

Class 11: Structural Bioinformatics pt 2

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Background

we saw last day that the PDB has 209,886 entries (Oct/Nov 2023). UniProtKB (i.e. protein sequence database) has 199,579,901 entries.

```
209886/199579901 * 100
```

```
[1] 0.1051639
```

so the PDB has only 0.1% coverage of the main sequence database.

Enter AlphaFold data base (AFDB) <<https://alphafold.ebi.ac.uk>> taht attempts to provide computed models for all sequences in UniProt.

accordidng to website, AlphaFold DB provides open access to over 200 million protein structure predictions to accelerate scientific research.

```
##AlphaFold
```

AlphaFold has 3 main outpouts - predicted coordinates (PDB files) -local quality score called pLDDDD (one for each amino-acid) - a second quality score PAE Predicted Aligned Error (for each pair of amino-acid)

We can run AlphaFold ourselves if we are not happy with AFDB (i.e. no coverage or poor model)

Interpreting/ analyzing AF results in R

```
results_dir <- "HIVPR_dimer_23119/"
```

```
pdb_files <- list.files(path=results_dir,  
                         pattern="*.pdb",  
                         full.names = TRUE)  
# Print our PDB file names  
basename(pdb_files)
```

```
[1] "HIVPR_dimer_23119_unrelaxed_rank_001_alphaFold2_multimer_v3_model_4_seed_000.pdb"  
[2] "HIVPR_dimer_23119_unrelaxed_rank_002_alphaFold2_multimer_v3_model_1_seed_000.pdb"  
[3] "HIVPR_dimer_23119_unrelaxed_rank_003_alphaFold2_multimer_v3_model_5_seed_000.pdb"  
[4] "HIVPR_dimer_23119_unrelaxed_rank_004_alphaFold2_multimer_v3_model_2_seed_000.pdb"  
[5] "HIVPR_dimer_23119_unrelaxed_rank_005_alphaFold2_multimer_v3_model_3_seed_000.pdb"
```

```
library(bio3d)  
  
# Read all data from Models  
# and superpose/fit coords  
pdbs <- pdbaln(pdb_files, fit=TRUE, exefile="msa")
```

Reading PDB files:

```
HIVPR_dimer_23119//HIVPR_dimer_23119_unrelaxed_rank_001_alphaFold2_multimer_v3_model_4_seed_000.pdb  
HIVPR_dimer_23119//HIVPR_dimer_23119_unrelaxed_rank_002_alphaFold2_multimer_v3_model_1_seed_000.pdb  
HIVPR_dimer_23119//HIVPR_dimer_23119_unrelaxed_rank_003_alphaFold2_multimer_v3_model_5_seed_000.pdb  
HIVPR_dimer_23119//HIVPR_dimer_23119_unrelaxed_rank_004_alphaFold2_multimer_v3_model_2_seed_000.pdb  
HIVPR_dimer_23119//HIVPR_dimer_23119_unrelaxed_rank_005_alphaFold2_multimer_v3_model_3_seed_000.pdb  
.....
```

Extracting sequences

```
pdb/seq: 1 name: HIVPR_dimer_23119//HIVPR_dimer_23119_unrelaxed_rank_001_alphaFold2_multimer_v3_model_4_seed_000.pdb  
pdb/seq: 2 name: HIVPR_dimer_23119//HIVPR_dimer_23119_unrelaxed_rank_002_alphaFold2_multimer_v3_model_1_seed_000.pdb  
pdb/seq: 3 name: HIVPR_dimer_23119//HIVPR_dimer_23119_unrelaxed_rank_003_alphaFold2_multimer_v3_model_5_seed_000.pdb  
pdb/seq: 4 name: HIVPR_dimer_23119//HIVPR_dimer_23119_unrelaxed_rank_004_alphaFold2_multimer_v3_model_2_seed_000.pdb  
pdb/seq: 5 name: HIVPR_dimer_23119//HIVPR_dimer_23119_unrelaxed_rank_005_alphaFold2_multimer_v3_model_3_seed_000.pdb
```

pdb\$

[Truncated_Name:1] HIVPR_dime	1	.	.	.	50
[Truncated_Name:2] HIVPR_dime	PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKPMIGGI				
[Truncated_Name:3] HIVPR_dime	PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKPMIGGI				
[Truncated_Name:4] HIVPR_dime	PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKPMIGGI				
[Truncated_Name:5] HIVPR_dime	PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKPMIGGI				
	*****	*****	*****	*****	*****
	1	.	.	.	50
[Truncated_Name:1] HIVPR_dime	51	.	.	.	100
[Truncated_Name:2] HIVPR_dime	GGFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFP				
[Truncated_Name:3] HIVPR_dime	GGFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFP				
[Truncated_Name:4] HIVPR_dime	GGFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFP				
[Truncated_Name:5] HIVPR_dime	GGFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFP				
	*****	*****	*****	*****	*****
	51	.	.	.	100
[Truncated_Name:1] HIVPR_dime	101	.	.	.	150
[Truncated_Name:2] HIVPR_dime	QITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKPMIGGI				
[Truncated_Name:3] HIVPR_dime	QITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKPMIGGI				
[Truncated_Name:4] HIVPR_dime	QITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKPMIGGI				
[Truncated_Name:5] HIVPR_dime	QITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKPMIGGI				
	*****	*****	*****	*****	*****
	101	.	.	.	150
[Truncated_Name:1] HIVPR_dime	151	.	.	.	198
[Truncated_Name:2] HIVPR_dime	GFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF				
[Truncated_Name:3] HIVPR_dime	GFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF				
[Truncated_Name:4] HIVPR_dime	GFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF				
[Truncated_Name:5] HIVPR_dime	GFIKVRQYDQILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNF				
	*****	*****	*****	*****	*****
	151	.	.	.	198

Call:

```
pdbaln(files = pdb_files, fit = TRUE, exefile = "msa")
```

Class:

```
pdb, fasta

Alignment dimensions:
  5 sequence rows; 198 position columns (198 non-gap, 0 gap)

+ attr: xyz, resno, b, chain, id, ali, resid, sse, call
```

RMSD is a standard measure of structural distance between coordinate sets. We can use the `rmsd()` function to calculate the RMSD between all pairs models.

```
rd <- rmsd(pdb, fit=T)
```

Warning in `rmsd(pdb, fit = T)`: No indices provided, using the 198 non NA positions

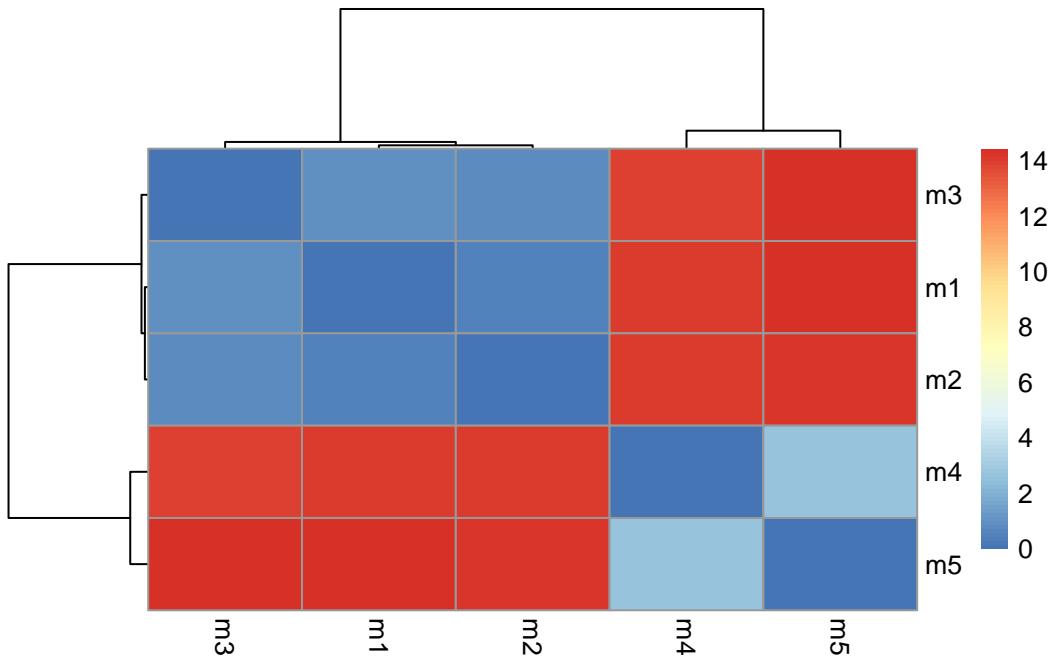
```
range(rd)
```

```
[1] 0.000 14.428
```

Draw a heatmap of these RMSD matrix values

```
library(pheatmap)

colnames(rd) <- paste0("m",1:5)
rownames(rd) <- paste0("m",1:5)
pheatmap(rd)
```



Alignment File

```
aln_file <- list.files(path=results_dir,
                        pattern=".a3m$",
                        full.names = TRUE)
```

```
[1] "HIVPR_dimer_23119//HIVPR_dimer_23119.a3m"
```