaconda	4.5.4
Python	3.6.5

**CHAPTER
ONE**

LABORATORY RESEARCH IN ENVIRONMENTAL ENGINEERING

CHAPTER
TWO

LABORATORY MANUAL



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2.1 Preface

Continued leadership in environmental protection requires efficient transfer of innovative environmental technologies to the next generation of engineers. Responding to this challenge, the Cornell Environmental Engineering faculty redesigned the undergraduate environmental engineering curriculum and created a new senior-level laboratory course. This laboratory manual is one of the products of the course development. Our goal is to disseminate this information to help expose undergraduates at Cornell and at other institutions to current environmental engineering problems and innovative solutions.

A major goal of the undergraduate laboratory course is to develop an atmosphere where student understanding will emerge for the physical, chemical, and biological processes that control material fate and transport in environmental and engineered systems. Student interest is piqued by laboratory exercises that present modern environmental problems to investigate and solve.

The experiments were designed to encourage the process of learning around the edges. The manifest purpose of an experiment may be a current environmental problem, but it is expected that students will become familiar with analytical methods in the course of the laboratory experiment (without transforming the laboratory into an exercise in analytical techniques). It is our goal that students employ the theoretical principles that underpin the environmental field in analysis of their observations without transforming the laboratories into exercises in process theory. As a result, students can experience the excitement of addressing a current problem while coincidentally becoming cognizant of relevant physical, chemical, and biological principles.

At Cornell, student teams of two or three carry out the exercises, maximizing the opportunity for a hands-on experience. Each team is equipped with modern instrumentation as well as experimental reactor apparatus designed to facilitate the study of each topic. Computerized data acquisition, instrument control, and process control are used extensively to make it easier for students to learn how to use new instruments and to eliminate the drudgery of manual data acquisition. Software was developed at Cornell to use computers as virtual instruments that interface with gas chromatographs (HP 5890A), UV-Vis Spectrophotometers (HP 8452) as well as a variety of sensors.

The development of this manual and the accompanying course would not have been possible without funds from the National Science Foundation, the DeFrees Family Foundation, the Procter and Gamble Fund, the School of Civil and Environmental Engineering and the College of Engineering at Cornell University.

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**CHAPTER
THREE**

LABORATORY SAFETY

3.1 Introduction

Safety is a collective responsibility that requires the full cooperation of everyone in the laboratory. However, the ultimate responsibility for safety rests with the person carrying out a given procedure. In the case of an academic laboratory, that person is usually the student. Accidents often result from an indifferent attitude, failure to use common sense, or failure to follow instructions. Each student should be aware of what the other students are doing because all can be victims of one individual's mistake. Do not hesitate to point out to other students that they are engaging in unsafe practices or operations. If necessary, report it to the instructor. In the final assessment, students have the greatest responsibility to ensure their own personal safety.

This guide provides a list of dos and don'ts to minimize safety and health problems associated with experimental laboratory work. It also provides, where possible, the ideas and concepts that underlie the practical suggestions. However, the reader is expected to become involved and to contribute to the overall solutions. The following are general guidelines for all laboratory workers:

1. Follow all safety instructions carefully.
2. Become thoroughly acquainted with the location and use of safety facilities such as safety showers, exits, and eyewash fountains.
3. Become familiar with the hazards of the chemicals being used, and know the safety precautions and emergency procedures before undertaking any work.
4. Become familiar with the chemical operations and the hazards involved before beginning an operation.

3.2 Personal Protection

3.2.1 Eye Protection

All people in the laboratory including visitors must wear eye protection at all times, even when not performing a chemical operation. Wearing of contact lenses in the laboratory is normally forbidden because contact lenses can hold foreign materials against the cornea. Furthermore, they may be difficult to remove in the case of a splash. Soft contact lenses present a particular hazard because they can absorb and retain chemical vapors. If the use of contact lenses is required for therapeutic reasons fitted goggles must also be worn. In addition, approved standing shields and face shields that protect the neck and ears as well as the face should be used when appropriate for work at reduced pressure or where there is a potential for explosions, implosions, or splashing. Normal prescription eyeglasses, though meeting the Food and Drug Administration's standards for shatter resistance, do not provide appropriate laboratory eye protection.

3.2.2 Clothing

Clothing worn in the laboratory should offer protection from splashes and spills, should be easily removable in case of accident, and should be at least fire resistant. Nonflammable, nonporous aprons offer the most satisfactory and the least expensive protection. Lab jackets or coats should have snap fasteners rather than buttons so that they can be readily removed.

High-heeled or open-toed shoes, sandals, or shoes made of woven material should not be worn in the laboratory. Shorts, cutoffs, and miniskirts are also inappropriate. Long hair and loose clothing should be constrained. Jewelry such as rings, bracelets, and watches should not be worn in order to prevent chemical seepage under the jewelry, contact with electrical sources, catching on equipment, and damage to the jewelry.

3.2.3 Gloves

Gloves can serve as an important part of personal protection when they are used correctly. Check to ensure the absence of cracks or small holes in the gloves before each use. In order to prevent the unintentional spread of chemicals, gloves should be removed before leaving the work area and before handling such things as telephones, doorknobs, writing instruments, computers, and laboratory notebooks. Gloves may be reused, cleaned, or discarded, consistent with their use and contamination.

A wide variety of gloves is available to protect against chemical exposure. Because the permeability of gloves of the same or similar material varies from manufacturer to manufacturer, no specific recommendations are given here. Be aware that if a chemical diffuses through a glove, that chemical is held against the workers hand and the individual may then be more exposed to the chemical than if the glove had not been worn.

3.3 Personal Hygiene

Everyone working in a chemistry laboratory should be aware of the dangers of ingesting chemicals. These common sense precautions will minimize the possibility of such exposure:

1. Do not prepare, store (even temporarily), or consume food or beverages in any chemical laboratory.
2. Do not smoke in any chemical laboratory. Additionally, be aware that tobacco products in opened packages can absorb chemical vapors.
3. Do not apply cosmetics in a laboratory.
4. Wash hands and arms thoroughly before leaving the laboratory, even if gloves have been worn.
5. Wash separately from personal laundry, lab coats or jackets on which chemicals have been spilled.
6. Never wear or bring lab coats or jackets into areas where food is consumed.
7. Never pipette by mouth. Always use a pipette aid or suction bulb.

3.4 Laboratory Protocol

The chemistry laboratory is a place for serious learning and working. Horseplay cannot be tolerated. Variations in procedures including changes in quantities or reagents may be dangerous. Such alterations may only be made with the knowledge and approval of the instructor.

3.4.1 Housekeeping

In the laboratory and elsewhere, keeping things clean and neat generally leads to a safer environment. Avoid unnecessary hazards by keeping drawers and cabinets closed while working. Never store materials, especially chemicals, on the floor, even temporarily. Work spaces and storage areas should be kept clear of broken glassware, leftover chemicals and scraps of paper. Keep aisles free of obstructions such as chairs, boxes, and waste receptacles. Avoid slipping hazards by keeping the floor clear of ice, stoppers, glass beads or rods, other small items, and spilled liquids. Use the required procedure for the proper disposal of chemical wastes and solvents.

3.4.2 Cleaning Containers

Clean containers at the laboratory sink or in laboratory dishwashers. Use hot water, if available, and soap or other detergent. If necessary, use a mild scouring powder. Wear appropriate gloves that have been checked to ensure that no holes are present. Use brushes of suitable stiffness and size. Avoid accumulating too many articles in the cleanup area. Usually work space around a sink is limited and piling up dirty or cleaned glassware leads to breakage. Remember that the turbid water in a sink may hide a jagged edge on a piece of broken glassware that was intact when put into the water. A pair of heavy gloves may be useful for removing broken glass, but care must be exercised to prevent glove contamination. To minimize breakage of glassware, sink bottoms should have rubber or plastic mats that do not block the drains.

Avoid the use of strong cleaning agents such as nitric acid, chromic acid, sulfuric acid, strong oxidizers, or any chemical with a per in its name (such as perchloric acid, ammonium persulfate, etc.) unless specifically instructed to use them, and then only when wearing proper protective equipment. A number of explosions involving strong oxidizing cleaning solutions, such as chromic sulfuric acid mixtures, have been reported. The use of flammable solvents should be minimized and, when they are used, appropriate precautions must be observed.

3.4.3 Unattended Operation of Equipment

Reactions that are left to run unattended overnight or at other times are prime sources for fires, floods, and explosions. Do not let equipment such as power stirrers, hot plates, heating mantles, and water condensers run overnight without fail-safe provisions and the instructors consent. Check unattended reactions periodically. Always leave a note plainly posted with a phone number where you and the instructor can be reached in case of emergency. Remember that in the middle of the night, emergency personnel are entirely dependent on accurate instructions and information.

3.4.4 Fume Hoods and Ventilation

A large number of common substances present acute respiratory hazards and should not be used in a confined area in large amounts. They should be dispensed and handled only where there is adequate ventilation, such as in a hood. Adequate ventilation is defined as ventilation that is sufficient to keep the concentration of a chemical below the threshold limit value or permissible exposure limit.

If you smell a chemical, it is obvious that you are inhaling it. However, odor does not necessarily indicate that a dangerous concentration has been reached. By contrast, many chemicals can be present at hazardous concentrations without any noticeable odor.

3.4.5 Refrigerators

Chemicals stored in refrigerators should be sealed, double packaged if possible, and labeled with the name of the material, the date placed in the refrigerator, and the name of the person who stored the material. A current inventory should be maintained. Old chemicals should be disposed of after a specified storage period. Household refrigerators should not be used for chemical storage.

If used for storage of radioactive materials, a refrigerator should be plainly marked with the standard radioactivity symbol and lettering, and routine surveys should be made to ensure that the radioactive material has not contaminated the refrigerator.

Food should never be stored in a refrigerator used for chemical storage. These refrigerators should be clearly labeled No Food. Conversely food refrigerators, which must be always outside of, and away from, the chemical work area, should be labeled Food OnlyNo Chemicals.

3.4.6 Working Alone

Avoid working alone in a building or in a laboratory.

3.5 Use of Chemicals

Before using any chemical you need to know how to safely handle it. The safety precautions taken are dependent on the exposure routes and the potential harmful effects.

3.5.1 Routes of Exposure

1. ingestion
2. inhalation
3. absorbed through skin
4. eye contact

Each potential exposure route requires different precautions. Chemical exposure may have acute (immediate, short term) or chronic (long term potentially cumulative) effects. Information on health hazards can be found on chemical labels and in Safety Data Sheets.

3.5.2 Safety Data Sheets (SDSs)

SDS can be found online with simple search queries. SDS provide extensive information on safe handling, first aid, toxicity, etc. The following is a list of terms commonly used in SDS:

TLV Threshold Limit Value are values for airborne toxic materials that are to be used as guides in control of health hazards. They represent concentrations to which nearly all workers (workers without special sensitivities) can be exposed to for long periods of time without harmful effect. TLVs are usually expressed as parts per million (ppm). TLVs are also expressed as mg of dust or vapor/m³ of air.

TDLo Toxic Dose Low the lowest dose of a substance introduced by any route, other than inhalation, over any given period of time and reported to produce any toxic effect in humans or to produce carcinogenic, neoplastigenic, or teratogenic effects in animals or humans.

TCLo Toxic Concentration Low the lowest concentration of a substance in air to which humans or animals have been exposed for any given period of time and reported to produce any toxic effect in humans or to produce carcinogenic, neoplastigenic, or teratogenic effects in animals or humans.

TDLo Lethal Dose Low the lowest dose (other than LD₅₀) of a substance introduced by any route, other than inhalation, over any given period of time in one or more divided portions and reported to have caused death in humans or animals.

LD50Lethal Dose Fiftya calculated dose of a substance that is expected to cause the death of 50% of an entire defined experimental animal population. It is determined from the exposure to the substance by any route other than inhalation of a significant number from that population.

LCLoLethal Concentration Lowthe lowest concentration of a substance in air, other than LC50, that has been reported to have caused death in humans or animals. The reported concentrations may be entered for periods of exposure that are less than 24 hours (acute) or greater than 24 hours (subacute and chronic).

LC50Lethal Concentration Fiftya calculated concentration of a substance in air, exposure to which for a specified length of time is expected to cause the death of 50% of an entire defined experimental animal population. It is determined from the exposure to the substance of a significant number from that population.

3.5.3 Chemical Labels

All chemicals must be labeled. Unlabeled containers of mystery chemicals or chemical solutions are a nightmare for disposal as well as a potential safety hazard. The OSHA Hazard Communication Standard and the OSHA Lab Standard have specific requirements for the labeling of chemicals. In a laboratory covered under the Lab Standard, if a chemical is designated as a hazardous material, that is having the characteristics of corrosivity, ignitability, toxicity (generally meaning a highly toxic material with an LD50 of 50 mg/kg or less), reactivity, etc., and if it is made into a solution or repackaged as a solid or liquid in a concentration greater than 1% (0.1% for a carcinogen) it needs to have a so called Right-To-Know (RTK) label that duplicates the hazard warnings, precautions, and first aid steps found on the original label. All other chemicals must have at minimum a label with the full chemical name (not just the chemical formula), concentration, and date prepared. Right-To-Know labels will be made available for your use when necessary.

National Fire Protection Association (NFPA) ratings indicate the types and severity of the hazards. The NFPA ratings are on a scale of 0-4 with 0 being nonhazardous and 4 being most hazardous. The ratings are described in

3.5.4 Chemical Storage

There has been much concern, and some confusion, about the proper storage of laboratory chemicals. Here proper means the storage of chemicals in such a manner as to prevent incompatible materials from being accidentally mixed together in the event of the breakage of one or more containers in the storage area or to prevent the formation of reactive vapors that may require vented chemical storage areas. Below is a concise guide to the storage of common laboratory chemicals.

1. Perchloric acid is separated from all other materials.
2. Hydrofluoric acid is separated from all other materials.
3. Concentrated nitric acid is separated from all other materials.
4. Highly toxic materials (LD50 of 50 mg/kg or less) are stored separately.
5. Carcinogenic chemicals are stored separately.
6. Inorganic acids (except for 1, 2, 3 above) are stored separately.
7. Bases are stored separately.
8. Strong oxidizing agents are stored separately.
9. Strong reducing agents are stored separately.
10. Water reactive, pyrophoric and explosive materials are stored separately.
11. Flammable organic materials (solvents, organic acids, organic reagents) are stored separately.

3.5.5 Guidelines for separating incompatible chemicals

1. Place the chemicals to be stored separately in a heavy gauge Nalgene (or similar plastic) tub. Plastic secondary containers must be compatible with the material being stored.
2. Strong acids, especially perchloric, nitric, and hydrofluoric are best stored in plastic containers designed to store strong mineral acids. These are available from lab equipment supply houses.
3. Bottle-in-a-can type of containers are also acceptable as secondary containment. Small containers of compatible chemicals may be stored in a dessicator or other secure container. Secondary containment is especially useful for highly toxic materials and carcinogens.
4. Dry chemicals stored in approved cabinets with doors may be grouped together by compatibility type on separate shelves or areas of shelves separated by taping off sections of shelving to designate where chemicals of one type are stored. Physically separated cabinets may be used to provide a barrier between groups of stored incompatible chemicals. Strong mineral acids may be stored in one cabinet and strong bases stored in a second cabinet, for example. Flammable solvents should be stored in a rated flammable storage cabinet if available.

If you are uncertain of the hazardous characteristics of a particular chemical refer to the SDS for that material. A good SDS will not only describe the hazardous characteristics of the chemical, it will also list incompatible materials.

3.5.6 Transporting Chemicals

Transport all chemicals using the container-within-a-container concept to shield chemicals from shock during any sudden change of movement. Large containers of corrosives should be transported from central storage in a chemically resistant bucket or other container designed for this purpose. Stairs must be negotiated carefully. Elevators, unless specifically indicated and so designated, should not be used for carrying chemicals. Smoking is never allowed around chemicals and apparatus in transit or in the work area itself.

When moving in the laboratory, anticipate sudden backing up or changes in direction from others. If you stumble or fall while carrying glassware or chemicals, try to project them away from yourself and others.

When a flammable liquid is withdrawn from a drum, or when a drum is filled, both the drum and the other equipment must be electrically wired to each other and to the ground in order to avoid the possible buildup of a static charge. Only small quantities should be transferred to glass containers. If transferring from a metal container to glass, the metal container should be grounded.

3.5.7 Chemical Disposal

The Environmental Protection Agency (EPA) classifies wastes by their reaction characteristics. A summary of the major classifications and some general treatment guidelines are listed below. Specific information may be found in the book, Prudent Practices in the Laboratory: Handling and Management of Chemical Hazards (National Academies Press, 2011), as well as other reference materials.

Ignitability: These substances generally include flammable solvents and certain solids. Flammable solvents must never be poured down the drain. They should be collected for disposal in approved flammable solvent containers. In some cases it may be feasible to recover and reuse solvents by distillation. Such solvent recovery must include appropriate safety precautions and attention to potentially dangerous contamination such as that due to peroxide formation.

Corrosivity: This classification includes common acids and bases. They must be collected in waste containers that will not ultimately corrode and leak, such as plastic containers. It often may be appropriate to neutralize waste acids with waste bases and where allowed by local regulations, dispose of the neutral materials via the sanitary sewer system. Again, the nature of the neutralized material must be considered to ensure that it does not involve an environmental hazard such as chromium salts from chromic acid neutralization.

Reactivity: These substances include reactive metals such as sodium and various water reactive reagents. Compounds such as cyanides or sulfides are included in this class if they can readily evolve toxic gases such as hydrogen cyanide.

Their collection for disposal must be carried out with particular care. When present in small quantities, it is advisable to deactivate reactive metals by careful reaction with appropriate alcohols and to deactivate certain oxygen or sulfur containing compounds through oxidation. Specific procedures should be consulted.

Toxicity: Although the EPA has specific procedures for determining toxicity, all chemicals may be toxic in certain concentrations. Appropriate procedures should be established in each laboratory for collection and disposal of these materials.

The handling of reaction byproducts, surplus and waste chemicals, and contaminated materials is an important part of laboratory safety procedures. Each laboratory worker is responsible for ensuring that wastes are handled in a manner that minimizes personal hazard and recognizes the potential for environmental contamination.

Most instructional laboratories will have clear procedures for students to follow in order to minimize the generation of waste materials. Typically reaction byproducts and surplus chemicals will be neutralized or deactivated as part of the experimental procedure. Waste materials must be handled in specific ways as designated by federal and local regulations. University guidelines for waste disposal can be found in Chapter 7 of the Cornell University [Chemical Hygiene Plan](#).

Some general guidelines are:

1. Dispose of pipette tips in the glass disposal container to prevent puncture of trash bags.
2. Dispose of waste materials promptly. When disposing of chemicals one basic principle applies: Keep each different class of chemical in a separate clearly labeled disposal container.
3. Never put chemicals into a sink or down the drain unless they are deactivated or neutralized and they are allowed by local regulation in the sanitary sewer system. [See Chemical Hygiene Plan for list of chemicals that can be safely disposed of in the sanitary sewer.]
4. Put ordinary waste paper in a wastepaper basket separate from the chemical wastes. If a piece of paper is contaminated, such as paper toweling used to clean up a spill, put the contaminated paper in the special container that is marked for this use. It must be treated as a chemical waste.
5. Broken glass belongs in its own marked waste container. Broken thermometers may contain mercury in the fragments and these belong in their own special sealed broken thermometer container.
6. Peroxides, because of their reactivity, and the unpredictable nature of their formation in laboratory chemicals, have attracted considerable attention. The disposal of large quantities (25 g or more) of peroxides requires expert assistance. Consider each case individually for handling and disposal.

A complete list of compounds considered safe for drain disposal can be found in Chapter 7 of the Cornell University [Chemical Hygiene Plan](#). Disposal techniques for chemicals not found in this list must be disposed of using techniques approved of by Cornell Environmental Health and Safety. When possible, hazardous chemicals can be neutralized and then disposed. When chemicals are produced that cannot be disposed of using the sanitary sewer, techniques to decrease the volume of the waste should be considered.

3.6 References

Safety in Academic Chemistry Laboratories. A publication of the American Chemical Society Committee on Chemical Safety. Fifth edition. 1990

3.7 Prelab Questions

1. Why are contact lenses hazardous in the laboratory?
2. What is the minimum information needed on the label for each chemical? When are Right-To-Know labels required?

3. Why is it important to label a bottle even if it only contains distilled water?
4. Find an SDS for sodium nitrate. a) Who created the SDS? b) What is the solubility of sodium nitrate in water? c) Is sodium nitrate carcinogenic? d) What is the LD50 oral rat? e) How much sodium nitrate would you have to ingest to give a 50% chance of death (estimate from available LD50 data). f) How much of a 1 M solution would you have to ingest to give a 50% chance of death? g) Are there any chronic effects of exposure to sodium nitrate?
5. You are in the laboratory preparing chemical solutions for an experiment and it is lunchtime. You decide to go to CTB to eat. What must you do before leaving the laboratory?

CHAPTER
FOUR

LABORATORY MEASUREMENTS AND PROCEDURES

4.1 Introduction

Measurements of masses, volumes, and preparation of chemical solutions of known composition are essential laboratory skills. The goal of this exercise is to gain familiarity with these laboratory procedures. You will use these skills repeatedly throughout the semester.

4.2 Theory

Many laboratory procedures require preparation of chemical solutions. Most chemical solutions are prepared on the basis of mass of solute per volume of solution (grams per liter or moles per liter). Preparation of these chemical solutions requires the ability to accurately measure both mass and volume.

Preparation of dilutions is also frequently required. Many analytical techniques require the preparation of known standards. Standards are generally prepared with concentrations similar to that of the samples being analyzed. In environmental work many of the analyses are for hazardous substances at very low concentrations (mg/L or $\mu\text{g}/\text{L}$ levels). It is difficult to accurately weigh a few milligrams of a chemical with an analytical balance. Often dry chemicals are in crystalline or granular form with each crystal weighing several milligrams making it difficult to get close to the desired weight. Thus it is often easier to prepare a low concentration standard by diluting a higher concentration stock solution. For example, 100 mL of a 10 mg/L solution of NaCl could be obtained by first preparing a 1 g/L NaCl solution (100 mg in 100 mL). One mL of the 1 g/L stock solution would then be diluted to 100 mL to obtain a 10 mg/L solution.

Absorption spectroscopy is one analytical technique that can be used to measure the concentration of a compound. Solutions that are colored absorb light in the visible range. The resulting color of the solution is from the light that is transmitted. According to Beers law the attenuation of light in a chemical solution is related to the concentration and the length of the path that the light passes through.

$$\log \left(\frac{P_o}{P} \right) = \varepsilon bc \quad (4.1)$$

where c is the concentration of the chemical species, b is the distance the light travels through the solution, ε is a constant, P_o is the intensity of the incident light, and P is the intensity of the transmitted light. Absorption, A , is defined as:

$$A = \log \left(\frac{P_o}{P} \right) \quad (4.2)$$

In practice P_0 is the intensity of light through a reference sample (such as deionized water) and thus accounts for any losses in the walls of the sample chamber. From equation (4.1) and (4.2) it may be seen that absorption is directly proportional to the concentration of the chemical species.

$$A = \varepsilon bc \quad (4.3)$$

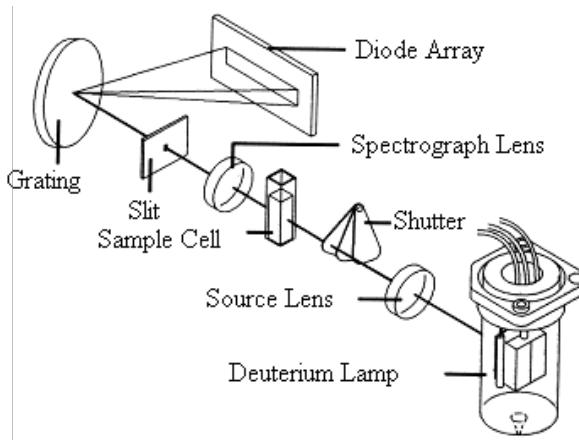


Fig. 4.1: Diagram of light path in diode array spectrophotometer

One instrument you may use to measure absorbance is a Hewlett Packard (HP) model 8452A diode array spectrophotometer. The diode array spectrophotometer uses a broad-spectrum source of incident light from a deuterium lamp. The light passes through the sample, 1 cm path length, and is split by a grating into a spectrum of light that is measured by an array of diodes. Each diode measures a bandwidth of 2 nm with 316 diodes covering the range from 190 nm to 820 nm. The wavelengths of light and their colors are given in Table 2- #. The light path for the diode array spectrophotometer is shown in Fig. 4.1.

The HP 8452A spectrophotometer has a photometric range of 0.002 - 3.3 absorbance units. In practice absorbance measurements greater than 2.5 are not very meaningful as they indicate that 99.7% of the incident light at that wavelength was absorbed. Conversely, an absorbance of 0.002 means that 0.5% of the incident light at that wavelength was absorbed.

4.3 Experimental Objectives

To gain proficiency in:

1. Calibrating and using electronic balances
2. Using signal conditioning boxes and data acquisition software
3. Digital pipetting
4. Preparing a solution of known concentration
5. Preparing dilutions
6. Measuring concentrations using a UV-Vis spectrophotometer

4.4 Experimental Methods

4.4.1 Mass Measurements

Mass can be accurately measured with an electronic analytical balance. Perhaps because balances are so easy to use it is easy to forget that they should be calibrated on a regular basis. It is recommended that balances be calibrated once a week, after the balance has been moved, or if excessive temperature variations have occurred. In order for balances

to operate correctly they also need to be level. Most balances come with a bubble level and adjustable feet. Before calibrating a balance verify that the balance is level.

The environmental laboratory is equipped with 200 g balances. As part of this exercise, we will calibrate the 200 g as follows:

1. Start with the balance off.
2. Press and hold the ON/ZERO key until the screen reads MENU.
3. Releasing the ON/ZERO key will take you to calibration mode indicated by .C.A.L. on the screen.
4. Press the ON/ZERO key to indicate Yes to calibration.
5. The balance will acquire the zero value (and read – C –).
6. Once the zero value is obtained, the balance screen will blink – 200 g – indicating that the 200 g mass will be used for the calibration.
7. Place the 200 g calibration mass on the pan (handle the calibration mass using a cotton glove or tissue paper) and press ON/ZERO key.
8. The balance will calibrate to the mass added. Remove the mass when the screen reads 200.00 g indicating the calibration is complete.
9. Measure the mass of a second calibration mass of different size (e.g., 100 g) to confirm calibration.
10. Record relevant data in the attached spreadsheet.

Dry chemicals can be weighed in disposable plastic weighing boats or other suitable containers. It is often desirable to subtract the weight of the container in which the chemical is being weighed. The weight of the chemical can be obtained either by weighing the container first and then subtracting, or by zeroing the balance with the container on the balance.

4.4.2 Temperature Measurement and ProCoDA

We will use a data acquisition system designed and fabricated in CEE at Cornell University. Each group has their own ProCoDA box and associated power supply and USB cable. The power supply and USB cable must be plugged into the ProCoDA box and then into the AC power on your lab bench and a USB port on your lab bench computer, respectively.

Use a thermistor to measure the temperature of distilled water. The thermistors are usually hanging on the rack to the right of the fume hoods (you should have one on your bench today). The thermistor has a 4-mm diameter metallic probe. Plug the thermistor into the red signal-conditioning box. The conditioned signal is connected to the ProCoDA box using a red cable. Connect the red cable to one of the sensor ports on the top row of the ProCoDA box.

1. Open ProCoDA II and *configure temperature monitoring*
2. Place the temperature probe in a 100-mL plastic beaker full of distilled water. Wait at least 15 seconds to allow the probe to equilibrate with the solution.
3. Record this temperature in the attached spreadsheet.

4.4.3 Pipette Technique

1. Use Fig. 4.2 or `density = pc.density_water(Temp)` to estimate the mass of 990 μL of distilled water (at the measured temperature).
2. Use a 100-1000 μL digital pipette to transfer 990 μL of distilled water to a tared weighing boat on a balance with mg resolution. Record the mass of the water and compare with the expected value (see Fig. 4.2). Repeat this step if necessary until your pipetting error is less than 2%, then measure the mass of 5 replicate 990 μL pipette

samples. Calculate the mean (\bar{x}), standard deviation (s), and coefficient of variation, $\frac{s}{\bar{x}}$, for your measurements. The coefficient of variation (c.v.) is a good measure of the precision of your technique. For this test a c.v. < 1% should be achievable.

4.4.4 Measure Density

1. Weigh a 100 mL volumetric flask with its cap (use 200 g balance with resolution of 0.001 g).
2. Prepare 100 mL of a 1 M solution of sodium chloride in the weighed flask. You can also dissolve the NaCl in a clean beaker and transfer to the volumetric flask. Make sure to mix the solution and then verify that you have **exactly 100 mL** of solution. Note that the combined **volume of NaCl and water decreases** as the salt dissolves.
3. Weigh the flask (with its cap) plus the sodium chloride solution and calculate the density of the 1 M NaCl solution.

```
""" importing """
from aide_design.play import*
Temp = np.linspace(0,30)*u.degC
density = pc.density_water(Temp)
fig, ax = plt.subplots()
ax.plot(Temp,density)
ax.set(xlabel='Temperature (degrees Celsius)', ylabel=r'Density of water (kg/$m^3$)')
fig.savefig('Laboratory_Measurements/Images/Density_vs_T')
plt.show()
```

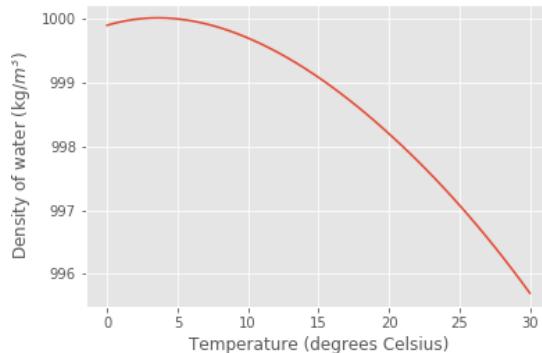


Fig. 4.2: Density of water vs. temperature.

4.4.5 Prepare red dye standards of several concentrations

A red dye stock solution of 10 g/L has been prepared.

1. Use the red dye stock solution to prepare 100 mL of each of the following concentrations: 1 mg/L, 2 mg/L, 5 mg/L, 10 mg/L, 20 mg/L, 50 mg/L, 100 mg/L, and possibly 200 mg/L. Record your calculations in the attached spreadsheet. Use pipettes and volumetric flasks to create accurate dilutions.
2. Note any errors in transfer of mass as you prepare these dilutions (the color will make it easy to see). Make sure to transfer every drop!

4.4.6 Create a standard curve and measure an unknown

1. Create a calibration curve using the standards created above and the *photometer calibration method*.   
 2. Make sure to both save AND export the calibration data from ProCoDA (   - 3. Was the calibration linear? If not, which standards caused it to depart from linearity? #. The high concentration standards may be beyond the linear range for the sensor. This can occur if the amount of light reaching the sensor is too low to create an output voltage that is proportional to the light. Remember that an absorbance of 2 means that 99% of the light is adsorbed by the dye! If the absorbance for the highest standard isn't in the line of the lower concentration samples, then delete the highest standards sequentially until you get a high correlation coefficient (R) as calculated by the ProCoDA photometer software. #. If the correlation coefficient is less than 0.99 then it suggests that your standards weren't accurately prepared. See if you can identify what you did incorrectly with your pipette or dilution technique and consider (talk with a TA) preparing new standards.
4. Save AND export the calibration data from ProCoDA (  

4.5 Prelab Questions

1. You need 100 mL of a $1 \mu M$ solution of zinc that you will use as a standard to calibrate an atomic absorption spectrophotometer. Find a source of zinc ions combined either with chloride or nitrate (you can use the internet or any other source of information). What is the molecular formula of the compound that you found? Zinc disposal down the sanitary sewer is restricted at Cornell and the solutions you prepare may need to be disposed of as hazardous waste. As an environmental engineering student you strive to minimize waste production. How would you prepare this standard using techniques readily available in the environmental laboratory so that you minimize the production of solutions that you don't need? Note that we have pipettes that can dispense volumes between $10 \mu L$ and 1 mL and that we have 100 mL and 1 L volumetric flasks. Include enough information so that you could prepare the standard without doing any additional calculations. Consider your ability to accurately weigh small masses. Explain your procedure for any dilutions. Note that the stock solution concentration should be an easy multiple of your desired solution concentration so you don't have to attempt to pipette a volume that the digital pipettes can't be set for such as $13.6 \mu L$.
2. The density of sodium chloride solutions as a function of concentration is approximately $0.6985C + \rho_{water}$. What is the density of a 1 M solution of sodium chloride?

4.6 Data Analysis and Questions

Submit one spreadsheet containing the data sheet, exported absorbance data, graphs and answers to the questions.

1. Fill out the Excel data sheet available from the course syllabus. Make sure that all calculated values are entered in the spreadsheet as equations. Failure to use the spreadsheet to do the calculations will not receive full credit. Note that this is likely the only assignment that we will do using Excel. All remaining analysis for the course will be done in Atom using Python!
2. Create a graph of absorbance vs. concentration of red dye \#40 in Atom using the exported data file. Does absorbance increase linearly with concentration of the red dye?
3. What is the value of the extinction coefficient, ϵ ?
4. Did you use interpolation or extrapolation to get the concentration of the unknown?

5. What measurement controls the accuracy of the density measurement for the NaCl solution?
6. What density did you expect (see prelab 2)?
7. Approximately what should the accuracy be?
8. Dont forget to write a brief paragraph on conclusions and on suggestions using Markdown.
9. Verify that your report and graphs meet the requirements as outlined in the course materials.

4.7 Lab Prep Notes

4.8 Setup

1. Prepare stock red dye #40 solution (10 g/L) and distribute to student workstations in 20 mL vials.
2. Prepare 1 L of unknown in concentration range of standards. Divide into six 150 mL bottles (one for each student bench (teams/2)).
3. Verify that balances calibrate easily.
4. Disassemble, clean, and lubricate all pipettes.
5. Distribute all supplies needed for the lab.

CHAPTER
FIVE

ACID PRECIPITATION AND REMEDIATION OF ACID LAKES

5.1 Introduction

Acid precipitation has been a serious environmental problem in many areas of the world for the last few decades. Acid precipitation results from the combustion of fossil fuels which produce oxides of sulfur and nitrogen that react in the earth's atmosphere to form sulfuric and nitric acid. One of the most significant impacts of acid rain is the acidification of lakes and streams. In some watersheds the soil doesn't provide ample acid neutralizing capacity to mitigate the effect of incident acid precipitation. These susceptible regions are usually high elevation lakes with small watersheds and shallow non-calcareous soils. The underlying bedrock of acid-sensitive lakes tends to be granite or quartz. These minerals are slow to weather and therefore have little capacity to neutralize acids. The relatively short contact time between the acid precipitation and the watershed soil system exacerbates the problem. Lakes most susceptible to acidification: 1) are located downwind, sometimes hundreds of miles downwind, from major pollution sources—electricity generation, metal refining operations, heavy industry, large population centers, etc.; 2) are surrounded by hard, insoluble bedrock with thin, sandy, infertile soil; 3) have a high runoff to infiltration ratio; 4) have a low watershed to lake surface area. Isopleths of precipitation pH are depicted in Fig. 5.1.



Fig. 5.1: The pH of precipitation in 2009.

In acid-sensitive lakes the major parameter of concern is pH ($pH = -\log\{H^+\}$), where $\{H^+\}$ is the hydrogen ion activity, and activity is approximately equal to concentration in moles/L). In a healthy lake, ecosystem pH should be in the range of 6.5 to 8.5. In most natural freshwater systems, the dominant pH buffering (controlling) system is the carbonate system. The carbonate buffering system is composed of four components: dissolved carbon dioxide ($CO_2 \text{ }_{aq}$), carbonic acid (H_2CO_3), bicarbonate (HCO_3^-), and carbonate (CO_3^{2-}). Carbonic acid exists only at very low levels in aqueous systems and for purposes of acid neutralization is indistinguishable from dissolved carbon dioxide. Thus to simplify things we define

$$[H_2CO_3^*] = [CO_2 \text{ }_{aq}] + [H_2CO_3] \quad (5.1)$$

The $[CO_2]_{aq} >> [H_2CO_3]$ and thus $[H_2CO_3^*] \cong [CO_2]_{aq}$ (all terms enclosed in [] are in units of moles/L).

The sum of the molar concentrations of all the components of the carbonate system is designated as C_T as shown in the equation below.

$$C_T = [H_2CO_3^*] + [HCO_3^-] + [CO_3^{2-}] \quad (5.2)$$

The carbonate system can be considered to be a volatile system or a non-volatile system depending on whether or not aqueous carbon dioxide is allowed to exchange and equilibrate with atmospheric carbon dioxide. Mixing conditions and hydraulic residence time determine whether an aquatic system is volatile or non-volatile relative to atmospheric carbon dioxide equilibrium. First, consider the non-volatile system.

5.1.1 Non-volatile System

For a fixed C_T , the molar concentration of each species of the carbonate system is determined by pH. Equations (5.3)-(5.8) show these functional relationships.

$$[H_2CO_3^*] = \frac{C_T}{1 + \frac{K_1}{[H^+]} + \frac{K_1 K_2}{[H^+]^2}} = \alpha_0 C_T \quad (5.3)$$

where

$$\alpha_0 = \frac{1}{1 + \frac{K_1}{[H^+]} + \frac{K_1 K_2}{[H^+]^2}} \quad (5.4)$$

$$[HCO_3^-] = \frac{C_T}{\frac{[H^+]}{K_1} + 1 + \frac{K_2}{[H^+]}} = \alpha_1 C_T \quad (5.5)$$

where

$$\alpha_1 = \frac{1}{\frac{[H^+]}{K_1} + 1 + \frac{K_2}{[H^+]}} \quad (5.6)$$

$$[CO_3^{2-}] = \frac{C_T}{\frac{[H^+]^2}{K_1 K_2} + \frac{[H^+]}{K_2} + 1} = \alpha_2 C_T \quad (5.7)$$

where

$$\alpha_2 = \frac{1}{\frac{[H^+]^2}{K_1 K_2} + \frac{[H^+]}{K_2} + 1} \quad (5.8)$$

K_1 and K_2 are the first and second dissociation constants for carbonic acid and α_0 , α_1 , and α_2 are the fraction of C_T in the form $H_2CO_3^*$, HCO_3^- , and CO_3^{2-} respectively. Because K_1 and K_2 are constants ($K_1 = 10^{-6.3}$ and $K_2 = 10^{-10.3}$), α_0 , α_1 , and α_2 are only functions of pH.

A measure of the susceptibility of lakes to acidification is the acid neutralizing capacity (ANC) of the lake water. In the case of the carbonate system, the ANC is exhausted when enough acid has been added to convert the carbonate species HCO_3^- and CO_3^{2-} to $H_2CO_3^*$. A formal definition of total acid neutralizing capacity is given by equation (5.9)

$$ANC = [HCO_3^-] + 2[CO_3^{2-}] + [OH^-] - [H^+] \quad (5.9)$$

ANC has units of equivalents per liter. The hydroxide ion concentration can be obtained from the hydrogen ion concentration and the dissociation constant for water, K_w .

$$[OH^-] = \frac{K_w}{[H^+]} \quad (5.10)$$

Substituting equations (5.5), (5.7), and (5.10) into equation (5.9), we obtain

$$ANC = C_T (\alpha_1 + 2\alpha_2) + \frac{K_w}{[H^+]} - [H^+] \quad (5.11)$$

For the carbonate system, ANC is usually referred to as alkalinity. Alkalinity can be expressed as equivalents/L or as mg/L (ppm) of $CaCO_3$. 50,000 mg/L $CaCO_3$ = 1 equivalent/L.

5.1.2 Volatile Systems

Now consider the case where aqueous CO_2 is volatile and in equilibrium with atmospheric carbon dioxide. Henrys Law can be used to describe the equilibrium relationship between atmospheric and dissolved carbon dioxide.

$$[CO_2]_{aq} = P_{CO_2} K_H \quad (5.12)$$

where K_H is Henrys constant for CO_2 in moles/L-atm and P_{CO_2} is partial pressure of CO_2 in the atmosphere ($K_H = 10^{-1.5}$ and $P_{CO_2} = 10^{-3.5}$). Because $[CO_2]_{aq}$ is approximately equal to $[H_2CO_3^*]$ and from equations (5.1) and (5.3)

$$P_{CO_2} K_H = \alpha_0 C_T \quad (5.13)$$

$$C_T = \frac{P_{CO_2} K_H}{\alpha_0} \quad (5.14)$$

Equation (5.14) gives the equilibrium concentration of carbonate species as a function of pH and the partial pressure of carbon dioxide.

The acid neutralizing capacity expression for a volatile system can be obtained by combining equations (5.14) and (5.11).

$$ANC = \frac{P_{CO_2} K_H}{\alpha_0} (\alpha_1 + 2\alpha_2) + \frac{K_w}{[H^+]} - [H^+] \quad (5.15)$$

In both non-volatile and volatile systems, equilibrium pH is controlled by system ANC. Addition or depletion of any ANC component in equation (5.11) or (5.15) will result in a pH change. Natural bodies of water are most likely to approach equilibrium with the atmosphere (volatile system) if the hydraulic residence time is long and the body of water is shallow.

Lake ANC is a direct reflection of the mineral composition of the watershed. Lake watersheds with hard, insoluble minerals yield lakes with low ANC. Typically watersheds with soluble, calcareous minerals yield lakes with high ANC. ANC of freshwater lakes is generally composed of bicarbonate, carbonate, and sometimes organic matter (A_{org}^-). Organic matter derives from decaying plant matter in the watershed. When organic matter is significant, the ANC becomes (from equations (5.11) and (5.15)):

$$ANC = C_T (\alpha_1 + 2\alpha_2) + \frac{K_w}{[H^+]} - [H^+] + [A_{org}^-] \quad (5.16)$$

$$ANC = \frac{P_{CO_2} K_H}{\alpha_0} (\alpha_1 + 2\alpha_2) + \frac{K_w}{[H^+]} - [H^+] + [A_{org}^-] \quad (5.17)$$

where equation (5.16) is for a non-volatile system and equation (5.17) is for a volatile system.

During chemical neutralization of acid, the components of ANC associate with added acid to form protonated molecules. For example:



or



In essence, the ANC of a system is a result of the reaction of acid inputs to form associated acids from basic anions that were dissolved in the lake water. The ANC (equation (5.9)) is consumed as the basic anions are converted to associated acids. This conversion is near completion at low pH (approximately pH 4.5 for the bicarbonate and carbonate components of ANC). Neutralizing capacity to another (probably higher) pH may be more useful for natural aquatic systems. Determination of ANC to a particular pH is fundamentally easy simply add and measure the amount of acid required to lower the sample pH from its initial value to the pH of interest. Techniques to measure ANC are described under the procedures section of this lab.

Neutralization of acid precipitation can occur in the watershed or directly in the lake. How much neutralization occurs in the watershed versus the lake is a function of the watershed to lake surface area. Generally, watershed neutralization is dominant. Engineered remediation of acid lakes has been accomplished by adding bases such as limestone, lime, or sodium bicarbonate to the watershed or directly to the lakes.

5.2 Reactor Theory Applied to Acid Lake Remediation

In this experiment sodium bicarbonate will be added to a lake to mitigate the deleterious effect of acid rain. Usually sodium bicarbonate is added in batch doses (as opposed to metering in). The quantity of sodium bicarbonate added depends on how long a treatment is desired, the acceptable pH range and the quantity and pH of the incident rainfall. For purposes of this experiment, a 15-minute design period will be used. That is, we would like to add enough sodium bicarbonate to keep the lake at or above its original pH and alkalinity for a period of 15 minutes (i.e., for one hydraulic residence time).

By dealing with ANC instead of pH as a design parameter, we avoid the issue of whether the system is at equilibrium with atmospheric carbon dioxide. Keep in mind that eventually the lake will come to equilibrium with the atmosphere. In practice, neutralizing agent dosages may have to be adjusted to take into account non-equilibrium conditions.

We must add enough sodium bicarbonate to equal the negative ANC from the acid precipitation input plus the amount of ANC lost by outflow from the lake during the 15-minute design period. Initially (following the dosing of sodium bicarbonate) the pH and ANC will rise, but over the course of 15 minutes, both parameters will drop. Calculation of required sodium bicarbonate dosage requires performing a mass balance on ANC around the lake. This mass balance will assume a completely mixed lake and conservation of ANC. Chemical equilibrium can also be assumed so that the sodium bicarbonate is assumed to react immediately with the incoming acid precipitation. Mass balance on the reactor yields:

$$Q(ANC_{in} - ANC_{out})V \frac{d(ANC)}{dt} \quad (5.20)$$

where:

ANC_{out} = ANC in lake outflow at any time t (for a completely mixed lake the effluent ANC is the same as the ANC in the lake)

ANC_{in} = ANC of acid rain input

V = volume of reactor

Q = acid rain input flow rate.

If the initial ANC in the lake is designated as ANC_0 , then the solution to the mass balance differential equation is:

$$ANC_{out} = ANC_{in} \cdot \left(1 - e^{-t/\theta}\right) + ANC_0 \cdot e^{-t/\theta} \quad (5.21)$$

where:

$$\theta = V/Q$$

We want to find ANC_0 such that $ANC_{out} = 50 \mu eq/L$ when t is equal to θ . Solving for ANC_0 we get:

$$ANC_0 = \left[ANC_{out} - ANC_{in} \cdot \left(1 - e^{-t/\theta}\right) \right] e^{t/\theta} \quad (5.22)$$

The ANC of the acid rain (ANC_{in}) can be estimated from its pH. Below pH 6.3 most of the carbonates will be in the form $H_2CO_3^*$ and thus for pH below about 4.3 equation (5.9) simplifies to

$$ANC \cong -[H^+] \quad (5.23)$$

An influent pH of 3.0 implies the $ANC_{in} = -[H^+] = -0.001\text{eq/L}$

Substituting into equation (5.22):

$$ANC_0 = [0.000050 + 0.001 \cdot (1 - e^{-1})] e^1 = 1.854\text{meq/L} \quad (5.24)$$

The quantity of sodium bicarbonate required can be calculated from:

$$[NaHCO_3]_0 = ANC_0 \quad (5.25)$$

where $[NaHCO_3]_0$ = moles of sodium bicarbonate required per liter of lake water

$$\frac{1.854 \text{ mmole } NaHCO_3}{\text{liter}} \times \frac{84 \text{ mg } NaHCO_3}{\text{mmole } NaHCO_3} \times 4 \text{ Liters} = 623 \text{ mg } NaHCO_3 \quad (5.26)$$

5.3 Experimental Objectives

Remediation of acid lakes involves addition of ANC so that the pH is raised to an acceptable level and maintained at or above this level for some design period. In this experiment sodium bicarbonate ($NaHCO_3$) will be used as the ANC supplement. Since ANC addition usually occurs as a batch addition, the design pH is initially exceeded. ANC dosage is selected so that at the end of the design period pH is at the acceptable level. Care must be taken to avoid excessive initial pH. High pH can be as deleterious as low pH.

The most common remediation procedure is to apply the neutralizing agent directly to the lake surface, instead of on the watershed. We will follow that practice in this lab experiment. Sodium bicarbonate will be added directly to the surface of the lake that has an initial ANC of $0\mu\text{eq/L}$ and is receiving acid rain with a pH of 3. After the sodium bicarbonate is applied, the lake pH and ANC will be monitored for over two approximately 20 minute periods.

5.4 Experimental Apparatus

The experimental apparatus consists of an acid rain storage reservoir, peristaltic pump, and lake (Fig. 5.2). The pH of the lake will be monitored using a pH probe connected to a signal-conditioning box that is connected to ProCoDA.

5.5 Procedures

The following directions are written for the use of ProCoDA II hardware and software for pH data collection and manual control of the peristaltic pump. It would also be possible to use automate the experiment and control the pump using the ProCoDA II hardware and software.

We will use a pH probe to measure pH in this experiment. The pH probes are stored in a small plastic box. Each bench has one pH probe. Plug the pH probe into the blue signal-conditioning box (it takes a push and a twist). Connect the cable to one of the sensor ports on your ProCoDA box.

1. *Setup and calibrate the pH probe*
2. Verify that the experimental setup is plumbed so that the acid rain is pumped directly into the lake. The lake outflow should discharge into the small drain on the side of your work bench.



Fig. 5.2: Schematic drawing of the experimental setup.

3. Organize the bench setup so that the metal tube discharging the acid rain into the lake is solidly touching the metal stand that is connected to the stirrer. This will ground the solution that is in the lake and reduce voltage fluctuations that are easily measured by the pH probe.
4. Preset pump to give desired flow rate of 267 mL/min (4 L/15 minutes) based on the size of pump tubing selected. Do not turn the pump on yet! For each tubing size, different pump speeds will correspond to different flow rates being output by the pump. The peristaltic tubing sizes are rather arbitrary and are labeled by numbers: 13, 14, 16, 17, and 18 in increasing order of size. If you have #18 tubing, you will want an RPM setting of $(267 \text{ mL/min}) / (3.8 \text{ mL/rev}) = 70.3 \text{ RPM}$ (see Table 5.1).
5. Fill lake with reverse osmosis water and verify that the outflow is set so the lake volume is approximately 4 L. Place the lake on top of a magnetic stirrer and add a stir bar.
6. Set stirrer speed to 8.
7. Add 1 mL of bromocresol green indicator solution to the lake.
8. Weigh out 623 mg (not grams!) NaHCO_3 .
9. Add NaHCO_3 to the lake.
10. After the lake is well stirred take a 100 mL sample from the lake in the plastic sample bottle on your bench. Dont forget to label the sample bottle (include the time of the sample).
11. Clip the pH probe to the side of your lake in a more quiescent zone, away from the influent and effluent.
12. We will continuously measure the pH of the effluent and log the data into a tab delimited file. Set the data



interval to 1 second. Begin logging data to file by clicking on the  button. Create a new file in S:\Courses\4530\Group #\Lab 2 - Acid Rain.

13. Prepare to write a comment in the file to mark the time when the pump starts by clicking on the  button. Type in a comment and then wait.
14. At time equal zero ($t=0$) start the peristaltic pump and click on the enter button in the comment dialog box.
15. Take 100-mL grab samples from the lake effluent at 5, 10, 15, and 20 minutes in the plastic sample bottle on your bench. Dont forget to label the sample bottle (include the time of the sample). The sample volumes do not

need to be measured exactly.

16. After the 20-minute sample, measure the flow rate by collecting effluent in a beaker for 30 seconds and measuring the volume collected (in a graduated cylinder for more accurate measurement).
17. Turn off the pump and stop measuring pH.
18. Measure the lake volume. This can be done in a large graduated cylinder OR by taking the mass of the water in the lake. Which would be more accurate?
19. Repeat the experiment and change one of the following parameters: stirring, initial ANC, ANC source (use $CaCO_3$ instead of $NaHCO_3$), or amount of ANC added.

Table 5.1: Pump tubing selection.

	Tubing Size	13	14	16	17	18
RPM/ID (mm)	0.8	1.6	3	6.3	8	
flow	1	0.0010	0.0035	0.0133	0.0467	0.0633
rate	50	0.0500	0.1750	0.6667	2.3333	3.1667
in	100	0.1000	0.3500	1.3333	4.6667	6.3333
mL/s	mL/rev	0.06	0.21	0.80	2.8	3.8

5.6 pH Measurement

pH. pH ($-\log \{H^+\}$) is usually measured electrometrically with a pH meter. The pH meter is a null-point potentiometer that measures the potential difference between an indicator electrode and a reference electrode. The two electrodes commonly used for pH measurement are the glass electrode and a reference electrode. The glass electrode is an indicator electrode that develops a potential across a glass membrane as a function of the activity (\sim molarity) of H^+ . Combination pH electrodes, in which the H^+ -sensitive and reference electrodes are combined within a single electrode body will be used in this lab. The reference electrode portion of a combination pH electrode is a $[Ag/AgCl/4M KCl]$ reference. The response (output voltage) of the electrode follows a Nernstian behavior with respect to H^+ ion activity.

$$E = E^0 + \frac{RT}{nF} \ln \left(\frac{[H^+]}{[H^{+0}]} \right) \quad (5.27)$$

where

R is the universal gas constant

T is temperature in Kelvin

n is the charge of the hydrogen ion,

F is the Faraday constant.

E^0 is the calibration potential (Volts),

E is the potential (Volts) measured by the pH meter between glass and reference electrode. The slope of the response curve is dependent on the temperature of the sample and this effect is normally accounted for with simultaneous temperature measurements.

The electrical potential that is developed between the glass electrode and the reference electrode needs to be correlated with the actual pH of the sample. The pH meter is calibrated with a series of buffer solutions whose pH values encompass the range of intended use. The pH meter is used to adjust the response of the electrode system to ensure a Nernstian response is achieved over the range of the calibration standards.

To measure pH the electrode(s) are submersed in at least 50 mL of a sample. Samples are generally stirred during pH reading to establish homogeneity, to prevent local accumulation of reference electrode filling solution at the interface near the electrode, and to ensure the diffusive boundary layer thickness at the electrode surface is uniform and small.

ANC. The most common method to determine ANC for aqueous samples is titration with a strong acid to an endpoint pH. A pH meter is usually used to determine the endpoint or equivalence point of an ANC titration. Determination of the endpoint pH is difficult because it is dependent on the magnitude of the sample ANC. Theoretically this endpoint pH should be the pH where all of the ANC of the system is consumed, but since the ANC is not known *a priori*, a true endpoint cannot be predetermined. However, if most of the ANC is composed of carbonate and bicarbonate this endpoint is approximately pH = 4.5 for a wide range of ANC values.

A 50 to 100-mL sample is usually titrated while slowly stirred by a magnetic stirrer. pH electrodes are ordinarily used to record pH as a function of the volume of strong acid titrant added. The volume of strong acid required to reach the ANC endpoint (pH 4.5) is called the equivalent volume and is used in the following equation to compute ANC.

$$ANC = \frac{(equivalent\ vol.)(normality\ of\ titrant)}{(vol.\ of\ sample)} \quad (5.28)$$

A more accurate technique to measure ANC is the Gran plot analysis. This is the subject of next weeks analysis. We will directly measure the ANC of the samples that were taken at t=0, 5, 10, 15, and 20 minutes in both of your experiments by means of a Gran plot analysis.

5.7 Prelab Questions

- How many grams of $NaHCO_3$ would be required to keep the ANC levels in a lake above $50 \mu eq/L$ for 3 hydraulic residence times given an influent pH of 3.0 and a lake volume of 4 L, if the current lake ANC is $0 \mu eq/L$?

5.8 Data Analysis

$$K_1 = 10^{-6.3}, K_2 = 10^{-10.3}, K_H = 10^{-1.5} \frac{mol}{Latm}, P_{CO_2} = 10^{-3.5} atm, \text{ and } K_w = 10^{-14}.$$

- Plot measured pH of the lake versus dimensionless hydraulic residence time (t/θ).
- Assuming that the lake can be modeled as a completely mixed flow reactor and that ANC is a conservative parameter, equation (5.22) can be used to calculate the expected ANC in the lake effluent as the experiment proceeds. Graph the expected ANC in the lake effluent versus the hydraulic residence time (t/θ) based on the completely mixed flow reactor equation with the plot labeled (in the legend) as conservative ANC.
- If we assume that there are no carbonates exchanged with the atmosphere during the experiment, then we can calculate ANC in the lake effluent by using equation (5.11) describing the ANC of a closed system. Calculate the ANC under the assumption of a closed system and plot it on the same graph produced in answering question #3 with the plot labeled (in the legend) as closed ANC.
- If we assume that there is exchange with the atmosphere and that carbonates are at equilibrium with the atmosphere, then we can calculate ANC in the lake effluent by using equation (5.15) describing the ANC of an open system. Calculate the ANC under the assumption of an open system and plot it on the same graph produced in answering question #3 with the plot labeled (in the legend) as open ANC.
- Analyze the data from the second experiment and graph the data appropriately. What did you learn from the second experiment?

5.9 Questions

- What do you think would happen if enough $NaHCO_3$ were added to the lake to maintain an ANC greater than $50 \mu eq/L$ for 3 residence times with the stirrer turned off? How much $NaHCO_3$ would need to be added?

2. What are some of the complicating factors you might find in attempting to remediate a lake using $CaCO_3$? Below is a list
- extent of mixing
 - solubility of $CaCO_3$ (find the solubility and compare with $NaHCO_3$)
 - density of $CaCO_3$ slurry (find the density of $CaCO_3$)

5.10 References

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5.11 Lab Prep Notes

Table 5.2: Reagent list.

Description	Supplier	Catalog number
HCL 5.0 N	Fisher Scientific	LC15360-2
H ₂ SO ₄ 5N	Fisher Scientific	LC25840-2
CaCO ₃	Fisher Scientific	C63-3
Na ₂ CO ₃	Fisher Scientific	S263-500
Buffer-Pac	Fisher Scientific	SB105
NaHCO ₃	Fisher Scientific	S233-500
Bromocresol Green	Fisher Scientific	B383-5
ethanol	Fisher Scientific	A962P-4

5.11.1 Bromocresol Green Indicating Solution

Prepare solution of 400 mg Bromocresol green/100 mL ethanol. Add 0.2 mL of indicator solution per liter of acid rain or lake.

5.11.2 Acid rain

Acid rain is at pH 3.0. Prepare from reverse osmosis water. Add 1 meq H_2SO_4/L ($[H^+]$ at pH 3.0) to obtain a pH of 3.0. To acidify 20 liters of distilled water using 10 N H_2SO_4 :

$$20 \text{ L} \bullet \frac{1 \text{ meq } H_2SO_4}{L} \bullet \frac{1}{10 \text{ N } H_2SO_4} \bullet \frac{1 \text{ N}}{1000 \text{ meq}} = 2 \text{ mL of } 10 \text{ N } H_2SO_4 \quad (5.29)$$

5.11.3 Flow Rate

The residence time of the lake should be 15 minutes. The lake volume is 4 L. thus the flow rate is 267 mL/min. Use # 18 PharMed tubing.

5.12 Setup

1. Prepare 20-L acid rain for each group.
2. Prepare bromocresol green solution if necessary.
3. Attach one Easy-Load pump head to the pump drives and plumb with #18 tubing.
4. Plumb Jerrican to pump to lake using quick connectors (see Fig. 5.2).
5. Verify that pH probes are operational, stable, and can be calibrated.
6.)Verify that buffers (pH = 4, 7, 10) are distributed to each student group.
7. Provide a effluent cup in which pH can be measured.

CHAPTER
SIX

ACID NEUTRALIZING CAPACITY

6.1 Introduction

Acid neutralizing capacity (ANC) is a measure of the ability of water to neutralize acid inputs. Lakes with high ANC (such as Cayuga Lake) can maintain a neutral pH even with some acid rain input whereas lakes with an ANC less than the acid input will not maintain a neutral pH. In the Adirondack region of New York State, lakes typically receive large inputs of acids during the spring thaw when the accumulated winter snow melts and runs off into the lakes. The ANC of Adirondack lakes is not always sufficient to neutralize these inputs.

6.2 Theory

The ANC for a typical carbonate-containing sample is defined as:

$$ANC = [HCO_3^-] + 2[CO_3^{2-}] + [OH^-] - [H^+] \quad (6.1)$$

Equation (6.1) can be derived from a charge balance if ANC is considered to be the cation contributed by a strong base titrant and if other ions present do not contribute significantly.

Determination of ANC or alkalinity involves determination of an equivalence point by titration with a strong acid. The equivalence point is defined as the point in the titration where titrant volume that has been added equals the equivalent volume (V_e). The equivalent volume is defined as:

$$V_e = \frac{V_s \cdot N_s}{N_t} \quad (6.2)$$

where:

N_s = normality (in this case alkalinity or ANC) of sample, equivalents/L

V_s = volume of sample, liters

N_t = normality of titrant, equivalents/L.

The titration procedure involves incrementally adding known volumes of standardized normality strong acid (or base) to a known volume of unknown normality base (or acid). When enough acid (or base) has been added to equal the amount of base (or acid) in the unknown solution we are at the equivalence point. The point at which we add exactly an equivalent or stoichiometric amount of titrant is the equivalence point. Experimentally, the point at which we estimate to be the equivalence point is called the titration endpoint.

There are several methods for determining V_e (or the equivalence point pH) from titration data (titrant volume versus pH). The shape of the titration curve (V_t versus pH) can reveal V_e . It can be shown that one inflection point occurs at $V_t = V_e$. In the case of monoprotic acids, there is only one inflection in the pH range of interest. Therefore, an effective method to find the equivalence volume is to plot the titration curve and find the inflection point. Alternately, plot the first derivative of the titration plot and look for a maximum.

6.2.1 Gran Plot

Another method to find the ANC of an unknown solution is the Gran plot technique. When an ANC determination is being made, titration with a strong acid is used to cancel the initial ANC so that at the equivalence point the sample ANC is zero. The Gran plot technique is based on the fact that further titration will result in an increase in the number of moles of H^+ equal to the number of moles of H^+ added. Thus after the equivalence point has been attained, the number of moles of H^+ added equals the number of moles of H^+ in solution. An equation describing this mass balance is provided as:

$$N_t(V_t - V_e) = (V_s + V_t)[H^+] \quad (6.3)$$

Solving for the hydrogen ion concentration:

$$[H^+] = \frac{N_t(V_t - V_e)}{(V_s + V_t)} \quad (6.4)$$

Equation (6.4) can also be solved directly for the equivalent volume.

$$V_e = V_t - \frac{[H^+](V_s + V_t)}{N_t} \quad (6.5)$$

Equation (6.5) is valid if enough titrant has been added to neutralize the ANC. A better measure of the equivalent volume can be obtained by rearranging equation (6.5) so that linear regression on multiple titrant volume - pH data pairs can be used.

$$\frac{(V_s + V_t)}{V_s}[H^+] = \frac{N_t V_t}{V_s} - \frac{N_t V_e}{V_s} \quad (6.6)$$

We define F1 (First Gran function) as:

$$F_1 = \frac{V_s + V_t}{V_s}[H^+] \quad (6.7)$$

If F_1 is plotted as a function of V_t the result is a straight line with slope $= \frac{N_t}{V_s}$ and abscissa intercept of V_e (Fig. 6.1).

The ANC is readily obtained given the equivalent volume. At the equivalence point:

$$V_s ANC = V_e N_t \quad (6.8)$$

Equation (6.8) can be rearranged to obtain ANC as a function of the equivalent volume.

$$ANC = \frac{V_e N_t}{V_s} \quad (6.9)$$

6.2.2 pH Measurements

The pH can be measured either as activity $\{H^+\}$ as measured approximately by pH meter or molar concentration $([H^+])$. The choice only affects the slope of F1 since $[H^+] = \{H^+\}/\gamma$.

$$F_1 = \frac{V_s + V_t}{V_s}[H^+] = \frac{V_s + V_t}{V_s} \frac{\{H^+\}}{\gamma} = N_t \frac{V_t - V_e}{V_s} \quad (6.10)$$

where γ is the activity correction factor and the slope is N_t/V_0 . If $[H^+]$ concentration is used then

$$F_1 = \frac{V_s + V_t}{V_s}\{H^+\} = \gamma N_t \frac{V_t - V_e}{V_s} \quad (6.11)$$

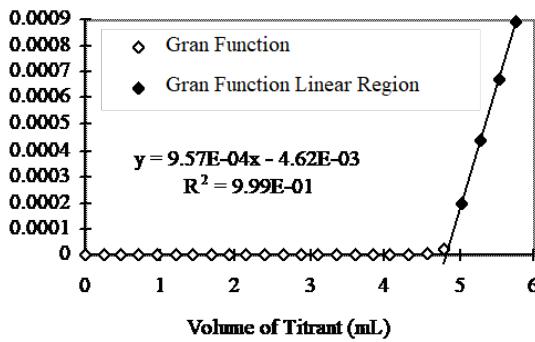


Fig. 6.1: Gran plot from titration of a weak base with 0.05 N acid. $C_T = 0.001$ moles of carbonate and sample volume is 48 mL. The equivalent volume was 4.8 mL. From equation (6.9) the ANC was 5 meq/L.

where the slope is $\frac{\gamma \cdot N_t}{V_s}$.

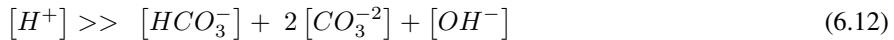
This analysis assumes that the activity correction factor doesn't change appreciably during the titration).

There are many other Gran functions that can be derived. For example, one can be derived for Acidity or the concentration of a single weak or strong acid or base.

To facilitate data generation and subsequent Gran plot construction and analysis pH versus titrant volume can be read directly into a computer, that can be programmed to analyze the data using the Gran plot theory. The program generates the Gran function for all data and then systematically eliminates data until the Gran function (plot) is as linear as possible. The line is then extrapolated to the abscissa to find the equivalent volume.

6.2.3 ANC Determination for Samples with $pH < 4$

After the equivalence point has been reached (adding more acid than ANC = 0) the only significant terms in equation (6.1) are $[H^+]$ and ANC.



When the pH is 2 pH units or more below the lowest pK_a of the bases in the system the only species contributing significantly to ANC is the hydrogen ion (equation (6.12)) and thus the ANC is simply

$$ANC = [H^+] \quad (6.13)$$

For a sample containing only carbonates, if the pH is below 4 the ANC is approximately equal to $-[H^+]$ and no titration is necessary.

6.2.4 Titration Techniques

Operationally, the first few titrant volumes can be relatively large increments since the important data lies at pH values less than that of the equivalence point (approximately pH = 4.5 for an Alkalinity titration). As the pH is lowered by addition of acid the ionic strength of the solution increases and the activity of the hydrogen ion deviates from the hydrogen ion concentration. This effect is significant below pH 3 and thus the effective linear range is generally between pH 4.5 and pH 3.0. The maximum incremental titrant volume (ΔV_a) that will yield n points in this linear region is obtained as follows.

If $V_s \gg V_t$ then equation (6.3) reduces to

$$N_t \cdot \frac{(V_t - V_e)}{V_s} \cong [H^+] \quad (6.14)$$

Let $[H^+]_e$ be the concentration of hydrogen ions at the equivalence point and $[H^+]_f$ be the final concentration of hydrogen ions at the end of the titration.

$$N_t \frac{(V_e - V_e) - (V_f - V_e)}{V_s} = [H^+]_e [H^+]_f \quad (6.15)$$

Thus the volume of acid added to go from $[H^+]_e$ to $[H^+]_f$ is

$$V_f - V_e = \frac{V_s ([H^+]_f - [H^+]_e)}{N_t} \quad (6.16)$$

To obtain n data points between $[H^+]_e$ - $[H^+]_f$ requires the incremental titrant volume (ΔV_t) be 1/n times the volume of acid added between the equivalence point and the final titrant volume. Thus by substituting $n\Delta V_t$, and typical hydrogen ion concentrations of $[H^+]_e = 10^{-4.5}$ and $[H^+]_f = 10^{-3.0}$ into equation (6.16) the maximum incremental titrant volume is obtained.

$$\Delta V_t \cong \frac{(0.001 - 0.00003)V_s}{n N_t} \cong \frac{0.001V_s}{n N_t} \quad (6.17)$$

6.3 Procedures

6.3.1 Determine ANC of Acid Rain Samples

Determine the ANC for all samples collected from the Acid Lake Remediation lab. Start with 50 mL from the t=0 sample and run through the procedure to learn how the software works. Then repeat the procedure with 50 mL from the t=0 sample and for remaining samples with the goal of making an accurate ANC measurement and creating an accurate titration curve by using 0.100 mL titrant increments throughout the entire titration. Remember that the biggest source of error for this lab will likely be poor pipette techniques.

1. Measure 50 mL of an acid lake sample in a graduated cylinder or using an electronic balance.
2. Add to a 100 mL beaker.
3. Place the beaker on the magnetic stirrer, add a stir bar and stir slowly.
4. Place the pH electrode in the solution.
5. If the initial pH is less than 4.5 no titration is necessary and equation (6.13) can be used to calculate the ANC.
6. Record the initial pH (prior to adding any titrant) and initial sample volume.
7. Analyze the sample using *Gran plot analysis*. Add 0.05 N HCl (the titrant) using a digital pipette in increments of 0.100 mL.
8. Save the Gran data to S:\Courses\4530\Group #\ANC\Gran by selecting . The data will be saved in a tab delimited format. You will use this data to plot a titration curve and to verify that the Gran technique accurately measures the ANC of a sample.
9. Record the ANC and the equivalent volume.

If the error is greater than 2% then check your pipette technique using a balance and then repeat the titration.

6.4 Prelab Questions

1. Compare the ability of Cayuga lake and Wolf pond (an Adirondack lake) to withstand an acid rain runoff event (from snow melt) that results in 20% of the original lake water being replaced by acid rain. The acid rain has a pH of 3.5 and is in equilibrium with the atmosphere. The ANC of Cayuga lake is 1.6 meq/L and the ANC of Wolf Pond is 70 $\mu\text{eq}/\text{L}$. Assume that carbonate species are the primary component of ANC in both lakes, and that they are in equilibrium with the atmosphere. What is the pH of both bodies of water after the acid rain input? Remember that ANC is the conservative parameter (not pH!). Hint: You can use the scipy optimize root finding function called brentq. Scipy cant handle units so the units must be removed using .magnitude.}
2. What is the ANC of a water sample containing only carbonates and a strong acid that is at pH 3.2? This requires that you inspect all of the species in the ANC equation (Equation (6.1)) and determine which species are important.
3. Why is $[H^+]$ not a conserved species?

6.5 Data Analysis

1. Plot the titration curve of the t=0 sample with 0.05 N HCl (plot pH as a function of titrant volume). Label the equivalent volume of titrant. Label the 2 regions of the graph where pH changes slowly with the dominant reaction that is occurring. (Place labels with the chemical reactions on the graph in the pH regions where each reaction is occurring.) Note that in a third region of slow pH change no significant reactions are occurring (added hydrogen ions contribute directly to change in pH).
2. Prepare a Gran plot using the data from the titration curve of the t=0 sample. Use linear regression on the linear region or simply draw a straight line through the linear region of the curve to identify the equivalent volume. Compare your calculation of V_e with that was calculated by ProCoDA.
3. Plot the measured ANC of the lake on the same graph as was used to plot the conservative, volatile, and non-volatile ANC models (see questions 2 to 5 of the Acid Precipitation and Remediation of an Acid Lake lab). Did the measured ANC values agree with the conservative ANC model?

6.6 References

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- Pankow, J.F. textit{Aquatic Chemistry Concepts}, Lewis Publishers (1991).
- Morel, F.M.M. and J.G. Hering textit{Principles and Applications of Aquatic Chemistry} Wiley-Interscience (1993).
- Stumm, W. and J.J. Morgan textit{Aquatic Chemistry} 2nd ed. Wiley Interscience (1981).

6.7 Lab Prep Notes

Table 6.1: Reagent list.

Description	Supplier	Catalog number
HCl 5.0 N	Fisher Scientific	LC15360-2
Buffer-Pac	Fisher Scientific	SB105

6.8 Setup

1. Verify that the pH probes are operational, stable, and can be calibrated.
2. Verify that buffers (pH = 4, 7, 10) are distributed to each student group

REACTOR CHARACTERISTICS

7.1 Introduction

Chemical, biological, and physical processes in nature and in engineered systems usually take place in what we call reactors. Reactors are defined by a real or imaginary boundary that physically confines the processes. Lakes, segments of a river, and settling tanks in treatment plants are examples of reactors. Most, but not all, reactors experience continuous flow (in and out). Sequencing batch reactors have a sequence of states including fill, react, and empty. It is important to know the mixing level and residence time in reactors, since they both affect the degree of process reaction that occurs while the fluid (usually water) and its components (often pollutants) pass through the reactor.

Chlorine contactor tanks are designed to maximize the contact time between chlorine and pathogens before the water is delivered to consumers. Thus the design objective is to maximize the time that it takes for water to travel from the tank influent to the effluent. Tracer studies can be used to determine the hydraulic characteristics of a reactor such as the disinfection contact tanks at water treatment plants. The results from tracer studies are used to obtain accurate estimates of the effective contact time. In this laboratory students will experiment with different reactor designs with the goal of maximizing the contact time.

7.2 Classifications

7.2.1 Dispersion

Mixing levels give rise to three categories of reactors; completely mixed flow reactors (CMFR), plug flow reactors (PFR) and flow with dispersion reactors (FDR). The plug flow reactor is an idealized extreme not attainable in practice. All real reactors fall under the category of FDR or CMFR.

7.2.2 Boundary Conditions

The reactor inlet and outlet boundary conditions significantly affect the reactor response to the addition of a pulse of tracer. If dispersion is possible across a boundary, then the boundary is open. An example of a reactor with open boundaries is a section of a river. If a tracer is added to a section of a river it is possible for some of the tracer to move upstream, illustrating that an arbitrary section of a river is an open reactor (Fig. 7.1). This is equivalent to letting the reactor be defined as a section of a long reactor. One of the characteristics of open boundaries is that some of the tracer introduced at the reactor inlet can be carried upstream and thus the residence time for a conservative tracer can be greater than the hydraulic residence time!

A closed reactor is one where the reactor has a diffusion or dispersion coefficient different than those of the entrance or exit (Fig. 7.2). Typically exit and entrance diffusion/dispersion are much less than the diffusion/dispersion in the reactor. An example of a reactor with closed boundaries is a tank with small inlet and outlet pipes.

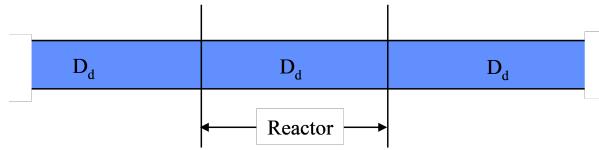


Fig. 7.1: Schematic of a reactor with open boundary conditions.

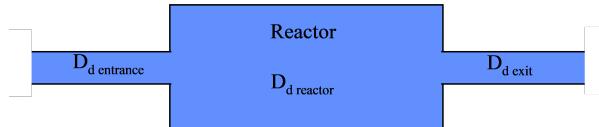


Fig. 7.2: Schematic of a reactor with closed boundary conditions.

7.2.3 Modeling

Reactors can be studied by measuring the effluent concentration after the addition of a spike or pulse of a tracer in the influent or after a step function change in input concentration. The resulting response curves can be made non-dimensional by plotting $\frac{C_t V_r}{C_{tr} V_{tr}}$ as a function of t/θ , where θ is the hydraulic residence time. The term t/θ will be defined as t^* . The non-dimensional response curves from pulse inputs and step inputs are known as E curves and F curves, respectively. The E curve is the exit age distribution and represents the actual output of tracer from a reactor. The F curve is the cumulative age distribution and represents the cumulative fraction of tracer that has exited a reactor at time t^* . The E and F curves are related by

$$F_{(t^*)} = \int_0^{t^*} E_{(t^*)} dt^* \quad (7.1)$$

The $F_{(t^*)}$ curve can either be obtained by integrating $E_{(t^*)}$ or by applying a step function change in influent concentration to a reactor and monitoring the effluent concentration. In that case $F_{(t^*)}$ is defined as

$$F_{(t^*)} = \frac{C_{(t^*)}}{C_{in}} \quad (7.2)$$

The following sections will include typical E and F curves for different reactor types.

7.2.4 Completely Mixed Flow Reactor

Completely mixed flow regimes can be approximated quite closely in practice. In the case of CMFRs, there is not an analytical solution to the advection-dispersion equation so we revert to a simple mass balance. For a completely mixed reactor a mass balance on a conservative tracer yields the following differential equation:

$$V_r \frac{dC}{dt} = (C_{in} - C) Q \quad (7.3)$$

where Q is the volumetric flow rate and V_r is the volume of the reactor.

Equation (7.3) can be used to predict a variety of effluent responses to tracer inputs. If a pulse of tracer is discharged directly into a reactor so that the initial concentration of tracer in the reactor is $C_0 = \frac{C_{tr} V_{tr}}{V_r}$ and the input concentration is zero ($C_{in} = 0$) the solution to the differential equation is:

$$E_{(t)} = \frac{C_t V_r}{C_{tr} V_{tr}} = e^{(-t/\theta)} \quad (7.4)$$

or in the dimensionless form

$$E_{(t^*)} = \frac{C_{(t^*)} V_r}{C_{tr} V_{tr}} = e^{(-t^*)} \quad (7.5)$$

where $E_{(t^*)}$ is the exit age or residence time distribution curve, V_r is the reactor volume, V_{tr} is the tracer volume, and C_{tr} is the tracer concentration. If a reactor has a completely mixed flow regime its response, $E_{(t^*)}$, to a pulse input should plot as a straight line on a semi-logarithmic plot. Response curves for a CMFR are shown in Fig. 7.3.

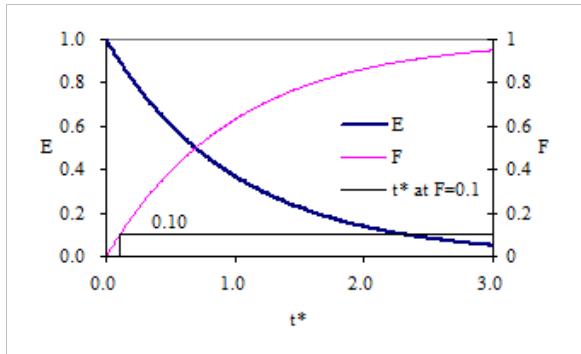


Fig. 7.3: Exit age (E) and Cumulative exit age (F) curves for completely mixed flow reactors.

The time for 10% of the pulse to arrive at the effluent of a CMFR is approximately $0.1 t^*$.

7.2.5 Plug Flow Reactor

Plug flow regimes are impossible to attain because mass transport must be by advection alone. There can be no differential displacement of tracer relative to the average advective velocity. In practice some mixing will occur due to molecular diffusion, turbulent dispersion, and/or fluid shear. For the case of the plug flow reactor the advection-dispersion equation in 1.7 reduces to:

$$\frac{\partial C}{\partial t} = -U \frac{\partial C}{\partial x} \quad (7.6)$$

The velocity, U, serves to transform the directional concentration gradient into a temporal concentration gradient. In other words, a conservative substance moves with the advective flow of the fluid. The solutions to this differential equation for a pulse input and for a step input are shown graphically in Fig. 7.4. The time for 10% of the pulse to arrive at the effluent of a PFR is t^* since all of the pulse arrives at the same time.

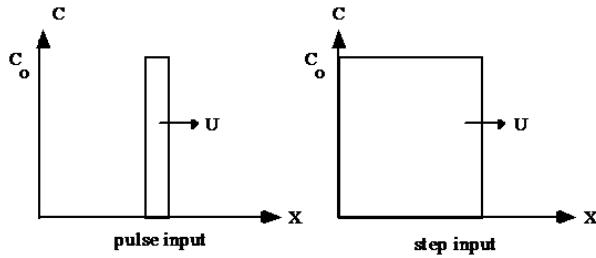


Fig. 7.4: Pulse and step input in a plug flow reactor.

7.2.6 Flow with Dispersion

Real flow reactors that aren't completely mixed exhibit behavior that is between plug flow and completely mixed. The reactor flow behavior is a function of the relative strength of the mixing process (or dispersion) compared to the advective flow. Analytical solutions that describe these real reactors are more difficult and in most cases a parameter

describing the dispersion is fit to the data rather than predicted *a priori*. Two models for arbitrary mixing levels are described below.

7.2.7 Open Boundary conditions: One Dimensional Advection-Dispersion Equation

The governing differential equation for a conservative (i.e., non-reactive) substance in a reactor that has advective transport (i.e., flow) and some mixing (dispersion) in the direction of flow (x - dimension) is given in equation (7.7).

$$\frac{\partial C}{\partial t} = -U \frac{\partial C}{\partial x} + D_d \frac{\partial^2 C}{\partial x^2} \quad (7.7)$$

Where

C = concentration of a conservative substance

U = average fluid velocity in the x direction

D_d = longitudinal dispersion coefficient

t = time

The dispersion is described by a single parameter that is the same in the reactor as it is at the inlet and outlet for open boundary conditions. The solution to (7.7) for the case of complete mixing in the y - z plane and advective and dispersive transport only in the x direction for any x and t (after $t=0$) gives:

$$C(x, t) = \frac{M}{A\sqrt{4\pi D_d t}} \exp\left[\frac{-x'^2}{4D_d t}\right] \quad (7.8)$$

Where

M = mass of conservative material in the spike

D_d = axial dispersion coefficient [L^2/T],

$x = x - Ut$, U = longitudinal advective velocity in the reactor,

A is the cross-sectional area of the reactor.

A measure of dispersion can be obtained directly from equation (7.8). From this equation we expect a maximum value of C at $t = x/U$. At this time $C(x, t) = \frac{M}{A\sqrt{4\pi D_d t}}$. If the mass of the tracer input (M) and reactor cross-sectional area (A) are known, then D_d can be estimated. Equation (7.8) can be made dimensionless to more clearly reveal the important parameters. The dispersion coefficient can be made dimensionless by dividing into a velocity and a length.

$$Pe = \frac{UL}{D_d} \quad (7.9)$$

where L is the length of the reactor and U is the mean advective velocity. The dimensionless parameter Pe (Peclet number) is used to characterize the level of dispersion in a reactor. The Peclet number is the ratio of advective to dispersive transport. In the limiting cases when $Pe = 0$ (very high dispersion) we have a completely mixed regime (CMFR) and when $Pe = \infty$, $D_d = 0$, no dispersion) we have a plug flow reactor (PFR).

The time can be normalized by dividing by the hydraulic residence time.

$$t^* = \frac{tU}{L} \quad (7.10)$$

Combining Equations (7.9) and (7.10) we obtain $D_d t = t^* Pe L^2$ for substitution into Equation (7.8). The concentration can be normalized by the mass of the tracer and the volume, V , of the reactor.

$$V = AL \quad (7.11)$$

$$E_{(t^*)} = \frac{C(x, t)V}{M} \quad (7.12)$$

The position in the reactor where the concentration measurements are made can be changed to the reactor effluent by substituting L for x. Substituting Equation (7.10) into the definition of x'^2 we obtain:

$$x'^2 = (L - Ut)^2 = (1 - t^*)^2 L^2 \quad (7.13)$$

The resulting dimensionless form of Equation (7.8) is given in Equation (7.14).

$$E_{(t^*)} = \sqrt{\frac{Pe}{4\pi t^*}} \exp \left[\frac{-(1-t^*)^2 Pe}{4t^*} \right] \quad (7.14)$$

Response curves for the advection-dispersion equation are shown in Fig. 7.5 and Fig. 7.6. The exit age curve is highly skewed for low Pe because tracer that is in the reactor longer has more time for dispersion. As the Peclet number increases the dispersion decreases and the response becomes closer to plug flow.

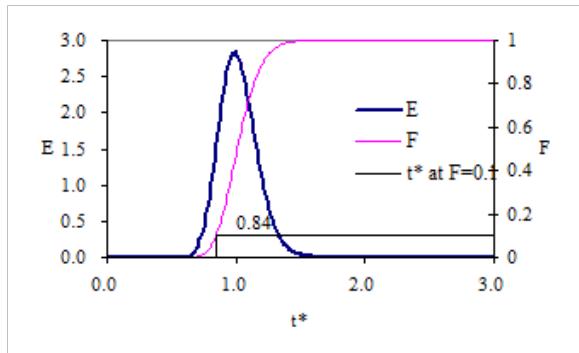


Fig. 7.5: Exit age (E) and cumulative exit age (F) curves for the advection-dispersion equation with Pe of 100.

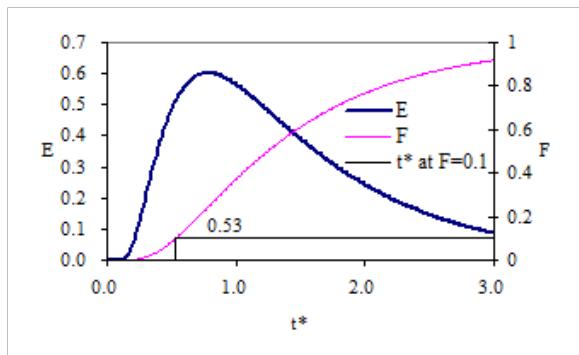


Fig. 7.6: Exit age (E) and cumulative exit age (F) curves for the advection-dispersion equation with Pe of 4.

Flow through porous media (such as groundwater through soil) is a type of flow with dispersion. The above equations can be applied by recognizing that the relevant water velocity is the pore water velocity. The pore water velocity is $U = \frac{Q}{A\phi}$ where A is the cross sectional area of the porous media and ϕ (volume of voids/total volume) is the porosity of the porous media.

The form of equation (7.8) is exactly like the normal distribution curve:

$$\frac{CA}{M} = \frac{1}{\sigma_x \sqrt{2\pi}} \exp \left[\frac{-x^2}{4\sigma_x^2} \right] \quad (7.15)$$

where

$$\sigma_x^2 = 2D_d t \quad (7.16)$$

Or in dimensionless form:

$$\frac{\sigma_x^2}{L^2} = 2t^*Pe \quad (7.17)$$

The variance in concentration over space (σ_x^2) is the variance in concentrations taken from many different positions in the reactor at some single moment in time, t . The variance in x (σ_x^2) has dimensions of length squared.

7.2.8 Closed Boundary Conditions: Completely Mixed Flow Reactor in Series

Completely mixed flow reactors are sometimes connected in series to create a reactor system with flow characteristics in between CMFR and PFR. CMFRs in series increase overall process efficiency because the reactants are at higher concentrations in the first reactors than they would be in a single large CMFR. Thus the total reactor volume can be smaller with a series of CMFRs. Similarly the reaction process can be made more efficient given the same reactor volume if it is divided into a series of CMFRs.

Non ideal reactors with mixing levels between that of CMFRs and PFRs and reactors containing baffles can also be modeled as a series of CMFRs. In this case the mixing level for the system is represented by the number of CMFRs that are used to represent the non-ideal reactor. The equation describing the concentration of a conservative tracer in the N^{th} reactor is given in equation (7.18).

$$\frac{C_N(t)V}{M} = \frac{N^N}{(N-1)!} \left(\frac{t}{\theta}\right)^{N-1} e^{\left(-\frac{Nt}{\theta}\right)} \quad (7.18)$$

The non-dimensional form of equation (7.18) is given in equation (7.19)

$$E_{N(t^*)} = \frac{N^N}{(N-1)!} (t^*)^{N-1} e^{(-Nt^*)} \quad (7.19)$$

It can be shown that equations (7.14) and (7.19) converge for large N when

$$Pe = 2N \quad (7.20)$$

Equation (7.18) is limited to integer values of N . The model can be extended to include non-integer values of N if the factorial function (i.e., $(N-1)!$) were replaced with the Γ function. This extension is particularly beneficial when fitting data to the models because the Γ function is continuous.

Response curves for the advection-dispersion equation are shown in Fig. 7.7. Notice that the effluent concentration begins to rise much more quickly in series CMFRs than in the advection-dispersion model for a single CMFR. This distinction between the two models lessens as the number of reactors increases and the Pe increases.

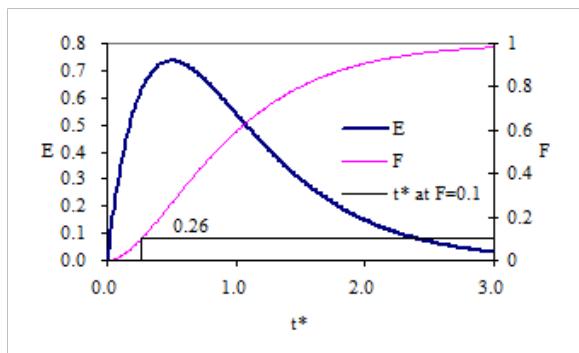


Fig. 7.7: Exit age (E) and cumulative exit age (F) curves for series CMFR with N of 2.

7.3 Reactor Studies

One of the easiest methods to determine the mixing (dispersion) characteristics of a reactor is to add a spike input of a conservative material and then monitor the concentration of the material in the reactor effluent. The variance of tracer concentration versus time (σ_t^2 , with dimensions of time squared) can be measured by sampling at a single point in the reactor at many different times and can be computed using the following equations.

$$\sigma_t^2 = \frac{\int_0^\infty C(t)(t - \bar{t})^2 dt}{\int_0^\infty C(t)dt} = \frac{\int_0^\infty t^2 \cdot C(t)dt}{\int_0^\infty C(t)dt} - \bar{t}^2 \quad (7.21)$$

where

$$\bar{t} = \frac{\int_0^\infty t \cdot C(t)dt}{\int_0^\infty C(t)dt} \quad (7.22)$$

For discrete data points:

$$\sigma_t^2 = \frac{\sum_{i=0}^n t_i^2 \cdot C_i \Delta t}{\sum_{i=0}^n C_i \Delta t} - \bar{t}^2 \quad (7.23)$$

and

$$\bar{t} = \frac{\sum_{i=0}^n t_i \cdot C_i \Delta t}{\sum_{i=0}^n C_i \Delta t} \quad (7.24)$$

Inlet and outlet boundary conditions affect the response obtained from a reactor. Closed reactors have little dispersion across their inlet and outlet boundaries whereas open reactors can have significant dispersion across their inlet and outlet boundaries. Typically open systems have no physical boundaries in the direction of flow. An example of an open system would be a river segment. Closed systems have small inlets and outlets that minimize dispersion across the inlet and outlet regions. An example of a closed system is a tank (or a lake) with a small inlet and outlet. The reactor used in the lab is closed. The \bar{t} in equation (7.24) is the measured average residence time for the tracer in the reactor. For ideal closed reactors the measured residence time, \bar{t} , is equal to the theoretical hydraulic residence time (t_h = reactor volume/flow rate). For open reactors \bar{t} can exceed the hydraulic residence time. If \bar{t} is less than the hydraulic residence time it may indicate that the reactor contains dead volume or short circuiting such that some volume of the reactor is effectively unused.

The above equations suggest that from the reactor response to a spike input we can compute the dispersion coefficient for the reactor. We have two options for measuring reactor response:

1. synoptic measurements: at a fixed time sampling many points along the axis of the reactor will yield a Gaussian curve of concentration vs. distance. In practice synoptic measurements are difficult because it requires sampling devices that are time-coordinated. By combining equations (7.16), (7.23), and (7.24) it is possible to estimate the dispersion coefficient from synoptic measurements.
2. single point sampling: measure the concentration at a fixed position along the x axis of the reactor for many times. If the reactor length is fixed at L and measurements are made at the effluent of the reactor (observe the concentration of a tracer at $x = L$ as a function of time) then x is no longer a variable and $C(x,t)$ becomes $C(t)$ only. The response curve obtained through single point sampling is skewed. The curve spread changes during the sampling period and the response curve is skewed.

7.3.1 Peclet Number Estimation

For single point sampling of the effluent response curve, skew increases as the dispersion level in the reactor increases. The degree of skew depends on the dispersion coefficient, the velocity in the x-direction, and the length of the reactor. Peclet values in the range $100 < Pe < \infty$ result in a symmetric response curve.

Response curve skew makes the assumption of a symmetrical normal distribution curve inappropriate and a new relationship between the variance and the dispersion coefficient (or Pe) has to be determined. Boundary conditions affect the determination of the dispersion coefficient. The relationship between the Peclet number and variance for open systems is given by

$$\sigma_t^2 = \left(\frac{2}{Pe} + \frac{8}{Pe^2} \right) \cdot \theta^2 \quad (7.25)$$

For closed systems the relationship is:

$$\sigma_t^2 = \left[\frac{2}{Pe} - \frac{2}{Pe^2} \cdot (1 - e^{-Pe}) \right] \cdot \theta^2 \quad (7.26)$$

The term $\frac{2}{Pe}$ in equations (7.25) and (7.26) is dominant for Peclet numbers much greater than 10 as is shown in Fig. 7.8. The additional terms in equations (7.25) and (7.26) are corrections for skewedness in the response curve. These skewedness corrections are not very significant for Peclet numbers greater than 10. Thus for Peclet numbers greater than 10 the Peclet number can be determined using equation (7.27) for both open and closed systems.

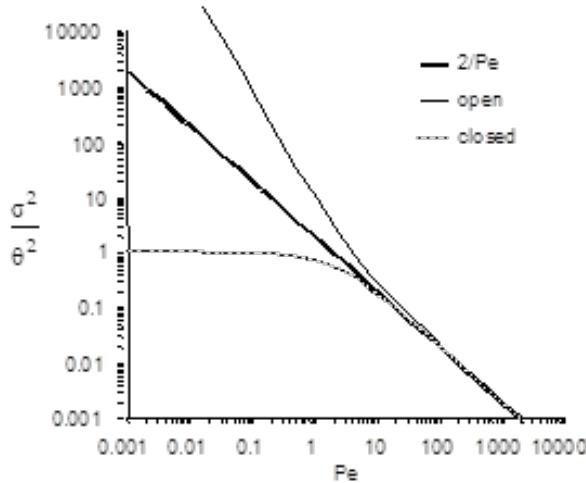


Fig. 7.8: Relationship between equations (7.25) through (7.25).

$$Pe = \frac{2\theta^2}{\sigma_t^2} \quad (7.27)$$

7.3.2 Mass Conservation

When a pulse of conservative tracer is added to a continuous flow reactor, all of the tracer is expected to leave the reactor eventually. The mass of a substance that has left the reactor is given in equation (7.28).

$$M_{out} = \sum_{i=0}^n QC_i \Delta t_i \quad (7.28)$$

where Q is the flow rate and M is the mass of any substance whose concentration is given by C . If Q and Δt are constant, then equation (7.28) can be rewritten as

$$M_{out} = Q \Delta t \sum_{i=0}^n C_i \quad (7.29)$$

Equation (7.29) can be used to determine if all of the tracer was measured in the reactor effluent. Alternately, mass conservation can be checked by plotting the F curve and checking if the plot approaches #. If some of the tracer is in the reactor at the end of the experiment then the following equation can be used to test for mass closure.

$$M_{in} = M_{out} + \Delta M_{stored} \quad (7.30)$$

7.4 Tracers

A conservative tracer will be used to characterize your reactors. Typical tracers include salt and various dyes. For this experiment we will use #40 Red Dye as the conservative tracer. The red dye concentration will be measured using an optical spectrophotometric probe. The linear range of operation for the probes is from about 0.01 mg/L to 30 mg/L. We will add an amount of red dye near the high end of this range to acquire a dataset that spans multiple orders of magnitude.

Additional constraints for the tracer are that if a highly concentrated tracer is used, then there will be a significant density difference between the tracer and the water in the reactor. If a very dilute tracer is used then a large volume of tracer will need to be added and the volume of tracer could become a large fraction of the reactor volume and thus the pulse could no longer be modeled as having negligible volume.

The reactors are 30 cm long and 15 cm wide. If the depth is limited to (a maximum of) 5 cm, then the total volume is 2.25 L and with a peristaltic pumping rate of approximately 380 mL/min, the residence time is approximately 6 minutes.

The tracer should be added directly into the first chamber of your reactor. The red dye will also make it possible to qualitatively observe the advective and dispersive transport in the reactors.

7.4.1 Mass balance

Collect data that makes it possible to perform a mass balance on red dye. Note that if it is expedient to shorten an experimental run such that there is a significant quantity of the tracer residual in the reactor, that the mass of the residual (stored) tracer can be measured by pouring the reactor contents into a container and measuring the mass or volume of the mixture. The concentration of red dye in the mixture can be measured by connecting back to the optical probe.

7.5 Setting up the reactor for experiments

See the section on [calibrate photometer](#) to set up the photometer

In these experiments, we will be pumping tap water from a 20 L Jerrican to the influent of your reactor. Place your reactor on a stir plate and make sure that the 3/8 push-connect fitting is on the effluent side of your reactor (some may have 3/8 holes on both sides). The effluent of the reactor should be a straight short tube to the drain.

Use a second pump head with #17 tubing to pull a sample from near the effluent weir through the photometer and then to the drain.

Before running an experiment, it is important to determine whether your setup works. Turn the pump to approximately 380 mL/min and get water flowing through the system and off to waste. Two important things to check:

1. Is your sensor reading a stable voltage of approximately +3.5V? If not, you might have some air bubbles trapped in the sensor cell. Tap it gently to release the air.
2. Is the effluent weir working properly or is the reactor gaining volume? We want to maintain a constant volume throughout the experiment, so let the pump run for ten minutes or so and confirm that the volume is stable (put a line or a piece of tape to mark the water level).

7.6 Testing protocol

To ensure that everything is working properly I recommend that you begin by first testing a CMFR. Use the CMFR data to verify that you obtain mass balance and that you obtain the expected exit age response.

For each test make sure that you accurately measure the reactor volume, residual reactor red dye concentration, and the flow rate. The reactor volume can be approximated by taking the mass of the reactor, as you did to estimate the volume of your lakes. Measure the residual concentration of red dye in the reactor by measuring the completely mixed concentration (this is just the concentration at the end of the experiment for the CMFR, but you will have to remove the baffles and mix for the baffled reactors). The flow rate can be accurately determined by measuring the volume of a timed sample from the pump.

1. Log data to file so that you have a record of the red dye concentration in the effluent of the reactor as a function of time.
2. Prepare to save a text note into the data file indicating the exact moment when you will actually start the experiment.
3. Add a volume of red dye #40 stock that will give a maximum concentration of *approximately* 30 mg/L near the influent of the reactor. Immediately enter the text note to record when the experiment began. For the CMFR you can easily calculate this. For other reactors you may need to try and if necessary repeat the experiment if the tracer concentration goes above the 50 mg/L or if the maximum tracer concentration is below 10 mg/L.
4. Collect data until the majority of the tracer has exited.
5. Stop the experiment and pour the contents of the reactor into a container. Weigh the container to determine the exact volume of the reactor.
6. Sample the contents of the mixed reactor with the photometer to obtain the average concentration in the reactor at the end of your experiment. This will enable you to do a mass balance on the red dye.



Fig. 7.9: Front view of the reactor setup.

The reactor setup is designed to maintain a constant depth in the tank with an overflow weir. The second pump channel is used to sample the effluent through the photometer without sending air bubbles into the photometer.

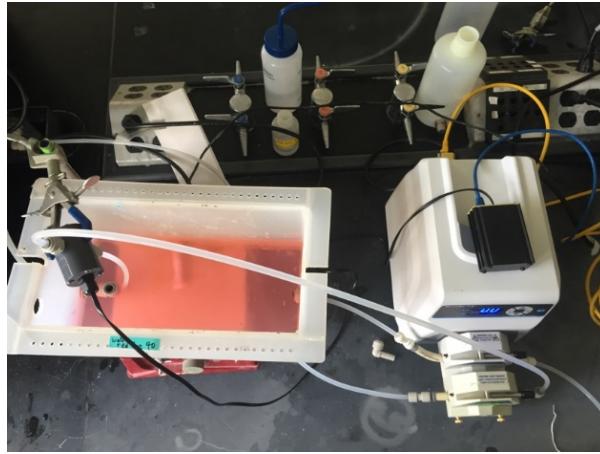


Fig. 7.10: Top view of the reactor setup.

7.7 Reactor Design

7.7.1 Baffle Design

Baffles can be added to tanks to decrease the ability of the solution to travel quickly from the tank inlet to the tank outlet. The goal of the baffle design depends on whether the tank is designed for a reaction or for separation. The baffles in reactors should generally be designed to achieve mixing in each of the baffled zones. The baffles in tanks designed for separation (clarifiers) should be designed to achieve uniform flow (a much more difficult goal to achieve). Mixing within each baffled zone could be achieved with a propeller mixer or by appropriately sized pores in the baffles. If pore mixing is desired the energy for mixing is provided by the kinetic energy of the fluid flowing through the pores.

The flow characteristics of a baffled tank are complex and would require computational fluid dynamics (CFD) to model. In lieu of developing a CFD model we will turn to dimensional analysis to help us characterize the mixing efficiency of a baffled reactor segment. The independent parameter that we will measure to characterize the reactor is its Peclet number or the number of N CMFRs in series.

The design goal is to determine the diameter and spacing of the pores required to achieve adequate mixing. The fluid flow out of the pores is jet flow. The jets entrain and mix with the surrounding fluid. To develop effective mixing the jet should be turbulent. The transition between turbulent and laminar behavior for jets occurs at a jet Reynolds number of a few hundred (Personal communication with Dr. Cowen).

$$\begin{aligned} Re_{jet} &= \frac{V_{jet}d_{jet}}{\nu} \\ Re_{jet} &= \frac{4Q_{jet}}{\pi d_{jet}\nu} \\ Re_{jet} &= \frac{4Q_{reactor}}{n_{ports}\pi d_{jet}\nu} \end{aligned} \tag{7.31}$$

The optimal pore spacing for maximum mixing could be based on the spread of the jet in the distance to the next baffle. Jets expand in the radial direction at a rate of approximately 10% of the distance the jet travels axially. Thus the pore spacing could be in a rectangular grid with the distance between pores equal to 10% of the distance between baffles. This spacing may be smaller than practical since it will require a large number of pores.

An additional constraint on pore design is that the head loss through the pores not be excessive. The hydraulic grade line (and thus depth of each baffled reactor) will drop by a distance equal to the head loss through the pores. The head loss through the pores is given by the orifice equation

$$Q_{orifice} = K_{orifice} A_{orifice} \sqrt{2g\Delta h} \tag{7.32}$$

where the orifice coefficient, $K_{orifice}$, has a value of approximately 0.6, the area of the orifice is $A_{orifice}$, and the head loss through the orifice is Δh .

$$Q_{orifice} = K_{orifice} \frac{\pi d_{orifice}^2}{4} \sqrt{2g\Delta h} \quad (7.33)$$

$$Q_{reactor} = n_{orifice} K_{orifice} \frac{\pi d_{orifice}^2}{4} \sqrt{2g\Delta h} \quad (7.34)$$

where $n_{orifice}$ is the number of pores. Solving for the pore diameter we obtain.

$$d_{orifice} = \sqrt{\frac{4Q_{reactor}}{\pi n_{orifice} K_{orifice} \sqrt{2g\Delta h}}} \quad (7.35)$$

7.7.2 Chlorine Contact Tank Design

The design objective for chlorine contact tanks is to maximize the inactivation of pathogens by maximizing the contact time between the chlorine and the pathogens before the water is sent to the distribution system. This objective has been codified by the EPA with the contact time for a tank defined as the time for the effluent of the tank to reach 10% of the influent value after a step change in the influent.

Table 7.1: Comparison of t^* at $F=0.1$ for non-ideal reactors (Malcolm Pirnie, 1991).

Baffling Condition	Guidance manual value of t^{star} at $F=0.1$	Extent of Baffles
Unbaffled (CMFR)	0.1	No baffles, agitated basin with low length to width ratio, high inlet and outlet flow velocities
Poorly baffled	0.3	Single or multiple unbaffled inlets and outlets, no intrabasin baffles
Average	0.5	Baffled inlet or outlet with some intrabasin baffles
Superior	0.7	Perforated inlet baffles, serpentine or perforated intrabasin baffles, outlet weir or perforated launders
Perfect (PFR)	1.0	Very high length to width ratio (pipeline flow), perforated inlet, outlet and intrabasin baffles

7.8 Procedures

The reactor you will be working with can be easily modified by installing various configurations of perforated or staggered baffles. Your objectives are

1. to modify the reactor to obtain a maximum value of t^* at $F = 0.1$.
2. to document your progress toward this goal by obtaining appropriate experimental data
3. to compare your experimental data with appropriate models

Note that you are allowed to collaborate with other teams to obtain the results from additional tests. All data sharing must be documented in your report.

7.8.1 Baffle installation

The baffles can be installed by rolling a 3 mm diameter roll of putty and applying the putty to the bottom and one side of the baffle. Carefully install the baffle by rotating it into place (so the putty on the one side doesn't have to slide) and then pressing it down (so the putty on the bottom doesn't have to slide). Install the two bolts with wing nuts to hold the baffle in place. Add a roll of putty to the corner formed at the unsealed side of the baffle and press the putty into place.

7.9 Prelab Questions

1. Calculate the **incremental** volume of a 100 g/L red dye stock that would need to be added to 1 L of water to produce 0, 1, 2, 5, 10, 20, 30, 40, and 50 mg/L calibration points. Calculate a numpy array containing the **cumulative** volume of red dye required. Strip the units from the array using .magnitude. Then create a copy of the array with a zero appended (np.append) in front and the last element deleted (np.delete). Then use numpy subtract to get the difference between the two arrays to calculate the incremental volume that you need to add.
2. Calculate the change in hydraulic grade line between baffled sections of a reactor with a flow rate of 380 mL/min. The reactor baffles are perforated with 6 holes 1 mm in diameter. Is the flow through these orifices in series or in parallel? Do you multiply the head loss for one orifice by the number of orifices to get the total head loss? Use the pc.head_orifice function to calculate the head loss through an orifice. The *vena contracta* for the orifice can be found at exp.RATIO_VC_ORIFICE. Why would 6 holes 1 mm in diameter not be a good design for this reactor?
3. On a single graph plot the exit age distribution ($E_{(t^*)}$) for a reactor that operates as a 1-dimensional advection-dispersion reactor with Peclet numbers of 1, 10, and 100 (there will be three plots on the graph and thus a legend is required). The x-axis should be t^* from 0.0 to 3.0. Comment on the shapes of the curves as a function of the Peclet number.

7.10 Data Analysis

You will analyze all of the datasets in the same way. Use a consistent set of units throughout your data analysis and include the units in your python code and discussion!

1. Use multivariable nonlinear regression to obtain the best fit between the experimental data and the two models by minimizing the sum of the squared errors. Use EPA.Solver_AD_Pe and EPA.Solver_CMFR_N. These functions will minimize the error by varying the values of average residence time, (mass of tracer/reactor volume), and either the number of CMFR in series or the Peclet number.
2. Generate a plot showing the experimental data as points and the model results as thin lines for each of your experiments. Explain which model fits best and discuss those results based on your expectations.
3. Compare the trends in the estimated values of N and Pe across your set of experiments. How did your chosen reactor modifications effect dispersion?
4. Report the values of t^* at $F = 0.1$ for each of your experiments. Do they meet your expectations?
5. Evaluate whether there is any evidence of dead volumes or short circuiting in your reactor.
6. Make a recommendation for the design of a full scale chlorine contact tank. As part of your recommendation discuss the parameter you chose to vary as part of your experimentation and what the optimal value was determined to be.

7.11 References

Weber, W. J. J. and F. A. Digiano. 1996. Process Dynamics in Environmental Systems. New York, John Wiley & Sons, Inc.

Malcolm Pirnie, Inc., and HDR Engineering, Inc. Guidance Manual for Compliance with the Filtration and Disinfection Requirements for Public Water Systems Using Surface Water Sources. AWWA, Denver CO, 1991.

7.12 Lab Prep Notes

1. Create 100 mL of 100 g/L red dye stock (thus use 10 g of red dye) and distribute in small vials to each lab bench.
2. Use # 18 tubing for inlet and #17 for photometer sampling.
3. The maximum concentration of red dye that the photometer can measure accurately is 50 mg/L.
4. Organize team research so they dont use the same materials and so they answer different questions.

**CHAPTER
EIGHT**

ADSORPTION

Water is sometimes called the universal solvent and that property has both positive and negative aspects. On the positive side, water's ability to serve as a carrier for many compounds is essential for life. On the negative side that same ability can result in water carrying substances that are harmful to life. Environmental engineers often have the challenge of removing harmful dissolved species from water. This task shows up when we are producing safe drinking water from a ground water source that is contaminated with arsenic, fluoride, nitrate, or atrazine. Surface water sources can be contaminated with naturally organic matter that interacts with disinfectants to produce toxic byproducts. Wastewater reuse requires removal of a long list of dissolved species that would otherwise accumulate. Dyeing of clothing produces an industrial waste water contaminated with high concentrations of dissolved dyes. As we develop improved analytical techniques we realize that many of the substances that we use including caffeine, cocaine, pharmaceuticals, herbicides, and pesticides are present in the environment.

Our strategies for removing dissolved species are limited to stopping the manufacture and use of the harmful compounds, encouraging reactions to transform the compounds into less harmful products, or removing the compounds from water. Of course, removing the compounds from water requires that we put the compounds somewhere else. We can either concentrate the compounds in water (reverse osmosis) or transfer the compounds into a gas (air stripping) or solid phase (adsorption).

Adsorption to a solid phase is commonly used in drinking water treatment plants. Aluminum and iron hydroxide coagulant nanoparticles are used for flocculation because they are sticky and readily attach to suspended particles. The coagulant nanoparticles also adsorb many dissolved species including naturally occurring dissolved organics. The coagulant nanoparticles are then concentrated in the solid phase of the sludge produced by water treatment plants. Activated carbon is also used in drinking water treatment to transfer dissolved species to the solid phase. This laboratory experiment will give us an opportunity to learn about the removal of dissolved species through adsorption to a solid phase.

8.1 Objectives

The objectives of this experiment are to:

1. Investigate the breakthrough characteristics of red dye #40 on activated carbon or on coagulant nanoparticles in a continuous-flow carbon contactor at a range of concentrations.
2. Determine the equilibrium partitioning of red dye #40 on activated carbon or on coagulant nanoparticles

8.2 Background

Adsorption is a unit operation in which surface-active materials in true solution are removed from the solvent by interphase transfer to the surface of an adsorbent particle. This process is employed in environmental engineering practice for removal of various pollutants such as soluble organics, dyes, pesticides, humic substances, etc., from wastewaters and for removal of color, taste, and odor-producing compounds from natural waters that are to be used as potable

water supplies. Adsorption onto granular activated carbon (GAC) also finds wide application in the remediation of groundwater contaminated with volatile and nonvolatile organic pollutants.

Under isothermal conditions the equilibrium partitioning between solution and solid phase is a function of concentration. In this experiment the partitioning will be analyzed using both Freundlich

$$q = K_f C^{\frac{1}{n}} \quad (8.1)$$

and Langmuir partitioning models

|Where| : $math : 'q' is the mass of adsorbate (species being removed from solution) per mass of adsorbent (solid phase)| : math : 'C'$

$$(8.2)$$

We will preload columns of sand (or glass beads) with the adsorbent and then pump a solution of red dye #40 through the column. We will measure the effluent concentration of the dye as a function of time using a photometer. In these experiments the column of sand is inert, but provides a mechanism to hold the adsorbent stationary (fixed bed) while the red dye solution flows through. The red dye will adsorb to the adsorbent and the effluent red dye concentration is expected to be very low for some time. Red dye will begin breaking through once the majority of the adsorbent reaches equilibrium with the influent concentration of the red dye.

In this analysis we are neglecting several aspects of the adsorption process. We assume that the time required to reach equilibrium partitioning is very small relative to the hydraulic residence time (θ_{HRT}) of the column. This assumption requires that both liquid phase and solid phase adsorbate mass transport mechanisms are relatively fast. Transport within the solid phase matrix is by the slow process of diffusion. Thus we expect diffusion to increase the time required to reach equilibrium for adsorbents that require diffusion over longer distances. Diffusion time will be greatest for Large diameter granular activated carbon, less for powdered activated carbon, and very fast for coagulant nanoparticles.

The mass of the adsorbate in the column per plan view area at breakthrough is given by

|where| : math : ‘ M_{AC} ‘and : math : ‘ M_{GAC} ‘are the masses of the heads or bents per plan view surface area of the column| : math : ‘(8.3)

The mass transfer zone travels at velocity v_{mtz} through the fixed bed as the absorbent slowly fills to the equilibrium density, q_0 , based on the influent concentration, C_0 .

The velocity of the mass transfer zone (mtz or the adsorption front) can be obtained by a mass balance on the system. If we set our frame of reference (and our control volume) to be centered on the mass transfer zone, then the average velocity (over the pore fraction of the control surface) of fluid entering the mtz is equal to pore water velocity minus the velocity of the mtz. The fluid phase concentration of the adsorbate entering the control surface is C_0 and the fraction of the control surface where fluid is passing through is the porosity, ϕ .

The average velocity of the solid phase exiting through the control surface is $-v_{mtz}$. The bulk density of the adsorbate is $q_0 \rho_{bulk\ adsorbent}$ where $\rho_{bulk\ adsorbent}$ is the mass of adsorbent per volume of the packed bed. The mass rate of adsorbate passing through the control surface in liquid phase must precisely balance the mass rate of adsorbate passing through the control surface in the solid phase because the mtz is stationary.

$$[(v_{pore} - v_{mtz})C_0\phi] - [(v_{mtz})q_0\rho_{bulk adsorbent}] = 0 \quad (8.4)$$

We can apply continuity to find the relationship between the velocity in the pores and velocity above the porous fixed bed. The plan view area of the fixed bed cancels out.

$$\phi v_{pore} = v_a \quad (8.5)$$

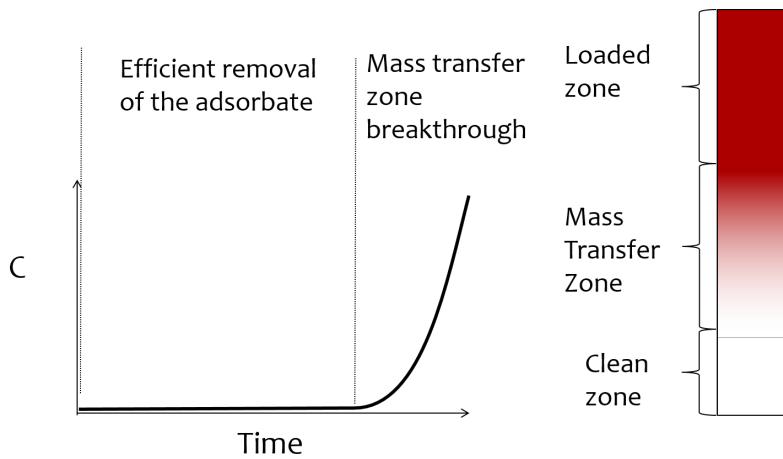


Fig. 8.1: Movie illustrating how the effluent concentration of the absorbate changes with time as the mass transfer zone moves through the fixed bed.

Eliminate v_{pore} from the equation

$$(v_a C_0 - v_{mtz} C_0 \phi) - [(v_{mtz}) q_0 \rho_{bulk adsorbent}] = 0 \quad (8.6)$$

Now solve for v_{mtz} .

$$v_{mtz} = \frac{v_a C_0}{C_0 \phi + q_0 \rho_{bulk adsorbent}} \quad (8.7)$$

In equation (8.7) the term $C_0 \phi$ represents the liquid phase mass of the adsorbate per unit volume of the fixed bed and the term $q_0 \rho_{bulk adsorbent}$ represents the solid phase mass of the adsorbate per unit volume of the fixed bed. The second term dominates for fixed bed adsorption reactors that are effective and thus equation (8.7) simplifies to:

$$v_{mtz} \cong \frac{v_a C_0}{q_0 \rho_{bulk adsorbent}} \quad (8.8)$$

The time until breakthrough can be obtained by dividing the length of the adsorption column (L_{column}) by the velocity of the mtz (equation (8.7))

$$\frac{L_{column}}{v_{mtz}} = \frac{L_{column} \phi}{v_a} + \frac{L_{column} q_0 \rho_{bulk adsorbent}}{v_a C_0} \quad t_{mtz} = t_{water} + t_{ads} \quad (8.9)$$

Thus the time to breakthrough is the time required for water to flow through the reactor plus the additional time required due to the adsorption process. The retardation factor is defined as the ratio of the time for the mass transfer zone to travel through the bed divided by the time for water to travel through the bed.

$$R_{adsorption} = \frac{t_{mtz}}{t_{water}} = \frac{v_{water}}{v_{mtz}} \quad R_{adsorption} \cong \frac{v_a q_0 \rho_{bulk adsorbent}}{\phi v_a C_0} = \frac{q_0 \rho_{bulk adsorbent}}{\phi C_0} \quad (8.10)$$

The effective bed porosity, ϕ can be calculated from

$$\phi = 1 - \frac{\rho_b}{\rho_{ac}} \quad (8.11)$$

where

ρ_b = apparent bulk density

$$\rho_{ac} = 2.1 \text{ g/cm}^3$$

From experiments conducted in the Cornell environmental laboratory around 2003 we have q_0 (50 mg/L) = 0.08. Our goal is to design a fixed bed reactor that has a t_{mtz} of about 30 minutes. With a 15 cm deep column at 1 mm/s and with a porosity of 0.4 the hydraulic residence time is 1 minute. Given a target retardation factor of 30 we can calculate the bulk density of carbon that we should have in the column. We can achieve this bulk density by diluting the activated carbon with sand.

$$\rho_{\text{bulk adsorbent}} \cong \frac{R_{\text{adsorption}} \phi C_0}{q_0} \quad (8.12)$$

Different teams can try different concentrations of red dye or different masses of activated carbon.

```
""" importing """
from aide_design.play import*
v_a = 1 * u.mm/u.s
porosity = 0.4
L_column = 10 * u.cm
C_0 = 50 * u.mg/u.L
q_0 = 0.08
t_water = (L_column*porosity/v_a).to(u.s)
t_mtz = 1800*u.s
# set the breakthrough time to 30 minutes = 1800 s
R_adsorption = t_mtz/t_water
Density_bulk = (R_adsorption * porosity * C_0/q_0).to(u.kg/u.m**3)
Density_bulk
D_column = 1*u.inch
A_column = pc.area_circle(D_column)
V_column = A_column * L_column
M_carbon = (V_column * Density_bulk).to(u.mg)
M_carbon
V_reddye = (v_a*A_column*t_mtz).to(u.L)
V_reddye
Q_reddye = (v_a*A_column).to(u.mL/u.min)
Q_reddye
density_sand = 2650 * u.kg/u.m**3
M_sand = (V_column*density_sand*(1-porosity)).to(u.g)
M_sand
```

8.3 Contactor Procedures

8.3.1 Carbon Contactor Setup

Assemble the system shown in Fig. 8.2. Use a peristaltic pump with #14 tubing at approximately 10 rpm. Prepare 20 L jerricans with 50 mg/L of Red dye #40. Use reverse osmosis water to dilute the dye. The carbon contactor will be operated in down flow mode. The specifications for the carbon contactors are given in Table Table 8.1.

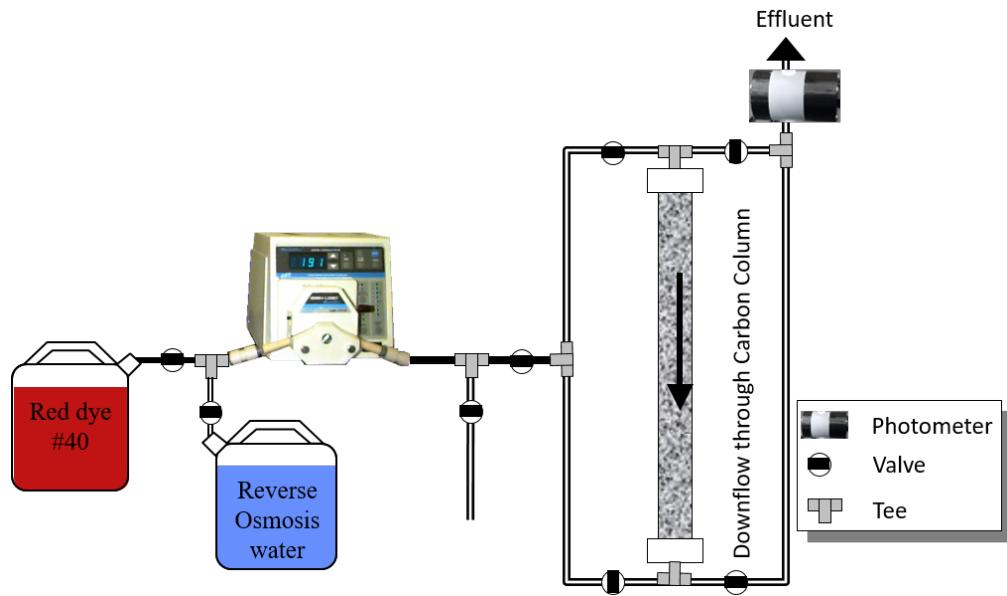


Fig. 8.2: Proposed design of the carbon column and feed system.

Table 8.1: Carbon contactor settings.

Parameter	Value
Influent red dye Concentration	0.050 g/L
Mass of red dye/20 L	1.00 g
Depth of fixed bed	15 cm
Mass of sand	80 g
Influent flow rate	30 mL/min (0.5 mL/s)
Column diameter	2.54 cm
$q(50\text{mg/L})$	0.080 g/g
Mass of carbon	0.2, 0.5, 1, 2, 5, or 10 g

8.3.2 Set up the Contactor

Work through this procedure twice. For the first test skip the activated carbon and thus measure the F curve (see *reactor modeling*) for the sand column. Rinse the column with RO water, remove the sand, and repeat the procedure with activated carbon.

1. Test column and pump and all tubing to ensure that it is leak tight using reverse osmosis water.
2. Remove top from column
3. Mix 80 g of sand and your teams assigned mass of activated carbon
4. Wet method
 1. Pour mixture of sand and activated carbon into a beaker containing reverse osmosis water.
 2. Swirl until most of the air is released.
 3. Use a funnel and a reverse osmosis water wash bottle to wash the mixture from the beaker into the column.
 4. Use a 50 mL syringe to remove excess water from the top of the column if necessary.

5. Use a long rod to gently stir activated carbon to help release air bubbles.
6. Assemble the column end fitting.
4. Dry method
 1. Use a funnel to pour dry mixture of sand and activated carbon into the column
 2. Assemble the column end fitting.
 3. Fill the column with water in up flow mode (at 5 mL/min - idea is to do this slowly so that air escapes)
 5. In up flow mode (at 30 mL/min), discharge the column effluent to waste until most of the fines are removed.
 6. Reverse the direction of flow to down flow and verify that the photometer is reading approximately 0 mg/L of red dye. This indicates that most of the activated carbon fines are removed from the column.
 7. Configure ProCoDA to log the concentration of red dye at 5 second intervals
 8. Start pumping Red Dye #40.
 9. Measure the flow rate using a balance to get mass of water in approximately 1 minute.
 10. It is impractical to try and achieve $C/C_0 = 1$, but run long enough to attain at least $C/C_0 = 0.6$.

8.3.3 Troubleshooting and Reflections

Spring 2019 is the first time that we are including this experiment. There are always multiple challenges associated with developing a new laboratory experiment and in this case it is possible to anticipate several potential problems with this experimental design.

1. Air bubbles! Air in the sand column or in the photometer will result in failure. Surface tension makes it difficult to remove the air bubbles from the activated carbon. The methods may need to be modified if air causes problems.
2. Mass transport of the red dye into the activated carbon pores is slow because it is a diffusion process. It is possible that this will result in premature breakthrough of red dye long before the activated carbon reaches equilibrium with the influent concentration of red dye.
3. The relatively small amount of activated carbon in the sand column may result in inefficient transport of the red dye to the activated carbon granules. This would also result in inefficient removal of red dye.

An alternative to granular activated carbon is Poly aluminum chloride (PACl - not to be confused with Powdered Activated Carbon or PAC). PACl will remove red dye by adsorption and given that PACl forms nanoparticles (rather than millimeter sized granules for GAC) it is possible that the mass transport of red dye to PACl will be much faster mass transport of red dye to GAC. This could be tested by substituting PACl for GAC in the column. We would need to develop a method to deliver the PACl flocs to the sand column.

8.3.4 Contactor Results and Analysis

1. Plot the breakthrough curves showing $\frac{C}{C_0}$ versus time.
2. Find the time when the effluent concentration was 50% of the influent concentration and plot that as a function of the mass of activated carbon used.
3. Calculate the retardation coefficient ($R_{adsorption}$) based on the time to breakthrough for the columns with and without activated carbon.

4. Calculate the quantity of Red Dye #40 that was transferred to the activated carbon based on the influent concentration, flow rate, and 50% breakthrough time.
5. Calculate the q_0 for each of the columns. Plot this as a function of the mass of activated carbon used.

What did you learn from this analysis? How can you explain the results that you have obtained? What changes to the experimental method do you recommend for next year (or for a project)?

8.4 Lab Prep Notes

Table 8.2: Reagent list. :header: Description, Supplier, Catalog number
:widths: 20, 20, 10 :align: center

activated carbon				
red dye #40				
1. Verify that all necessary supplies are in place for the pumps	tanks	column	valves	and tubing.
1. Prepare the Red Dye #40 stock solution.				
1. Prepare a 5% bleach solution (5 mL bleach diluted to 100 mL with distilled water) for cleaning the photometer sample cell and sample lines.				

8.4.1 Procedure to remove air from the top of the column

1. Close the Red Dye #40 influent valve.
2. Open the distilled water influent valve.
3. Wait for the influent line to clear of Red Dye #40.
4. Turn off the pump.

5. Reverse the column flow direction.
6. Turn on the pump until the air is removed.
7. Turn off the pump.
8. Reverse the column flow direction.
9. Turn on the pump and switch the influent to Red Dye #40.

8.5 References

Lawler, D. F., & Benjamin, M. M. (2013). Water quality engineering: physical / chemical treatment processes. Hoboken, N.J.: Wiley. Retrieved from <http://search.ebscohost.com/login.aspx?direct=true&scope=site&db=nlebk&db=nlabk&AN=631668>

**CHAPTER
NINE**

GAS TRANSFER

9.1 Introduction

Exchange of gases between aqueous and gaseous phases is an essential element of many environmental processes. Wastewater treatment plants require enhanced transfer of oxygen into activated sludge tanks to maintain aerobic degradation. Water treatment plants require gas transfer to dissolve chlorine gas or ozone. Gas transfer can also be used to remove unwanted volatile chemicals such as carbon tetrachloride, tetrachloroethylene, trichloroethylene, chloroform, bromodichloromethane, and bromoform from water (Zander et al., 1989). Exchange of a dissolved compound with the atmosphere is controlled by the extent of mixing in the aqueous and gaseous phase, the surface area of the interface, the concentration of the compound in the two phases, and the equilibrium distribution of the compound. Technologies that have been developed to enhance gas transfer include: aeration diffusers, packed-tower air stripping, and membrane stripping. Each of these technologies creates a high interface surface area to enhance gas transfer.

9.2 Theory

Oxygen transfer is important in many environmental systems. Oxygen transfer is controlled by the partial pressure of oxygen in the atmosphere (0.21 atm) and the corresponding equilibrium concentration in water (approximately 10 mg/L). According to Henry's Law, the equilibrium concentration of oxygen in water is proportional to the partial pressure of oxygen in the atmosphere.

```
""" importing """
from aide_design.play import*
# the code below will eventually be in the AguaClara core and should be called_
    ↪directly
def O2_sat(P_air, temp):
    """This equation returns saturated oxygen concentration in mg/L. It is valid
    for 278 K < T < 318 K
    Parameters
    -----
    Pressure_air : float
        air pressure with appropriate units.
    Temperature :
        water temperature with appropriate units
    Returns
    -----
    Saturated oxygen concentration in mg/L
    Examples
    -----
    >>> O2_sat(1*u.atm , 300*u.kelvin)
    8.093157231428425 milligram/liter
    """
```

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```

fraction_O2 = 0.21
P_O2 = P_air * fraction_O2
return ((P_O2.to(u.atm).magnitude) *
        u.mg/u.L*np.exp(1727 / temp.to(u.K).magnitude - 2.105))

P_air = 101.3*u.kPa
temp = np.linspace(0, 40)*u.degC
C_Oxygen = O2_sat(P_air,temp)

fig, ax = plt.subplots()
ax.plot(temp,C_Oxygen)
ax.set(xlabel='Temperature (degrees Celsius)', ylabel='Oxygen concentration (mg/L)')
fig.savefig('Gas_Transfer/Images/Oxygen_vs_T')
plt.show()

```

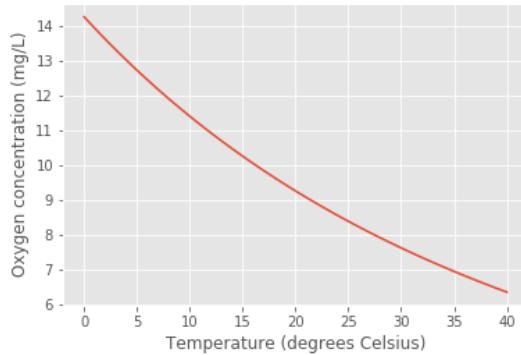


Fig. 9.1: Dissolved oxygen concentrations in equilibrium with the atmosphere.

Natural bodies of water may be either supersaturated or undersaturated with oxygen depending on the relative magnitude of the sources and sinks of oxygen. Algae can be a significant source of oxygen during active photosynthesis and can produce supersaturation. Algae also deplete oxygen levels during the night.

At high levels of supersaturation dissolved gas will form microbubbles that eventually coalesce, rise, and burst at the water surface. The bubbles provide a very efficient transfer of supersaturated dissolved gas to the gaseous phase, a process that can be observed when the partial pressure of carbon dioxide is decreased by opening a carbonated beverage. Bubble formation by supersaturated gasses also occurs in the environment when cold water in equilibrium with the atmosphere is warmed rapidly. The equilibrium dissolved oxygen concentration decreases as the water is warmed (Fig. 9.1).

Supersaturation of dissolved gases can also occur when water carrying gas bubbles from a waterfall or spillway plunges into a deep pool. The pressure increases with depth in the pool and gasses carried deep into the pool dissolve in the water. When the water eventually approaches the surface the pressure decreases and the dissolved gases come out of solution and form bubbles. Bubble formation by supersaturated gases can kill fish (similar to the bends in humans) as the bubbles form in the bloodstream.

9.2.1 Gas Transfer Coefficient

The gas transfer rate can be modeled as the product of a driving force (the difference between the equilibrium concentration and the actual concentration) and an overall volumetric gas transfer coefficient (a function of the geometry,

mixing levels of the system and the solubility of the compound). In equation form

$$\frac{dC}{dt} = \hat{k}_{v,l} (C^* - C) \quad (9.1)$$

where C is the dissolved gas concentration, C^* is the equilibrium dissolved gas concentration, and $\hat{k}_{v,l}$ is the overall volumetric gas transfer coefficient. Although $\hat{k}_{v,l}$ has dimensions of $1/T$, it is a function of the interface surface area (A), the liquid volume (V), the oxygen diffusion coefficient in water (D), and the thickness of the laminar boundary layer (δ) through which the gas must diffuse before the much faster turbulent mixing process can disperse the dissolved gas throughout the reactor.

$$\hat{k}_{v,l} = f(D, \delta, A, V) \quad (9.2)$$

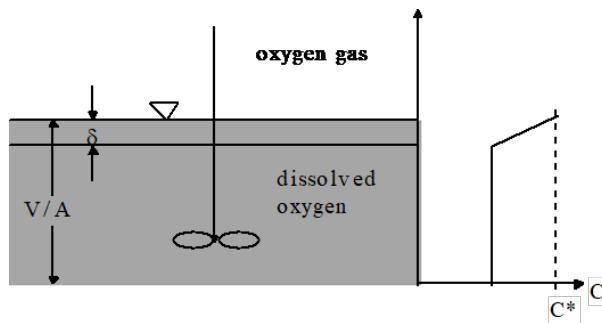


Fig. 9.2: Single film model of interphase mass transfer of oxygen.

The overall volumetric gas transfer coefficient is system specific and thus must be evaluated separately for each system of interest (Weber and Digiano, 1996).

A schematic of the gas transfer process is shown in Fig. 9.2. Fickian diffusion controls the gas transfer in the laminar boundary layer. The oxygen concentration in the bulk of the fluid is assumed to be homogeneous due to mixing and the oxygen concentration above the liquid is assumed to be that of the atmosphere.

The gas transfer coefficient will increase with the interface area and the diffusion coefficient and will decrease with the reactor volume and the thickness of the boundary layer. The functional form of the relationship is given by

$$\hat{k}_{v,l} = \frac{AD}{V\delta} \quad (9.3)$$

Equation (9.1) can be integrated with appropriate initial conditions to obtain the concentration of oxygen as a function of time. However, care must be taken to ensure that the overall volumetric gas transfer coefficient is not a function of the dissolved oxygen concentration. This dependency can occur where air is pumped through diffusers on the bottom of activated sludge tanks. Rising air bubbles are significantly depleted of oxygen as they rise through the activated sludge tank and the extent of oxygen depletion is a function of the concentration of oxygen in the activated sludge. Integrating equation (9.1) with initial conditions of $C = C_0$ at $t = t_0$

$$\int_{C_0}^C \frac{dC}{C^* - C} = \int_{t_0}^t \hat{k}_{v,l} dt \quad (9.4)$$

$$\ln \frac{C^* - C}{C^* - C_0} = -\hat{k}_{v,l}(t - t_0) \quad (9.5)$$

Equation (9.5) can be evaluated using linear regression so that $\hat{k}_{v,l}$ is the slope of the line.

The simple gas transfer model given in equation (9.5) is appropriate when the gas transfer coefficient is independent of the dissolved gas concentration. This requirement can be met in systems where the gas bubbles do not change concentration significantly as they rise through the water column. This condition is met when the water column is shallow, the bubbles have large diameters, or the difference between the concentration of dissolved gas and the equilibrium concentration is small.

9.2.2 Oxygen Transfer Efficiency

An important parameter in the design of aeration systems for the activated sludge process is the energy cost of compressing air to be pumped through diffusers. The pumping costs are a function of the pressure and the airflow rate. The pressure is a function of the hydrostatic pressure (based on the depth of submergence of the diffusers) and the head loss in the pipes and through the diffuser. The required airflow rate is a function of the BOD of the wastewater and the efficiency with which oxygen is transferred from the gas phase to the liquid phase. This oxygen transfer efficiency (OTE) is a function of the type of diffuser, the diffuser depth of submergence, as well as temperature and ionic strength of the activated sludge. Oxygen transfer is a remarkably inefficient process; only a small fraction of the oxygen carried by the rising bubbles diffuses into the activated sludge. The most efficient systems use membrane diffusers and achieve an OTE of approximately 10%.

The manufacturer typically provides oxygen transfer efficiency for a specific diffuser. In this laboratory we will measure oxygen transfer efficiency for the aeration stone that we will be using in an activated sludge tank. The molar transfer rate of oxygen through the diffuser is

$$\dot{n}_{\text{gas } \text{O}_2} = \frac{Q_{\text{air}} P_{\text{air}} f_{\text{O}_2}}{RT} \quad (9.6)$$

where

- f_{O_2} is the molar fraction of air that is oxygen (0.21),
- Q_{air} is the volumetric flow rate of air into the diffuser,
- P_{air} is the air pressure immediately upstream from the diffuser,
- R is the universal gas constant and
- T is absolute temperature.

If the airflow rate is already given with units of moles/s then the molar transfer rate of oxygen can be obtained by multiplying by the molar fraction of air that is oxygen.

The molar rate of dissolution into the aqueous phase is

$$\dot{n}_{\text{aq } \text{O}_2} = \frac{V}{MW_{\text{O}_2}} \frac{dC}{dt} \quad (9.7)$$

where

- MW_{O_2} is the molecular weight of oxygen,
- V is the reactor volume, and
- $\frac{dC}{dt}$ is the change in aqueous oxygen concentration with time.

The rate of change of oxygen concentration is a function of the dissolved oxygen concentration and is a maximum when the dissolved oxygen concentration is zero. Oxygen transfer efficiency could be measured for any dissolved oxygen concentration. A better method of analysis is to substitute the right side of equation (9.1) for $\frac{dC}{dt}$.

$$\dot{n}_{\text{aq } \text{O}_2} = \frac{V \hat{k}_{v,l} (C^* - C)}{MW_{\text{O}_2}} \quad (9.8)$$

The oxygen transfer efficiency is the ratio of equation (9.8) to equation (9.6).

$$OTE = \frac{\hat{k}_{v,l} (C^* - C) VRT}{MW_{\text{O}_2} Q_{\text{air}} P_{\text{air}} f_{\text{O}_2}} \quad (9.9)$$

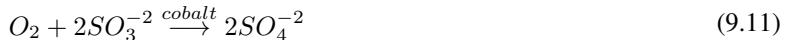
Measurement of OTE using equation (9.9) requires that the gas transfer coefficient, air flow rate, air pressure, and the air temperature be measured. (P_{air} and Q_{air} have to correlate and in this experiment the best combination is atmospheric pressure and the flow rate given by the pump.)

If the molar airflow rate is controlled then OTE is based on the ratio of equation (9.8) to the molar transfer rate of supplied oxygen.

$$OTE = \frac{\dot{n}_{\text{aq } \text{O}_2}}{f_{\text{O}_2} \dot{n}_{\text{air}}} = \frac{V \hat{k}_{v,l} (C^* - C)}{f_{\text{O}_2} \dot{n}_{\text{air}} MW_{\text{O}_2}} \quad (9.10)$$

9.2.3 Deoxygenation

To measure the reaeration rate it is necessary to first remove the oxygen from the reactor. This can be accomplished by bubbling the solution with a gas that contains no oxygen. Nitrogen gas is typically used to remove oxygen from laboratory reactors. Alternately, a reductant can be used. Sulfite is a strong reductant that will reduce dissolved oxygen in the presence of a catalyst.



The mass of sodium sulfite required to deoxygenate 1 mg of oxygen is calculated from the stoichiometry of equation (9.1).

$$\frac{\text{mole } O_2}{32000 \text{ mg } O_2} \cdot \frac{2 \text{ mole } Na_2SO_3}{\text{mole } O_2} \cdot \frac{126,000 \text{ mg } Na_2SO_3}{\text{mole } Na_2SO_3} = \frac{7.875 \text{ mg } Na_2SO_3}{\text{mg } O_2} \quad (9.12)$$

If complete deoxygenation is desired a 10% excess of sulfite can be added. The sulfite will continue to react with oxygen as oxygen is transferred into the solution. The oxygen concentration can be measured with a dissolved oxygen probe or can be estimated if the temperature is known and equilibrium with the atmosphere assumed (Fig. 9.1).

9.3 Experimental Objectives

The objectives of this lab are to:

1. Illustrate the dependence of gas transfer on gas flow rate.
2. Develop a functional relationship between gas flow rate and gas transfer.
3. Measure the oxygen transfer efficiency of a coarse bubble diffuser.
4. Explain the theory and use of dissolved oxygen probes.

A small reactor that meets the conditions of a constant gas transfer coefficient will be used to characterize the dependence of the gas transfer coefficient on the gas flow rate through a simple diffuser. The gas transfer coefficient is a function of the gas flow rate because the interface surface area i.e., the surface area of the air bubbles) increases as the gas flow rate increases.

9.4 Dissolved Oxygen Probes

9.4.1 Theory

The dissolved oxygen probes make use of the fact that an applied potential of 0.8 V can reduce O_2 to H_2O :



The cell is separated from solution by a gas permeable membrane that allows O_2 to pass through. The concentration of O_2 in the cell is kept very low by reduction to H_2O . The rate at which oxygen diffuses through the gas permeable membrane is proportional to the difference in oxygen concentration across the membrane. The concentration of oxygen in the cell is ≈ 0 and thus the rate at which oxygen diffuses through the membrane is proportional to the oxygen concentration in the solution.

Oxygen is reduced to water at a silver (Ag) cathode of the probe. Oxygen reduction produces a current that is measured by the meter.

9.4.2 Calibration

ref Calibrate the dissolved oxygen probe <heading_ProCoDA_Dissolved_Oxygen> after you have assembled the apparatus.

9.5 Experimental Methods

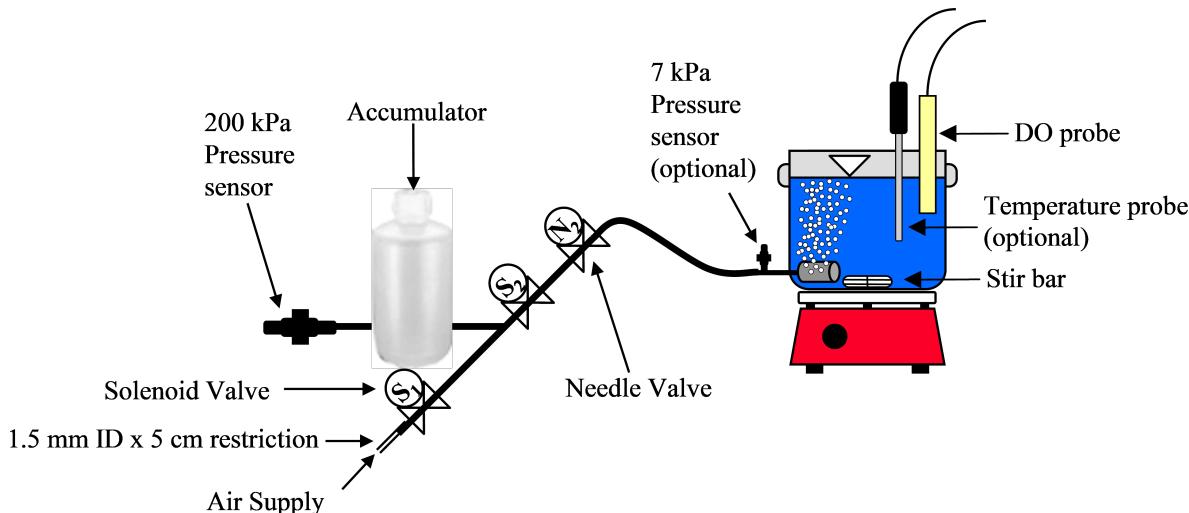


Fig. 9.3: Apparatus used to measure reaeration rate.

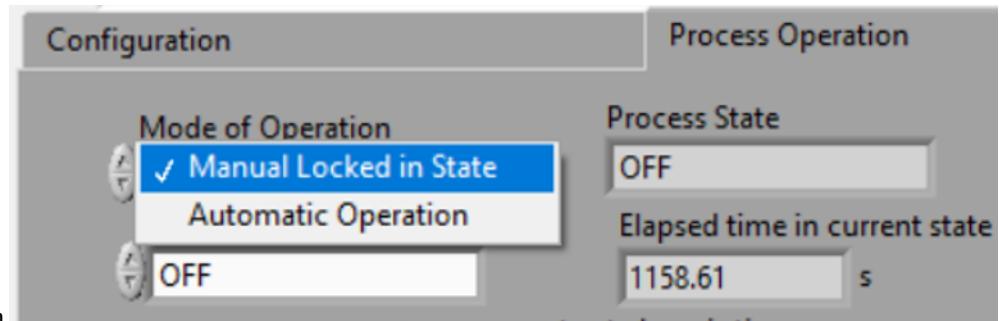
The reactors are 600 mL containers (Fig. 9.3). The DO probe should be placed in a location so as to minimize the risk of air bubbles lodging on the membrane on the bottom of the probe. The aeration stone is connected to a source of regulated air flow. A 7-kPa pressure sensor (optional) can be used to measure the air pressure immediately upstream from the diffuser stone. A 200-kPa pressure sensor is used to measure the air pressure in the accumulator.

9.5.1 Initial Setup

Follow these steps to set up the experiment.

1. Assemble the apparatus (don't forget the 1.5 mm x 5 cm restriction).
2. Install the head loss orifice as close to the valve as possible (plug it directly into the valve!).
3. The ProCoDA II software will be used to control the air flow rate for the aeration experiment. The software will use external code to calculate the calibration constant for the flow restriction, to control valve 1 (the air supply valve), and to regulate the flow of air into the accumulator. The calibration uses the ideal gas law to determine the flow rate as a function of the difference in pressure between the source and the accumulator. Once this calibration is obtained a separate code will set the fraction of time that valve 1 needs to be open to obtain the desired flow rate of air into the accumulator.
4. Use the on the ProCoDA configuration tab to load a method file containing the configuration necessary to control airflow. The file is at S:\Courses\4530\GasTransfer2.pcm. You will need to adjust the channels for the accumulator pressure and the DO probe to match where you plugged them in your ProCoDA box. You will also need to make sure that your valves are connected to the correct ports on the ProCoDA box.
5. Navigate to the Process Operation tab.

6. Set the **operator selected state** to toggle. The solenoid valves should click rhythmically if they are working properly.
7. Install a membrane on the oxygen probe.
8. Add 4 L of tap water to the reactor.



9. Set the mode of operation to automatic operation and the *operator selected state* to prepare to calibrate. The software should quickly cycle through the calibration step and then begin attempting to control the air flow rate to the target value. Note: the purpose of the prepare to calibrate state is to vent excess pressure from the accumulator. The state will not change to calibrate until the pressure drops below a predefined threshold. To speed this up, you may open the top of the air accumulator to release the air *before starting the automatic calibration*.
10. Set the stirrer speed to achieve a vortex on the surface of the water.
11. Calibrate the DO probe if you havent already. Use 22°C as the temperature.

9.5.2 Test the air flow controller

In the following test, the air flow controller should provide a constant flow of air into the accumulator. You can assess how well the air flow controller is working based on the slope of the pressure as a function of time.

1. Set the **mode of operation** to Manual Locked in State.
2. Set the **operator selected state** to off
3. Open the accumulator cap to empty the accumulator.
4. Close the accumulator cap.
5. Close the needle valve.
6. We can set the air flow rate based on our calibration be navigating to the Configuration tab and selecting edit rules. We want to control the air flow rate, so select air flow rate from the set points and variables list. Set the air flow rate to a value of $200\mu\text{M/s}$ ($200 \mu\text{M/s}$).
7. Begin logging data from the 200kPa pressure sensor (accumulator pressure) at a 1 s interval using the datalog button on the configuration tab. Data is being logged when the icon is green.
8. Navigate back to process operation tab and set the **operator selected state** to aerate.
9. End logging data when the accumulator pressure is approximately equal to the source pressure.
10. Analyze the data to see if the airflow rate is close to the expected value. This can be done using the data obtained and the ideal gas law. Plot the accumulator air pressure as a function of time. The slope of the best fit linear line is in units of Pa/s. The volume of the accumulator is 1 liter. Solving the ideal gas law for n gives a result in units of moles/s. You set the air flow rate for $200 \mu\text{M/s}$ and that is what you are expecting from this calculation.
11. If the error is greater than 20% look for leaks and recalibrate the airflow controller.

9.5.3 Measure the Gas Transfer

1. Call the instructor and/or TA to check the system configuration.
2. The instructor or TA will add 0.1 mg $CoCl_2 \cdot 6H_2O$ (note this only needs to be added once because it is the catalyst). A stock solution of $CoCl_2 \cdot 6H_2O$ (1 mg/mL – thus add 100 μL) has been prepared to facilitate measurement of small cobalt doses. (Use gloves when handling cobalt!)
3. Prepare to record the dissolved oxygen concentration using ProCoDA software. Use 5-second data intervals and log the data to S:\Courses\4530\Group #\gastran\x where x is the flow rate in $\mu M/s$ for later analysis. Include the actual flow rate in the file name.
4. Set the airflow rate to the desired flow rate. Each group will investigate six flowrates. The instructor will assign the flowrates on the day of the lab exercise.
5. Set the **operator selected state** to aerate.
6. Set the needle valve so the pressure in the accumulator is approximately 75% of the source pressure.
7. Wait until the accumulator pressure reaches steady state.
8. Turn the air off by changing the operator selected state to OFF.
9. Add enough sodium sulfite to deoxygenate the solution. A stock solution of sodium sulfite (100 mg/mL) has been prepared to facilitate measurement of small sulfite doses. Calculate this dose based on the measured dissolved oxygen concentration. ($4 \text{ L of water at } C_{\text{oxygen}} \text{ mg O}_2/\text{L} = 4 C_{\text{oxygen}} \text{ mg O}_2$, therefore add $4(7.875)C_{\text{oxygen}}$ mg sodium sulfite or $4(7.875)(C_{\text{oxygen}})/100mL$ of stock solution.)
10. Turn the air on by changing the **operator selected state** to Aerate.
11. Monitor the dissolved oxygen concentration until it reaches 50% of saturation value or 10 minutes (whichever is shorter).
12. Repeat steps 3-11 to collect data from at least two additional flow rates.
13. Consolidate the files into one spreadsheet file with a separate sheet for each flow rate.
14. Collect data from the whole class to analyze the full spectrum of flow rates investigated.

9.6 Prelab Questions

1. Calculate the mass of sodium sulfite needed to reduce all the dissolved oxygen in 600 mL of pure water in equilibrium with the atmosphere and at $22^\circ C$.
2. Describe your expectations for dissolved oxygen concentration as a function of time during a reaeration experiment. Assume you have added enough sodium sulfite to consume all of the oxygen at the start of the experiment. What would the shape of the curve look like?
3. Why is $\hat{k}_{v,l}$ not zero when the gas flow rate is zero? How can oxygen transfer into the reactor even when no air is pumped into the diffuser?
4. Describe your expectations for $\hat{k}_{v,l}$ as a function of gas flow rate. Do you expect a straight line? Why?
5. A dissolved oxygen probe was placed in a small vial in such a way that the vial was sealed. The water in the vial was sterile. Over a period of several hours the dissolved oxygen concentration gradually decreased to zero. Why? (You need to know how dissolved oxygen probes work!)

9.7 Data Analysis

This lab requires a significant amount of repetitive data analysis. Plan how you will organize the analysis to be as easy as possible.

1. Calculate the air flow rate from testing the air flow controller and compare with the target value.
2. Eliminate the data from each data set when the dissolved oxygen concentration was less than 0.5 mg/L. This will ensure that all of the sulfite has reacted.
3. Plot a representative data set showing dissolved oxygen vs. time.
4. Calculate C^* based on the average water temperature, barometric pressure, and the following equation. $C^* = P_{O_2} e^{(\frac{1727}{T} - 2.105)}$ where T is in Kelvin, P_{O_2} is the partial pressure of oxygen in atmospheres, and C^* is in mg/L. This equation is valid for $278 \text{ K} < T < 318 \text{ K}$.
5. Estimate $\hat{k}_{v,l}$ using linear regression and equation (9.5) for each data set.
6. Create a graph with a representative plot showing the linearized data, $\left(\ln \frac{C^* - C}{C^* - C_0} \right)$ vs. time, and the best-fit line.
7. Plot the reaeration model on the same graph as the dissolved oxygen vs. time data. This is done by solving equation for C.
8. Plot $\hat{k}_{v,l}$ as a function of airflow rate ($\mu\text{mole}/\text{s}$).
9. Look at each dataset and if necessary (to make more linear plots) eliminate more data from the beginning (or end) of the dataset. You will be able to see when the oxygen level is affected by residual sulfite at the beginning of the experiments.
10. Plot OTE as a function of airflow rate (?mole/s) with the oxygen deficit ($C^* - C$) set at 6 mg/L.
11. Plot the molar rate of oxygen dissolution into the aqueous phase ($\mu\text{mole}/\text{s}$) as a function of airflow rate ($\mu\text{mole}/\text{s}$).
12. Comment on results and compare with your expectations and with theory.
13. Verify that your report and graphs meet the requirements.

9.8 References

Weber, W. J. J. and F. A. Digiano. 1996. Process Dynamics in Environmental Systems. New York, John Wiley & Sons, Inc.
 Zander, A. K.; M. J. Semmens and R. M. Narbaitz. 1989. **Removing VOCs by membrane stripping** American Water Works Association Journal 81: 76-81.

9.9 Lab Prep Notes

Table 9.1: Reagent list.

Description	Supplier	Catalog number
Na_2SO_3	Fisher Scientific	S430-500
$CoCl_2 \cdot 6H_2O$	Fisher Scientific	C371-100

9.9.1 Setup

1. Prepare the sodium sulfite immediately before class and distribute to groups in 15 mL PP bottles to minimize oxygen dissolution and reaction with the sulfite.
2. The cobalt solution can be prepared anytime and stored long term.
3. Install the membranes on the DO probes and confirm that all probes are operational.
4. Provide clamps to mount DO probes on magnetic stirrers.
5. Determine maximum airflow rate that is reasonable given small reactor volume.

Table 9.2: Suggested flow rates.

Group Flows	($\mu M/s$)
1	200, 250, 300
2	350, 400, 450
3	500, 600, 700
4	800, 900, 1000
5	1200, 1500, 2000
6	3000, 4000, 5000

9.9.2 Class Plan

1. Show how to calibrate DO probe using Calibrator.
2. Assign groups different gas flow rates

9.10 Airflow Control

The ProCoDA software can be configured to control the flow of air into the reactor. The hardware required is shown in Fig. 9.3. The control algorithm is based on the theoretical relationship between head loss and flow rate for the air flowing into the accumulator. We can empirically measure the head loss coefficient and then use the theoretical relationship to determine what fraction of time the influent valve should be open to obtain the desired flow rate. We can use the change in pressure in the accumulator when the influent valve is open to determine how fast air was flowing into the accumulator. In order to develop an appropriate head loss model we need to know if the flow into the accumulator is laminar or turbulent.

$$Re = \frac{\rho V D}{\mu} \quad (9.14)$$

$$Re = \frac{4\rho Q}{\pi d \mu} \quad (9.15)$$

If we hold pressure and temperature constant and then take the derivative of the ideal gas law we obtain.

$$P \dot{V} = \dot{n} R T \quad (9.16)$$

and since change in volume with respect to time is a flow rate we have

$$Q = \frac{\dot{n} R T}{P} \quad (9.17)$$

Density of an ideal gas is given by

$$\rho = \frac{P M_{gas}}{R T} \quad (9.18)$$

Substituting these relationships into equation (9.14) for Reynolds number we obtain

$$Re = \frac{4\dot{n}M_{gas}}{\pi d\mu} \quad (9.19)$$

For the air flow controller used in the lab the following values are obtained

\dot{n} max flow is about 10,000 $\mu M/s$ M_{gas} is 0.029 kg/M, and μ is $1.8 \times 10^{-5} Ns/m^2$.

$$Re = \frac{4(10000 \times 10^{-6} \frac{M}{s})(0.029 \frac{kg}{M})}{\pi(1 \times 10^{-3} m)(1.8 \times 10^{-5} \frac{Ns}{m^2})} = 20,500 \quad (9.20)$$

The flow into the air accumulator will almost certainly be turbulent and thus we can use the turbulent flow equations for minor losses to describe head loss. The equation for minor losses is:

$$h_{minor} = K \frac{8Q^2}{g\pi^2 D^4} \quad (9.21)$$

To use equation (9.21) for air we substitute pressure change for h_{minor} , equation (9.17) for flow rate, and equation (9.18) for density.

$$\Delta p = K \frac{8M_{gas}RT\dot{n}^2}{\pi^2 D^4 P} \quad (9.22)$$

This change of pressure is occurring between the air supply and the accumulator. The pressure, P , in equation (9.22) helps determine the velocity of the air and thus head loss is a function of the pressure. The pressure varies between the pressure of the lab air supply, P_s , and the pressure in the air accumulator, P_a . As a reasonable first approximation we use the average pressure of the supply and the accumulator for P , the difference in pressure for Δp , and solve equation (9.22) for the molar flow rate.

$$\dot{n} = \frac{\pi D^2}{4\sqrt{KM_{gas}RT}} \sqrt{(P_s - P_a)(P_s + P_a)} \quad (9.23)$$

where \dot{n} is the molar flow rate.

Multiplying terms and noting that the supply pressure is relatively constant, but that the accumulator pressure varies as it charges according to the ideal gas law we obtain.

$$\frac{dn}{dt} = \frac{\pi D^2}{4\sqrt{KM_{gas}RT}} \sqrt{P_s^2 - \frac{n^2 R^2 T^2}{V^2}} \quad (9.24)$$

Separating terms and integrating from an initial condition with n_1 moles to a final condition with n_2 moles in the accumulator.

$$\frac{V}{RT} \int_{n_1}^{n_2} \frac{dn}{\sqrt{\frac{P_s^2 V^2}{R^2 T^2} - n^2}} = \int_0^t \frac{\pi D^2}{4\sqrt{KM_{gas}RT}} dt \quad (9.25)$$

After integrating we obtain the following equation.

$$\frac{V}{RT} \left(\sin^{-1} \frac{n_2 RT}{P_s V} - \sin^{-1} \frac{n_1 RT}{P_s V} \right) = \frac{\pi D^2 t}{4\sqrt{KM_{gas}RT}} \quad (9.26)$$

$$t = \frac{4\sqrt{KM_{gas}RT}}{\pi D^2} \left(\sin^{-1} \frac{n_2 RT}{P_s V} - \sin^{-1} \frac{n_1 RT}{P_s V} \right) \frac{V}{RT} \quad (9.27)$$

Since we will be measuring the pressure in the accumulator we can now substitute that pressure for the terms containing moles of air to obtain an equation that is in a linear form such that a single term containing K and D can be obtained by linear regression.

$$t = \frac{4\sqrt{KM_{gas}RT}}{\pi D^2} \left(\sin^{-1} \frac{P_{a2}}{P_s} - \sin^{-1} \frac{P_{a1}}{P_s} \right) \frac{V}{RT} \quad (9.28)$$

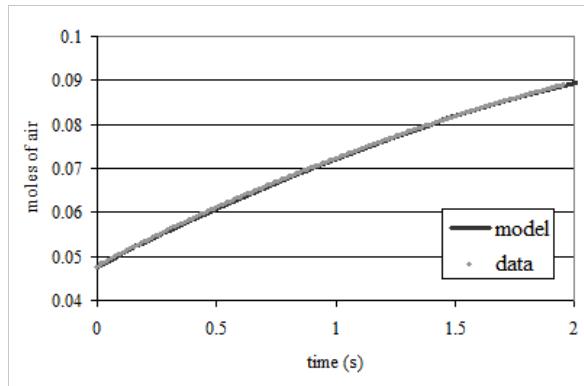


Fig. 9.4: The model describing the filling of the accumulator fits the data very well.

Taking a data set obtained by filling the accumulator, finding the unknown term $\frac{4\sqrt{KM_{gas}RT}}{\pi D^2}$ by linear regression and then plotting the resulting model next to the data we obtain Fig. 9.4.

The final step is to calculate the fraction of time that the valve must be open in order to obtain a desired flow rate into the accumulator. Take the target air flow rate \dot{n}_{target} and divide by the molar flow rate given by equation (9.23) to get the fraction of time the valve must be open to get the desired average flow rate.

$$f_{valve} = \frac{\dot{n}_{target}}{\dot{n}} = \frac{\dot{n}_{target}}{\frac{\pi D^2}{4\sqrt{KM_{gas}RT}} \sqrt{P_s^2 - P_a^2}} \quad (9.29)$$

Equation (9.29) assumes that inertial effects during flow startup are not significant. Application of equation (9.29) results in slightly more air being delivered than requested. The reason for this error is that when the valve is closed the volume between the location of the head loss and the valve fills to the pressure of the source. This volume of air quickly discharges through the valve as soon as the valve is opened. This error can be minimized by using small valves and by keeping the head loss orifice as close to the valve as possible.

Equation (9.29) is used by the air flow control.vi to calculate the fraction of time that the valve should be open. The ability of the control algorithm to create a desired flow rate can be measured by setting the flow rate and closing the effluent valves from the accumulator. The result is that the accumulator will gradually fill and as it fills f_{valve} will gradually increase so the flow rate into the accumulator remains constant. The slope of the pressure vs. time line is proportional to the flow rate.

NUTRIENT REMOVAL WITH SEQUENCING BATCH REACTORS

10.1 Process Control

An introduction to automated process control is available in pages 1681 to 1703 of (Metcalf & Eddy, Tchobanoglous et al. 2003).

10.1.1 Sensors

Sensors that can be used to monitor the status of the sequencing batch reactor include pressure, temperature, dissolved oxygen, pH, and turbidity. The pressure sensors are quite versatile and can be used to measure airflow, water flow, volume of water in the reactor, as well as head loss through the coarse bubble diffuser.

10.2 Measure Oxygen Uptake

One of the objectives of a wastewater treatment plant is to reduce the Biochemical Oxygen Demand (BOD). The minimum national standard for secondary wastewater treatment is that the average 30-day concentration of BOD_5 be less than 30 mg/L. Biochemical oxygen demand is difficult to measure since it takes 5 days for a test. The long test period also precludes the possibility of using BOD as a control parameter in operating a WWTP. Most WWTPs don't have the luxury of knowing the concentration of influent BOD. For the NRP the composition and properties of the synthetic feed are known. Thus it should be possible to estimate the BOD removal and the residual BOD by measuring the oxygen uptake rate. Temporarily increasing the oxygen concentration in the sequencing batch reactor, turning the airflow off, and then measuring the decrease in oxygen concentration with time can measure the oxygen uptake rate. The aeration rate with the airflow turned off is insignificant and thus the rate of oxygen consumption is equal to the rate of change of the oxygen concentration.

10.3 References

- Cicek, N., J. P. Franco, et al. (1998). Using a Membrane bioreactor to reclaim wastewater. Journal American Water Works Association 90(11): 105-113. Metcalf & Eddy, I., G. Tchobanoglous, et al. (2003). Wastewater Engineering: Treatment and Reuse. New York, McGraw Hill. Rittmann, B. E. and P. L. McCarty (2001). Environmental Biotechnology: Principles and Applications. New York, McGraw Hill. R. Mikler, W. Kramer, O. Doblhoff - Dier, K. Bayer (04/07/95) Strategies For Optimal Dissolved Oxygen (Do) Control <https://doi.org/10.1016/B978-0-08-042377-7.50059-3>

10.4 Lab Setup

For 6 weeks of operation of 4 plants prepare 20 L of 100x organic stock. Mix in a 20 L Jerrican and use a power mixer inserted through the opening to mix after the addition of each chemical. Store 100x stock in a refrigerator.

Table 10.1: List of chemicals needed for synthetic waste

Compound	Chemical Formula	Molecular Weight (g/mol)	Concentration (mg/L)	total grams required (g)	stock
Starch		40,000	84.40	162.048	1
Casein		30,000	125.00	240	1
Sodium acetate	$C_2H_3O_2Na \cdot 3H_2O$	136.1	31.90	61.248	1
Capric acid	$C_{10}H_{20}O_2$	172.3	11.60	22.272	1
Ammonium chloride	NH_4Cl	53.5	75.33	144.6336	1
Potassium phosphate	K_2HPO_4	174.2	6.90	13.248	1
Sodium hydroxide	$NaOH$	40.0	1.75	3.36	1
Glycerol	$C_3H_8O_3$	92.1	12.00	23.04	1
Magnesium sulfate	$MgSO_4 \cdot 7H_2O$	246.5	69.60	133.632	2
Sodium molybdate	$NaMoO_4 \cdot 2H_2O$	241.9	0.15	0.288	2
Manganese sulfate	$MnSO_4 \cdot H_2O$	169.0	0.13	0.2496	2
Cupric sulfate	$CuSO_4 \cdot 4H_2O$	249.7	0.08	0.1536	2
Zinc sulfate	$ZnSO_4 \cdot 7H_2O$	287.5	0.48	0.9216	3
Calcium chloride	$CaCl_2 \cdot 2H_2O$	147.0	22.50	43.2	3
Iron chloride	$FeCl_3 \cdot 6H_2O$	270.3	18.33	35.1936	3
Cobalt chloride	$CoCl_2 \cdot 6H_2O$	237.9	0.42	0.8064	3

PROCODA: PROCESS CONTROL AND DATA ACQUISITION

11.1 Abstract

ProCoDA is easily configured process control and data acquisition software and hardware that significantly enhances experimental evaluation of unit processes and that permits fully automated parametric studies. The process control software is a LabVIEW executable and incorporates sensors, states, rules, an external microprocessor for controls, the ability to link to external LabVIEW code to add new features, and data logging. The composite system is flexible and user friendly. First year undergraduates have used the software to control a bench top drinking water treatment plant and both undergraduate and graduate students have used the software to conduct parametric studies of methane generation, the activated sludge process, flocculation, and porous media filtration. The system offers the user the ability to program an entire series of experimental runs including variation of a parameter over a series of values and inclusion of replicate tests to confirm repeatability of the results. The automated parametric testing has greatly increased our ability to study the response of complex systems. Evaluation of colloidal particle flocculation and porous media filtration in bench-scale experiments are presented here as example applications of the ProCoDA software.

11.2 Introduction

Process control is commonly used in undergraduate teaching laboratories in electrical and mechanical engineering with an emphasis on the PID (proportional, derivative, integral) control algorithm (Braatz and Johnson, 1998). Process control laboratory experiences typically focus on control of a single parameter such as flow rate, temperature, liquid level, pressure (Rehg et al., 1999), or motor angular velocity (Choi, 2004). Control of real processes generally requires integration of individual process control components into a larger system that includes data acquisition and data logging, as well as the concepts of process state and operating rules. Although researchers have frequently created customized process control code for specific applications ProCoDA is a highly configurable system that can be used to automate the control of parametric experiments or complex processes.

Experimental work is often characterized by repetitive experiments and the need for an individual to configure each of those experiments and then measure and record the data. In the case of engineered environmental systems we are often interested in optimizing the performance of unit processes and exploring the response of processes to a range of inputs. It is often necessary to cycle a selected process through a sequence of operational stages or states, to change process related variables, and to subject the process to a range of inputs. For example, operation of porous media filtration requires implementation of backwash, optional pretreatment, and particle challenge states. Filtration process variables might include flow velocities or the duration of a state, and inputs might include variation in the type and concentration of colloidal particles or coagulant type and dose. In conventional jar tests an experimenter would evaluate flocculation efficiency based on the turbidity of settled water or measure the critical sedimentation velocity by taking a batch sample of the flocs and measuring the turbidity as the sample settles. The measurement of settled water turbidity requires first loading the sample vial with flocs, then measuring turbidity as the flocs settle, and finally purging the sample vial with clean water. Although the states to control porous media filtration and to measure the effectiveness of flocculation could be implemented by a human operator, they are well suited for automation.

We have taken the automation components that are commonly used to control industrial and municipal processes and adapted them for use in the laboratory. Our objective was to create process control software that was highly configurable and link it to sensors and outputs for control. The ProCoDA software began as data acquisition software capable of remote monitoring of experiments with easily configured sensors. The ability to control on/off devices (valves, pumps, mixers, etc.) and the ability to control continuously variable devices such as a variable flow peristaltic pumps was then added. The process control and data acquisition components were integrated and a user friendly programming environment that is highly configurable was created that incorporates the concepts of states, rules, constants, variables, and customizable external functions.

The guiding principle in developing the ProCoDA code was to create an application that was general and easily configured to handle a wide variety of challenges. Thus the code is not designed for a particular unit process. The goal of designing code that was highly adaptable required implementing features that make it possible for users to customize the software for specific tasks. The ProCoDA software includes a built in programming environment for defining variables, constants, and sensors and for writing the logic that controls moving from one process state to the next. To extend the capabilities of the software without making the interface overly complex the ability to use external code to define new functions was added. Examples of external code that we have used to extend the capabilities of the ProCoDA software include proportional integral derivative control algorithms, acquisition of data from turbidimeters and electronic balances over USB ports, functions that incrementally vary a parameter over a specified range and functions that take a flow rate and tubing size and calculate the peristaltic pump speed.

The ProCoDA software was written using LabVIEW. The high level predefined functions, the ease of developing graphical user interfaces, and the integration with input and output devices made LabVIEW the preferred programming environment. In addition to the software we designed hardware interfaces to simplify the use of sensors and controls.

The ProCoDA software runs five parallel processes during normal operation. The five processes are:

1. Data acquisition from the ProCoDA hardware
2. Calculation of output values, processing of rules to determine state changes, sending commands to the ProCoDA hardware, and logging of state changes and the rule that caused the state to change
3. Data logging of sensor data as well as all user defined variables
4. Graphical display of all sensor data as well as all user defined variables
5. An event loop that responds to 20 user events to initiate processes such as sensor configuration, rule editing, pump calibration, and hardware selection.

The parallel processes are asynchronous and share data through the use of code objects (LabVIEW SubVIs) that contain both data and methods.

A core feature of the ProCoDA software is the user programmable feature that facilitates customization of the control logic for specific tasks. The rule editor provides a programming environment for setting up states, control logic, set points, variables defined by links to external code, and selecting which user defined parameter controls each output in each state.

Sensor signal wires and power supply connections have been integrated in a single cable to make sensors easy to use in a student laboratory environment. The sensors use a standardized wiring scheme so that a variety of sensors can be used in a connect-calibrate-measure approach. The elimination of power supplies at each experiment location and all of the associated wiring has made sensor use easy for students. The process control software interface for sensors allows adding and configuring sensors while simultaneously collecting data.

The ProCoDA hardware/software can vary the state of on/off devices as well as control devices that require a 4-20 mA signal. The on/off outputs are 24 V with sufficient current to operate small solenoid valves or to switch relays for controlling the power to any number of devices. The 4-20 mA analog outputs are used to control peristaltic pumps. External code is already provided for communication with digital devices including HF scientific turbidimeters, HF scientific AccUVView UV meter. We expect to add external code to control digital peristaltic pumps in the near future.

ProCoDA software allows the user to automate data collection and to control many parts of the experimental apparatus. This enables researchers to focus on experimental design and data analysis. Process control software can control the

speed of peristaltic pumps, regulate flow through solenoid valves, and measure (or control with feedback) water quality parameters such as temperature, pressure, and dissolved oxygen.

The ProCoDA software is designed to allow complete automation of experiments including running a series of tests where a parameter is adjusted to a different value for each test. ProCoDA is a state machine. That means that you can create a series of states. Each state has its own logic for which state to go to next and what to do while in that state. A simple example of a state machine is a washing machine. Washing machine states could include

Table 11.1: Washing machine states

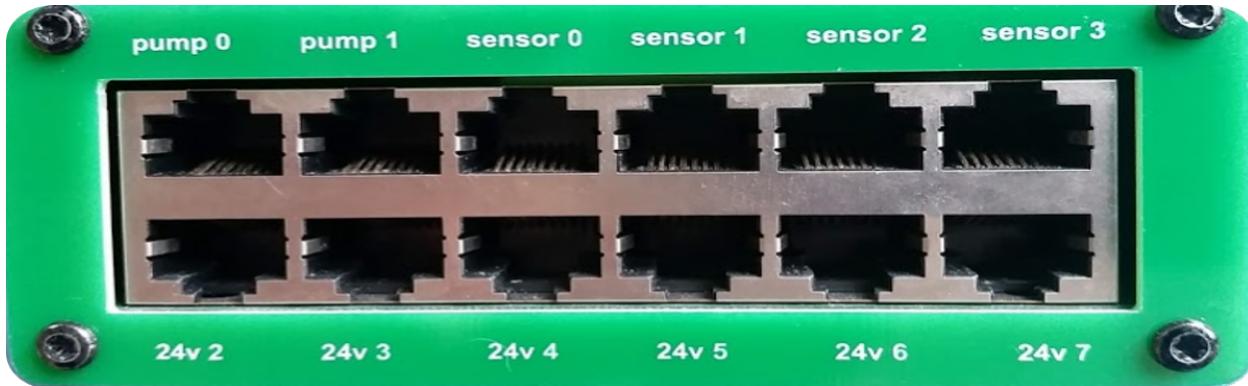
State	What it does	Exit rule
Fill	Fills tub with selected amount of water at the requested temperature	Switch to <i>Agitate</i> state when pressure sensor reaches target value
Agitate	Agitate to clean clothing	Switch to <i>Spin to remove dirty water</i> after target time is reached
Spin to remove dirty water	open drain and spin	Switch to <i>Rinse</i> after target time is reached
Rinse	Add water to target level while agitating	Switch to <i>Spin dry</i> when target time is reached.
Spin dry	Spin and drain water	Switch to <i>off</i> when target time is reached.

- Setpoints: There are two types of setpoints in ProCoDA, constants and variables. These are the numbers that ProCoDA uses to run any of its functions (These are things like warm or cold water temperatures)
- States: States are the different ways ProCoDA can run (These are the steps of a laundry cycle like agitate, rinse, spin)
- Rules: These control when ProCoDA changes between states (These are like going from rinse to spin)

11.2.1 General Notes

- When you input a value that it a decimal it will convert it into a form without the decimal (i.e. 0.3 will become 300m)
- The default unit of time is seconds and we recommend using seconds consistently
- ProCoDA doesn't actually use units internally. The units are only used for documentation. It is the users responsibility to make sure that you are using a consistent set of units.
- The required set points give the order of the set points and the set points must be in the same order when you input them
- All added set points should be added after ON and OFF,
- As a general rule, add new set points and states at the bottom of the list using add after. This is because ProCoDA only tracks set points and states as elements in an array and thus if the array elements are shifted ProCoDA will refer to the wrong element.

The hardware consists of a box with a National Instruments data acquisition board that connects to the computer via a USB port. The box has 12 ports.



1. pump 0 and 1: Ports that control on/off, direction, and speed of peristaltic pumps. The on/off and direction controls are based on digital outputs from the data acquisition board. The speed control uses an analog output from the data acquisition board that is converted into a 4-20 mA signal.
2. sensor 0 to 3 measure voltages from sensors. The ports also contain +5V, -5V, +10V, +15V, -15V power supplies that are used to power sensors (pressure, photometer) or is used to power signal conditioning circuits that are required by some sensors (pH, dissolved oxygen, temperature, photometer).
3. 24V 2 to 7 provide 24 volt outputs that can be used to power solenoid valves or relays that can then power larger electrical loads.

The distinction between **inputs and outputs** is **critical**. The pump and 24V ports are all outputs. They **control devices**. The sensor ports are inputs that are used to **measure**. Dont plug sensors into outputs! Dont plug pumps into 24 volt ports or into sensors! Dont plug solenoid valves into pump or sensor ports! Dont confuse in and out!

The software combines 3 elements: sensors (inputs from the real world), set points (inputs from the plant operator and calculated values based on sensors and other set points), and logic (rules that govern how the plant should operate given the sensor data and set points). The software contains a graphical user interface where you can edit, save, and open files containing sensor information and files containing the set point and logic information.

11.3 Data Acquisition

The ProCoDA software is designed to receive data from two types of digital sources. Sensors or meters that have their own digital communication interface can transmit their data to the ProCoDA software through external code that is written specifically for each meter. For example this method is used to acquire data from HF Scientific turbidimeters. Sensors that can be configured to produce an analog voltage such as pressure, pH and dissolved oxygen are monitored with a National Instruments USB data acquisition board.

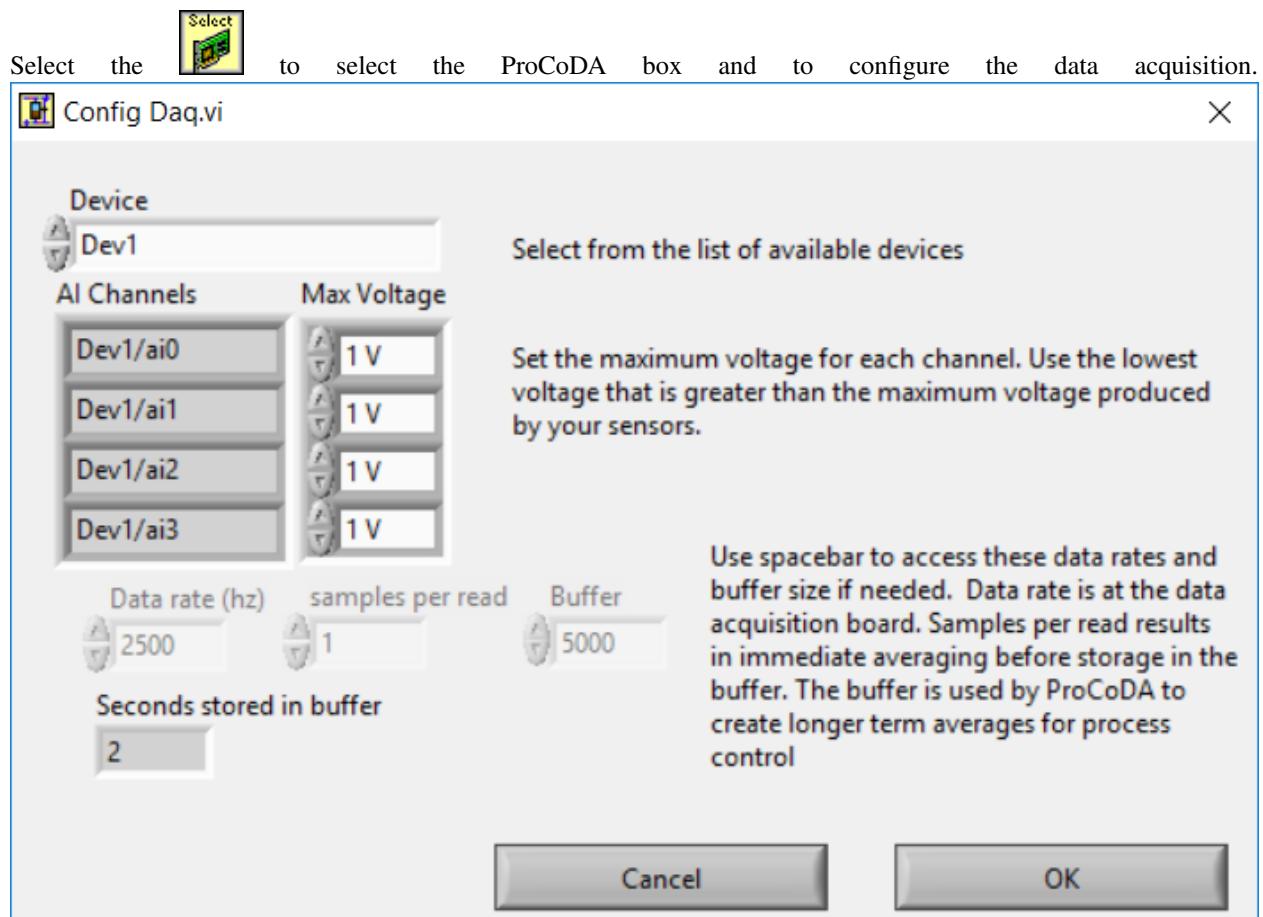
Sensors are wired using Category 5 or 6 Ethernet cable using our own *analog* wiring standard. Two wires are used to carry the sensor output voltage designated as S- and S+. Five wires are used for different voltage power supplies, and one wire is used for the power supply ground. Sensors are wired to connect to the power supply that they require. Thus, the sensor is powered and monitored through a single port. This scheme is used for a variety of sensors including pressure, strain gage, pH, dissolved oxygen, photometer, and temperature. For several of the sensors the power supplies are also used to power a signal conditioning circuit to transform the sensor output into a voltage that can easily be monitored with a data acquisition system.

Table 11.2: Wiring standard used for combining power supplies and analog data acquisition in a Category 5 Ethernet cable

T-568B standard	T-568A standard	voltage
white/orange	white/green	S-
white/green	white/orange	ground
white/brown	white/brown	-15 V
white/blue	white/blue	+5 V
orange	green	S+
green	orange	+10 V
brown	brown	+15V
blue	blue	-5 V

11.4 Configure

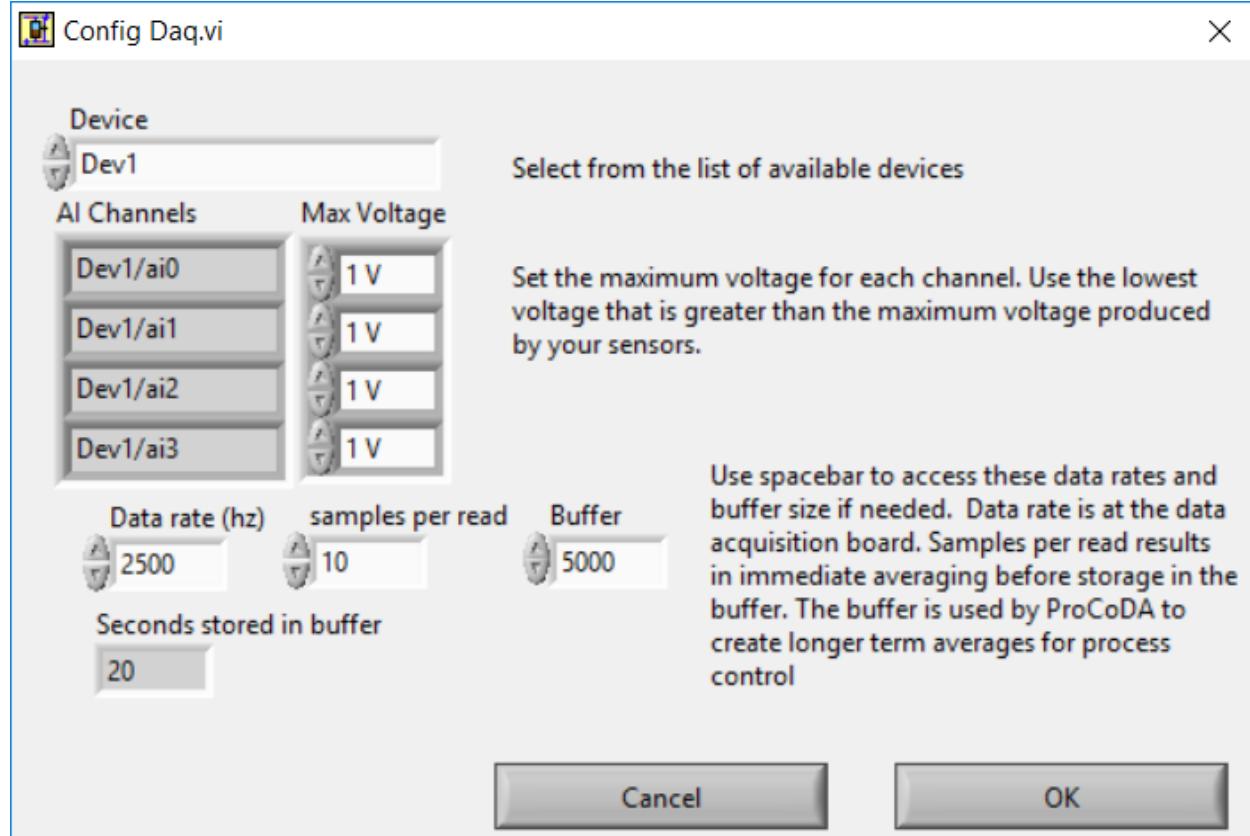
The configure tab of ProCoDA is used to select the ProCoDA box that will be controlled by the software. It is possible to connect more than one ProCoDA box to a single computer and have multiple instances of the ProCoDA software running at the same time. But that is a capability that we havent truly tested and most users want to control a single experiment with one ProCoDA box.



The available analog input (AI) channels (for sensors) are displayed along with the maximum voltage that can be measured. Most of our sensors have an output voltage of less than 1 volt and thus the maximum voltage can be set to 1 volt. The exception is the photometer that has an output maximum voltage of 5 V.

ProCoDA keeps a small amount of data in memory at all time that can be used for making decisions. This is critical because sensors area always noisy and thus it is poor practice to make decisions based on instantaneous measurements. Instead we use an average of recent data and select the amount of averaging based on the requirements for the control system. The length (in seconds) of the data record that is available in the buffer is set by the size of the buffer and the rate of sampling. The highest rate of sampling is 2500 Hz. We recommend that sampling be as fast as possible and that data averaging be used to smooth the data.

Data averaging is implemented by pressing the spacebar and then editing the number of samples per read. In the example below we have set samples per read to 10. In this case the data is read at 250 Hz in 10 sample chunks. Each chunk of 10 samples is averaged on its way into the ProCoDA program. Thus the data is smoothed and results in less noisy signals.



The ProCoDA circuitry that sets the peristaltic pump speed can be calibrated (in software) with . This calibration ensures that the pump actually rotates at the speed set by ProCoDA. This calibration only needs to be done once for each pump that is controlled.

ProCoDA has the ability to access data from other ProCoDA software on other computers if there is a shared server

where the data can be shared. This data sharing makes it possible for multiple users to have access to data that is being logged at one location. In the AguaClara laboratory researchers can access the laboratory water temperature using this system. In the Environmental Engineering teaching laboratory this feature is used to enable all of the workstations to access the pressure of the air supply that is used for the gas transfer experiments.

11.4.1 Methods

ProCoDA is highly configurable (it is after all, a friendly programming environment for laboratory automation) and those configurations or methods are saved in files. ProCoDA automatically saves ALL changes in configuration as they happen in C:\ProCoDA Data\ProCoDA 0.pcm where the integer refers to the instance of the ProCoDA software if multiple instances are being used. This method file is automatically loaded when ProCoDA is launched. Very occasionally ProCoDA crashes and creates a corrupted method file and thus refuses to launch properly. In that case simply delete this file.

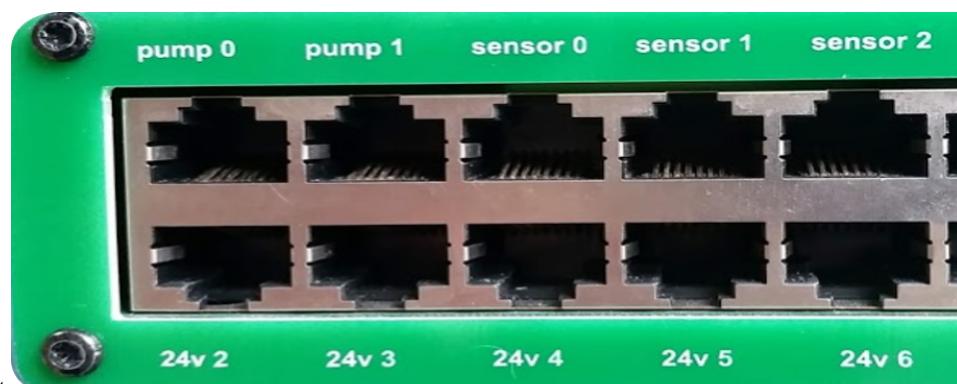
Given that ProCoDA automatically saves the method file that means that any mistakes in editing are immediately saved as well. To safeguard against this ProCoDA also saves a copy of the method file in the folder location where the data is being logged. We recommend that copies of the ProCoDA method also be saved in a secure location by the researcher as a third level of safety. Use the buttons to save the current method, retrieve a method from file, or export the method in a tab delimited file in a human readable format.

11.5 Sensors

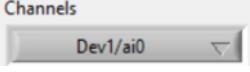
Monitoring sensors requires conversion of the measured voltage into a physically meaningful unit. The data acquisition module of the ProCoDA software uses conversion files to implement a variety of conversion algorithms including polynomials and correspondence tables as well as the calibration algorithms required for pH, dissolved oxygen, and photometers. The pressure sensor conversion files make it easy to use pressure sensors to measure pressures in various physical units, to measure reactor volumes (of known cross sectional area), and flow rates (through devices with known relationships between flow and pressure drop). In addition to the application of a conversion to a physical unit it is possible to calibrate the pressure sensor output to a specific value by changing an offset.

The sensor part of ProCoDA includes software that eliminates the need for pH meters, dissolved oxygen meters, temperature meters. A software interface for a single wavelength photometer is also included. Any sensor that has a voltage output can be monitored.

Add a sensor to the list of sensors by either inserting a new unconfigured sensor, , or by selecting a configured sensor that you want to duplicate, . The will automatically increment the channel that the sensors are connected to. You can always any channels that you dont want.



Each sensor must be connected to a sensor port

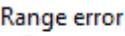
using  Note that it is possible to monitor the same port more than once using ProCoDA. This would be useful if you wanted to simultaneously log both the raw voltage and a calibrated physical unit from the same sensor. This would be particularly useful if you want the option to change the conversion from voltage to physical units.

Sensor that have simple linear relationships between voltage and calibrated output can be easily adjusted

 For example, a pressure sensor can be calibrated by setting its output to an independently measured value  Or the pressure could be zeroed  under conditions of no flow (if you are measuring head loss). The offsets can be cleared, , to return to the original uncalibrated sensor output.

 The  can be used to view how the voltage is being converted to a physical unit. This makes it possible to edit the conversion values and can be used to create conversion files for new sensors. New conversion files can be saved  for use later.

One of the failure modes with ProCoDA occurs when a sensor produces a voltage that is outside of the range 

 that was set when configuring the data acquisition board (see *ProCoDA Configure*). It is critical that the  sensor voltage not be out of range. Sensors that are out of range provide useless data! Your goal is to see .

11.5.1 Pressure Measurement

We use 1 psi (7 kPa) and 30 psi (200 kPa) pressure sensors (Omega sensor models PX26-001DV, and PX26-030DV) in our laboratory to measure water depth, reactor volumes, flow rates, and head loss. These sensors have maximum output voltages of 16.7 mV and 100 mV respectively with a power supply of 10 V. We use the differential pressure model since the sensors can be used to measure gage pressure or differential pressure. The sensors can directly measure water pressure although the electrical connections **must be kept dry**. The ProCoDA software converts the voltage output from the pressure sensors into the physical units of water column height or pressure using linear conversion algorithms. The sensors can also be zeroed or set to a measured value using a one point calibration.

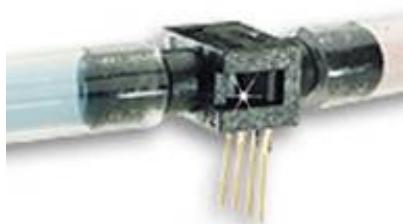


Fig. 11.1: Differential pressure sensors are used to measure water depth, head loss, and air pressure.

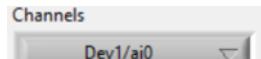
Steps to set up a pressure sensor.

1. Navigate to the Configuration tab

2. Click the  button to select and configure your sensor (thermistor).



3. Click to add a sensor to your list.



4. Now you need to tell ProCoDA where the pressure sensor is plugged in. In the pull-down menu, select the address of the pressure sensor.
5. Finally, you need to tell the software to convert the signal into a pressure. This is done with a calibration file. Click and select the folder named with the pressure range of the sensor you are using. Then select the pressure units you would like to use.
6. You should now be reading pressure. Verify that the pressure sensor is working by gently pushing on one of the pressure ports. The goal is shove some of your skin into the port to increase the pressure! One port should respond with a positive pressure and the other port should respond with a negative pressure.
7. Use what you learned about positive and negative ports to make sure that you connect the pressure sensor to your experimental apparatus correctly.

11.5.2 Temperature Measurement

We use a linear temperature sensor coupled with a simple voltage dividing circuit such as the Omega sensor model OL703. The voltage output is converted to a temperature using a linear equation.

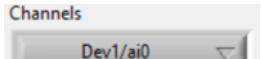
1. Navigate to the Configuration tab



2. Click the button to select and configure your sensor (thermistor).



3. Click to add a sensor to your list.



4. Now you need to tell the software where your sensor is plugged in. In the pull-down menu, select the address of your sensor. All addresses begin with a Dev/ai prefix. The number in the address refers to the number on the
5. Finally, you need to tell the software to convert the signal into temperature units. This is done with a calibration file. Click and select the calibration file named thermistor.smc.
6. You should now be reading temperature in units of degrees Celsius. Verify that you are monitoring the correct temperature probe by holding the temperature probe in your hand and warming it up. Does the temperature reading respond?



11.5.3 pH Measurement

pH sensors produce a voltage output in the range that would normally be easy to measure using standard data acquisition hardware. Unfortunately, the impedance requirement for a pH sensor is orders of magnitude higher than the inputs of standard data acquisition hardware and thus a signal conditioning circuit must be used to amplify the pH sensor output. The circuit consists of unity gain amplifiers that have less than 0.1 pA input leakage current.

pH measurements require calibration in known buffers.

1. Open the ProCoDA II software.

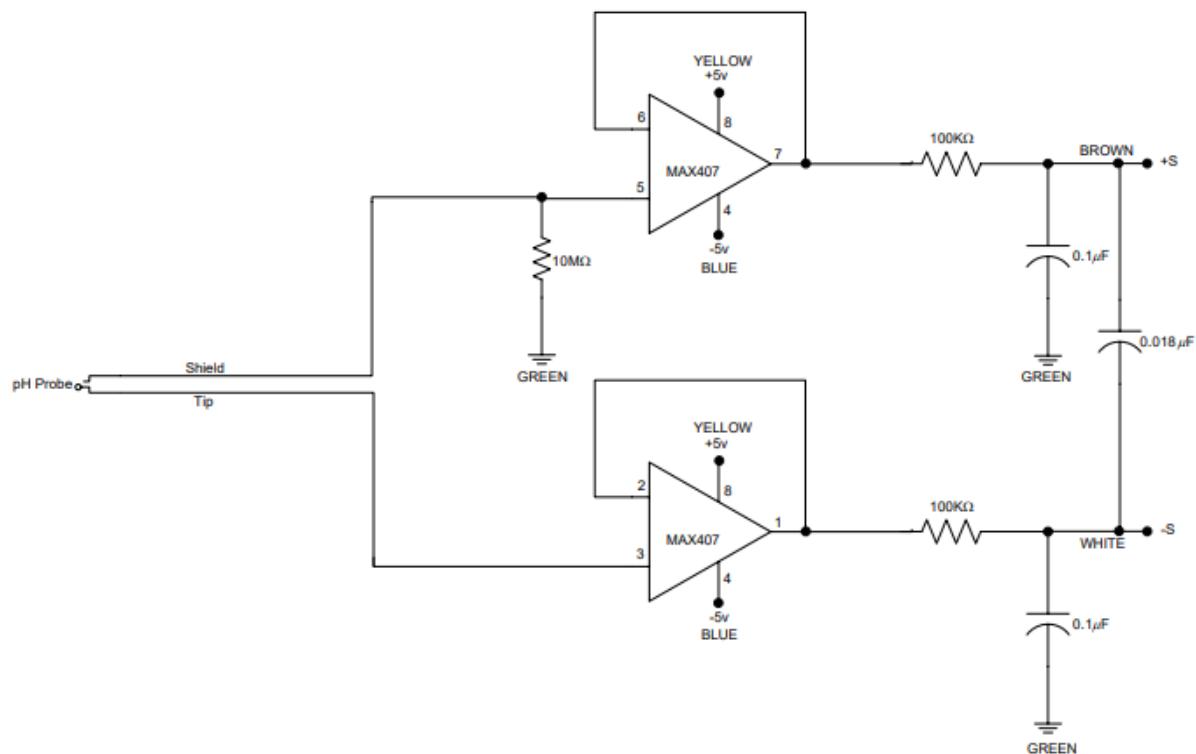


Fig. 11.2: Circuit diagram for the signal conditioning circuit that takes the output from a pH (or other ion selective) electrode and amplifies the signal so that it can be measured by the data acquisition system. This is required because pH probes have a very high impedance that is too high for standard data acquisition systems to measure.



2. Navigate to the Configuration tab and select the button.



3. Insert a new sensor at the bottom of the sensor list using the button.

4. Select the appropriate channel based on in which sensor port you plugged you pH probe.



5. Select .

6. The pH probe should never be dry and is therefore stored with a small vial of pH 4.0 buffer screwed onto the tip. Unscrew the storage vial cap and place the vial in a place where it will not be tipped over (the cap can stay on the probe).

7. Rinse the pH probe with DI water (use a squeeze bottle) into a beaker.

8. To calibrate the pH probe, we will use three pH buffer solutions with known pH (red=4.0, yellow=7.0, and blue=10.0). After rinsing the pH probe, place it into the pH=4.0 buffer. Stir gently and wait for the pH reading on the software to stabilize. Once stabilized, press the button. Rinse the pH probe with DI water and repeat for the pH=7.0 and pH=10.0 buffer solutions.



9. When you have tested all calibration buffers, click, OK to exit . Click OK again to exit .

The option is used if you have additional buffers that you want to use to calibrate a pH probe. The list of buffers can also be cleared, , and recreated by adding new buffers.

11.5.4 Gran Plot

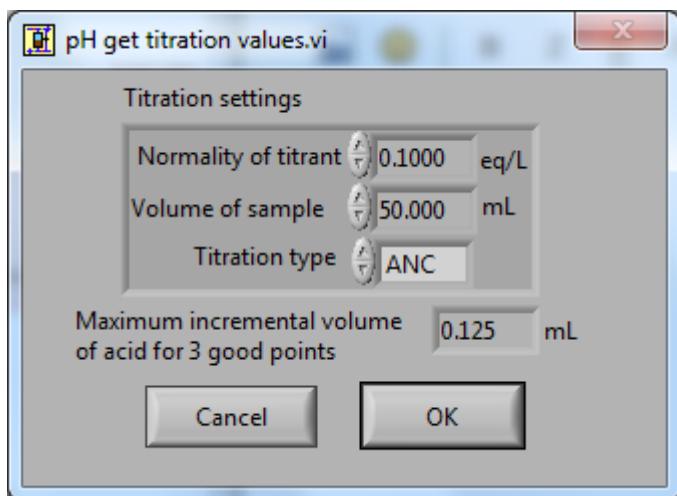
The Gran plot is used to measure the acid neutralizing capacity or the alkalinity of a water sample.

1. Open the ProCoDA II software.



2. navigate to configuration, select , select , and click on .

3. You will be prompted for the normality of titrant and the volume of sample. You can also choose to measure ANC (acid neutralizing capacity) or BNC (base neutralizing capacity). If you are measuring BNC you will need to titrate with a strong base. After entering the normality of acid (or base) and the sample volume the computer will suggest an incremental volume of titrant that will produce a good Gran plot. Smaller incremental titrant volumes can be used, but will require more time to titrate the sample. After entering the values, exit the dialog box by clicking on the OK button. It will look like this:



incremental
titrant added

4. The Gran Plot analysis uses 3 controls: , **Accept pH Value (F11)**, and **End Titration (F12)**.
- incremental
titrant added

The incremental titrant added is the amount of acid added since the previous time the **Accept pH Value (F11)** button was clicked. For the first data point if no titrant was added the incremental titrant added should be set to zero. For subsequent readings, change the incremental titrant added to the volume you are adding, add the titrant with a digital pipette, wait for the pH to stabilize and then click on **Accept pH Value (F11)**. Any amount of titrant can be added at each step, but it is important that below pH 5 the titrant volumes be smaller than the recommended value so that sufficient data points are obtained in the linear region.

5. There is no way to delete unwanted data points after they are accepted. Therefore, make sure you only press the enter button once after each addition of titrant.
6. Continue adding titrant until a line is fit through the linear region of the data. When the line is drawn through the linear region press **End Titration (F12)**. Note that **End Titration (F12)** accepts the last data point and ends the titration. **End Titration (F12)** is pressed after the last addition of acid INSTEAD of pressing **Accept pH Value (F11)**.
7. The equivalent volume (V_e) is given in the same units as were used for the titrant and sample volumes. The equivalent volume is the abscissa intercept of the line fit to the data in the region of constant slope. The ANC is given in equivalents per liter.



8. If desired the titration data can be saved in tab delimited format by selecting . You will be prompted for a file name and location.

11.5.5 Dissolved Oxygen



Dissolved oxygen diffuses across a gas-permeable membrane into a solution where all oxygen is immediately converted to water by an electrolysis circuit.



The current required to reduce the oxygen that is diffusing across the membrane is measured by a circuit. Dissolved oxygen probes produce a current in the pA range that is proportional to the oxygen concentration in the bulk solution. The signal conditioning circuit is designed to convert this very small current into a measurable voltage and to isolate the probe from the effects of fluctuations in the voltage level of the solution containing the probe. This isolation is critical if the solution is monitored with additional probes or if the solution is electrically connected to a building plumbing system or to any other voltage source.

When using the DO probe make sure that there *aren't any air bubbles* on the probe membrane. If you are aerating the sample place the probe as far from the air bubbles as possible. Air bubbles on the membrane will cause inaccurate readings.

1. Connect a DO probe to one of the sensor ports on the ProCoDA box using the gold signal conditioning box.



2. Navigate to the ProCoDA Configuration tab and select to configure the dissolved oxygen channel(s).
3. **Select the DO probe from the sensor list** (This is important! Otherwise you will turn a different sensor into a DO probe!) and point the channel to the correct sensor port.



4. Click on to calibrate the DO probe.
5. Enter the temperature of the sample. This can be measured by using a thermistor or a thermometer. A good estimate is 22°C. If you have a thermistor connected to ProCoDA you can configure the oxygen probe to incorporate continuous temperature readings into the calculation of the dissolved oxygen concentration.
6. Create a zero oxygen solution (50 mL is sufficient) by adding cobalt chloride as a catalyst and enough sodium sulfite to react with all of the dissolved oxygen.
7. Wait for the oxygen level to drop to zero and for the probe readings to stabilize. Then click on .
8. The current atmospheric pressure is required so that ProCoDA can calculate the equilibrium concentration in saturated water. The local air pressure can be obtained from the [National Weather Service](#) Be careful with the [Barometric pressure](#)
 101.3 kPa. Atmospheric pressure is always close to 100 kPa.
9. Place the probe in oxygen saturated water (use the air jet on your bench to bubble air into water in a small container). The voltage from the DO probe should be between 0.17 V and 0.23 V if the probe is working correctly. If the voltage is lower than 0.17 V it may be time to replace the membrane or the solution may not be saturated with oxygen.
10. Select to calibrate the DO sensor.
11. Select OK when you are satisfied with the calibration.
12. If desired you may save the calibration for later use . However, it is not necessary to save the calibration to use the calibration because it is automatically saved as part of the ProCoDA method file..

11.5.6 Photometer



The photometer is a flow cell with an optical path length of 19 mm. The flow cell has 1/8 inch NPT threads for connections to experimental or sample streams from processes. The photometer uses an [LED](#) with an emission peak at 465 nm as its light source. The spectral bandwidth defined by 50% of the dominant wavelength is 25 nm.

```
""" importing """
from aide_design.play import*
b_cell = (3/4 * u.inch).to(u.mm)
print(b_cell)
```



Fig. 11.3: The photometer flow cell is a sealed chamber with round glass plates on both sides. There is a 465 nm LED (blue light) on the right of this image. A photodetector on the left produces a voltage that varies linearly with the intensity of the light that passes through the sample cell.



Fig. 11.4: The photometer must always be held in a vertical orientation to ensure that air bubbles are carried out of the sample cell. The sensor output is conditioned for monitoring by ProCoDA in the black box.

To calibrate the photometer, you will need to connect the peristaltic pump, a 1 L bottle, and the photometer in a closed loop. Use enough tubing so that the flow path of the photometer is oriented in the vertical direction with flow **up** through the photometer (this ensures that air bubbles are carried up and out of the photometer). Add 1 L (or a known volume) of tap water to the bottle and turn the pump on at 380 mL/min. The high flow rate is to speed up the response time when the concentration is changed. The goal is to have a known volume of solution circulating through the calibration system.

Calibration steps

1. Connect the photometer probe to one of the sensor ports of your ProCoDA box.
2. The photometer will read out voltages in the range of +/- 5V, so we will need to let the software know to expect that range. On the Configuration Tab in the ProCoDA II software, find the section for the NI Input/Output device  and hit the  button. Set the maximum voltage for channel with the photometer to 5V.



Fig. 11.5: Experimental setup for calibrating photometer. Flow must be **up** through the photometer to ensure that any air bubbles are removed. It may be necessary to lightly tap the photometer to help release any trapped air bubbles.

3. Navigate to the ProCoDA configuration tab and then select
4. Select the sensor in the sensor list that you want to configure as a photometer.
5. Make sure that the sensor channel is set correctly.

6. Select . You will see a voltage reading in the top right corner. When the LED light in the photometer is off (toggle switch in the middle) the voltage should read approximately -1.3 V. When the LED is light in the photometer is on (toggle switch to the left or right) the voltage should read approximately +3.5 V. Verify that this range is being measured and is stable in the off and on configuration. If the voltage is -1 V or +1 V, check to make sure that the voltage range for the sensor was set correctly (set step 2). If the voltage is zero, check the ProCoDA power supply. If the voltage with the LED on is less than +3.5 V, then pump water up through the photometer and tap it gently to release the air bubble that is reflecting light in the sample cell.

7. Turn the LED off and when the voltage is stable and approximately -1.3 V click on

8. Turn the LED on with the blank solution in the photometer and click on . Remember the voltage should be approximately +3.5 V.

An example continuous flow calibration routine is given below. We suggest preparing a 40 g/L stock solution of Red Dye #40 to make a calibration curve for the photometer. Calculate the volume of red dye that will be needed to generate a calibration with points at 0, 1, 2, 5, 10, 20, 30, 40, and 50 mg/L. Remember that you will be adding the dye cumulatively and thus you need to calculate the incremental volumes. The first calibration point is 0 mg/L. This is the same as the blank.

Sipper cell flow Calibration

1. Insert the sipper tube into the standard (x mg/L) and use the syringe to pull the standard through the photometer.
2. Make sure that any air bubbles have been dislodged from the photometer and the voltage reading is stable.
3. Click on read standard next to the x mg/L row.

4. Use the syringe to push the standard back into the bottle
5. Rinse the photometer with the same water that is used for the blank if sample carry-over is a concern
6. Repeat for the other standards. The R squared value should be greater than 0.99.



7. When you are done, click on the save icon in to save the calibration as a file. This calibration should be good for as long as the LED lasts, which should be a very long time!
8. Use the export icon to create a tab delimited file containing all of the calibration data.

Continuous flow Calibration

1. Click on read standard next to the 0 mg/L row before adding any red dye.
2. Add red dye to make the concentration in the calibration system be 1 mg/L. If necessary, hit Add Standard and enter the concentration of the standard you are reading. This approach allows you to see how well the data is fitting to a straight line as you add the standards.
3. Continue to add dye, add standard, equilibrate, read standard until you have a full calibration and all of the standards have been read. The R squared value should be greater than 0.99.
4. When you are done, click on the save icon in to save the calibration as a file. This calibration should be good for as long as the LED lasts, which should be a very long time!
5. Use the export icon to create a tab delimited file containing all of the calibration data.

The photometer calibrator calculates the absorbance using the equation.

$$A = -\log \frac{V_{Sample} - V_{Dark}}{V_{Blank} - V_{Dark}} \quad (11.2)$$

This equation can be used to convert raw voltage data into absorbance readings. The absorbance is converted into a concentration by using Beers law.

$$A = \varepsilon b C \quad (11.3)$$

where

ε is the extinction coefficient for that particular wavelength and that particular dissolved species

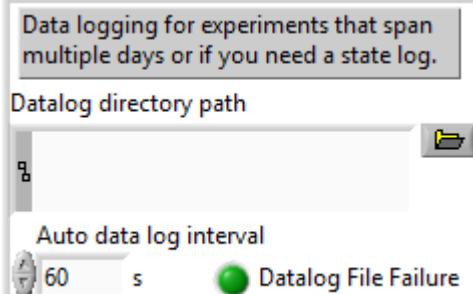
b is the optical path length

C is the concentration of the dissolved species

Given that A is dimensionless the extinction coefficient takes on whatever units are required.

11.6 Logging Data

ProCoDA offers two distinct methods of logging data. The first method is accessed by selecting a folder (not a file!)

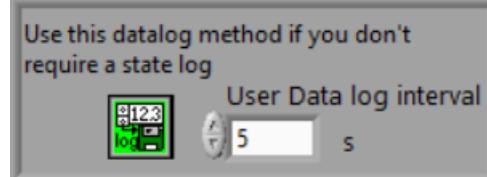


where you would like to save data.

In that system ProCoDA automatically creates 3 different types of files (data log, state log, and method file) and saves them. This approach is ideal for

long term experiments that span multiple days. Every day at midnight ProCoDA starts a new data log and state log file. The daily saving and closing of the files reduces the risk of data loss due to a power failure or file corruption. The Datalog File Failure indicator is the one (and only) red indicator light that can be safely ignored while use ProCoDA! The state logs are particularly useful when ProCoDA is used to cycle through a series of experiments or through a series of states and thus the data may only be of interest in one of those states.

The second method of saving data only creates a data log file. This can be most convenient for short duration experiments where the researcher is present during the experiment. Data is being logged when the data log icon is green.



The data interval can be set for both data logging methods. The data from the data buffer is averaged according to the user selected data log interval. It is important to recognize that the logged data is **not** the same as the data that is used by ProCoDA to make decisions. The averaging interval used to make decisions and the averaging interval used to log data are both user selected values and are independent.

11.7 Logic, States, and Outputs

Edit rules

ProCoDAs ability to quickly set up a state machine is all contained inside the rule editor.

11.7.1 Rule Editor

The programming environment for creating rules that determine exit conditions for states and which state to go to readily facilitates setting up the algorithms for controlling simple repetitive processes such as a sequencing batch reactors or rapid sand filters. For experimental purposes it is desirable to have the capability to systematically vary a parameter to test the performance of the process over a range of input values. This is accomplished via an external code that compares the number of specified replicates to a parameter that increments when the ProCoDA enters a specified state. The output parameter can be used to control pump speeds, times, or can be an input to subsequent calculations.

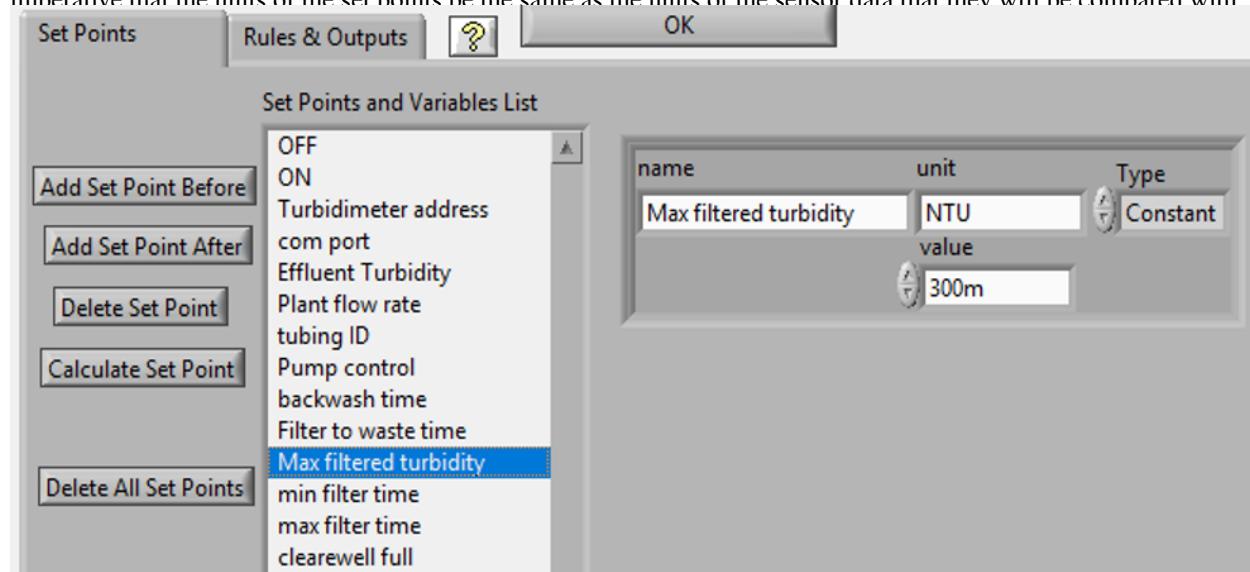
A word of caution. The sensors, set points, and states are used within the rule editor. If new sensors, set points or states are added in the middle of their respective lists or deleted from their lists any rules that were created previously may be incorrect. The software does not attempt to correct for changes in the lists of set points and states. It is your

responsibility to verify that all rules are correct when making changes to the configuration. You can minimize this problem by adding states, set points, and sensors at the end of their respective lists.

11.7.2 Set Points

The rule editor provides a graphical user interface where the operator can completely configure the control logic for the plant. Begin by creating the set points for the process. Set points can include time, parameters that can be compared with sensor values, parameters that are required inputs for external code, and parameters that are outputs of external code. Set points that are outputs of external code are designated as variables. The only constraint on developing the list of set points is that set points that are inputs to external code must be in the same order (although they don't have to be adjacent) in the list of set points as they are expected by the external code.

When adding a new set point the name, value and unit of the set point can be edited in the Set Points control. The list of the Current set points functions as the index to the array of Set Points, thus allowing the operator to select and edit any of the set points. The unit field is not used by the process controller, but is a reminder for the operator. It is imperative that the units of the set points be the same as the units of the sensor data that they will be compared with.



When configuring a set point as a variable calculated by external code first load the code by clicking on the folder icon. If the code fails to load it is either because the external code doesn't have the correct connector pane or because the external code has sub VIs that aren't in the same folder as the external code. If the code loads correctly it will display the list of needed inputs at the bottom of the dialog box.

11.7.3 External code

LabVIEW executables can be enabled to connect to external code. This capability makes it possible to easily extend the capabilities of the ProCoDA software. The external code must be designed to meet specific requirements for the data types of inputs and outputs. An external code interface has been created to take a variable number of numeric inputs and produce a single numeric output. The external code can be used for a wide variety of functions including simple math functions, a specialized function (such as one which sets a coagulant dose based on raw water turbidity) proportional-integral-derivative control that can be used to force a controlled parameter to a desired set point, data acquisition functions that acquire digital data from instruments, and control functions that set the speed of peristaltic pumps that are connected to a USB port.

11.7.4 Meters

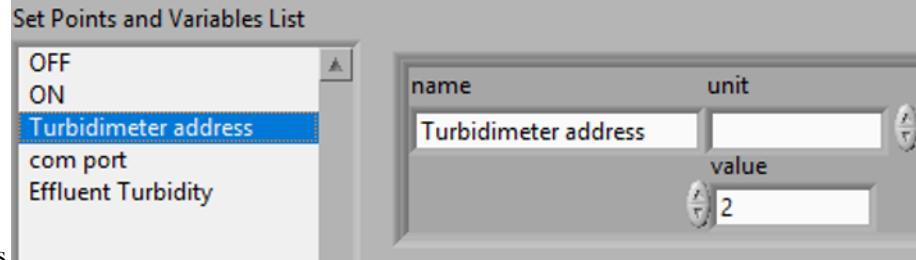
Turbidimeters, electronic balances, etc. can communicate with ProCoDA through a USB or serial port. These devices are treated like functions and their data is accessed with an external function call in the set point list (Accessed through



).

11.7.5 Connect an HF scientific MicroTol turbidimeter

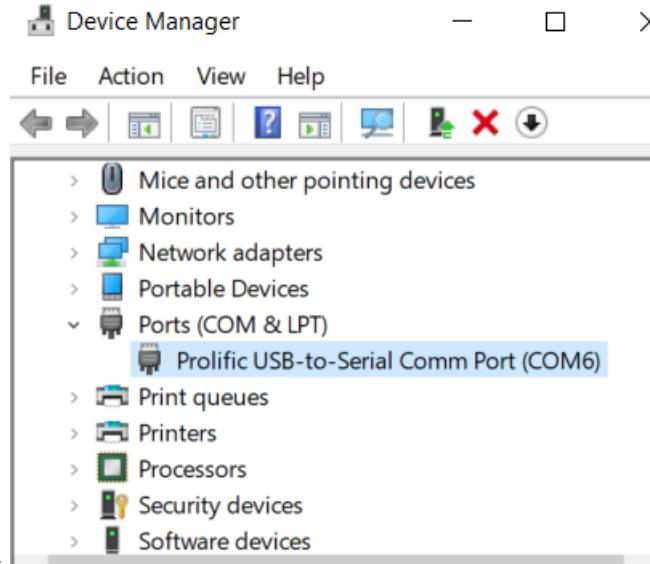
1. Edit Rules



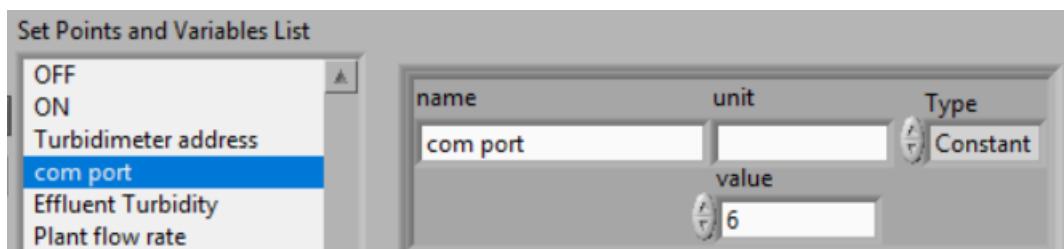
2. Add a set point for the Turbidimeter address

3. Check the unit ID on the turbidimeter (press twice to select the config option. Then press twice to select ADDR. You can adjust the unit ID using the up or down arrows. Press once more to exit and return to the turbidity view screen.)

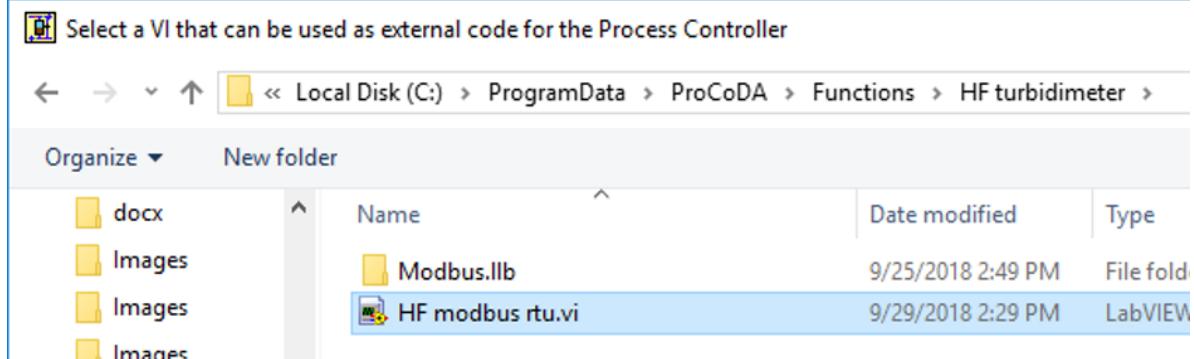
4. Use the windows Device manager to check which com ports are being used on your computer. In this example

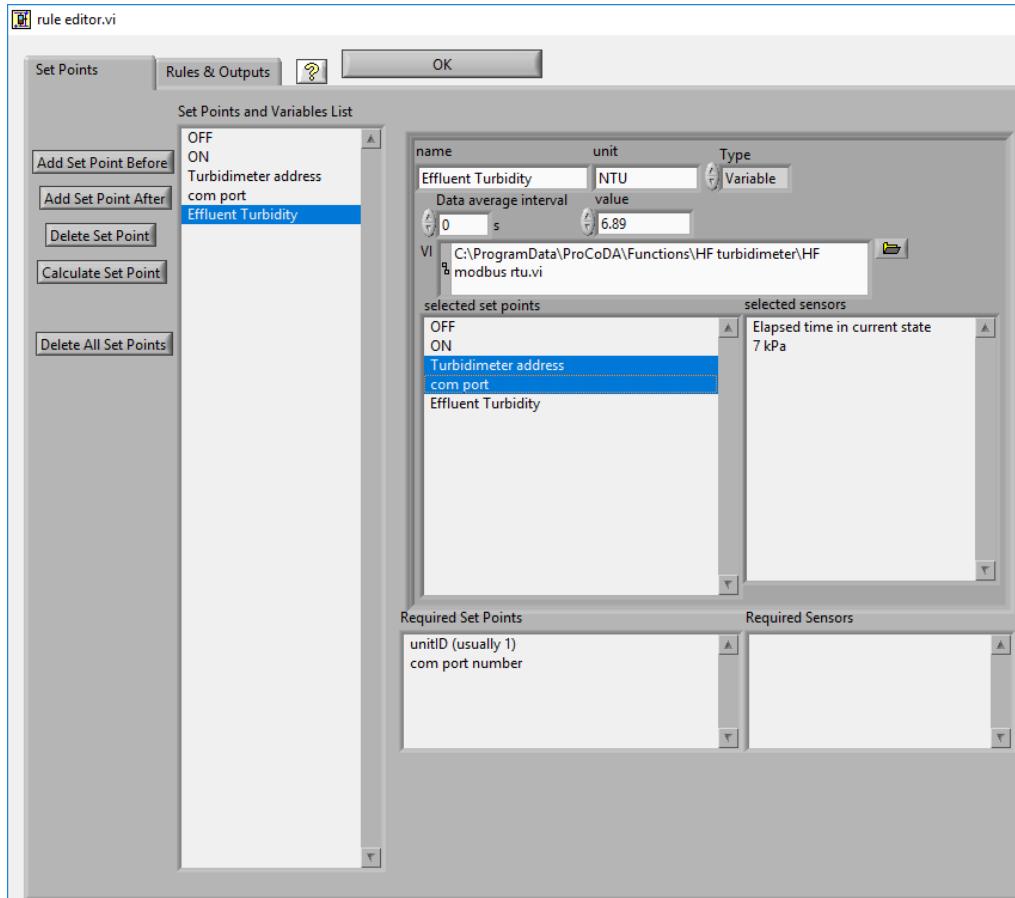


5. Add a second setpoint for the com port on the computer that the turbidity meter is connected to.



6. Add a third setpoint that will be the measured turbidity.
7. Change the third setpoint from a constant to a variable
8. Now more options will appear click on the folder which will open a window
9. Click on HF modbus rtu (the communication protocol used by HF and many other devices)





10. Select the required set points.

If successful the turbidity displayed on the meter should show up as the value. If there is a communication error you will get a -999.

11.7.6 Increment functions

A common experimental task is to systematically vary a parameter (for example, coagulant dose) over a wide range to measure the response (for example, settled water turbidity) of a system. ProCoDA external functions provide linear and power law options for incrementing over a range of values.

$$y_{\text{linear}} = \text{slope} \cdot x + \text{intercept} \quad (11.4)$$

$$y_{\text{power}} = \text{coefficient} \cdot \text{base}^x \quad (11.5)$$

The two increment functions are dependent on the state cycles. In the example shown in Fig. 11.6 the state was set to cycle between states 1, 2, and 3 with the exception of a manual reset to state 0 and then to state 1 to illustrate how the increment function is reset. The power law function is useful when it is desirable to explore a larger parameter space. However, care must be taken to ensure that the controlled processes have the ability to deliver the desired range of the varied parameter.

ProCoDA can be configured to stop an experiment after the reaching the maximum value of the parameter by using the external code called count states. Count states counts the number of times a state to count has been entered. This can be used to set an exit condition on a state. If no exit condition is created to end an experiment the increment function will reset the parameter to its initial value and begin the increment process again.

It is possible to systematically vary more than one parameter. To do this it is essential that different copies of the increment function code be used for each parameter. This is because increment function code includes a memory to

track its own state and if it is used by ProCoDA to do multiple things it will give unexpected results!

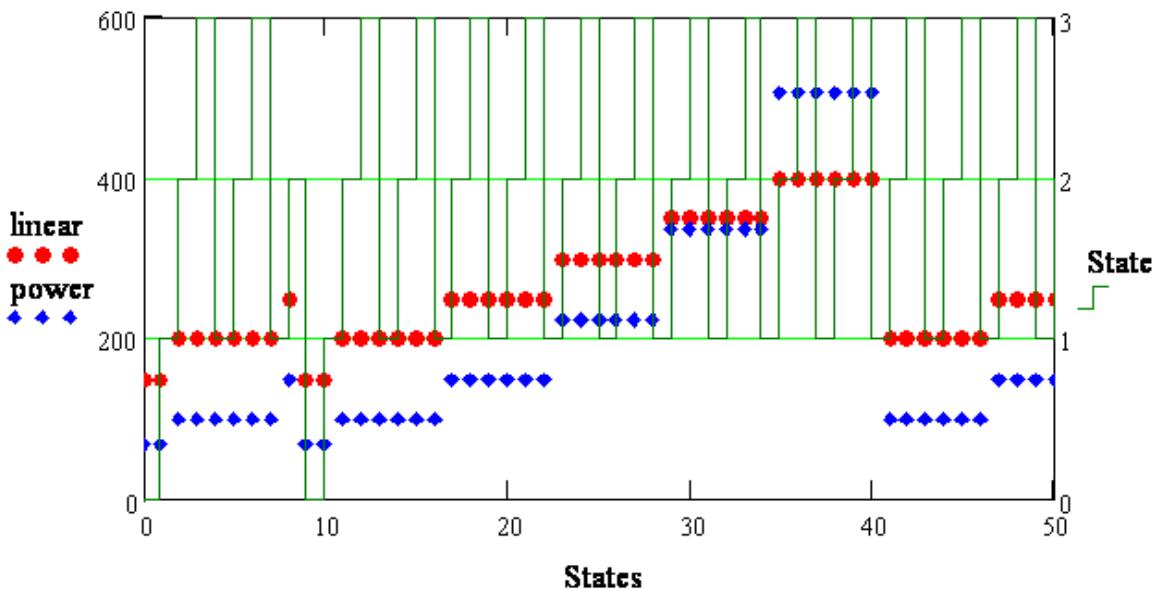


Fig. 11.6: Increment functions showing how the output varies as a function of the state. In this example the state cycled between states 1, 2, and 3. The increment state was 2, the number of replicates was 2, the reset state was 0, the y intercept was 200, the slope was 50, and the maximum value of x was 4. The power law relationship used a coefficient of 100 and a base of 1.5.

11.7.7 States

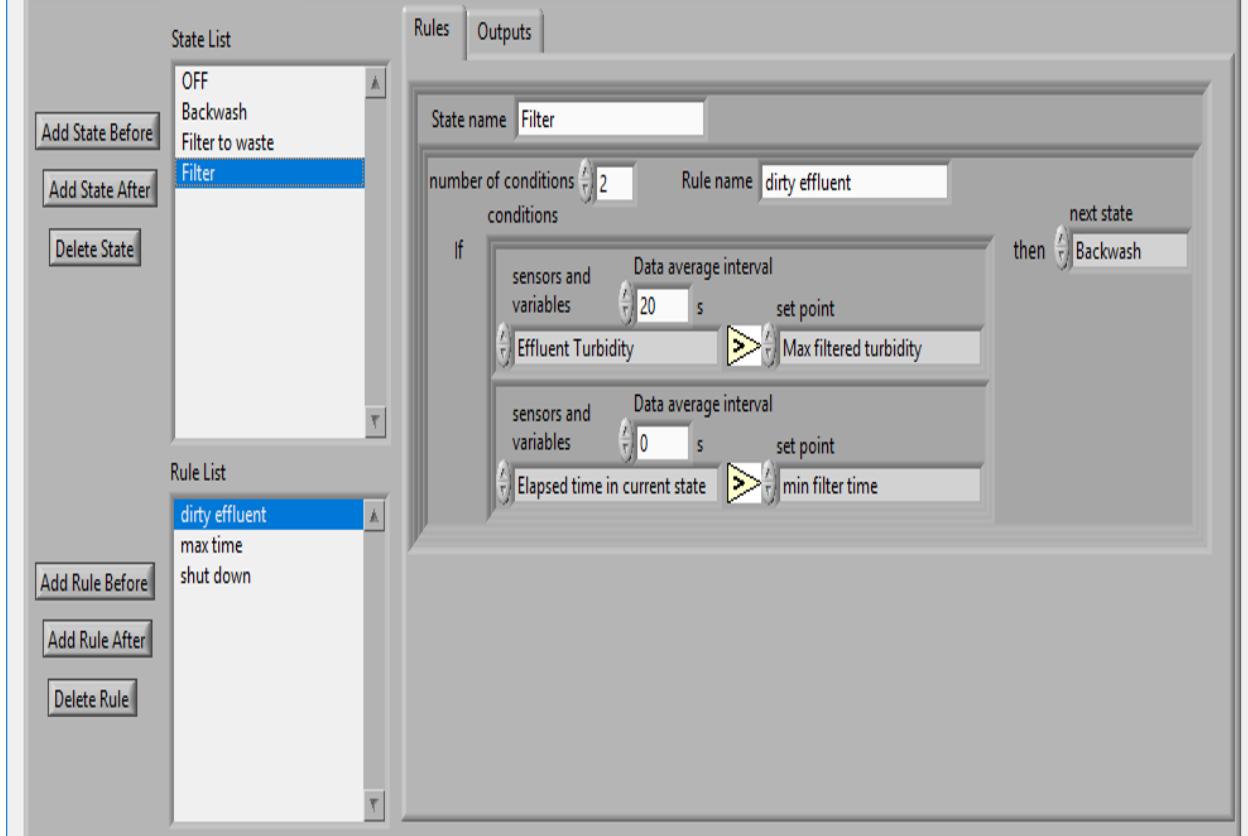
The list of states is created by adding new states and then naming the states in the Rules control. The states don't necessarily have to be listed in the order of the cycle. The first state should be the default off state. This state is used by the software as the default when the process is first turned on. The off state is also used as an emergency shutdown in case of a data acquisition failure.

The control output settings for each state indicate which set point is used to control that state. All set points used for control should be in the range of 0 to 1. Fractional values for the outputs 0 to 5 will cause the output to cycle and thus control the duty cycle. Fractional values for the pump speed will control the pump speed.

11.7.8 Rules

**Edit
rules**

The list of rules is created by adding new rules and then naming each rule in the Rules control. The rules are the logical comparisons that determine if the process should change to a new state. The logic for changing process state can include a requirement of meeting several conditions simultaneously (conditions that are anded). There may also be more than one rule that can cause the process to change state. These two options are incorporated into the rule editor. The anded rules have multiple conditions and multiple rules can be created that are essentially or'd.



Rules that have multiple conditions that must be fulfilled simultaneously are created by increasing the number of conditions. Each conditional test will have its own logical comparison. An example of a completed rule containing two conditions is if the effluent turbidity exceeds regulations AND the time in the filtration state is greater than a minimum amount of time, then switch to backwash. Each logical comparison consists of a measured value (either a sensor value or the elapsed time in the current state) that can be selected from the drop down menu. The measured value is compared with a set point that can also be selected from a drop down menu. The type of comparison can either be less than or greater than.

Each rule can select which state the process should go to next. Thus it is possible that within the same state different rules would cause the process to change to different states. In the example here the shut down rule will end filtration if the clearwell is full.

The rules are implemented in the order indicated by the Current Rule control. If two rules would cause a state change at the same time, the first rule is used.

11.7.9 Outputs

The ProCoDA hardware is designed and fabricated around an NI USB data acquisition board is used for on/off control of up to six devices and for variable control of up to two peristaltic pumps. The on/off devices are controlled with 24 V outputs that can be used to control solenoid valves, pinch valves, relays, or other low current devices.

Connecting a pump that is controlled through pump 0 or pump 1 ports.

There are many ways to connect a pump, I am going to stick to one pump head because it is easier and adding more is fairly str

1. Add a constant set point with the flow in mL/s

2. Add a constant set point with the tubing size that you will be using

3. Add a variable set point and call it something like pump speed control

4. Click on the folder and open the peristaltic folder and choose the right code you want to use for this tutorial it is single head pump control (mL per second, Tubing Size)

5. Select the set points

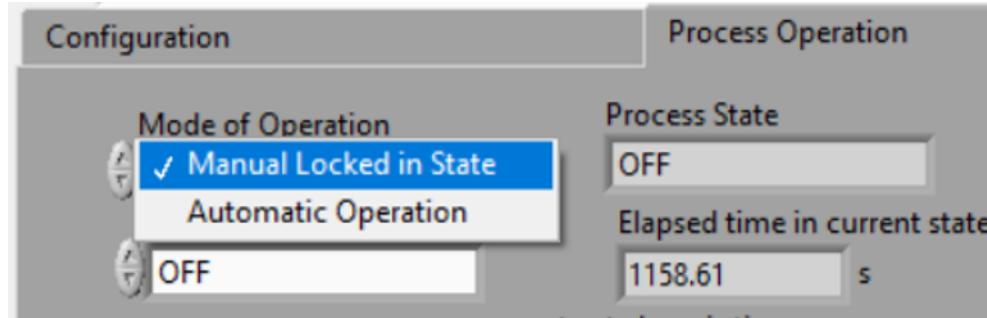
6. It should display the speed of the pump as a fraction of

7. Navigate to the Rules & Outputs tab and select Outputs
 8. Select (or create) a state where you want the pump to be running
 9. Select On for the pump off/on value
 10. Select either Off for clockwise rotation or On for counterclockwise rotation
 11. Select the variable that you created (pump speed control) for the pump speed.
 12. To test the pump go to process operation and change the state to the state that has the pump running

13. If your pump is not running at the speed ProCoDA displays you can calibrate it using the  button

11.8 Process Operation

ProCoDA can operate in either a manual or an automated mode. In Manual Locked in State mode the user sets the state and ProCoDA stays in that state forever. In Automatic Operation the user sets the starting state and then ProCoDA uses the rule logic to move from state to state.



11.8.1 How to set up a timed test

1. Edit Rules

2. Create a constant set point with the value as the number of SECONDS you want to test to go on for
3. Go to Rules & Outputs
4. Click on the state that runs the test then click on rules
5. Create a rule: If elapsed time in current state > run time of test then go to next state (the next state could be OFF if you want a simple timed test)
6. You can also do many other conditional statements with this function like have states change based on any sensor data or other variable.

11.9 Troubleshooting

This is complex software with lots of capabilities and thus there are many opportunities for failure. The goal is to recognize failure quickly and then fix it!

1. States change unexpectedly. Use data averaging on sensor and meter data to prevent noisy data signals from causing state changes.`lData_Average`
2. Sensor out of range (change the voltage range on the *data acquisition system*)
3. Red LED indicators indicate failures. Make sure there are no red LEDs on the configuration tab. The only allowable red LED is the Datalog File Failure if you don't want to log data using that method.



4. A red LED next to the on the configure tab indicates that the computer is not connected to a ProCoDA box. Check the USB connections. If this is the first time using ProCoDA on this computer, then make sure you have followed all of the *installation steps* because data acquisition won't work without the data acquisition drivers.
5. Sensors give zero or close to zero response. Check the 24 volt power supply for the ProCoDA box. Check to ensure that the power supply LED lights on the back end of the ProCoDA box are lit. ProCoDA boxes built in 2014 didn't have automatic overload protection and thus it is possible for an internal fuse to blow.
6. Software is slow and data acquisition is slow. External code that uses serial or USB port communication (turbidimeters, balances, etc.) may be incorrectly configured. If serial communication fails to these devices ProCoDA software waits until a serial port timeout occurs and during that wait everything slows down. Always switch external code that accesses meters back to constant from variable when the meter is no longer accessible by ProCoDA.

11.10 References

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PDF and LaTeX versions¹.

Notes

¹ PDF and LaTeX versions may contain visual oddities because it is generated automatically. The website is the recommended way to read this textbook. Please visit our [GitHub](#) site to submit an issue, contribute, or comment.