

# 1 Supporting Information

## 2 SI 1: Implementation of the Crofton Method

3 The algorithm for fitting the Crofton Method (Crofton 1971) proceeds as follows.  
4 First, obtain a dataset with  $n$  hosts where each host has some parasite intensity 0  
5 to  $p_{max}$ . Starting with the full dataset, guess a vector of pre-mortality parameters  
6  $(N_{p1}, \mu_{p1}, k_{p1})$  where  $N_{p1}$  is the total number of hosts before mortality,  $\mu_{p1}$   
7 is the parasite intensity before mortality, and  $k_{p1}$  is the parasite aggregation  
8 before mortality. Given these parameters, use a negative binomial distribution  
9 to calculate the predicted number of hosts with  $0, 1, 2, \dots, p_{max}$  parasites. Compare  
10 the expected number of hosts with  $0, 1, 2, \dots, p_{max}$  parasites to the observed  
11 number hosts with  $0, 1, 2, \dots, p_{max}$  parasites and calculate the  $\chi^2$ -squared statistic  
12 associated with the observed and predicted vectors. In reality, one often has to bin  
13 the parasite intensity data because all parasite intensities are not represented in  
14 the dataset. Continue to guess  $(N_{p1}, \mu_{p1}, k_{p1})$  vectors until a set of parameters is  
15 found that minimizes the  $\chi^2$ -squared statistic.

16 Second, choose a truncation value ( $t_2$ ) such that  $t_2 < p_{max}$ . Truncate the  
17 data such that  $\text{data}_{\text{truncated}} \leq t_2$  and repeat the above iterative procedure to  
18 calculate another set of parameters  $(N_{p2}, \mu_{p2}, k_{p2})$  that minimizes the  $\chi^2$ -squared  
19 statistic on the truncated data. Choose a new truncated value  $t_3 < t_2$  and repeat  
20 the first two steps. Continue to truncate the dataset until it only contains hosts  
21 with 0, 1, and 2 parasites (or 3 bins). Because the method attempts to estimate  
22 three parameters, at least 3 classes are needed for all 3 parameters to be identifiable  
23 (Royce and Rossignol 1990).

24 Once the iterative procedure has been completed, parasite-induced host  
25 mortality is traditionally identified by plotting the different truncation values  $t_i$   
26 against the different values of  $N_{pi}$  and looking for a distinct “kink” in the resulting

plot. Once the “kink” as occurred, the values of  $N_{pi}$  will typically remain close to constant as  $t_i$  is decreased further. The “true” pre-mortality parameters  $N_{pt}$ ,  $\mu_{pt}$ , and  $k_{pt}$  are taken to be at the point where the “kink” occurs.

We provide an implementation and unit tests of the Crofton Method in Supplementary Information 4. Figure 10 visually shows that our implementation of the Crofton Method agrees with results previously published by Crofton (1971).

## SI 2: Implementation of the Adjei Method

The Adjei Method for estimating PIHM has two steps (Adjei et al. 1986). The first step is to estimate the parameters of the pre-mortality host-parasite distribution using the Crofton Method (see *SI 1*). The three parameters estimated are the total number of hosts before mortality  $N_p$ , the mean number of parasites per host before mortality  $\mu_p$ , and the aggregation of parasites before mortality given by the parameter  $k_p$  from a negative binomial distribution. When  $k_p$  is small, parasites are highly aggregated among hosts and when  $k_p$  is large parasites are more evenly distributed across hosts (Wilson et al. 2002). The implementation of the Crofton Method has been discussed at length elsewhere (e.g. Royce and Rossignol 1990; Lester 1984, and in *SI 1*) and we provide a tested implementation of the method in *SI 4*.

The second step of the Adjei Method is to make the assumption that infection, host mortality, and sampling occur in that order and are temporally Adjei et al. (1986). Next, Adjei et al. assume that the host survival function follows the logistic form

$$h(\text{survival}; x, a, b) = \frac{\exp(a - b \log(x))}{1 + \exp(a - b \log(x))} \quad (1)$$

where  $x$  is the parasite intensity in a given host and  $a$  and  $b$  are the two parameters

of the logistic function. Generally, a larger  $a$  allows for hosts to tolerate larger parasite intensities before experiencing parasite-induced mortality and a larger  $b$  leads to a more rapid decline in the probability of host survival as parasite intensity increases. The value  $\exp(a/b)$  is referred to as the  $LD_{50}$ . Individuals with loads higher than this will have a greater than 50% chance of death.

By taking the first and second derivatives of equation 1, one can easily find that the maximum rate of decline in host survival probability with increasing parasite intensity occurs at the  $LD_{50}$  and has a value of  $b/4$ . This is in many ways analogous to the parasite pathogenicity parameter  $\alpha$  given in classic macroparasite models, which specifies the slope of the linear relationship between host death rate and parasite intensity (Anderson and May 1978; Isham 1995). The parameter  $a$  is easily interpreted by holding  $b$  constant and looking at how a one unit change in  $a$  affects the log parasite intensity at which some percentage  $p$  of hosts experience mortality. Letting  $a_1$  and  $a_2$  be two different values of  $a$  and  $x_1$  and  $x_2$  be two different parasite intensities, a bit of rearranging of equation 1 gives.

$$\begin{aligned}\log \frac{p}{1-p} &= a_1 - b \log x_1 \\ -\log \frac{p}{1-p} &= a_1 - b \log x_2 \\ 0 &= a_1 - a_2 - b \log x_1 + b \log x_2 \\ a_2 - a_1 &= b(\log x_2 - \log x_1)\end{aligned}$$

If  $a_2 - a_1 = 1$ , then the change in log parasite intensity at which  $p$  percentage of hosts survive is  $1/b$ .

To estimate the parameters in equation 1, the Adjei Method first calculates the expected number of hosts with a given parasite load  $x$  by using the equation  $g(x; \mu_p, k_p) * N_p$ , where  $g(x; \mu_p, k_p)$  is the negative binomial pre-mortality

70 distribution. Second, the observed and predicted number of hosts with  $x$  parasites  
71 are paired as a single data point and the method then assumes that this data  
72 point follows a binomial distribution with the total number of “trials” equal to  
73 the predicted number of hosts and the total number of “successes” equal to the  
74 observed number of hosts. In some cases, the observed number of hosts is greater  
75 than the expected number of hosts and the Adjei Method alters the data so that  
76 the observed is equal to the predicted (Adjei et al. 1986). After this questionable  
77 manipulation, the (observed, predicted) pairs are fit to a standard Generalized  
78 Linear Model (McCullagh and Nelder 1989) with a binomial response variable  
79 and a logistic link function given by equation 1. This model provides estimates for  
80 parameters  $a$ ,  $b$  and  $LD_{50}$ .

81 While not included in the original implementation of the Adjei Method,  
82 a  $\chi^2$  test with a degrees of freedom of 1 can be used to assess whether a GLM  
83 model that includes parasite intensity as a predictor of host survival probability is  
84 a “better” model than a GLM without this predictor. This allows the Adjei Method  
85 to determine whether PIHM is a significant factor in a host-parasite system.

86 The Adjei Method’s most glaring deficiency is the need to alter the observed  
87 data in order to fit the model into the binomial GLM framework. A second more  
88 subtle problem with the Adjei Method is the potential need to bin data in order  
89 to predict greater than one host in a given parasite intensity class. For example,  
90 if the total number of hosts pre-mortality was 50, the mean number of parasites  
91 per host pre-mortality was 100 and the aggregation parameter was 1, applying  
92 the equation  $g(x; \mu_p = 100, k_p = 1) * 50$  would result in less than 1 individual in  
93 all parasite intensities  $x$ . In other words, the Adjei Method cannot be applied to  
94 samples with either very high mean parasite loads, small sample sizes, or both  
95 without some sort of binning of the data. While this is not a flaw *per se*, it does  
96 add a certain level of subjectivity (i.e. which bins should you use?) to a method  
97 that already has serious potential issues. In this analysis, we always assume the

98 Adjei Method is not binning the data, though we provide code for applying the  
99 binning method in SI 4.

## 100 **SI 3: Additional Figures**

101 See Figures 1 - 9.

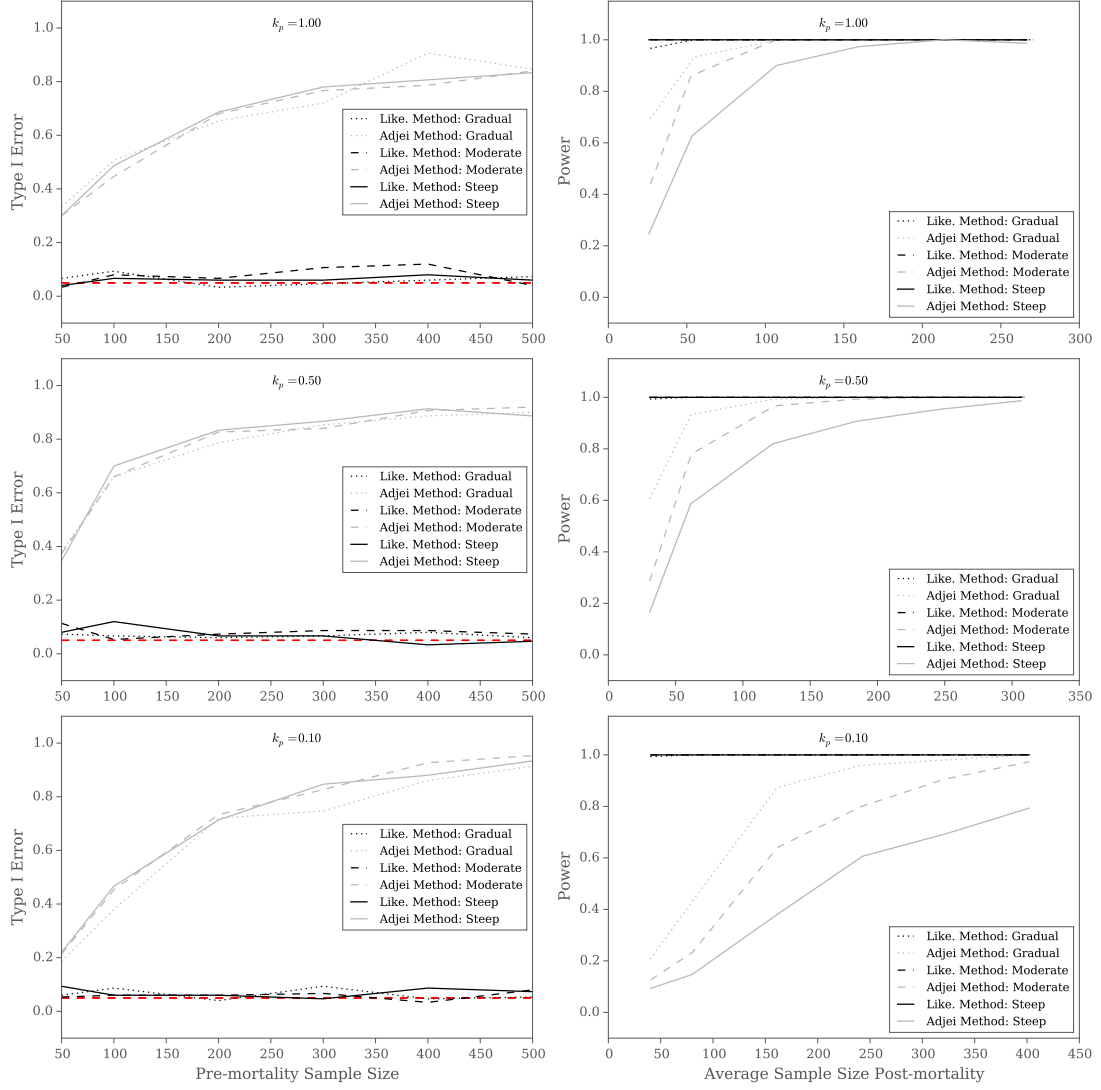
## 102 **SI 4: Code and unit tests for estimating parasite-induced host** 103 **mortality**

104 Python code, unit tests, and a help file for the Crofton Method, the Adjei Method  
105 and the Likelihood Method can be found at [https://github.com/mqwilber/](https://github.com/mqwilber/parasite_mortality)  
106 `parasite_mortality`

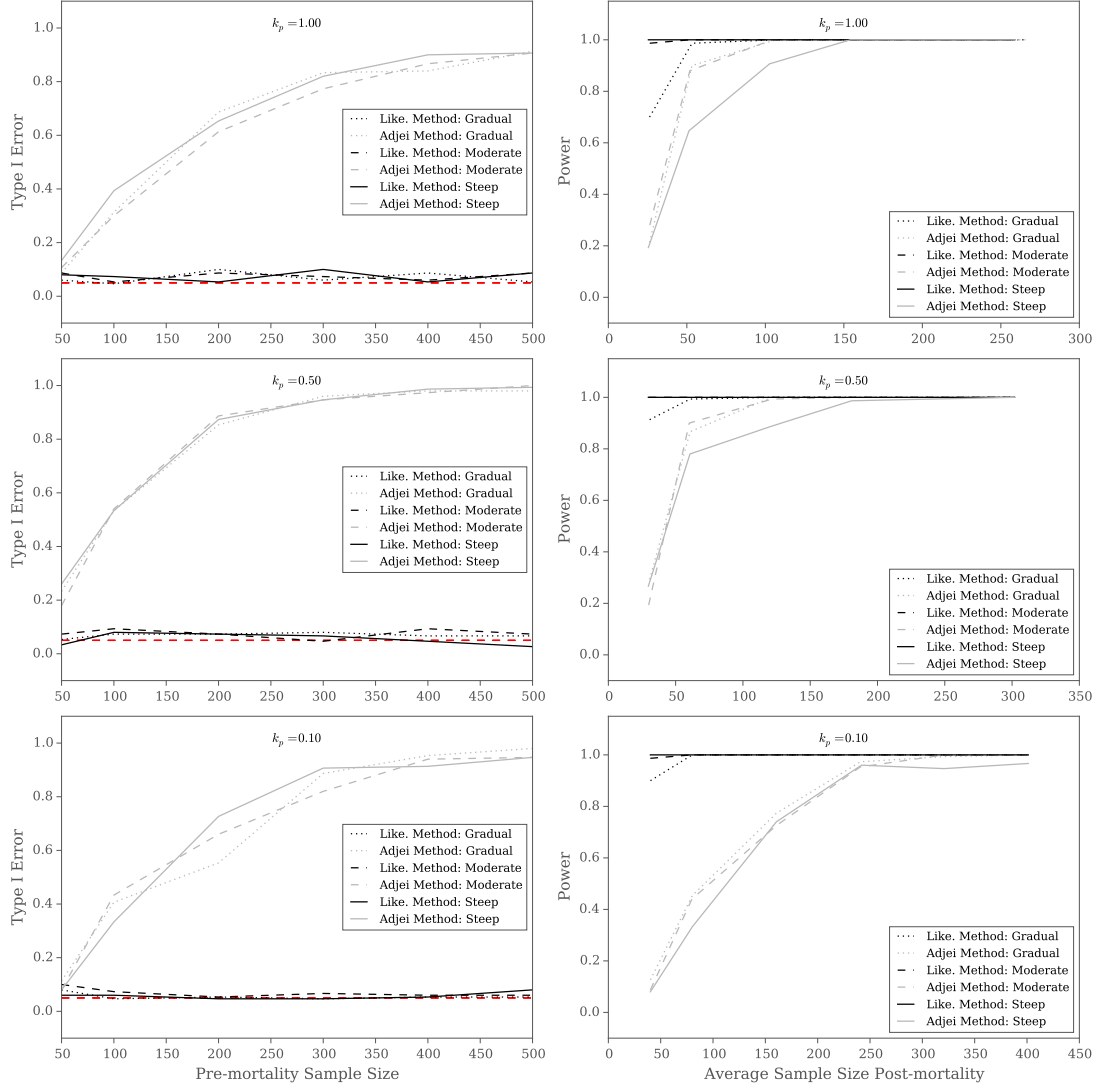
## 107 **References**

- 108 1.  
109 Adjei, E.L., Barnes, A., Lester, R.J.G., 1986. A method for estimating possible  
110 parasite-related host mortality, illustrated using data from *Callitetrarhynchus*  
111 *gracilis* (Cestoda: Trypanorhyncha) in lizardfish (*Saurida* spp.). *Parasitology*. 92,  
112 227–243.
- 113 2.  
114 Anderson, R.M., May, R.M., 1978. Regulation and stability of host-parasite  
115 interactions: I. Regulatory processes. *J. Anim. Ecol.* 47, 219–247.
- 116 3.  
117 Crofton, H.D. 1971. A quantitative approach to parasitism. *Parasitology*. 62,  
118 179–193.
- 119 4.  
120 Isham, V. 1995. Stochastic models of host-macroparasite interaction. *Ann. Appl.*  
121 *Probab.* 5, 720–740.
- 122 5.  
123 Lester, R.J.G. 1984. A review of methods for estimating mortality due to parasites  
124 in wild fish populations. *Helgolander Meeresun.* 37, 53–64.
- 125 6.  
126 McCullagh, P., Nelder, J.A., 1989. *Generalized Linear Models* 2nd edn. Chapman  
127 & Hall, New York.

- 128 7.  
129 Royce, L.A., Rossignol, P., 1990. Epidemiology of honey bee parasites. *Parasitol.*  
130 *Today.* 6, 348–353.
- 131 8.  
132 Wilson, K., Bjoernstad, O.N., Dobson, A.P., Merler, S., Poglayen, G., Read, A.F.,  
133 Skorpington, A., 2002. Heterogeneities in macroparasite infections: patterns and  
134 processes. In: Hudson, P.J., Rizzoli, A., Grenfell, B., Heesterbeek, H., Dobson,  
135 A., (Eds.), *The Ecology of Wildlife Diseases*. Oxford University Press, Oxford, pp.  
136 6–44.

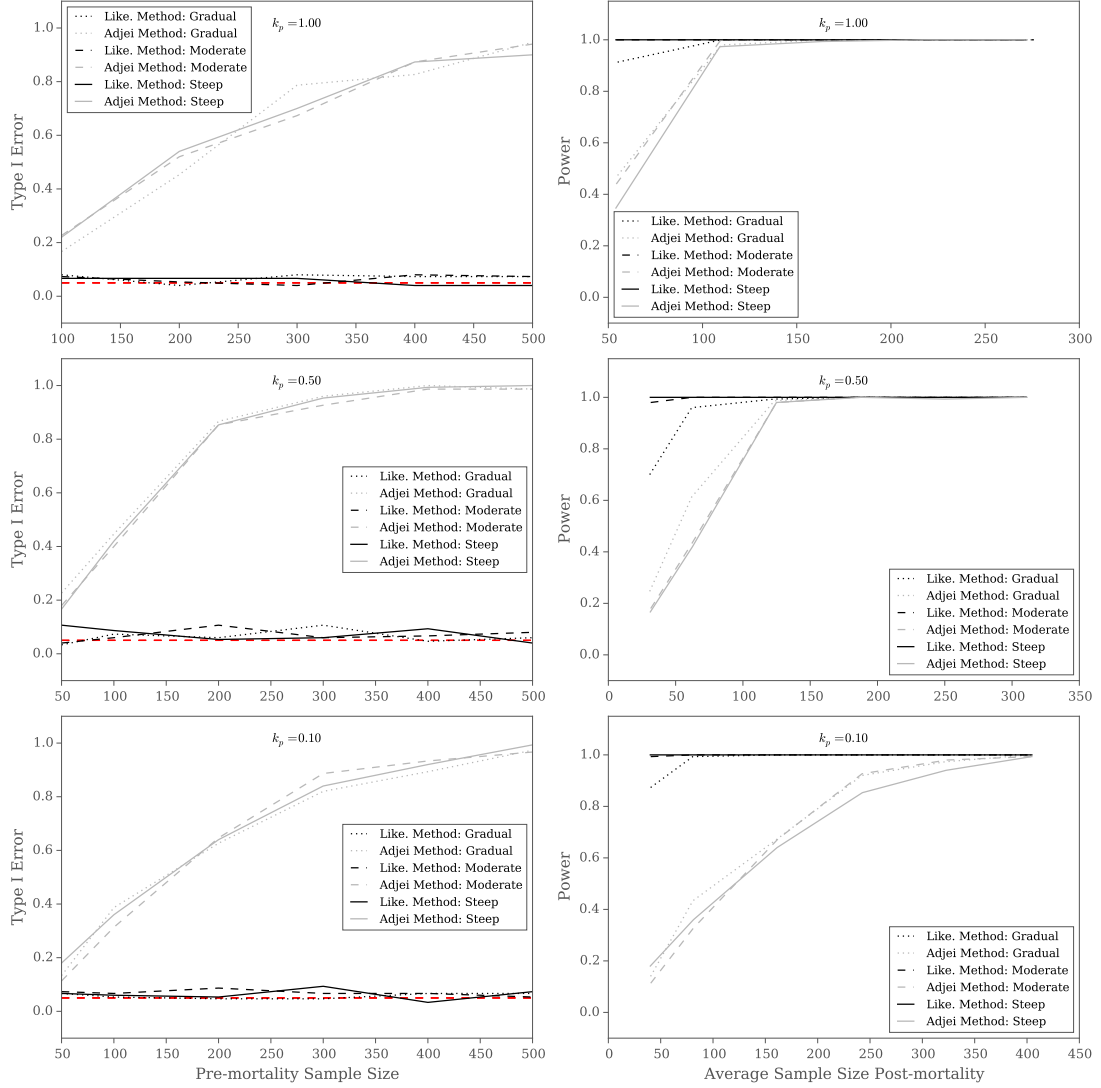


**Figure 1:** The type I error rate and the power of the Likelihood Method (solid line) and the Adjei Method (dashed lines) when  $\mu_p = 10$  for various shapes of the host survival function and levels of aggregation  $k_p$ . The first column gives the type I error rate of each method for falsely detecting PIHM when none is present. The red line gives the the pre-set type I error rate of  $\alpha = 0.05$ . The second column gives the power of a given method to detect PIHM when it is actually occurring.

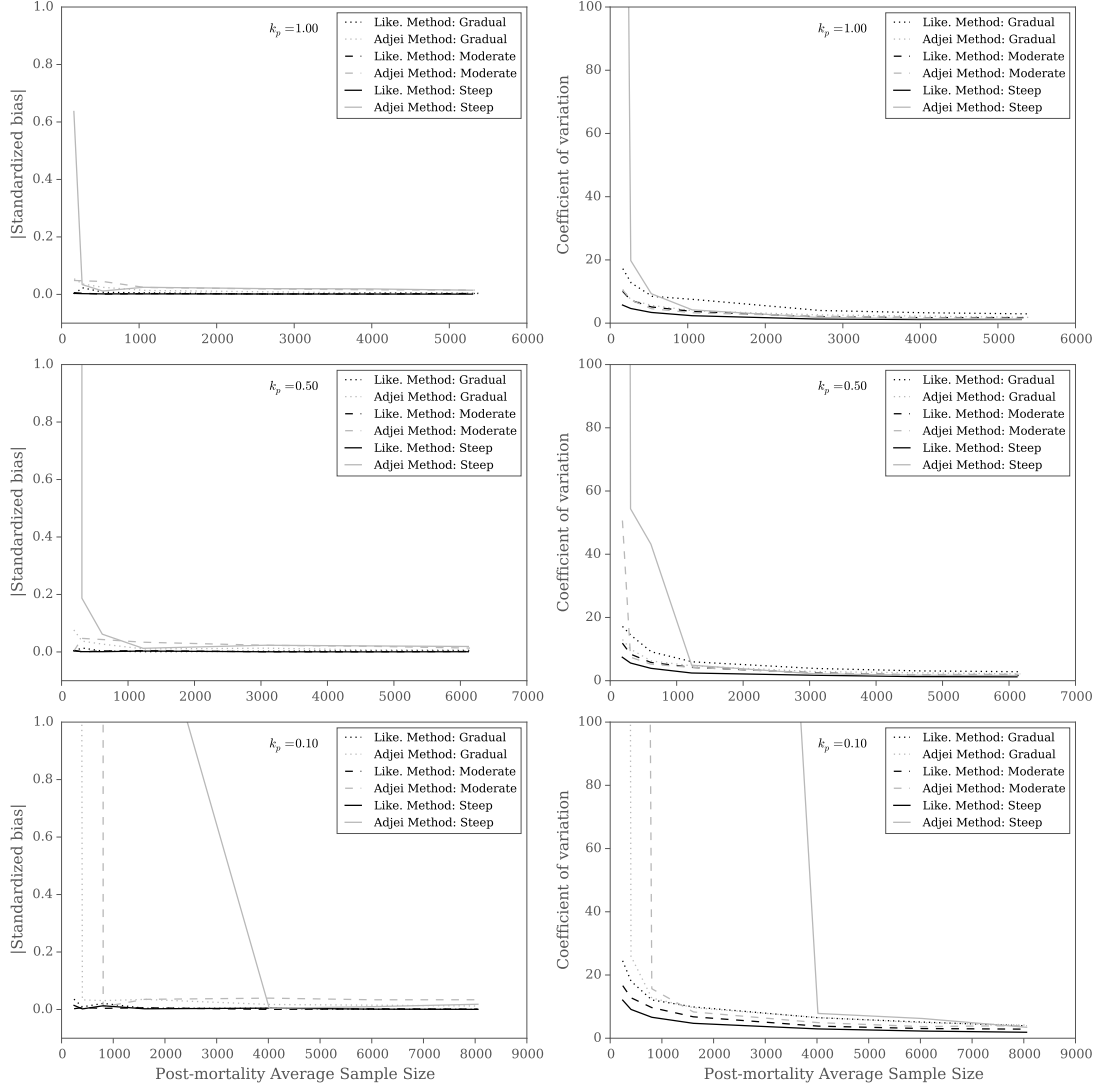


**Figure 2:** The type I error rate and the power of the Likelihood Method (solid line) and the Adjei Method (dashed lines) when  $\mu_p = 50$  for various shapes of the host survival function and levels of aggregation  $k_p$ . The first column gives the type I error rate of each method for falsely detecting PIHM when none is present. The red line gives the the pre-set type I error rate of  $\alpha = 0.05$ . The second column gives the power of a given method to detect PIHM when it is actually occurring.

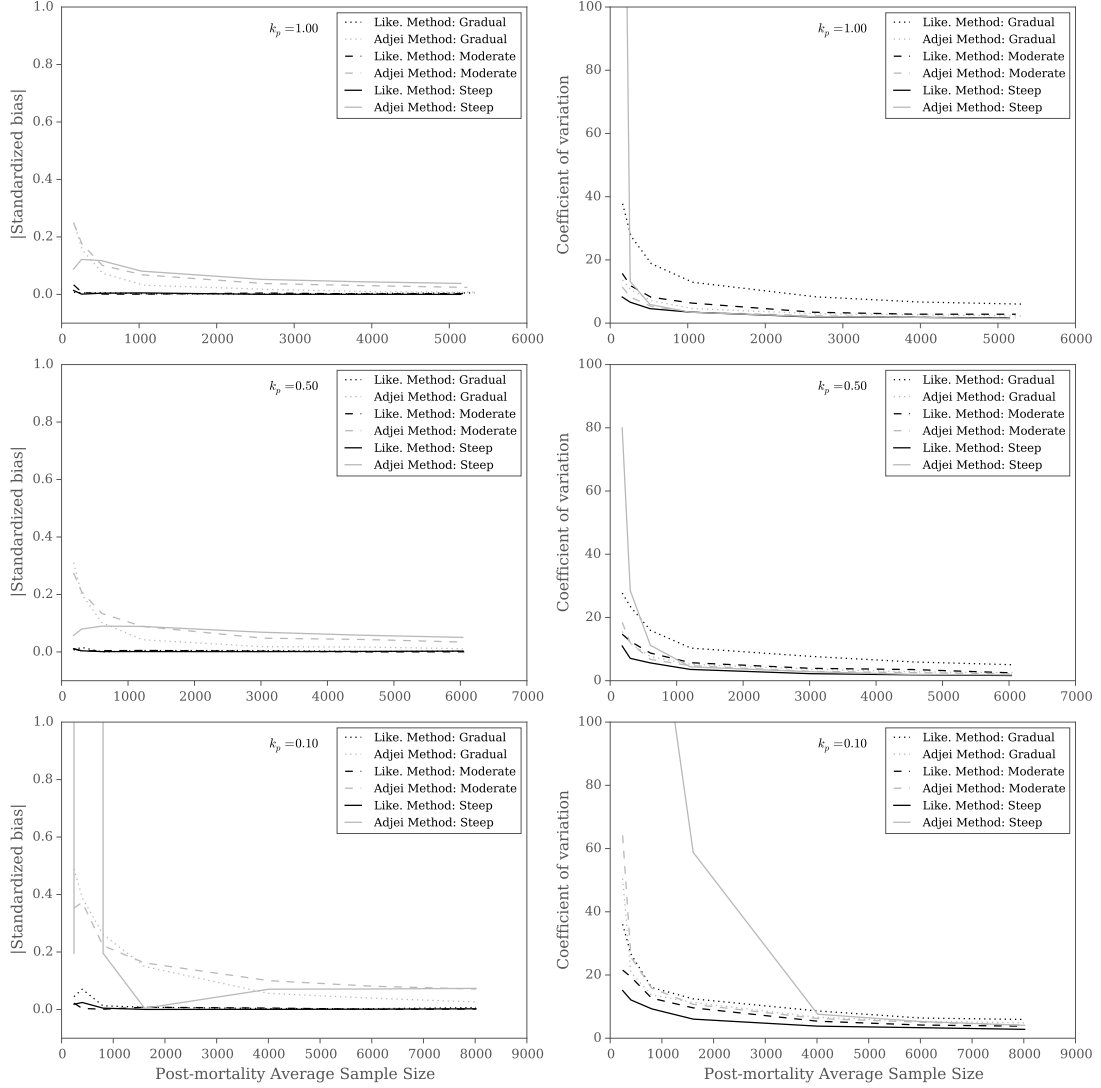




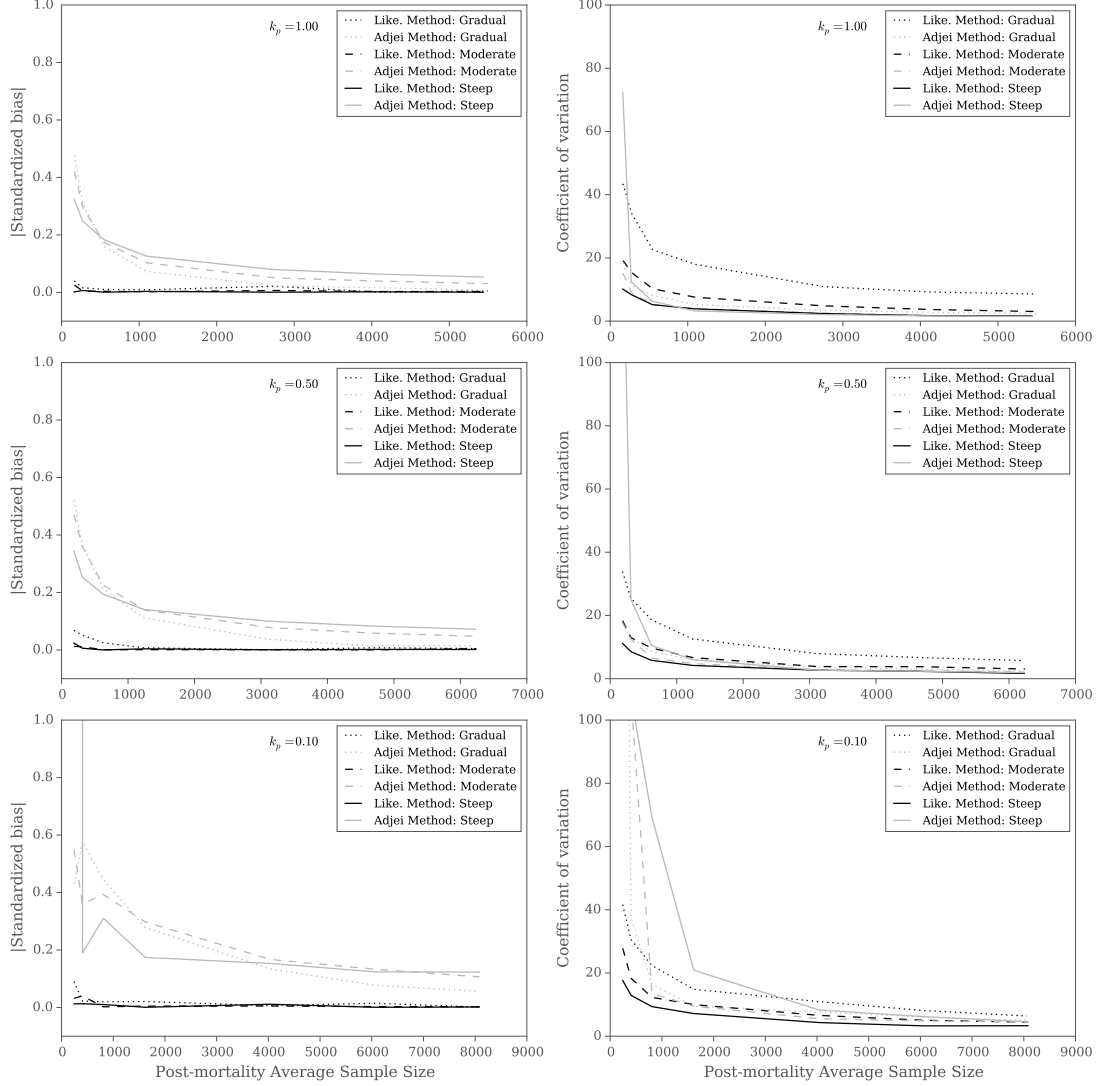
**Figure 3:** The type I error rate and the power of the Likelihood Method (solid line) and the Adjei Method (dashed lines) when  $\mu_p = 100$  for various shapes of the host survival function and levels of aggregation  $k_p$ . The first column gives the type I error rate of each method for falsely detecting PIHM when none is present. The red line gives the the pre-set type I error rate of  $\alpha = 0.05$ . The second column gives the power of a given method to detect PIHM when it is actually occurring.



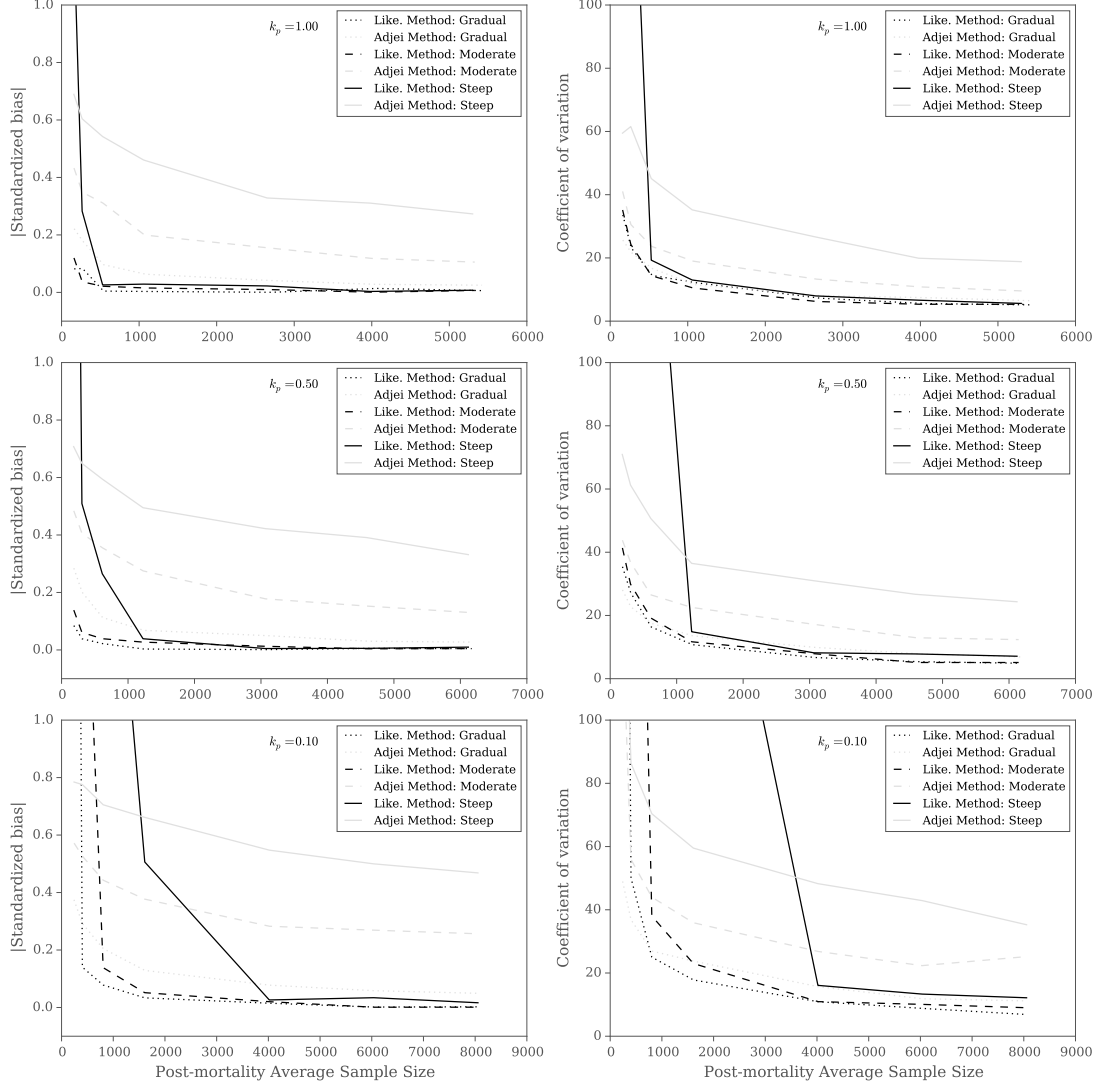
**Figure 4:** The bias and the precision of the Likelihood Method (solid line) and the Adjei Method (dashed lines) when  $\mu_p = 10$  for various shapes of the host survival function and levels of aggregation  $k_p$  when estimating  $LD_{50}$ . The first column gives the bias of each method's  $LD_{50}$  estimate over 150 simulations. The second column gives the precision of each method's  $LD_{50}$  estimate over 150 simulations.



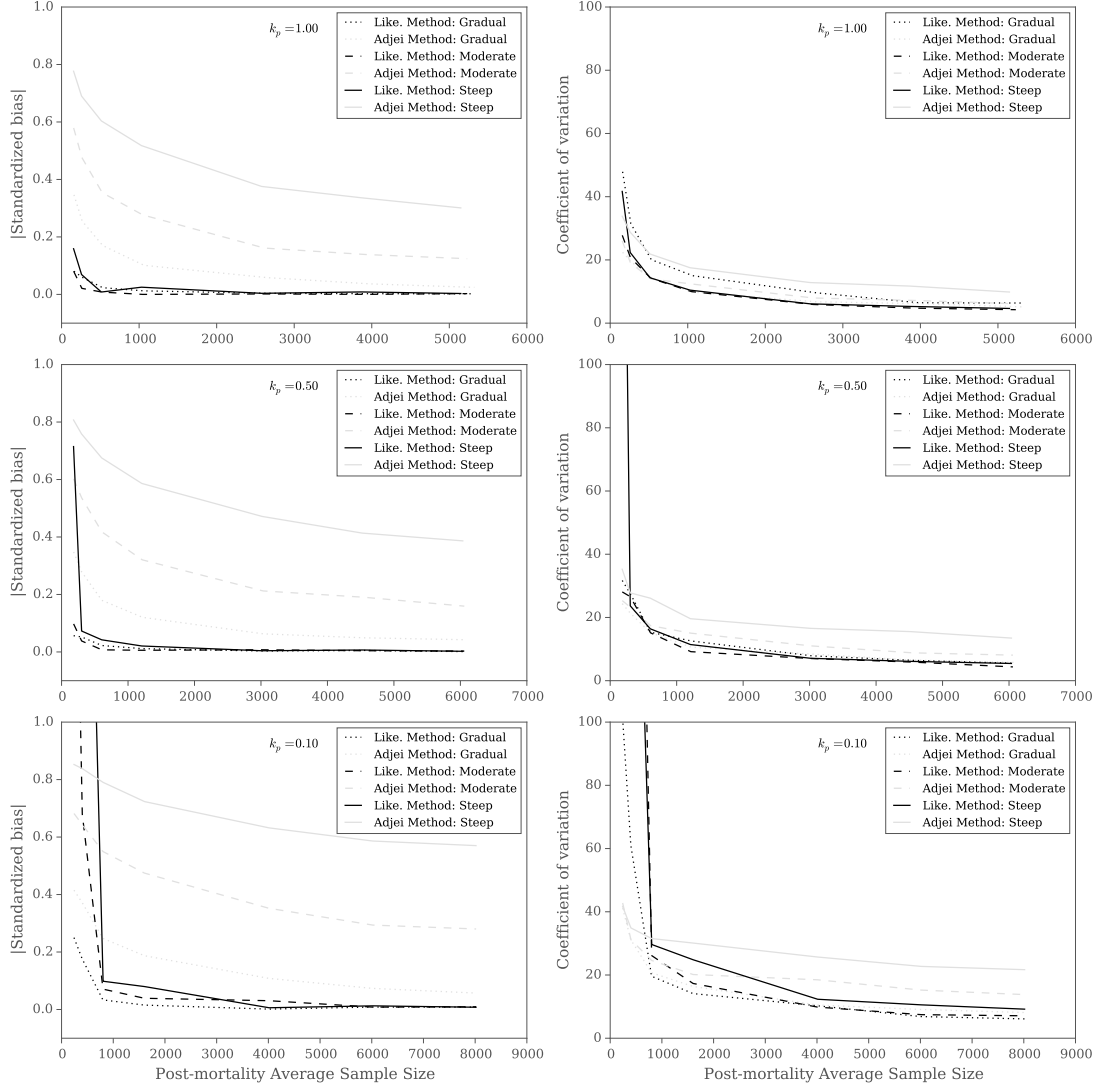
**Figure 5:** The bias and the precision of the Likelihood Method (solid line) and the Adjei Method (dashed lines) when  $\mu_p = 50$  for various shapes of the host survival function and levels of aggregation  $k_p$  when estimating  $LD_{50}$ . The first column gives the bias of each method's  $LD_{50}$  estimate over 150 simulations. The second column gives the precision of each method's  $LD_{50}$  estimate over 150 simulations.



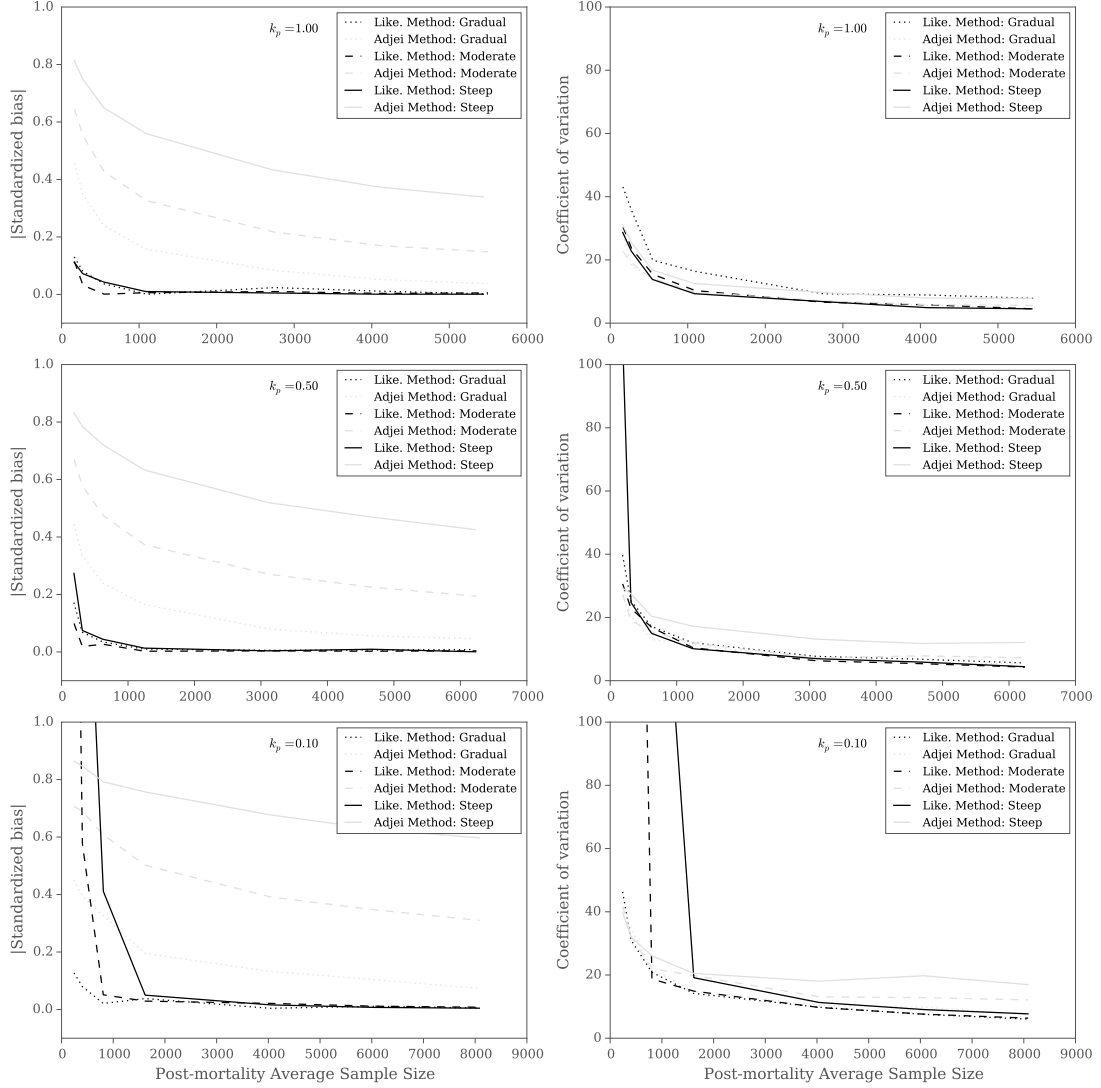
**Figure 6:** The bias and the precision of the Likelihood Method (solid line) and the Adjei Method (dashed lines) when  $\mu_p = 100$  for various shapes of the host survival function and levels of aggregation  $k_p$  when estimating  $LD_{50}$ . The first column gives the bias of each method's  $LD_{50}$  estimate over 150 simulations. The second column gives the precision of each method's  $LD_{50}$  estimate over 150 simulations.



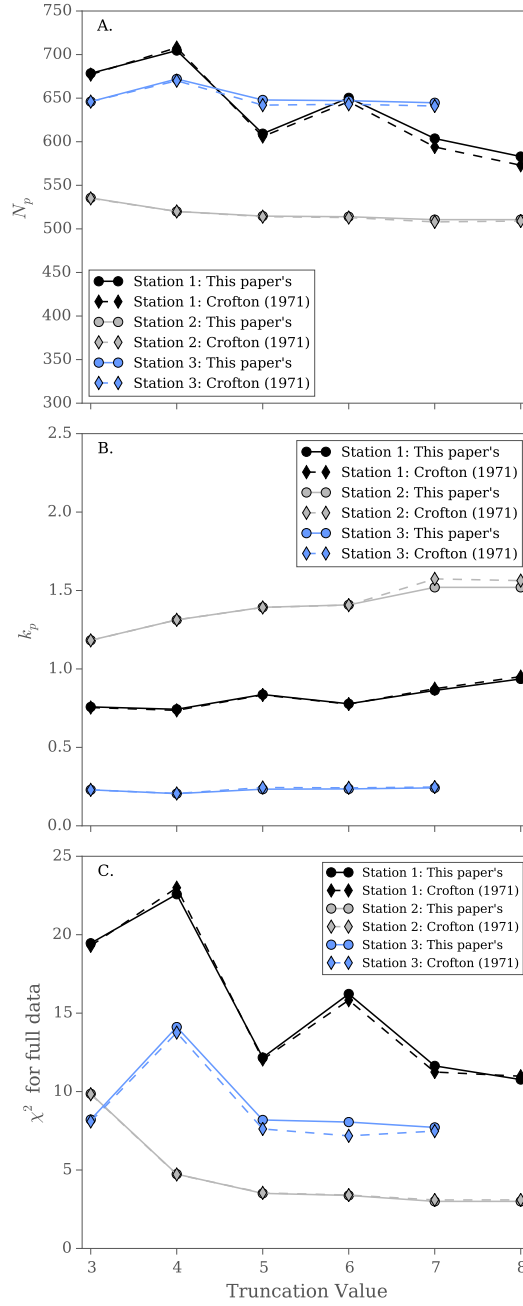
**Figure 7:** The bias and the precision of the Likelihood Method (solid line) and the Adjei Method (dashed lines) when  $\mu_p = 10$  for various shapes of the host survival function and levels of aggregation  $k_p$  when estimating the  $a$  parameter of the host survival function. The first column gives the bias of each method's  $a$  estimate over 150 simulations. The second column gives the precision of each method's  $a$  estimate over 150 simulations.



**Figure 8:** The bias and the precision of the Likelihood Method (solid line) and the Adjei Method (dashed lines) when  $\mu_p = 50$  for various shapes of the host survival function and levels of aggregation  $k_p$  when estimating the  $a$  parameter of the host survival function. The first column gives the bias of each method's  $a$  estimate over 150 simulations. The second column gives the precision of each method's  $a$  estimate over 150 simulations.



**Figure 9:** The bias and the precision of the Likelihood Method (solid line) and the Adjei Method (dashed lines) when  $\mu_p = 100$  for various shapes of the host survival function and levels of aggregation  $k_p$  when estimating the  $a$  parameter of the host survival function. The first column gives the bias of each method's  $a$  estimate over 150 simulations. The second column gives the precision of each method's  $a$  estimate over 150 simulations.



**Figure 10:** A comparison of this paper's implementation (solid line, circles) of the Crofton Method with the results given in Crofton (1971) (dashed line, diamonds). Figure A compares the predicted number of hosts in a population pre-mortality ( $N_p$ ). Figure B compares the predicted parasite aggregation pre-mortality ( $k_p$ ). Figure C compares the  $\chi^2$  statistic for each implementation. Three of the 6 stations fit by Crofton are shown here and all show that our implementation gives very similar results to those given by Crofton.