

9. T. A. White *et al.*, *PLOS Pathog.* **6**, e1001249 (2010).
10. G. Hu, J. Liu, K. A. Taylor, K. H. Roux, *J. Virol.* **85**, 2741–2750 (2011).
11. P. D. Kwong *et al.*, *Nature* **393**, 648–659 (1998).
12. M. Pancera *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* **107**, 1166–1171 (2010).
13. E. E. Tran *et al.*, *PLOS Pathog.* **8**, e1002797 (2012).
14. Y. Mao *et al.*, *Nat. Struct. Mol. Biol.* **19**, 893–899 (2012).
15. A. Harris *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* **108**, 11440–11445 (2011).
16. C. G. Moscoso *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* **108**, 6091–6096 (2011).
17. S. R. Wu *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* **107**, 18844–18849 (2010).
18. R. Roy, S. Hohng, T. Ha, *Nat. Methods* **5**, 507–516 (2008).
19. Z. Zhou *et al.*, *ACS Chem. Biol.* **2**, 337–346 (2007).
20. C. W. Lin, A. Y. Ting, *J. Am. Chem. Soc.* **128**, 4542–4543 (2006).
21. Materials and Methods are available as supplementary materials on Science Online.
22. Single-letter abbreviations for the amino acid residues are as follows: A, Ala; C, Cys; D, Asp; E, Glu; F, Phe; G, Gly; H, His; I, Ile; K, Lys; L, Leu; M, Met; N, Asn; P, Pro; Q, Gln; R, Arg; S, Ser; T, Thr; V, Val; W, Trp; and Y, Tyr.
23. Q. Zheng *et al.*, *Chem. Soc. Rev.* **43**, 1044–1056 (2014).
24. N. G. Mukherjee, L. A. Lyon, J. M. Le Doux, *Nanotechnology* **20**, 065103 (2009).
25. K. Henzler-Wildman, D. Kern, *Nature* **450**, 964–972 (2007).
26. P. D. Kwong *et al.*, *Nature* **420**, 678–682 (2002).
27. U. Olshesky *et al.*, *J. Virol.* **64**, 5701–5707 (1990).
28. F. Qin, *Biophys. J.* **86**, 1488–1501 (2004).
29. H. Haim *et al.*, *PLOS Pathog.* **5**, e1000360 (2009).
30. N. Sullivan *et al.*, *J. Virol.* **72**, 4694–4703 (1998).
31. J. Arthos *et al.*, *J. Biol. Chem.* **277**, 11456–11464 (2002).
32. A. L. DeVico, *Curr. HIV Res.* **5**, 561–571 (2007).
33. T. Zhou *et al.*, *Science* **329**, 811–817 (2010).
34. L. M. Walker *et al.*, *Science* **326**, 285–289 (2009).
35. L. M. Walker *et al.*, *Nature* **477**, 466–470 (2011).
36. M. Pancera *et al.*, *Nat. Struct. Mol. Biol.* **20**, 804–813 (2013).
37. J.-P. Julien *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* **110**, 4351–4356 (2013).
38. J.-P. Julien *et al.*, *PLOS Pathog.* **9**, e1003342 (2013).
39. P. D. Kwong, J. R. Mascola, G. J. Nabel, *Nat. Rev. Immunol.* **13**, 693–701 (2013).
40. Z. Li *et al.*, *Antimicrob. Agents Chemother.* **57**, 4172–4180 (2013).
41. J. B. Munro, K. Y. Sanbonmatsu, C. M. Spahn, S. C. Blanchard, *Trends Biochem. Sci.* **34**, 390–400 (2009).
42. Subsequent to the submission of this manuscript, the structure of a trimeric prefusion HIV-1 Env (43) was determined in complex with broadly neutralizing antibodies PGT122 (35) and 35022 (44). To determine the conformational state these antibodies captured in the crystal lattice, we measured smFRET on labeled JR-FL virions in the presence of either PGT122 or 35022 or both. These data indicated that PGT122 strongly stabilized the ground state. In contrast, 35022 had little effect on Env conformation. HIV-1 Env complexed with both antibodies exhibited slight ground state stabilization.
43. M. Pancera *et al.*, *Nature* **514**, 455–461 (2014).
44. J. Huang *et al.*, *Nature* 10.1038/nature13601 (2014).
45. J. S. McLellan *et al.*, *Nature* **480**, 336–343 (2011).

ACKNOWLEDGMENTS

We thank J. Jin, L. Agosto, T. Wang, R. B. Altman, and M. R. Wasserman for assistance; C. Walsh, J. Binely, A. Trkola, and M. Krystal for reagents; and members of the Structural Biology Section, Vaccine Research Center, for critically reading the manuscript. We thank I. Wilson and J. Sodroski for encouraging us to extend our approach to a R5-tropic Env. The data presented in this paper are tabulated in the main paper and in the supplementary materials. This work was supported by NIH grants R21 AI100696 to W.M. and S.C.B.; P01 56550 to W.M., S.C.B., and A.B.S.; and R01 GM098859 to S.C.B.; by the Irvington Fellows Program of the Cancer Research Institute to J.B.M.; by a fellowship from the China Scholarship Council–Yale World Scholars to X.M.; by grants from the International AIDS Vaccine Initiative's (IAVI) Neutralizing Antibody Consortium to D.R.B., W.C.K., and P.D.K.; and by funding from the NIH Intramural Research Program (Vaccine Research Center) to P.D.K. IAVI's work is made possible by generous support from many donors including: the Bill & Melinda Gates Foundation and the U.S. Agency for International Development (USAID). This study is made possible by the generous support of the American people through USAID. The contents are the responsibility of the authors and do not necessarily reflect the views of USAID or the U.S. government. Reagents from the NIH are subject to nonrestrictive material transfer

agreements. Patent applications pertaining to this work are the U.S. and World Application US2009/006049 and WO/2010/053583, Synthesis of JRC-II-191 (A.B.S., J.R.C.), U.S. Patent Application 13/202,351, Methods and Compositions for Altering Photophysical Properties of Fluorophores via Proximal Quenching (S.C.B., Z.Z.); U.S. Patent Application 14/373,402 Dye Compositions, Methods of Preparation, Conjugates Thereof, and Methods of Use (S.C.B., Z.Z.); and International and US Patent Application PCT/US13/42249 Reagents and Methods for Identifying Anti-HIV Compounds (S.C.B., J.B.M., W.M.). S.C.B. is a co-founder of Lumidyne Corporation.

SUPPLEMENTARY MATERIALS

www.sciencemag.org/content/346/6210/759/suppl/DC1
Materials and Methods
Figs. S1 to S15
Tables S1 to S3
References (46–56)

4 April 2014; accepted 15 September 2014
Published online 8 October 2014;
10.1126/science.1254426

INSECT PHYLOGENOMICS

Phylogenomics resolves the timing and pattern of insect evolution

Bernhard Misof,^{1,*} Shanlin Liu,^{2,3,*} Karen Meusemann,^{1,4,*} Ralph S. Peters,^{5,*} Alexander Donath,^{1,*} Christoph Mayer,^{1,*} Paul B. Frandsen,^{6,*} Jessica Ware,^{7,*} Tomáš Flouri,^{8,*} Rolf G. Beutel,^{9,*} Oliver Niehuis,^{1,*} Malte Petersen,^{1,*} Fernando Izquierdo-Carrasco,^{8,*} Torsten Wappler,^{10,*} Jes Rust,^{10,*} Andre J. Aberer,⁸ Ulrike Aspöck,^{11,12} Horst Aspöck,¹³ Daniela Bartel,¹² Alexander Blanke,^{1,18} Simon Berger,⁸ Alexander Böhm,¹² Thomas R. Buckley,¹⁴ Brett Calcott,¹⁵ Junqing Chen,³ Frank Friedrich,¹⁶ Makiko Fukui,¹⁷ Mari Fujita,¹⁸ Carola Greve,¹ Peter Grobe,¹ Shengchang Gu,³ Ying Huang,^{2,3} Lars S. Jermiin,¹⁹ Akito Y. Kawahara,²⁰ Lars Krogmann,²¹ Martin Kubiak,¹⁶ Robert Lanfear,^{22,23,24} Harald Letsch,²⁵ Yiyuan Li,^{2,3} Zhenyu Li,³ Jiguang Li,³ Haorong Lu,³ Ryuichiro Machida,¹⁸ Yuta Mashimo,¹⁸ Pashalia Kapli,^{8,26} Duane D. McKenna,²⁷ Guanliang Meng,^{2,3} Yasutaka Nakagaki,¹⁸ José Luis Navarrete-Heredia,²⁸ Michael Ott,²⁹ Yanxiang Ou,³ Günther Pass,¹² Lars Podsiadlowski,³⁰ Hans Pohl,⁹ Björn M. von Reumont,³¹ Kai Schütte,³² Kaoru Sekiya,¹⁸ Shota Shimizu,¹⁸ Adam Slipinski,⁴ Alexandros Stamatakis,^{8,33} Wenhui Song,^{2,3} Xu Su,^{2,3} Nikolaus U. Szucsich,¹² Meihua Tan,^{2,3} Xuemei Tan,³ Min Tang,^{2,3} Jingbo Tang,³ Gerald Timelthaler,¹² Shigekazu Tomizuka,¹⁸ Michelle Trautwein,³⁴ Xiaoli Tong,³⁵ Toshiki Uchifune,^{18,36} Manfred G. Walz,¹² Brian M. Wiegmann,³⁷ Jeanne Wilbrandt,¹ Benjamin Wipfler,⁹ Thomas K. F. Wong,¹⁹ Qiong Wu,^{2,3} Gengxiong Wu,³ Yinlong Xie,³ Shenzhou Yang,^{2,3} Qing Yang,^{2,3} David K. Yeates,⁴ Kazunori Yoshizawa,³⁸ Qing Zhang,^{2,3} Rui Zhang,^{2,3} Wenwei Zhang,³ Yunhui Zhang,³ Jing Zhao,^{2,3} Chengran Zhou,^{2,3} Lili Zhou,^{2,3} Tanja Ziesmann,¹ Shijie Zou,³ Yingrui Li,³ Xun Xu,³ Yong Zhang,^{2,3} Huanming Yang,³ Jian Wang,³ Jun Wang,^{3,39,40,41,42,*} Karl M. Kjer,^{43,*} Xin Zhou,^{2,3,*}

Insects are the most speciose group of animals, but the phylogenetic relationships of many major lineages remain unresolved. We inferred the phylogeny of insects from 1478 protein-coding genes. Phylogenomic analyses of nucleotide and amino acid sequences, with site-specific nucleotide or domain-specific amino acid substitution models, produced statistically robust and congruent results resolving previously controversial phylogenetic relationships. We dated the origin of insects to the Early Ordovician [~479 million years ago (Ma)], of insect flight to the Early Devonian (~406 Ma), of major extant lineages to the Mississippian (~345 Ma), and the major diversification of holometabolous insects to the Early Cretaceous. Our phylogenomic study provides a comprehensive reliable scaffold for future comparative analyses of evolutionary innovations among insects.

Insects (*I*) were among the first animals to colonize and exploit terrestrial and freshwater ecosystems. They have shaped Earth's biota, exhibiting coevolved relationships with many groups, from flowering plants to humans. They were the first to master flight and establish social societies. However, many aspects of insect evolution are still poorly understood (2). The oldest known fossil insects are from the Early Devonian [~412 million years ago (Ma)], which has led to the hypothesis that insects originated in the Late Silurian with the earliest terrestrial ecosystems (3). Molecular

data, however, point to a Cambrian or at least Early Ordovician origin (4), which implies that early diversification of insects occurred in marine or coastal environments. Because of the absence of insect fossils from the Cambrian to the Silurian, these conclusions remain highly controversial. Furthermore, the phylogenetic relationships among major clades of polyneopteran insect orders—including grasshoppers and crickets (Orthoptera), cockroaches (Blattodea), and termites (Isoptera)—have remained elusive, as has the phylogenetic position of the enigmatic Zoraptera. Even the closest extant relatives of Holometabola (e.g.,

beetles, moths and butterflies, flies, sawflies, wasps, ants, and bees) are unknown. Thus, in order to understand the origins of physiological and morphological innovations in insects (e.g., wings and

metamorphosis), it is important to reliably reconstruct the tempo and mode of insect diversification. We therefore conducted a phylogenomic study on 1478 single-copy nuclear genes obtained from genomes and transcriptomes representing key taxa from all extant insect orders and other arthropods (144 taxa) and estimated divergence dates with a validated set of 37 fossils (5).

Phylogenomic analyses of transcriptome and genome sequence data (6) can be compromised by sparsely populated data matrices, gene paralogy, sequence misalignment, and deviations from the underlying assumptions of applied evolutionary models, which may result in biased statistical confidence in phylogenetic relationships and temporal inferences. We addressed these obstacles by removing confounding factors in our analysis (5) (fig. S2).

We sequenced more than 2.5 gigabases (Gb) of cDNA from each of 103 insect species, which represented all extant insect orders (5). Additionally, we included published transcript sequence data that met our standards (table S2) and official gene sets of 14 arthropods with sequenced draft genomes (5), of which 12 served as references during orthology prediction of transcripts (tables S2 and S4). Comparative analysis of the reference species' official gene sets identified 1478 single-copy nuclear genes present in all these species (tables S3 and S4). Functional annotation of these genes revealed that many serve basic cellular functions (tables S14 and S15 and figs. S4 to S6). A graph-based approach using the best reciprocal genome- and transcriptome-wide hit criterion identified, on average, 98% of these genes in the 103 de novo sequenced transcriptomes, but only 79% and 62% in the previously published transcriptomes of in- and out-group taxa, respectively (tables S12 and S13).

After transcripts had been assigned and aligned to the 1478 single-copy nuclear-encoded genes, we checked for highly divergent, putatively misaligned transcripts. Of the 196,027 aligned transcripts, 2033 (1%) were classified as highly divergent. Of these, 716 were satisfactorily realigned with an automated refinement. However, alignments of 1317 transcripts could not be improved, and these transcripts were excluded from our analyses (supplementary data file S5, <http://dx.doi.org/10.5061/dryad.3c0f1>).

Nonrandom distribution of missing data among taxa can inflate statistical support for incorrect tree topologies (7). Because we detected a nonrandom distribution of missing data, we only considered data blocks if they contained information from at least one representative of each of the 39 predefined taxonomic groups of undisputed monophyly (table S6). In this representative data set, the extent of missing data was still between 5 and 97.7% in pairwise sequence comparisons, with high percentages primarily because of the data scarcity in some previously published out-group taxa (table S19 and figs. S7 to S10).

We inferred maximum-likelihood phylogenetic trees (Fig. 1) with both nucleotide (second-codon positions only and applying a site-specific rate model) and amino acid–sequence data (applying

a protein domain–based partitioning scheme to improve the biological realism of the applied evolutionary models) from the representative data set (5) (figs. S21, S22, and S23, A and D). Trees from both data sets were fully congruent. The absence of taxa that cannot be robustly placed on the tree (rogue taxa) in the amino acid–sequence data set and the presence of a few rogue taxa that did not bias tree inference in the nucleotide sequence data set (5) indicated a sufficiently representative taxonomic sampling.

To detect confounding signal derived from nonrandom data coverage, we randomized amino acids within taxa, while preserving the distribution of data coverage in the representative data set (5). This approach revealed no evidence of biased node support that could be attributed to nonrandom data coverage (5) (figs. S11 and S12 and table S20). Phylogenomic data may violate the assumption of time-reversible evolutionary processes, irrespective of what partition scheme one applies, which could lead to incorrect tree estimates and biased node support. Because sections in the amino acid–sequence alignments of the representative data set violating these assumptions were present, we tested whether the observed compositional heterogeneity across taxa biased node support but found no evidence for this (5) (fig. S20). We next discarded data strongly violating the assumption of time-reversible evolutionary processes (tables S21 and S22, data files S6 to S8, and figs. S13 to S19). Results from phylogenetic analysis of this filtered data set (5) were fully congruent with those obtained from analyzing the unfiltered representative data set. The nucleotide sequence data of the representative data set containing also first and third codon positions strongly violated the assumption of time-reversible evolutionary processes, but still supported largely congruent topologies (fig. S23, B to D). In summary, our phylogenetic inferences are unlikely to be biased by any of the above-mentioned confounding factors.

Our phylogenomic study suggests an Early Ordovician origin of insects (Hexapoda) at ~479 Ma [confidence interval (CI), 509 to 452 Ma] and a radiation of ectognathous insects in the Early Silurian ~441 Ma (CI 465 to 421 Ma) (Figs. 1 and 2). These estimates imply that insects colonized land at roughly the same time as plants (8), in agreement with divergence date estimates on the basis of other molecular data (4).

The early diversification pattern of insects has remained unclear (2, 7, 9). We received support for a monophyly of insects, including Collembola and Protura as closest relatives (10), and Diplura as closest extant relatives of bristletails (Archaeognatha), silverfish (Zygentoma), and winged insects (Pterygota) (Fig. 1). Furthermore, our analyses corroborate Remipedia, cave-dwelling crustaceans, as the closest extant relatives of insects (11, 12).

A close phylogenetic relationship of bristletails to a clade uniting silverfish and winged insects (Dicondylia) is generally accepted. However, the monophyly of silverfish has been questioned, with the relict *Tricholepidion gertschi* considered more distantly related to winged insects than

¹Zoologisches Forschungsmuseum Alexander Koenig (ZFMK)/Zentrum für Molekulare Biodiversitätsforschung (ZMB), Bonn, Germany. ²China National GeneBank, BGI-Shenzhen, China. ³BGI-Shenzhen, China. ⁴Australian National Insect Collection, Commonwealth Scientific and Industrial Research Organization (Australia) (CSIRO), National Research Collections Australia, Canberra, ACT, Australia. ⁵Abteilung Arthropoda, Zoologisches Forschungsmuseum Alexander Koenig (ZFMK), Bonn, Germany. ⁶Department of Entomology, Rutgers University, New Brunswick, NJ 08854, USA. ⁷Department of Biological Sciences, Rutgers University, Newark, NJ 08854, USA. ⁸Scientific Computing, Heidelberg Institute for Theoretical Studies (HITS), Heidelberg, Germany. ⁹Institut für Spezielle Zoologie und Evolutionsbiologie mit Phyletischem Museum Jena, FSU Jena, Germany. ¹⁰Steinmann-Institut, Bereich Paläontologie, Universität Bonn, Germany. ¹¹Zoologische Abteilung (Insekten), Naturhistorisches Museum Wien, Vienna, Austria. ¹²Department of Integrative Zoology, Universität Wien, Vienna, Austria. ¹³Institut für Spezifische Prophylaxe und Tropenmedizin, Medizinische Parasitologie, Medizinische Universität Wien (MUW), Vienna, Austria. ¹⁴Manaaki Whenua Landcare Research, Auckland, New Zealand. ¹⁵Center for Advanced Modeling, Emergency Medicine Department, Johns Hopkins University, Baltimore, MD 21209, USA. ¹⁶Biozentrum Grindel und Zoologisches Museum, Universität Hamburg, Hamburg, Germany. ¹⁷Evolutionary Morphology Laboratory, Graduate School of Science and Engineering, Ehime University, Japan. ¹⁸Sugadaira Montane Research Center/Hexapod Comparative Embryology Laboratory, University of Tsukuba, Japan. ¹⁹Land and Water Flagship, CSIRO, Canberra, ACT, Australia. ²⁰Florida Museum of Natural History, University of Florida, Gainesville, FL 32611, USA. ²¹Entomology, Staatliches Museum für Naturkunde Stuttgart (SMNS), Germany. ²²Ecology Evolution and Genetics, Research School of Biology, Australian National University, Canberra, ACT, Australia. ²³National Evolutionary Synthesis Center, Durham, NC 27705, USA. ²⁴Department of Biological Sciences, Macquarie University, Sydney, Australia. ²⁵Department für Botanik und Biodiversitätsforschung, Universität Wien, Vienna, Austria. ²⁶Natural History Museum of Crete, University of Crete, Post Office Box 2208, GR-71409, Iraklio, and Biology Department, University of Crete, Iraklio, Crete, Greece. ²⁷Department of Biological Sciences and Feinstone Center for Genomic Research, University of Memphis, Memphis, TN 38152, USA. ²⁸Centro Universitario de Ciencias Biológicas y Agropecuarias, Centro de Estudios en Zoología, Universidad de Guadalajara, Zapopan, Jalisco, México. ²⁹Leibniz Supercomputing Centre of the Bavarian Academy of Sciences and Humanities, Garching, Germany. ³⁰Institute of Evolutionary Biology and Ecology, Zoology and Evolutionary Biology, University of Bonn, Bonn, Germany. ³¹Department of Life Sciences, The Natural History Museum London, London, UK. ³²Abteilung Entomologie, Biozentrum Grindel und Zoologisches Museum, Universität Hamburg, Hamburg, Germany. ³³Fakultät für Informatik, Karlsruher Institut für Technologie, Karlsruhe, Germany. ³⁴California Academy of Sciences, San Francisco, CA 94118, USA. ³⁵Department of Entomology, College of Natural Resources and Environment, South China Agricultural University, China. ³⁶Yokosuka City Museum, Yokosuka, Kanagawa, Japan. ³⁷Department of Entomology, North Carolina State University, Raleigh, NC 27695, USA. ³⁸Systematic Entomology, Hokkaido University, Sapporo, Japan. ³⁹Department of Biology, University of Copenhagen, Copenhagen, Denmark. ⁴⁰Princess Al Jawhara Center of Excellence in the Research of Hereditary Disorders, King Abdulaziz University, Jeddah, Saudi Arabia. ⁴¹Macau University of Science and Technology, Avenida Wai Long, Taipa, Macau, China. ⁴²Department of Medicine, University of Hong Kong, Hong Kong. ⁴³Department of Ecology, Evolution, and Natural Resources, Rutgers University, New Brunswick, NJ 08854, USA.

*Major contributors.
†Corresponding author. E-mail: xinzhou@genomics.cn (X.Z.), b.misof.zfmk@uni-bonn.de (B.M.), kjer@aesop.rutgers.edu (K.M.K.), wangj@genomics.cn (J.W.)

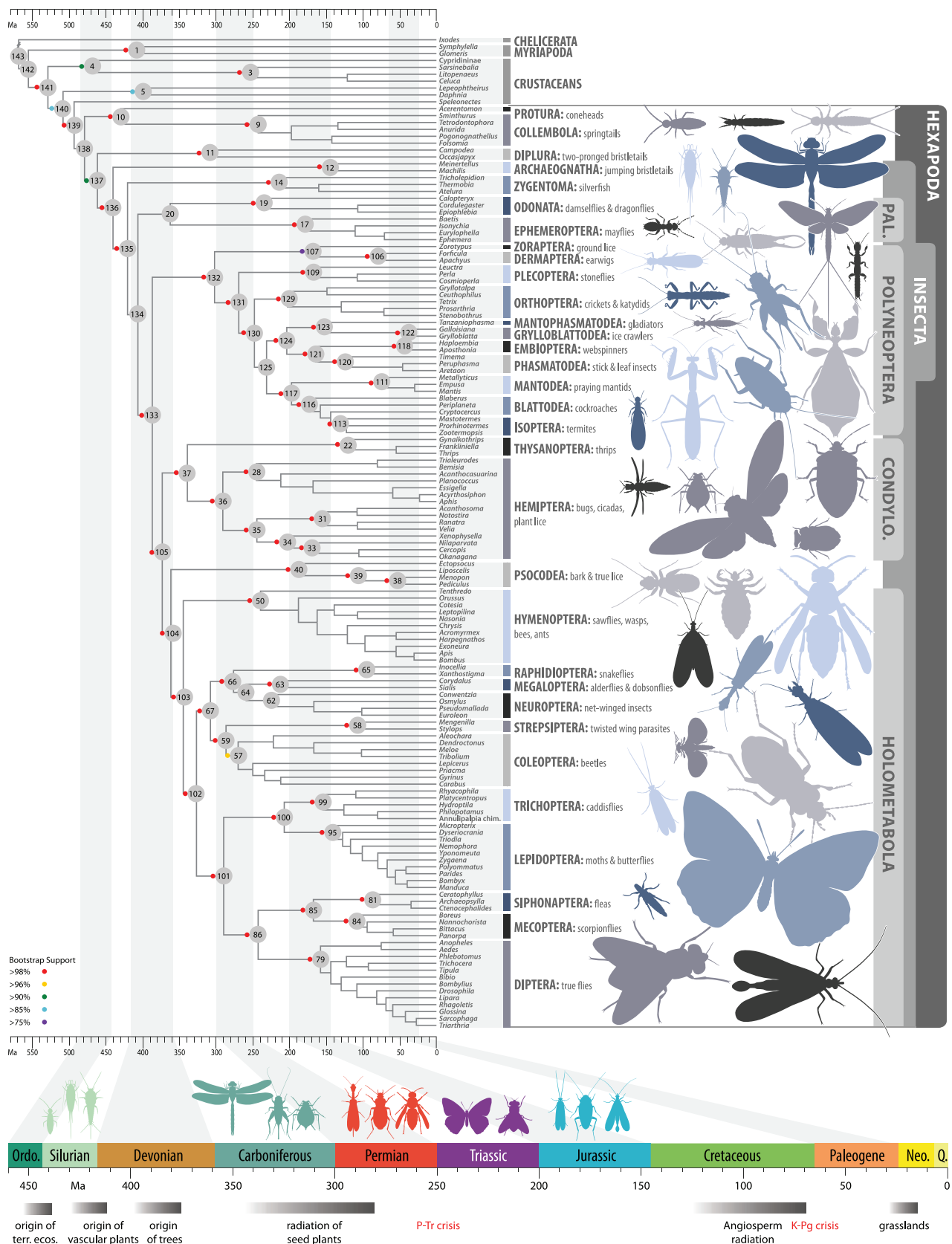


Fig. 1. Dated phylogenetic tree of insect relationships. The tree was inferred through a maximum-likelihood analysis of 413,459 amino acid sites divided into 479 metapartitions. Branch lengths were optimized and node ages estimated from 1,050,000 trees sampled from trees separately generated for 105 partitions that included all taxa (5). All nodes up to orders are labeled with numbers (gray circles). Colored circles indicate bootstrap support (5) (left key). The time line at the bottom of the tree relates the geological origin of insect clades to major geological and biological events. CONDYLLO, Condylgnatha; PAL, Palaeoptera.

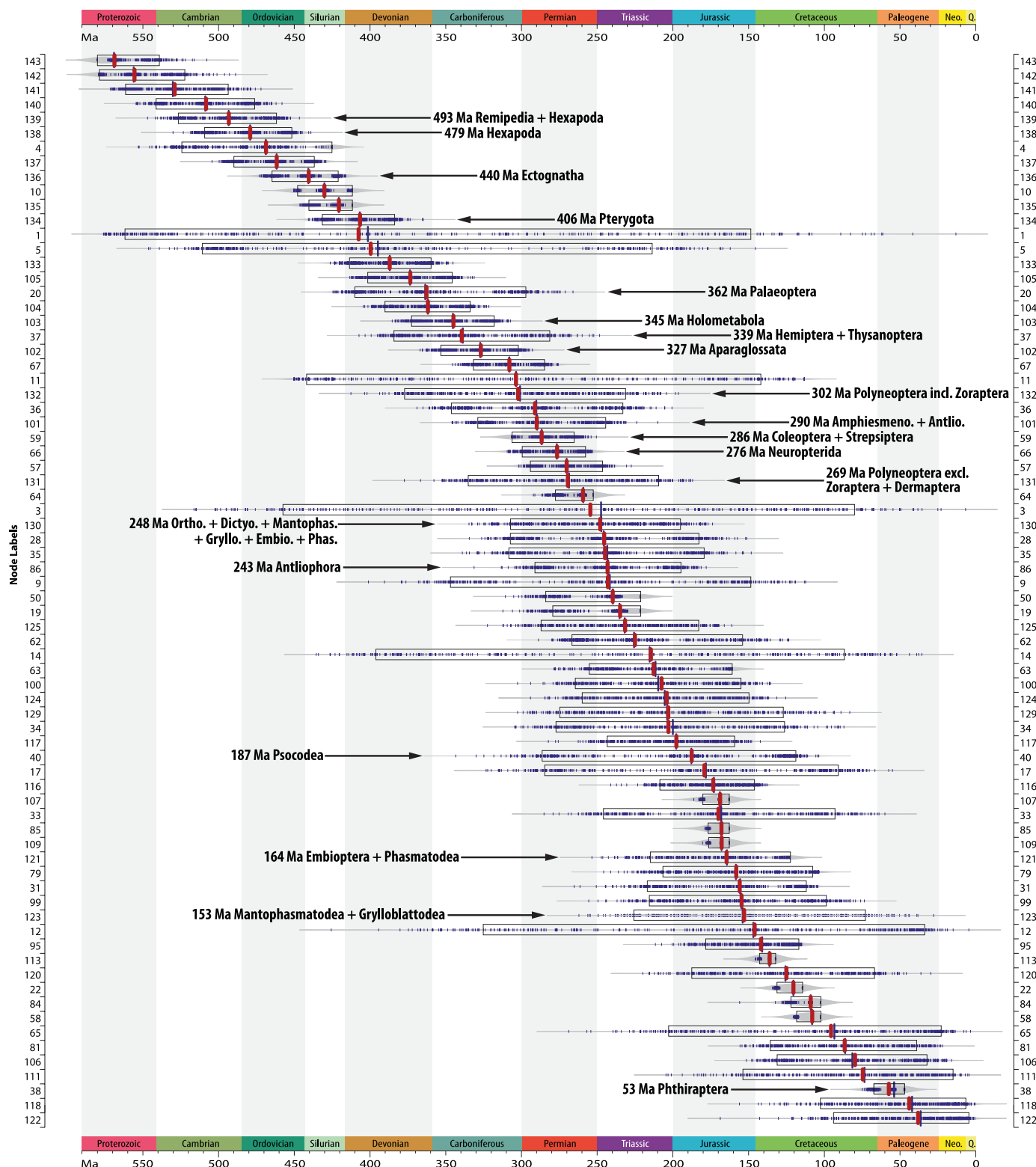


Fig. 2. Sorted ordinal and interordinal node age estimates. For each labeled node (numbers on the left and right of the figure correspond to the node labels in the tree of Fig. 1), the median (red bar), and the range of the upper and lower confidence interval (black rectangle) of age estimates are illustrated. These medians and upper and lower confidence intervals are derived from uniformly sampled trees over all 105 metapartitions (5). Additionally, we present medians

of age estimates separately derived from each metapartition. Within the bean plot (gray scale), blue bars indicate the distribution of median age estimates, large blue bars indicate the inferred median of medians. All node age estimates refer to the estimated common origin of included species. Stem-lineage representatives can, of course, be older. The maximum root age of the tree was set to 580 Ma to coincide with the oldest Ediacaran fossils (5).

other silverfish (13). We find that silverfish are monophyletic, consistent with recently published morphological studies (14), and estimate that *Tricholepidion* diverged from other silverfish in the Late Triassic (~214 Ma) (Figs. 1 and 2). This result implies parallel and independent loss of the ligamentous head endoskeleton, abdominal styli, and coxal vesicles in winged insects and silverfish (5).

The diversification of insects is undoubtedly related to the evolution of flight. Fossil winged insects exist from the Late Mississippian (~324 Ma) (15), which implies a pre-Carboniferous origin of insect flight. The description of †*Rhynognatha* (~412 Ma) from a mandible, potentially indicative of a winged insect, suggested an Early Devonian to Late Silurian origin of winged insects (3). Our results corroborate an origin of winged insect lineages during this time period (16) (Figs. 1 and 2), which implies that the ability to fly emerged after the establishment of complex terrestrial ecosystems.

Ephemeroptera and Odonata are, according to our analyses, derived from a common ancestor. However, node support is low for Palaeoptera (Ephemeroptera + Odonata) and for a sister group relationship of Palaeoptera to modern winged insects (Neoptera), which indicates that additional evidence, including extensive taxon sampling and the analysis of genomic meta-characters (17), will be necessary to corroborate these relationships.

We find strong support for the monophyly of Polyneoptera, a group that comprises earwigs, stoneflies, grasshoppers, crickets, katydids (Orthoptera), Embioptera, Phasmatodea, Mantophasmatodea, Grylloblattodea, cockroaches, mantids, termites, and Zoraptera (18–20). We estimated the origin of the polyneopteran lineages at ~302 Ma (CI 377 to 231 Ma) in the Pennsylvanian (Figs. 1 and 2), consistent with the idea that at least part of the rich Carboniferous neopteran insect fauna was of polyneopteran origin. Finally, our analyses suggest that the major diversity within living cockroaches, mantids, termites, and stick insects evolved after the Permian mass extinction.

Given that the oldest known fossil hemipterans date to the Middle Pennsylvanian (~310 Ma) (21), it had been thought that the stylet marks on liverworts from the Late Devonian (~380 Ma) (22) could not have been of hemipteran origin. Our study indicates that true bugs (Hemiptera) and their sister lineage, thrips (Thysanoptera), all of which possess piercing-sucking mouthparts, orig-

inated ~373 Ma (CI 401 to 346 Ma), which gives support to the possibility of a hemipteroid origin of Early Paleozoic stylet marks.

True bugs, thrips, bark lice (Psocoptera), and true lice (Phthiraptera) (together called Acercaria) were thought to be the closest extant relatives of Holometabola (Acercaria + Holometabola = Eumetabola) (10). However, convincing morphological features and fossil intermediates supporting a monophyly of Acercaria are lacking (13). We recovered bark and true lice (Psocodea) as likely closest extant relatives of Holometabola (5), which suggests that both groups started to diverge in the Devonian-Mississippian ~362 Ma (CI 390 to 334 Ma) (Figs. 1 and 2). However, this result did not receive support in all statistical tests and, therefore, should be further investigated in future studies that embrace additional types of characters (17).

We estimated that the radiation of parasitic lice occurred ~53 Ma (CI 67 to 46 Ma), which implies that they diversified well after the emergence of their avian and mammalian hosts in the Late Cretaceous–Early Eocene and contradicts the hypothesis that parasitic lice originated on feathered theropod dinosaurs ~130 Ma (23).

Within Holometabola, our study recovered phylogenetic relationships fully congruent with those suggested in recent studies (2, 24, 25). Although we estimated the origin of stem lineages of many holometabolous insect orders in the Late Carboniferous, we dated the spectacular diversifications within Hymenoptera, Diptera, and Lepidoptera to the Early Cretaceous, contemporary with the radiation of flowering plants (21, 26). The almost linear increase in interordinal insect diversity suggests that the process of diversification of extant insects may not have been severely affected by the Permian and Cretaceous biodiversity crises (Fig. 2).

With this study, we have provided a robust phylogenetic backbone tree and reliable time estimates of insect evolution. These data and analyses establish a framework for future comparative analyses on insects, their genomes, and their morphology.

REFERENCES AND NOTES

1. The term “insects” is used here in a broad sense and synonymous to Hexapoda (including the ancestrally wingless Protura, Collembola, and Diplura).
2. M. D. Trautwein, B. M. Wiegmann, R. Beutel, K. M. Kjer, D. K. Yeates, *Annu. Rev. Entomol.* **57**, 449–468 (2012).
3. M. S. Engel, D. A. Grimaldi, *Nature* **427**, 627–630 (2004).
4. O. Rota-Stabelli et al., *Curr. Biol.* **23**, 392–398 (2013).

5. Materials and methods are available as supplementary material on Science Online.
6. Transcriptome refers to the sequencing of all of the mRNAs of an individual or many individuals present at the time of preservation.
7. E. Dell’Ampio et al., *Mol. Biol. Evol.* **31**, 239–249 (2014).
8. C. C. Labandeira, *Arthro. Syst. Phylo.* **64**, 53–94 (2006).
9. K. Meusemann et al., *Mol. Biol. Evol.* **27**, 2451–2464 (2010).
10. W. Hennig, *Die Stammesgeschichte der Insekten* (Kramer, Frankfurt am Main, 1969).
11. With the notable exception of Cephalocarida, for which RNA-sequencing data were unavailable to us.
12. B. M. von Reumont et al., *Mol. Biol. Evol.* **29**, 1031–1045 (2012).
13. N. P. Kristensen, *Annu. Rev. Entomol.* **26**, 135–157 (1981).
14. A. Blanke, M. Koch, B. Wipfler, F. Wilde, B. Misof, *Front. Zool.* **11**, 16 (2014).
15. J. Prokop, A. Nel, I. Hoch, *Geobios* **38**, 383–387 (2005).
16. These estimates are robust whether or not †*Rhynognatha* (Pragian stage, ~412 Ma) is used as a calibration point.
17. O. Niehuis et al., *Curr. Biol.* **22**, 1309–1313 (2012).
18. H. Letsch, S. Simon, *Syst. Entomol.* **38**, 783–793 (2013).
19. K. Ishiwata, G. Sasaki, J. Ogawa, T. Miyata, Z.-H. Su, *Mol. Phylogenet. Evol.* **58**, 169–180 (2011).
20. K. Yoshizawa, *Syst. Entomol.* **36**, 377–394 (2011).
21. A. Nel et al., *Nature* **503**, 257–261 (2013).
22. C. C. Labandeira, S. L. Tremblay, K. E. Bartowski, L. VanAller-Hernick, *New Phytol.* **202**, 247–258 (2014).
23. V. S. Smith et al., *Biol. Lett.* **7**, 782–785 (2011).
24. R. G. Beutel et al., *Cladistics* **27**, 341–355 (2011).
25. B. M. Wiegmann et al., *BMC Biol.* **7**, 34 (2009).
26. J. A. Doyle, *Annu. Rev. Earth Planet. Sci.* **40**, 301–326 (2012).

ACKNOWLEDGMENTS

The data reported in this paper are tabulated in the supplementary materials and archived at National Center for Biotechnology Information, NIH, under the Umbrella BioProject ID PRJNA183205 (“The 1KITE project: evolution of insects”). Supplementary files are archived at the Dryad Digital Repository <http://dx.doi.org/10.5061/dryad.3c0f1>. Funding support: China National GeneBank and BGI-Shenzhen, China; German Research Foundation (NI 1387/1-1; MI 649/6, MI 649/10, RE 345/1-2, BE1789/8-1, BE 1789/10-1, STA 860/4, Heisenberg grant WA 1496/8-1); Austria Science Fund FWF; NSF (DEB 0816865); Ministry of Education, Culture, Sports, Science and Technology of Japan Grant-in-Aid for Young Scientists (B 22770090); Japan Society for the Promotion of Science (P14071); Deutsches Elektronen-Synchrotron (I-20120065); Paul Scherrer Institute (20110069); Schlinger Endowment to CSIRO Ecosystem Sciences; Heidelberg Institute for Theoretical Studies; University of Memphis-FedEx Institute of Technology; and Rutgers University. The authors declare no conflicts of interest.

SUPPLEMENTARY MATERIALS

www.sciencemag.org/content/346/6210/763/suppl/DC1
Materials and Methods
Supplementary Text
Figs. S1 to S27
Tables S1 to S26
Data File Captions S1 to S14
References (27–187)

17 June 2014; accepted 23 September 2014
10.1126/science.1257570