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quasispectral

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Using quasispectral

To download the library, first install the "devtools" package, which will allow you to install packages from github; this only needs to be done once:

```
install.packages("devtools")
```

Next, install the edgeR package. To to this, you will need to first install the "biocLite" package, which allows you to access code from bioconductor:

```
source("https://bioconductor.org/biocLite.R")
```

Now you can get edgeR as follows:

```
biocLite("edgeR")
```

When asked, allow R to compile and install the package. This will take a few minutes.

Next, install the fdrtools package, which computes qvalues:

`install.packages("fdrtools")'

Then install the quasispectral package from github:

```
devtools::install_github("mooredf22/quasispectral")
```

This will make the functions and data in the package available to you in R.

Next, load the libraries; the libraries QuasiSeq and fdrtool must have been previously installed from CRAN.

```
library(QuasiSeq)
library(fdrtool)
library(quasispectral)
```

The first few lines of the data may be examined:

head(spectralData_CLN1_late)

```
#> geneName CLN1.1 CLN1.2 CLN1.3 CLN1.4 CLN1.5 CLN1.6 wt.1 wt.2 wt.3 wt.4
#> 1 Aga 149 142 67 135 150 142 135 155 159 202
#> 2 Arsa 577 542 314 601 569 590 602 639 617 599
#> 3 Arsb 466 482 258 409 465 465 624 599 645 688
#> 4 Arsg 68 65 41 55 69 63 87 84 46 98
#> 5 Arsk 59 60 22 39 45 54 75 75 80 107
#> 6 Asah1 806 905 533 729 748 827 865 812 887 861
```

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```
#> wt.5 wt.6
#> 1 195 181
#> 2 613 631
#> 3 662 667
#> 4 107 99
#> 5 77 79
#> 6 918 904
```

The first column is the gene name, the next 6 columns are spectral counts for mutant animals, and the final 6 columns are counts for wildtype animals.

Finally, call the package as follows, and look at the first few lines of output:

```
result.QLfit <- quasiSpectral(spectralDataAll=spectralData_CLN1_late, n.mut=6, n.wt=6)
#> Currently, bias reduction has not yet been implemented for Poisson model.
#> Spline scaling factor: 0.983018553187791
#> Warning in fdrtool(p.values, statistic = "pvalue"): There may be too few
#> input test statistics for reliable FDR calculations!
#> Step 1... determine cutoff point
#> Step 2... estimate parameters of null distribution and eta0
#> Warning: Censored sample for null model estimation has only size 8 !
#> Step 3... compute p-values and estimate empirical PDF/CDF
#> Step 4... compute q-values and local fdr
#> Step 5... prepare for plotting
QLfit.out <- result.QLfit$QLfit
head(QLfit.out)
    geneName CLN1.1 CLN1.2 CLN1.3 CLN1.4 CLN1.5 CLN1.6 wt.1 wt.2 wt.3 wt.4
                                   135 150 142 135 155 159 202
#> 1
               149
        Aga
                      142
                             67
#> 2
       Arsa
               577 542 314 601 569 590 602 639 617 599
#> 3
       Arsb
               466
                      482 258
                                   409 465
                                                465 624 599 645
                                                                   688
#> 4
       Arsg
                68
                       65
                             41
                                   55
                                        69
                                                63
                                                      87
                                                           84
                                                                46
                                                                    98
#> 5
       Arsk
                59
                       60
                             22
                                   39
                                          45
                                                 54
                                                      75
                                                           75
                                                                80 107
       Asah1
                      905
                             533
                                   729
                                          748
                                                827 865 812 887 861
#> 6
                806
    wt.5 wt.6 coef.main p.values p.values.bonf p.values.holm
#> 1 195 181 0.3129084 1.254920e-03 1.016486e-01 4.141237e-02
#> 2 613 631 0.1382393 5.303151e-03 4.295553e-01 1.198925e-01
#> 3 662 667 0.5354857 4.922626e-11 3.987327e-09 3.790422e-09
#> 4 107 99 0.4545213 3.430625e-03 2.778806e-01 8.919292e-02
     77
          79 0.7465593 6.204245e-06 5.025439e-04 2.978038e-04
#> 6 918 904 0.1314973 1.497866e-02 1.000000e+00 2.845946e-01
     q.values.BH q.values.strimmer
#> 1 2.074460e-03
                     2.914893e-04
#> 2 7.159254e-03
                    1.029129e-03
#> 3 7.974655e-10
                     1.120724e-10
#> 4 4.875099e-03 7.007858e-04
```

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```
#> 5 1.478070e-05 2.104575e-06
#> 6 1.925828e-02 2.698284e-03
```

The first columns give the gene names and spectral count data that were input into the quasispectral function. The column "coef.main" is the log2 ratio of the estimated mean for wild type animals compared to the estimated mean for mutant animals.

To see the estimated proportion of null samples in the population, from fdrtool, look at eta0:

```
result.QLfit$eta0  # estimated proportion of null samples in population
#> [1] 0.143748
```