

Benchmark test with mrgsolve and NONMEM

Metrum Research Group

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1 Introduction

This document runs simulations from a pharmacokinetic model using both NONMEM and mrgsolve and compares the results.

All of the relevant code is presented so that the user can trace how the simulations are performed. The complete source code can be viewed [here](#).

The bottom line results are presented in graphical form [here](#) and numeric form [here](#).

2 Setup

```
Sys.setenv(RSTUDIO_PANDOC = "/usr/lib/rstudio-server/bin/pandoc")
```

```
.libPaths("/data/Rlibs")  
library(mrgsolve)  
library(dplyr)  
library(readr)  
library(ggplot2)  
library(parallel)  
library(purrr)  
library(tidyr)
```

```
carry <- c("cmt", "amt", "ii", "addl", "rate", "evid", "ss")
```

3 Functions

These functions assemble data sets, run simulations, and gather outputs. All scenarios are handled in exactly the same way.

3.1 Save mrgsim output as a nonmem input data set

```
to_data_set <- function(x, id=NULL) {  
  x <- as.data.frame(x)  
  x <- mutate(x, C = '.', DV = '.', cmt = if_else(cmt==0, 2, cmt))  
  x <- dplyr::select(x, "C", everything())  
  if(is.numeric(id)) x <- mutate(x, ID = id)  
  x  
}
```

3.2 Save the nonmem input data set

```
sv <- function(x, file) {  
  write.csv(file = file, row.names = FALSE, quote = FALSE, x)  
}
```

3.3 Run nonmem

```
run <- function(number) {  
  metrumrg::NONR(number, project = "model",  
                 command = "/opt/NONMEM/nm74/nmqual/autolog.pl",  
                 checkrunno=FALSE)  
  return(tabread(number))  
}
```

3.4 Read in nonmem simulation results

```
tabread <- function(number) {  
  tab <- file.path("model", number, "TAB")  
  if(file.exists(tab)) return(read_table(tab, skip=1))  
  stop("the run failed")  
}
```

3.5 Simulate a scenario with mrgsim

```
sim <- function(x, e,...) {  
  mrgsim(x, events = e, carry.out = carry, digits = 5, recsort=3, ...)  
}
```

4 The mrgsim model

```
code <- '  
$SET req = ""  
$PARAM CL = 1.1, V = 20, KA = 1.5  
LAGT = 0, MODE = 0, DUR2 = 2, RAT2 = 10, BIOAV = 1  
  
$PKMODEL cmt = "GUT CENT", depot = TRUE  
  
$MAIN  
  
F_CENT = BIOAV;  
ALAG_CENT = LAGT;  
  
if(MODE==1) R_CENT = RAT2;  
if(MODE==2) D_CENT = DUR2;  
  
$TABLE  
capture DV = (CENT/(V/1000));  
capture CP = DV;  
  
$CAPTURE LAGT MODE DUR2 RAT2 BIOAV  
'
```

```
mod <- mcode_cache("tests1", code)
```

```
. Building tests1 ... done.
```

```
mod <- update(mod, end=130, delta = 1)
```

5 Assemble the scenarios

There is a lot of code here. See the results section to see input data objects next to simulated data output from mrgsolve and NONMEM.

- Doses into cmt 2 are intravascular and doses into cmt 1 are extravascular
- LAGT sets the dosing lag time
- BIOAV sets the bioavailability fraction

```
env <- new.env()
env$ev <- list()
env$descr <- list()
push_back <- function(env, ev, descr) {
  n <- length(env$ev)+1
  m <- length(env$descr)+1
  env$ev[[n]] <- ev
  env$descr[[m]] <- descr
}
```

```
ev <- ev(amt = 100, ii = 24, addl = 3)
push_back(env, ev, "Bolus with additional")
```

```
ev <- ev(amt = 100, ii = 24, addl = 3, LAGT = 12.13, BIOAV = 2.23, cmt = 2)
push_back(env, ev, "Bolus with lag time and bioav")
```

```
ev <- ev(amt = 100, ii = 24, addl = 3, rate = 100/10, cmt = 2)
push_back(env, ev, "Infusion with additional")
```

```
ev <- ev(amt = 100, ii = 24, addl = 3, rate = 100/12, cmt = 1)
push_back(env, ev, "Infusion doses to depot, with additional")
```

```
ev <- ev(amt = 100, ii = 24, addl=3, rate = 100/10, LAGT = 4.15, cmt = 2)
push_back(env, ev, "Infusion doses, with additional and lag time")
```

```
ev <- ev(amt = 100, ii = 24, addl = 3, rate = 100/10, LAGT = 3.25, BIOAV = 0.412, cmt = 2)
push_back(env, ev, "Infusion doses, with lag time and bioav factor")
```

```
ev <- ev(amt = 100, ii = 24, addl = 3, rate = 100/10, LAGT = 3.16, BIOAV = 0.412, ss = 1, cmt = 2)
push_back(env, ev, "Infusion doses, with lag time and bioav factor")
```

```
ev <- ev(amt = 100, ii = 12, addl = 4, rate = 100/50, BIOAV = 0.812, ss = 1, cmt = 2)
push_back(env, ev, "Infusion doses at steady-state, with lag time and bioav factor")
```

```
ev <- ev(amt = 100, ii = 12, addl = 3, rate = 100/50, ss = 1, cmt = 2)
push_back(env, ev, "Infusion doses, with lag time and bioav factor")
```

```
ev <- ev(amt = 100, ii = 6, addl = 12, rate = signif(100/12,5), ss = 1, cmt = 2)
push_back(env, ev, "Infusion doses at steady state, II < DUR, no bioav factor")
```

```

ev <- ev(amt = 100, ii = 10, addl = 8, rate = 0.412*100/10, BIOAV = 0.412, ss = 1, cmt = 2)
push_back(env, ev, "Infusion doses at steady state where II == DUR, with bioav factor")

ev <- ev(amt = 100, ii = 10, addl = 8, rate = 100/10, ss = 1, cmt = 2)
push_back(env, ev, "Infusion doses at steady state, where II == DUR")

ev <- ev(amt = 100, ii = 24, addl = 3, LAGT = 4, BIOAV = 0.412, ss = 1, cmt = 2)
push_back(env, ev, "Bolus doses at steady state, with bioav factor and lag time")

ev <- ev(amt = 100, ii = 24, addl = 3, LAGT = 5, BIOAV = 0.412, cmt = 2)
push_back(env, ev, "Bolus doses with lag time and bioavailability factor")

ev <-
  ev(amt = 100, cmt = 2, LAGT = 1) +
  ev(time = 13, amt = 50, ii = 24, addl = 2, rate = 24)
push_back(env, ev, "Bolus then infusion")

ev <- ev(amt = 100, rate = -2, DUR2 = 9, MODE = 2, cmt = 2, ii = 24, addl = 3, LAGT = 5, BIOAV = 0.61)
push_back(env, ev, "Infusion with modeled duration, lag time, and bioav factor")

ev <- ev(amt = 100, rate = -2, DUR2 = 9, MODE = 2, cmt = 2, ii = 24, addl = 3, ss = 1, BIOAV = 0.61)
push_back(env, ev, "Infusion with modeled duration, at steady state with bioav factor")

ev <-
  ev(amt = 100, ii = 12, addl = 2, rate = 50, BIOAV = 0.61) +
  ev(amt = 120, evid = 4, time = 50, BIOAV = 0.5, ii = 12, addl = 3)
push_back(env, ev, "Reset and dose (EVID 4) with additional")

ev <-
  ev(amt = 100, ii = 12, addl = 3, rate = 50, BIOAV = 0.61) +
  ev(amt = 0, evid = 3, time = 50, cmt = 2, BIOAV=1) +
  ev(amt = 120, ii = 16, addl = 2, time = 54, BIOAV=1)
push_back(env, ev, "Reset (EVID 3) with additional")

ev <-
  ev(amt = 100, ii = 24, addl = 3, ss = 1) +
  ev(amt = 50, ii = 24, addl = 3, ss = 2, time = 12)
push_back(env, ev, "Steady state 1 and 2")

ev <- ev(amt = 0, rate = 100, ss=1)
push_back(env, ev, "Steady state infusion")

update_id <- function(ev, id) mutate(ev, ID = id)

runs <- tibble(ev = env$ev, descr = env$descr)
runs <- mutate(runs, ID = seq(n()))
runs <- mutate(runs, ev = map2(ev, ID, update_id))
runs <- mutate(runs, sims = mclapply(ev, sim, x = mod))

runs <- mutate(runs, data = map(sims, to_data_set))

data <- runs[["data"]] %>% bind_rows()

sv(data, "data/1001.csv")

```

6 Simulate with nonmem

```
out <- run(1001)

. Run 1001 complete.
. NONR complete.
. Parsed with column specification:
. cols(
.   ID = col_double(),
.   TIME = col_double(),
.   CP = col_double()
. )
```

7 Numeric Summary

Look at the difference between simulated values from mrgsolve and NONMEM.

```
runs <- mutate(runs, out = split(out,out$ID))

runs <- mutate(
  runs,
  comp = map2(out,sims, .f=function(out,sims) {
    tibble(
      ID = out$ID,
      time = sims$time,
      MRGSIM = sims$CP,
      NONMEM = out$CP,
      diff = MRGSIM-NONMEM)
  })
)

comp <- runs %>% select(ID,comp) %>% unnest()
```

7.1 Overall

This is the nonmem minus mrgsim summary

```
summary(comp$diff)
```

```
.   Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
.      0      0      0      0      0      0
```

7.2 Summary by scenario number

diff is the simulated CP from nonmem minus the simulated CP from mrgsim

```
group_by(comp,ID) %>% summarise(mean = mean(diff), max = max(diff), min = min(diff)) %>%
  as.data.frame
```

```

.      ID mean max min
. 1    1    0  0  0
. 2    2    0  0  0
. 3    3    0  0  0
. 4    4    0  0  0
. 5    5    0  0  0
. 6    6    0  0  0
. 7    7    0  0  0
. 8    8    0  0  0
. 9    9    0  0  0
. 10   10    0  0  0
. 11   11    0  0  0
. 12   12    0  0  0
. 13   13    0  0  0
. 14   14    0  0  0
. 15   15    0  0  0
. 16   16    0  0  0
. 17   17    0  0  0
. 18   18    0  0  0
. 19   19    0  0  0
. 20   20    0  0  0
. 21   21    0  0  0

```

```

comp_plot <- function(comp) {
  id <- comp$ID[1]
  ggplot(data = comp) +
    ggtitle(label=NULL,subtitle=paste0("ID: ", id, "; line: mrgsolve, point: NONMEM")) +
    geom_point(aes(time,NONMEM),color = "firebrick") +
    geom_line(aes(time,MRGSIM,group = ID)) +
    theme_bw() + ylab("Simulated value") + xlab("Time") +
    scale_x_continuous(breaks = seq(0,130,24))
}

runs <- mutate(runs, plot = map(comp, comp_plot))

```

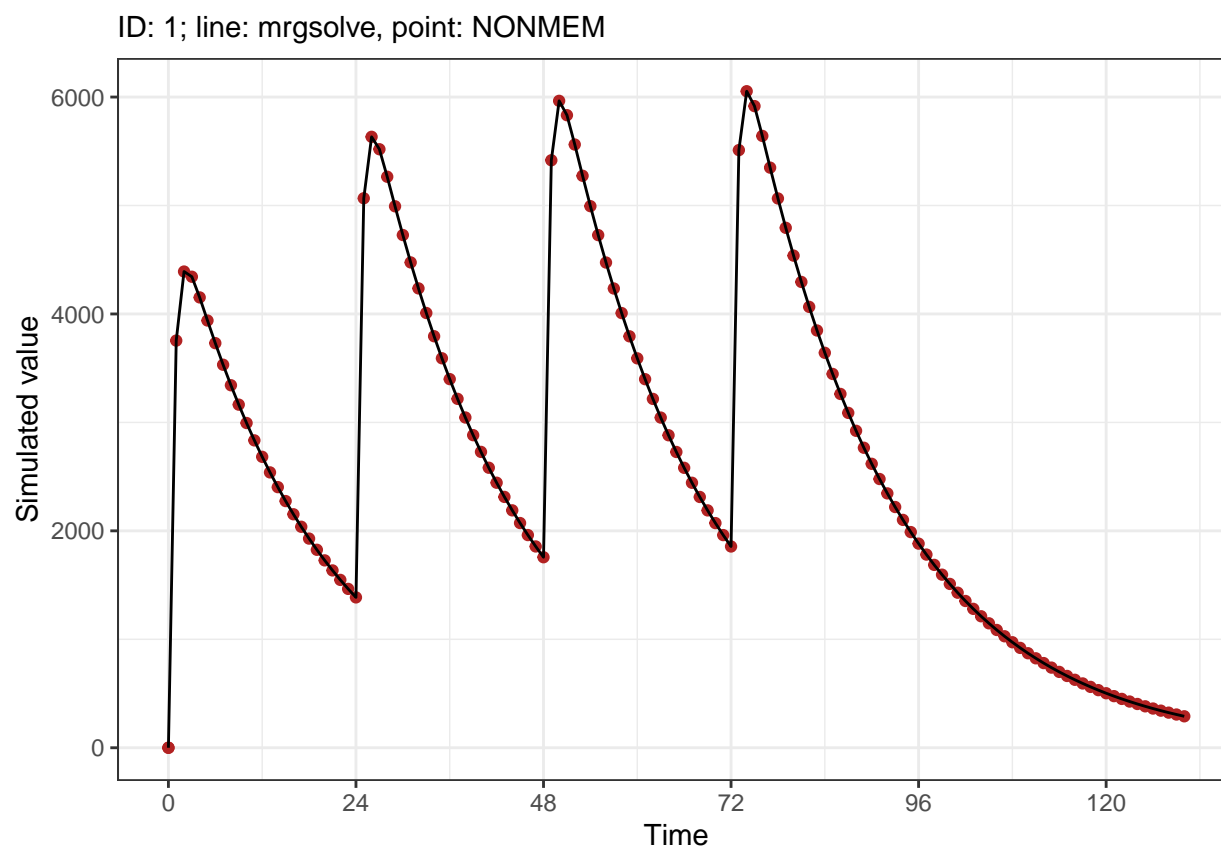
8 Results

8.1 1: Bolus with additional

```

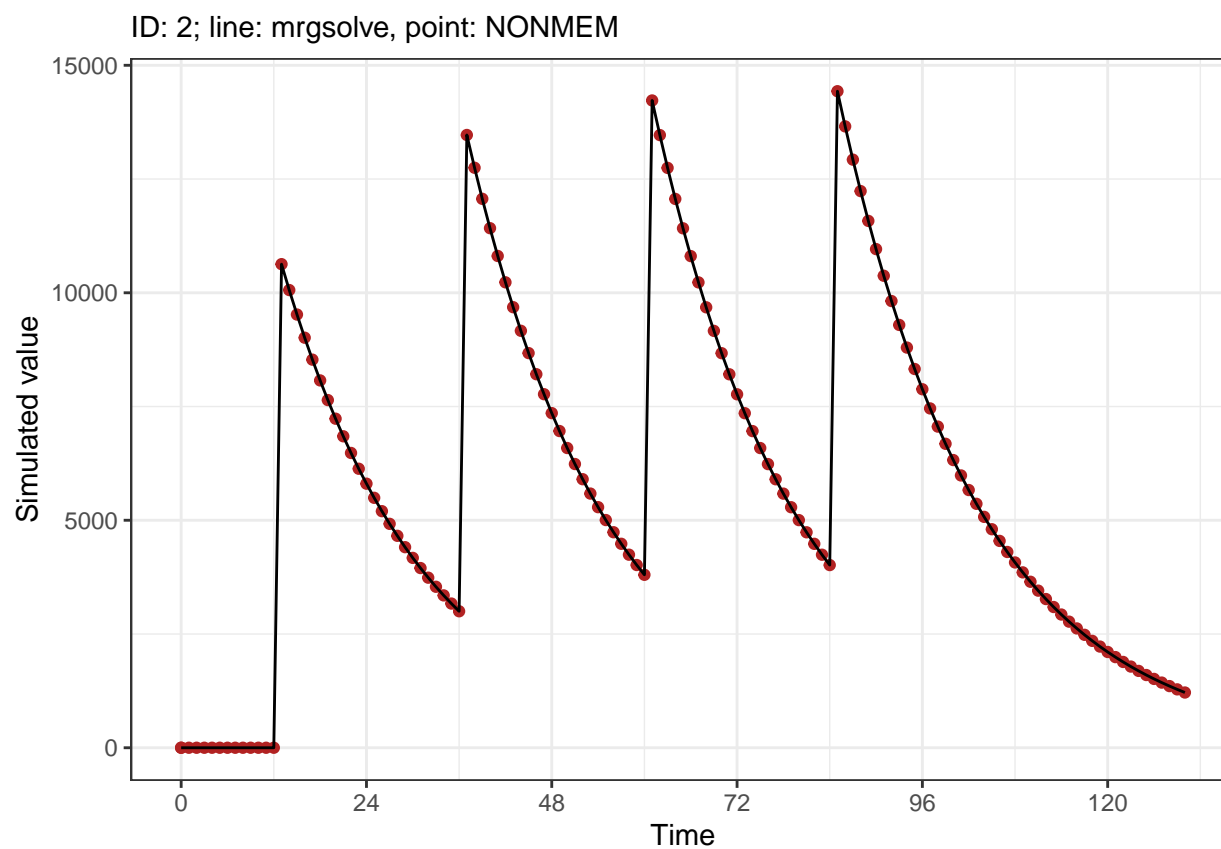
. $ev
. Events:
.   ID time amt ii addl cmt evid
. 1 1    0 100 24   3   1    1
.
. $plot

```



8.2 2: Bolus with lag time and bioav

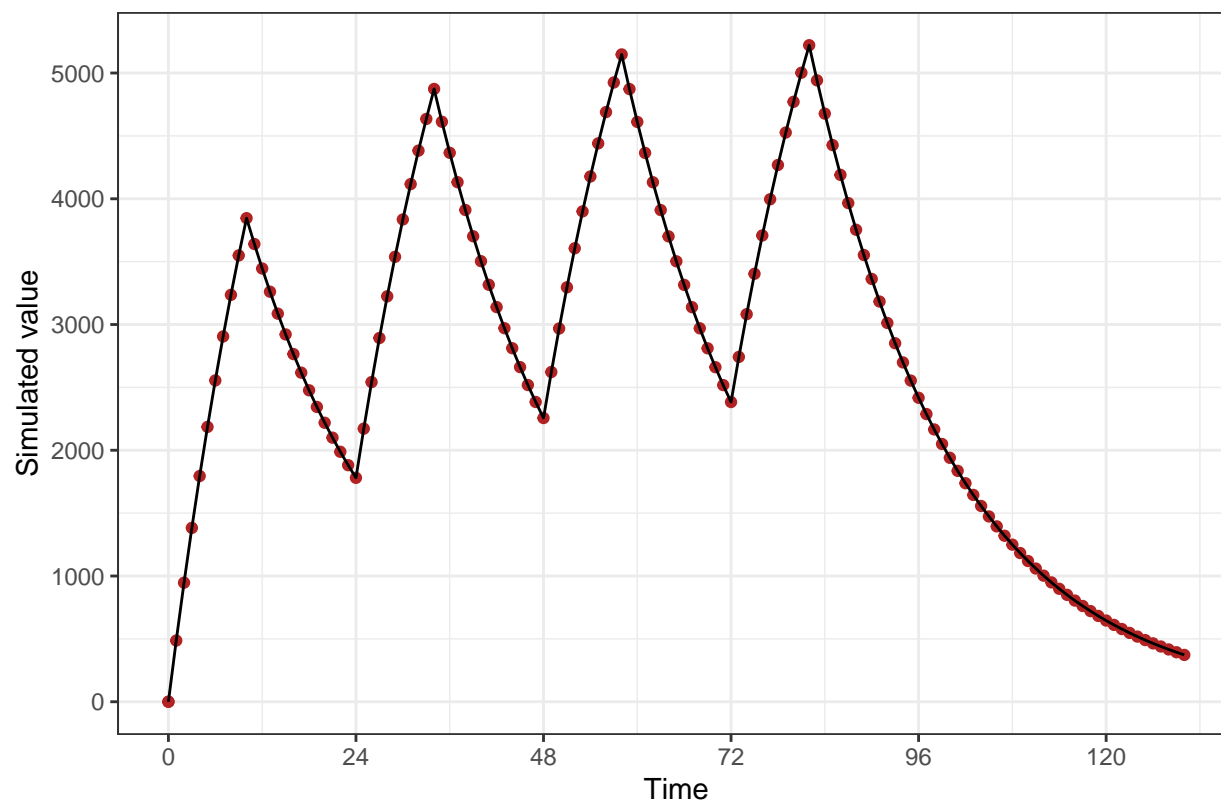
```
. $ev
. Events:
.   ID time amt ii addl cmt evid  LAGT BIOAV
. 1 2 0 100 24 3 2 1 12.13 2.23
.
. $plot
```

8.3 3: Infusion with additional

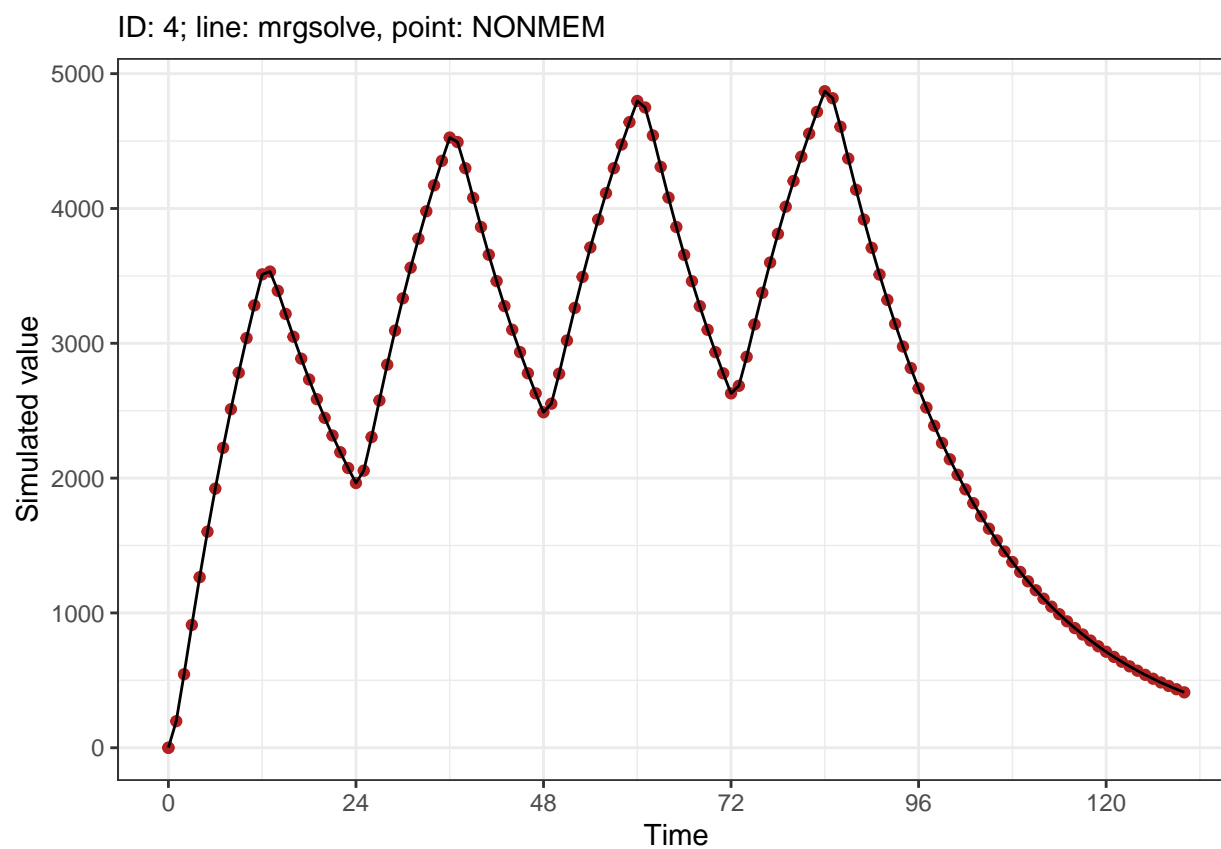
```
. $ev
. Events:
.   ID time amt rate ii addl cmt evid
. 1 3 0 100 10 24 3 2 1
.
. $plot
```

ID: 3; line: mrgsolve, point: NONMEM



8.4 4: Infusion doses to depot, with additional

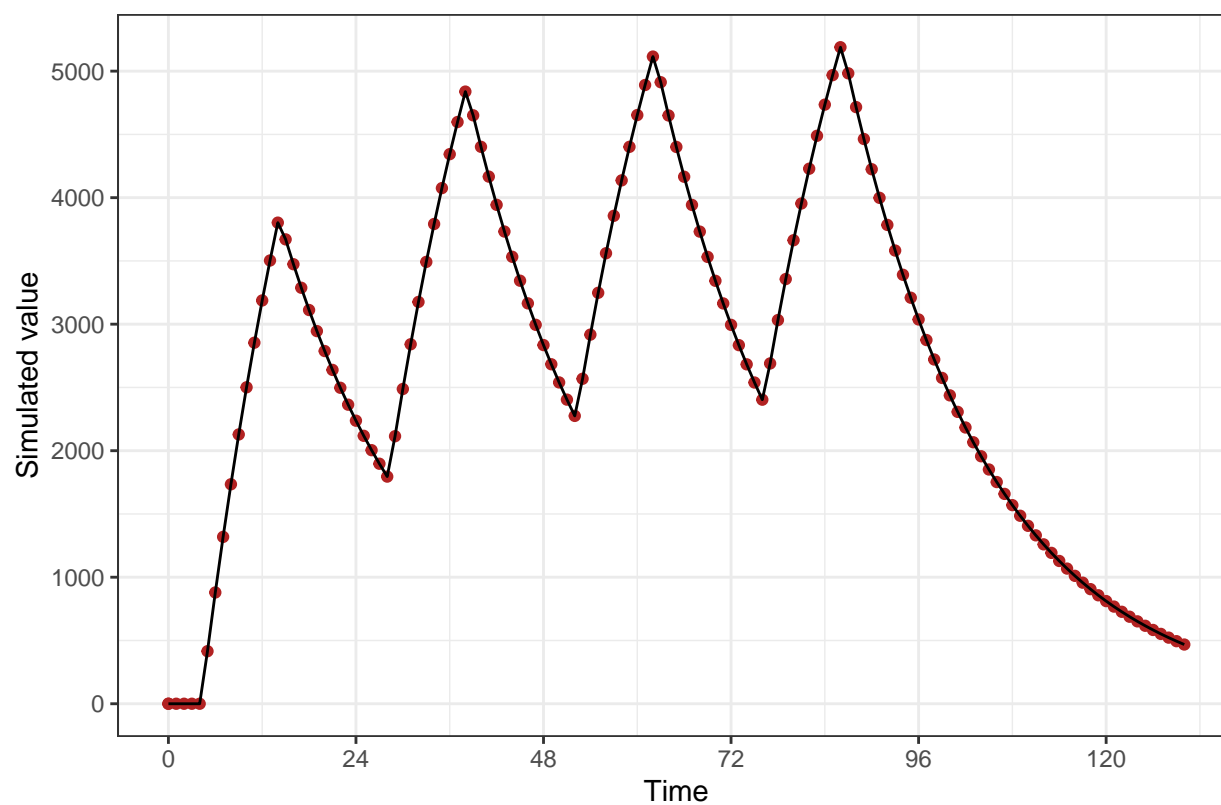
```
. $ev
. Events:
.   ID time amt      rate ii addl cmt evid
.   1  4   0 100 8.333333 24   3   1   1
.
. $plot
```



8.5 5: Infusion doses, with additional and lag time

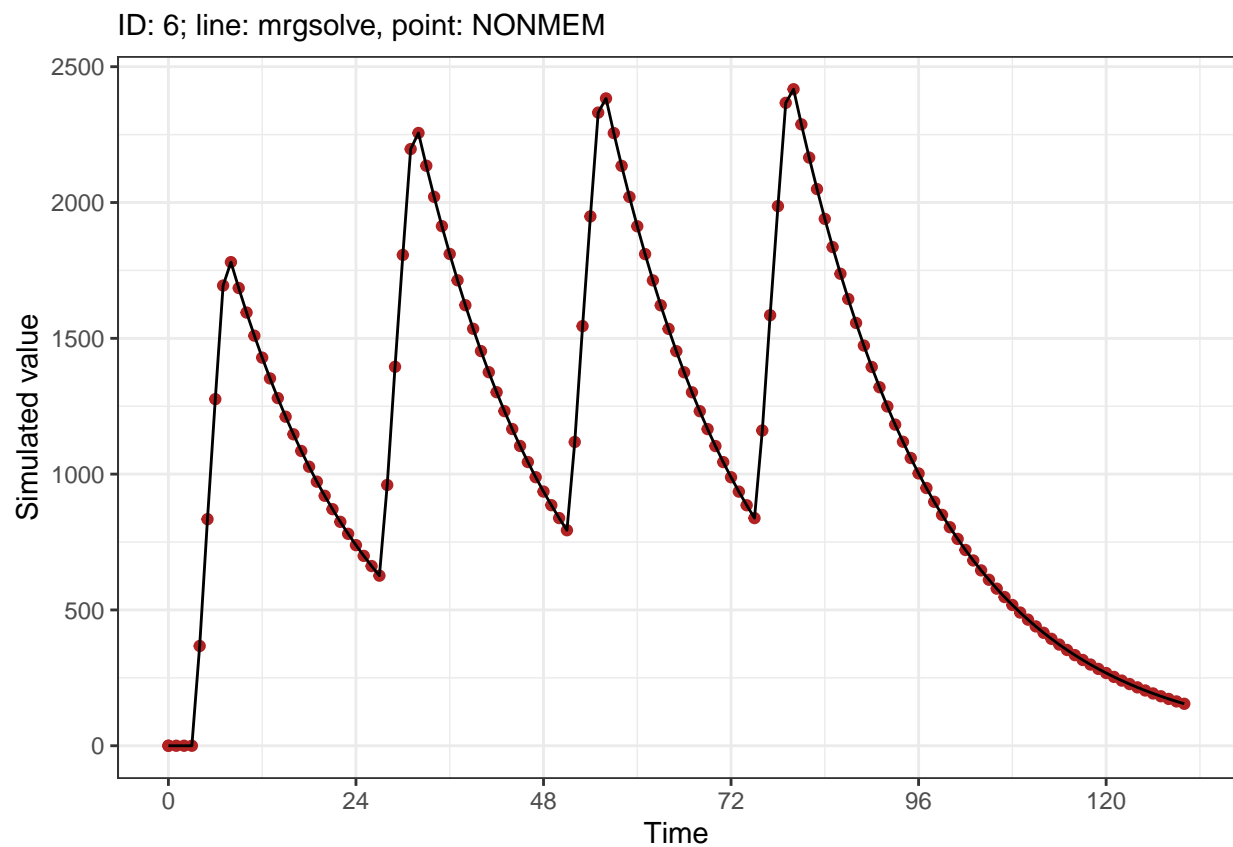
```
. $ev
. Events:
.   ID time amt rate ii addl cmt evid LAGT
. 1 5 0 100 10 24 3 2 1 4.15
.
. $plot
```

ID: 5; line: mrgsolve, point: NONMEM



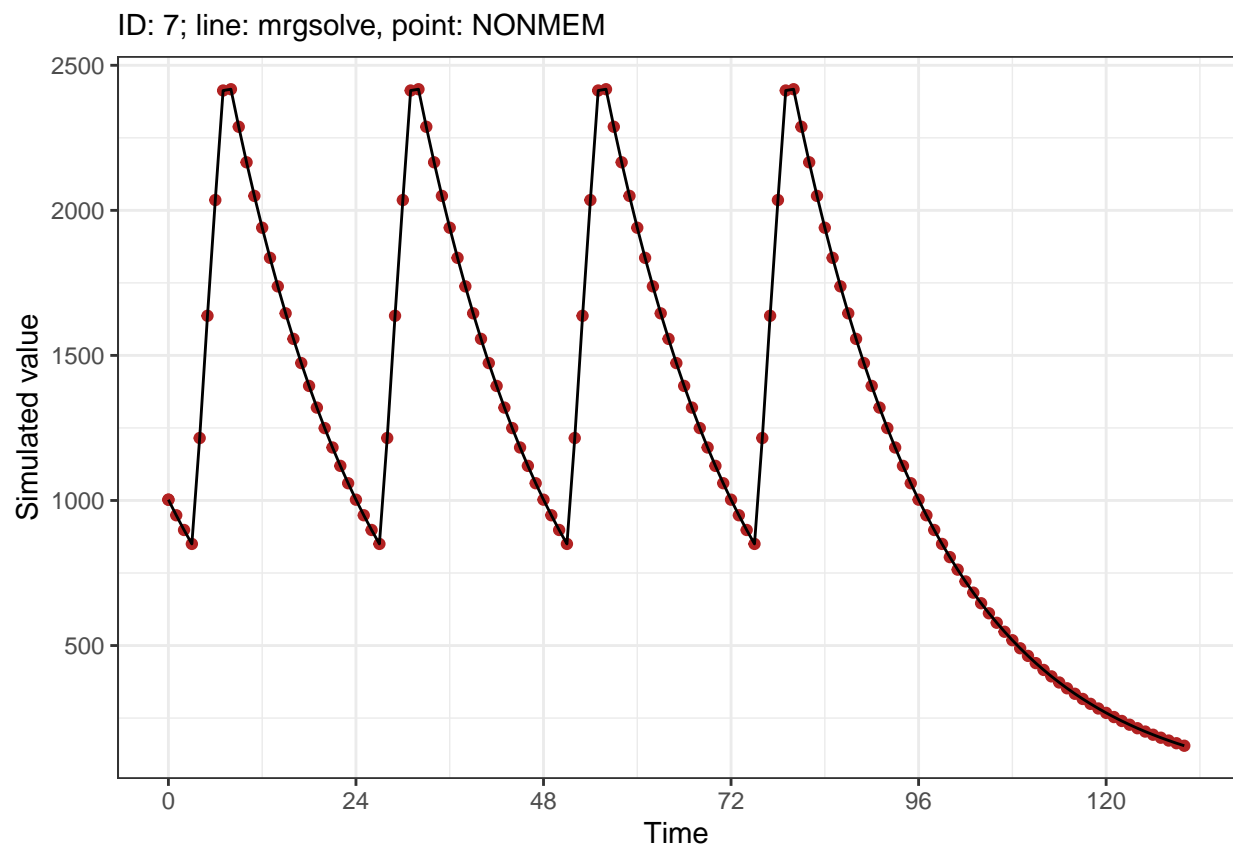
8.6 6: Infusion doses, with lag time and bioav factor

```
. $ev
. Events:
.   ID time amt rate ii addl cmt evid LAGT BIOAV
. 1 6 0 100 10 24 3 2 1 3.25 0.412
.
. $plot
```



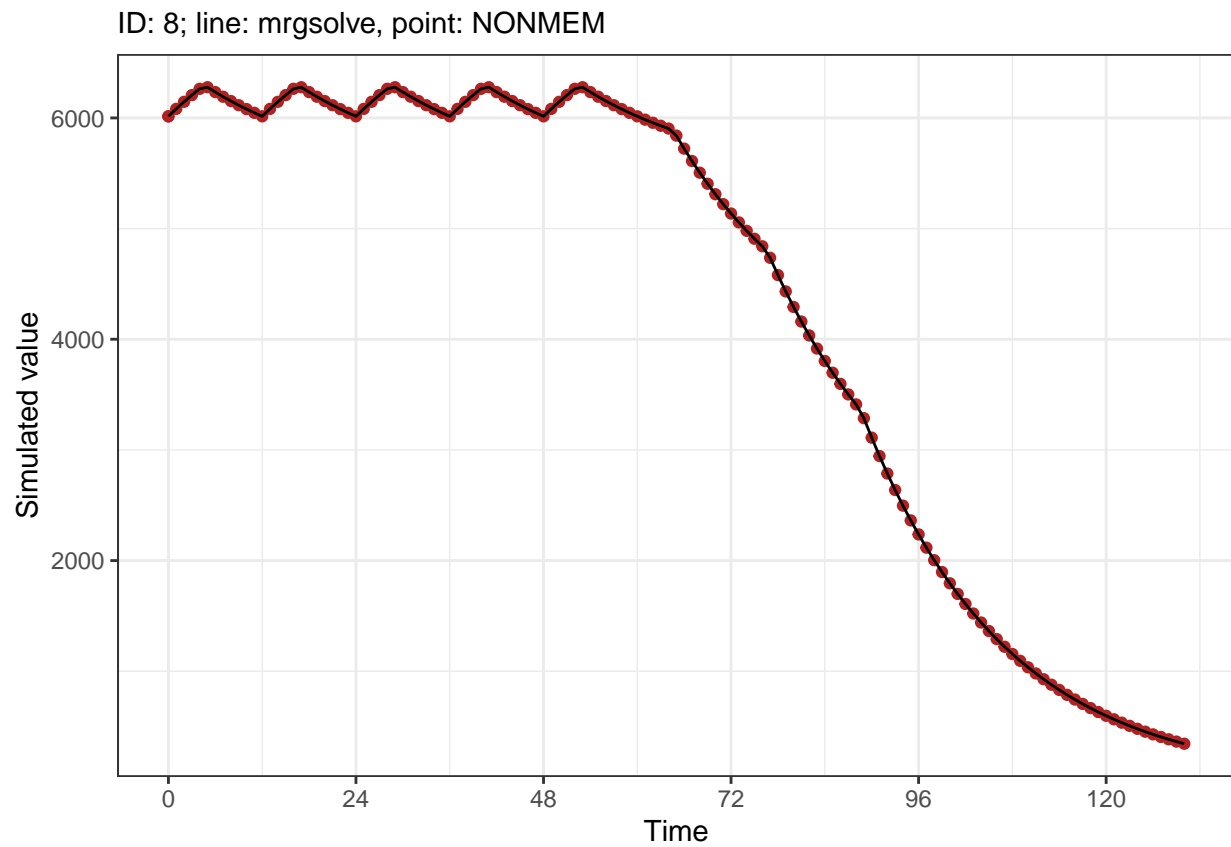
8.7 7: Infusion doses, with lag time and bioav factor

```
. $ev
. Events:
.   ID time amt rate ii addl cmt evid ss LAGT BIOAV
. 1 7 0 100 10 24 3 2 1 1 3.16 0.412
.
. $plot
```



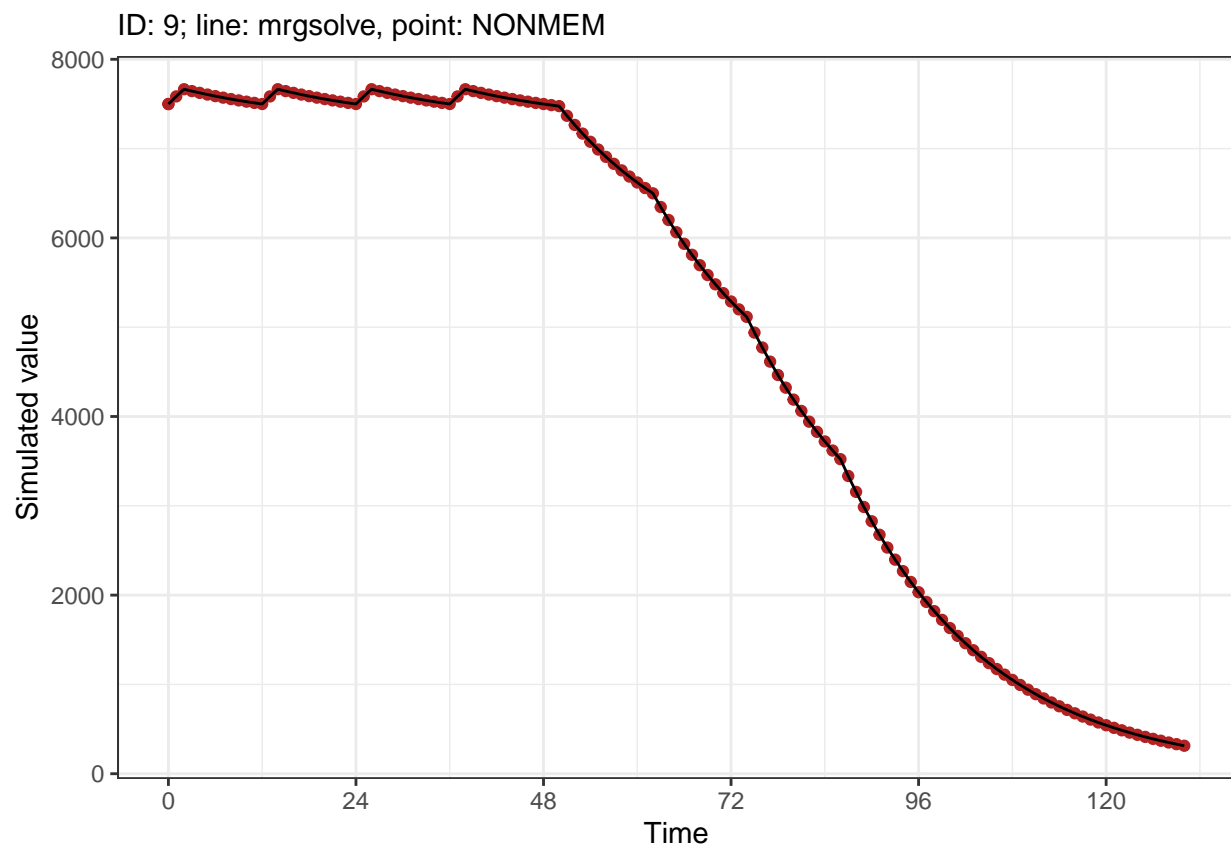
8.8 8: Infusion doses at steady-state, with lag time and bioav factor

```
. $ev
. Events:
.   ID time amt rate ii addl cmt evid ss BIOAV
. 1 8 0 100 2 12 4 2 1 1 0.812
.
. $plot
```



8.9 9: Infusion doses, with lag time and bioav factor

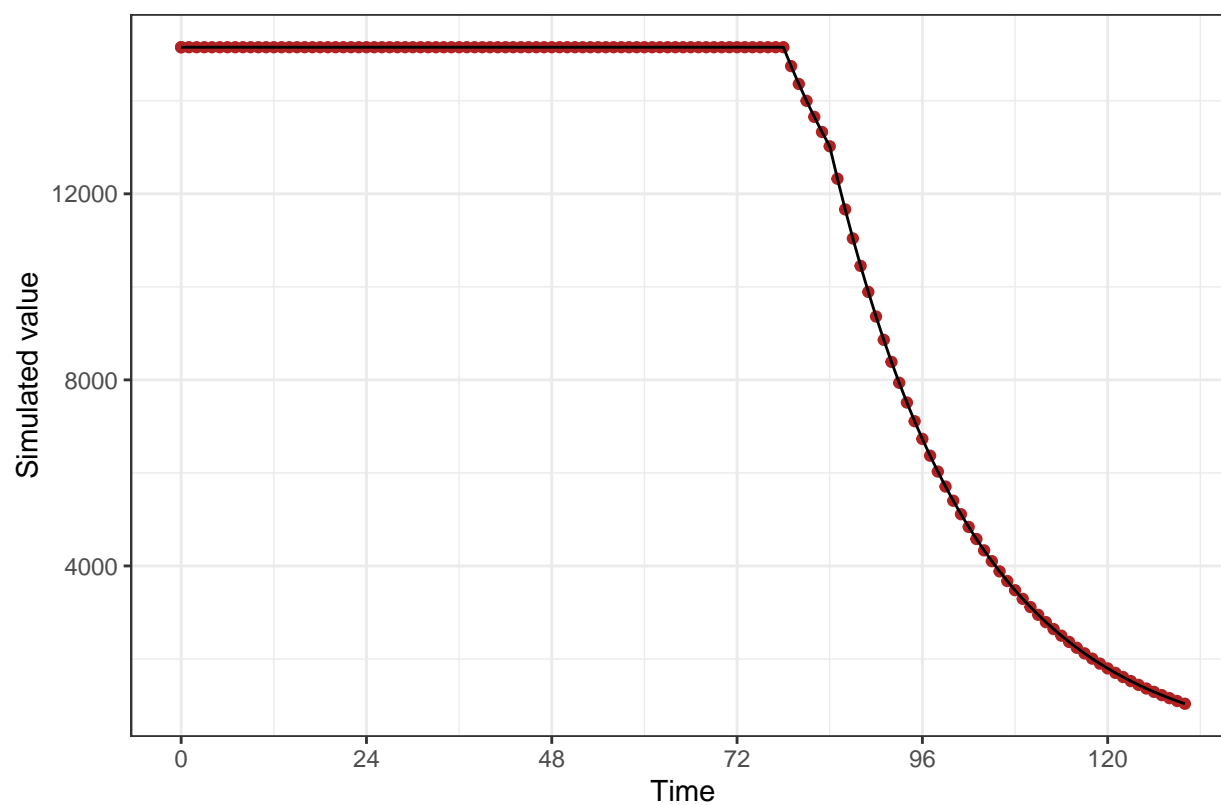
```
. $ev
. Events:
.   ID time amt rate ii addl cmt evid ss
. 1 9 0 100 2 12 3 2 1 1
.
. $plot
```



8.10 10: Infusion doses at steady state, $II < DUR$, no bioav factor

```
. $ev
. Events:
.   ID time amt   rate ii addl cmt evid ss
.   1 10   0 100 8.3333 6 12 2 1 1
.
. $plot
```

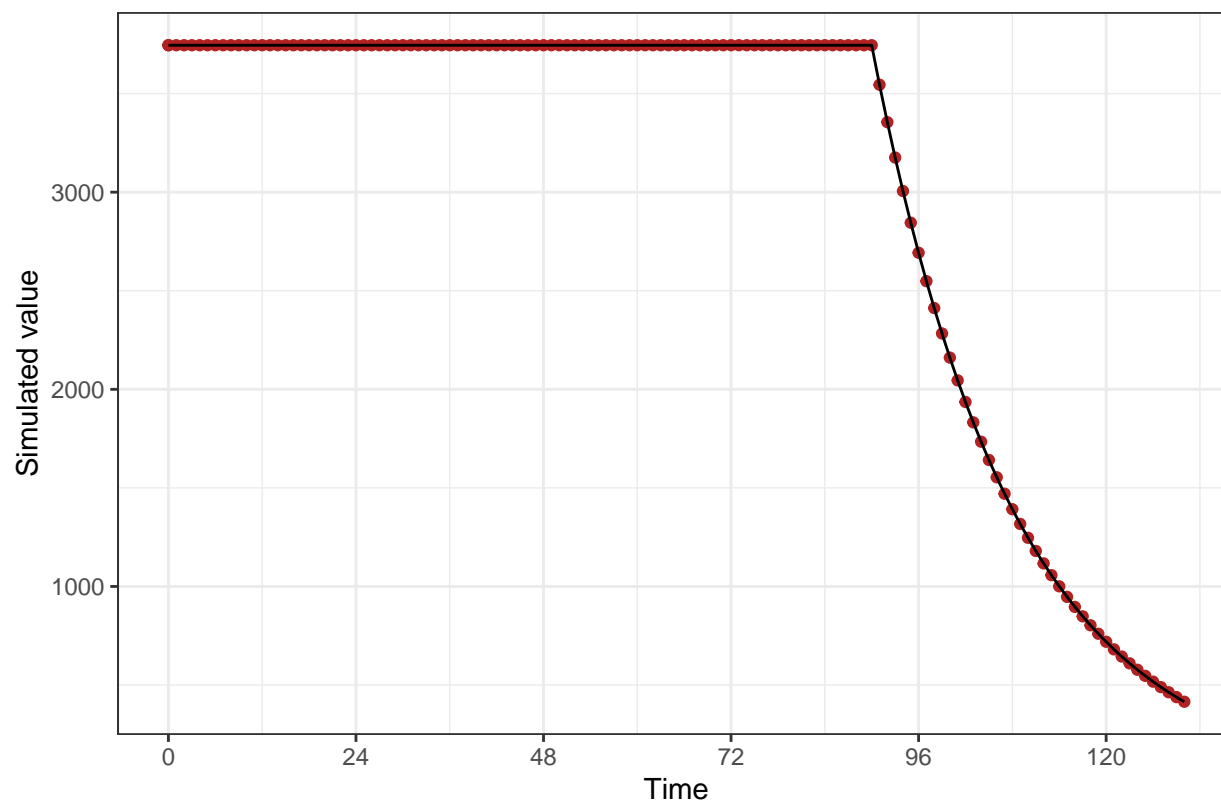

ID: 10; line: mrgsolve, point: NONMEM



8.11 11: Infusion doses at steady state where $II == DUR$, with bioav factor

```
. $ev
. Events:
.   ID time amt rate ii addl cmt evid ss BIOAV
. 1 11    0 100 4.12 10    8   2    1  1 0.412
.
. $plot
```

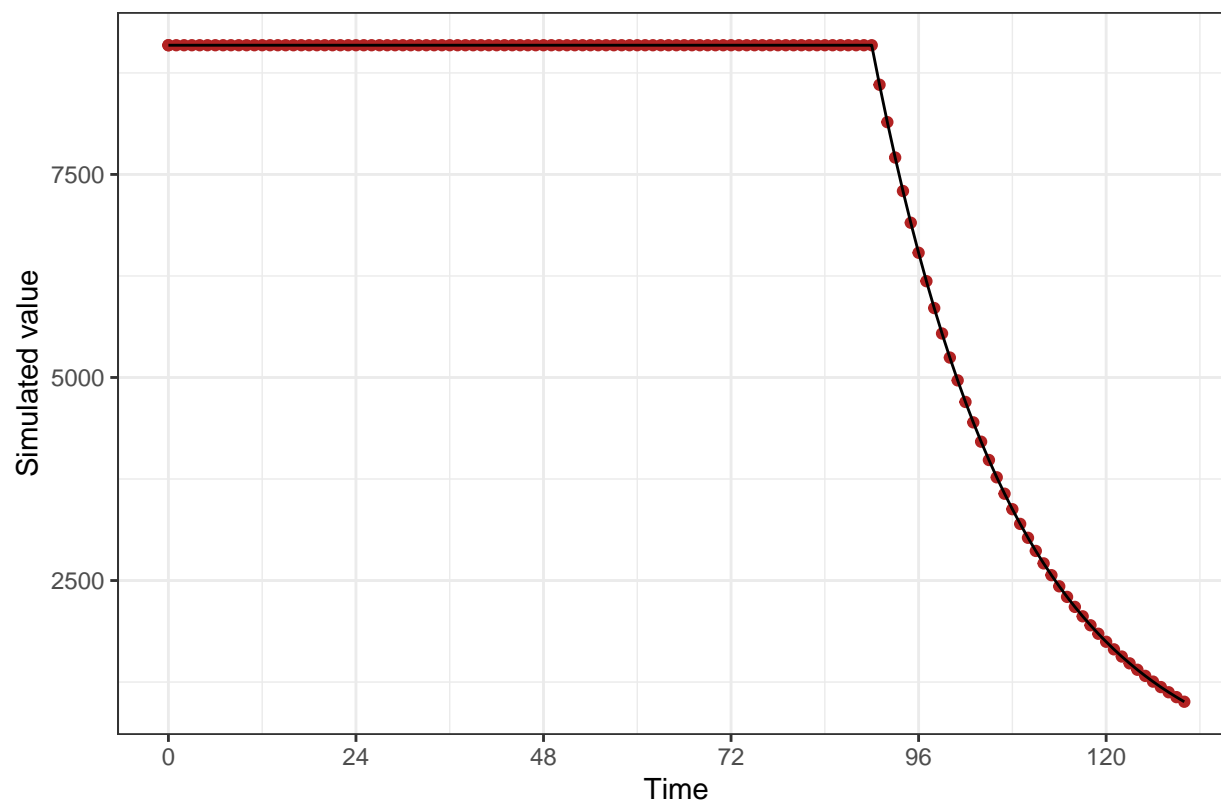
ID: 11; line: mrgsolve, point: NONMEM



8.12 12: Infusion doses at steady state, where $II == DUR$

```
. $ev
. Events:
.   ID time amt rate ii addl cmt evid ss
. 1 12 0 100 10 10 8 2 1 1
.
. $plot
```

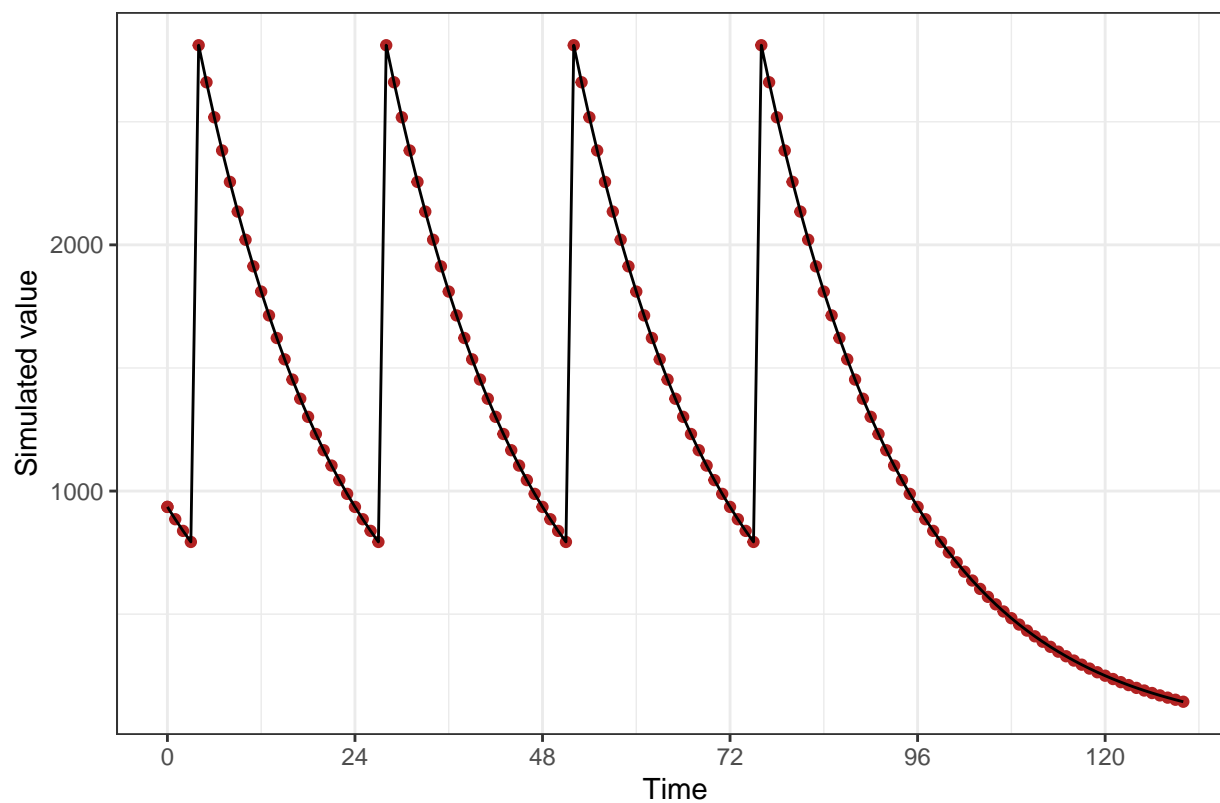
ID: 12; line: mrgsolve, point: NONMEM



8.13 13: Bolus doses at steady state, with bioav factor and lag time

```
. $ev
. Events:
.   ID time amt ii addl cmt evid ss LAGT BIOAV
. 1 13   0 100 24   3   2   1   1   4 0.412
.
. $plot
```

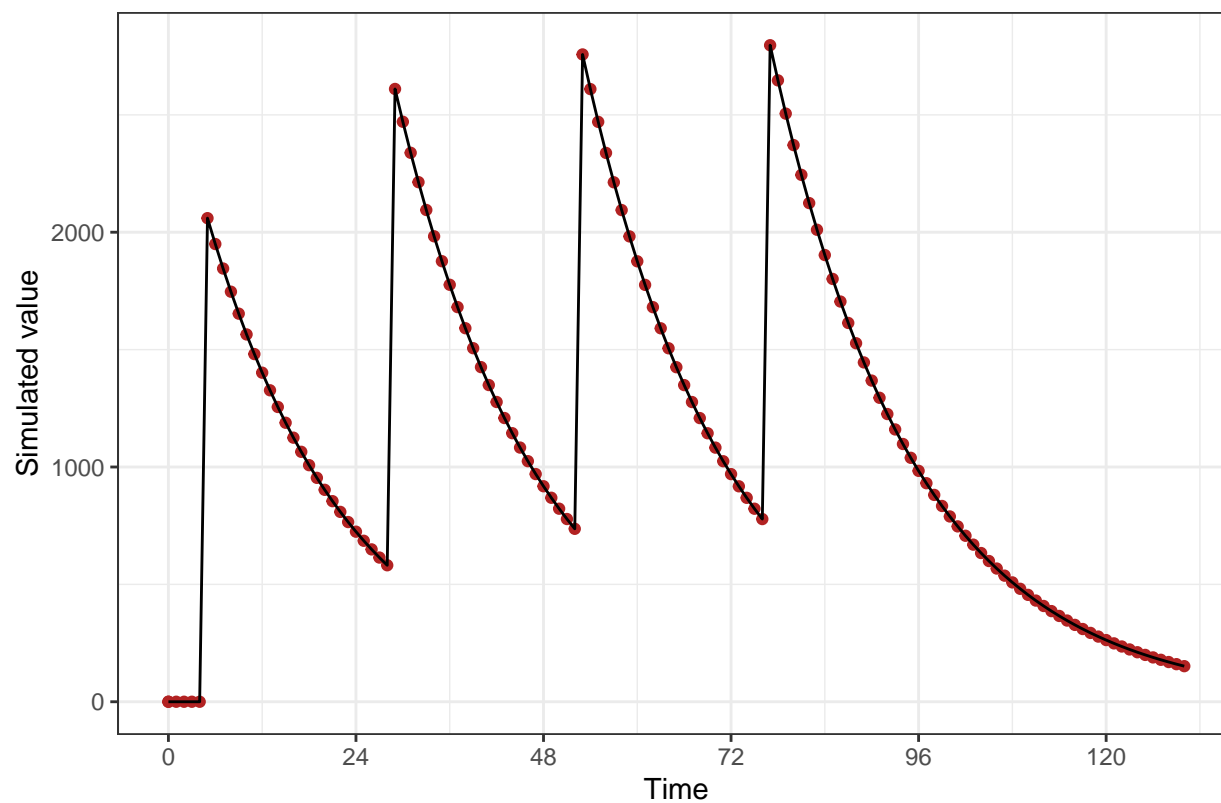
ID: 13; line: mrgsolve, point: NONMEM



8.14 14: Bolus doses with lag time and bioavailability factor

```
. $ev
. Events:
.   ID time amt ii addl cmt evid LAGT BIOAV
. 1 14   0 100 24   3   2   1   5 0.412
.
. $plot
```

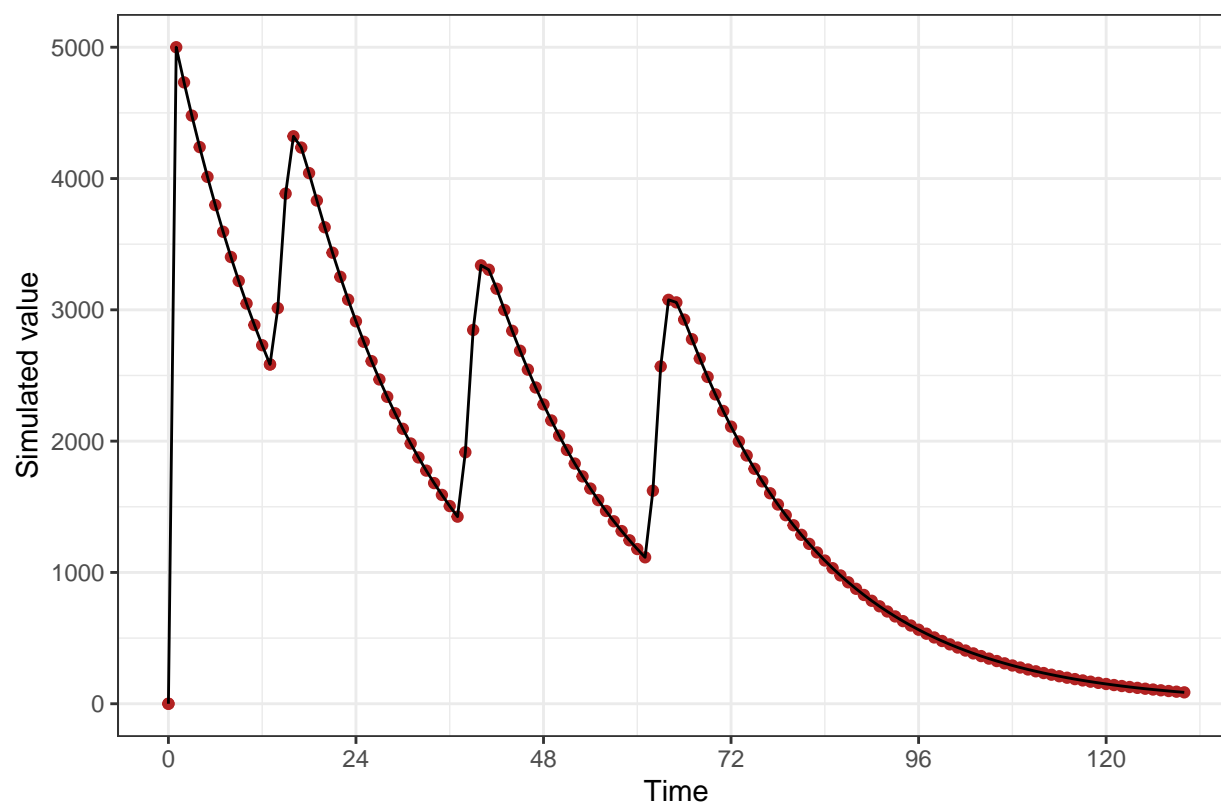
ID: 14; line: mrgsolve, point: NONMEM



8.15 15: Bolus then infusion

```
. $ev
. Events:
.   ID time amt rate ii addl cmt evid LAGT
. 1 15    0 100    0 0    0 2    1    1
. 2 15   13  50   24 24    2 1    1    0
.
. $plot
```

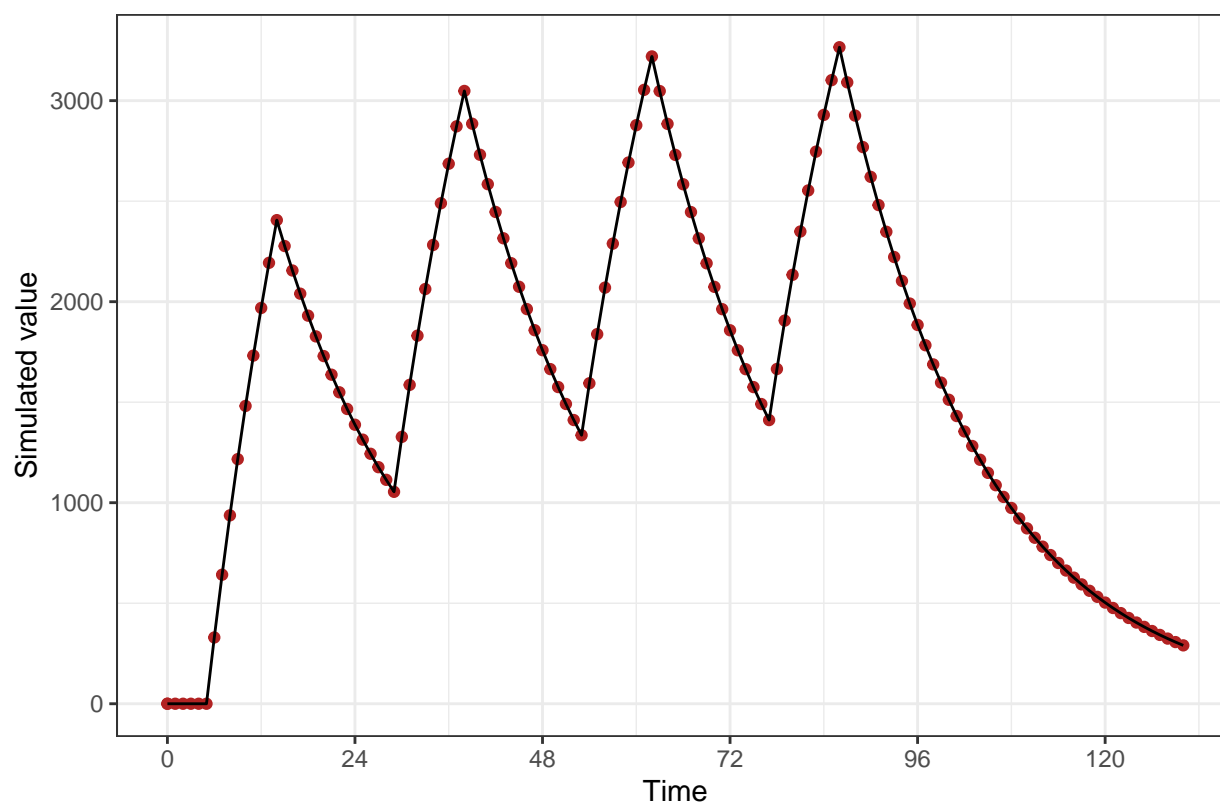
ID: 15; line: mrgsolve, point: NONMEM



8.16 16: Infusion with modeled duration, lag time, and bioav factor

```
. $ev
. Events:
.   ID time amt rate ii addl cmt evid DUR2 MODE LAGT BIOAV
. 1 16 0 100 -2 24 3 2 1 9 2 5 0.61
.
. $plot
```

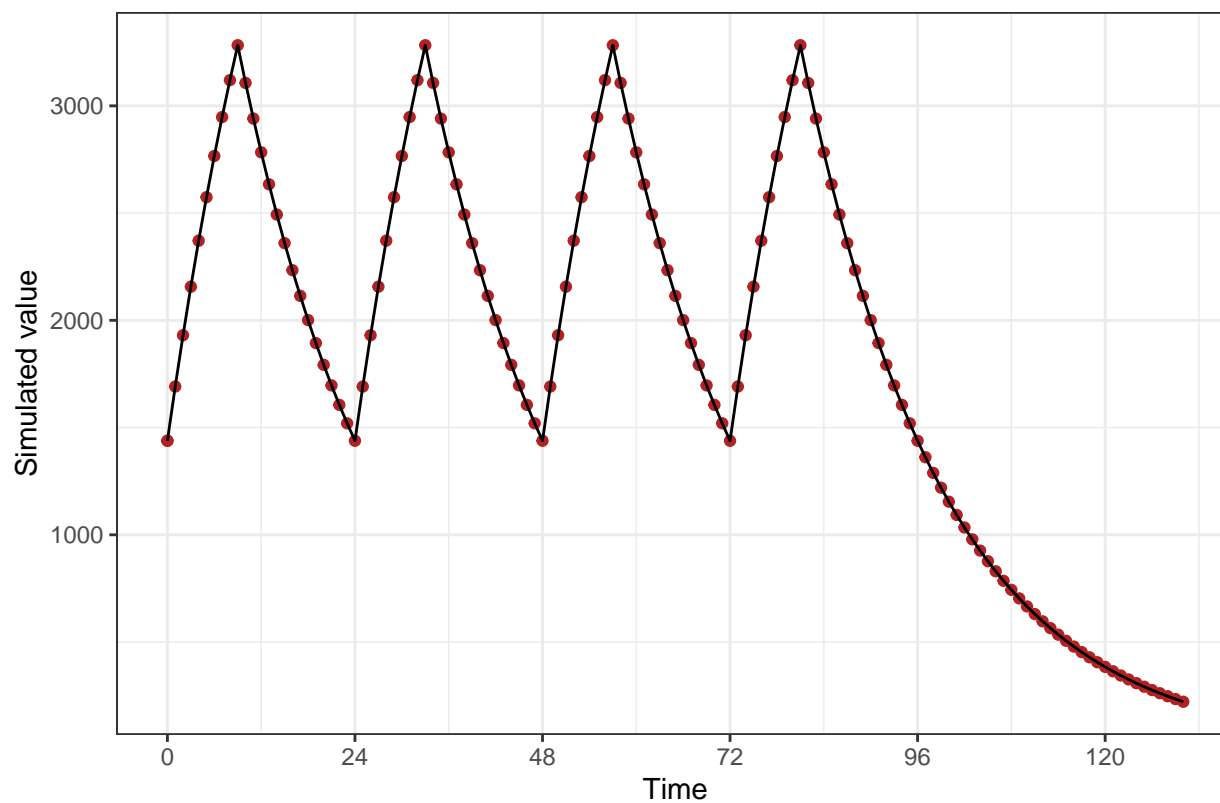
ID: 16; line: mrgsolve, point: NONMEM



8.17 17: Infusion with modeled duration, at steady state with bioav factor

```
. $ev
. Events:
.   ID time amt rate ii addl cmt evid ss DUR2 MODE BIOAV
. 1 17 0 100 -2 24 3 2 1 1 9 2 0.61
.
. $plot
```

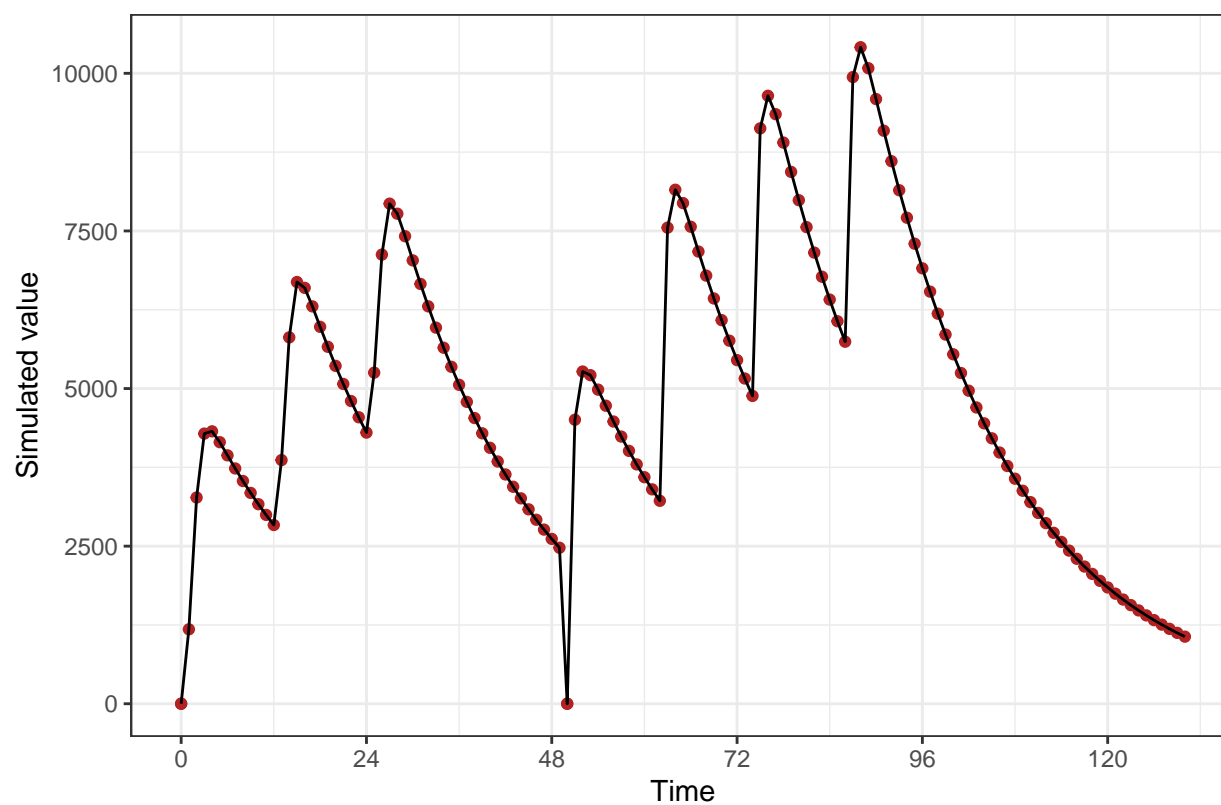
ID: 17; line: mrgsolve, point: NONMEM



8.18 18: Reset and dose (EVID 4) with additional

```
. $ev
. Events:
.   ID time amt rate ii addl cmt evid BIOAV
. 1 18   0 100   50 12   2   1   1 0.61
. 2 18   50 120    0 12   3   1   4 0.50
.
. $plot
```

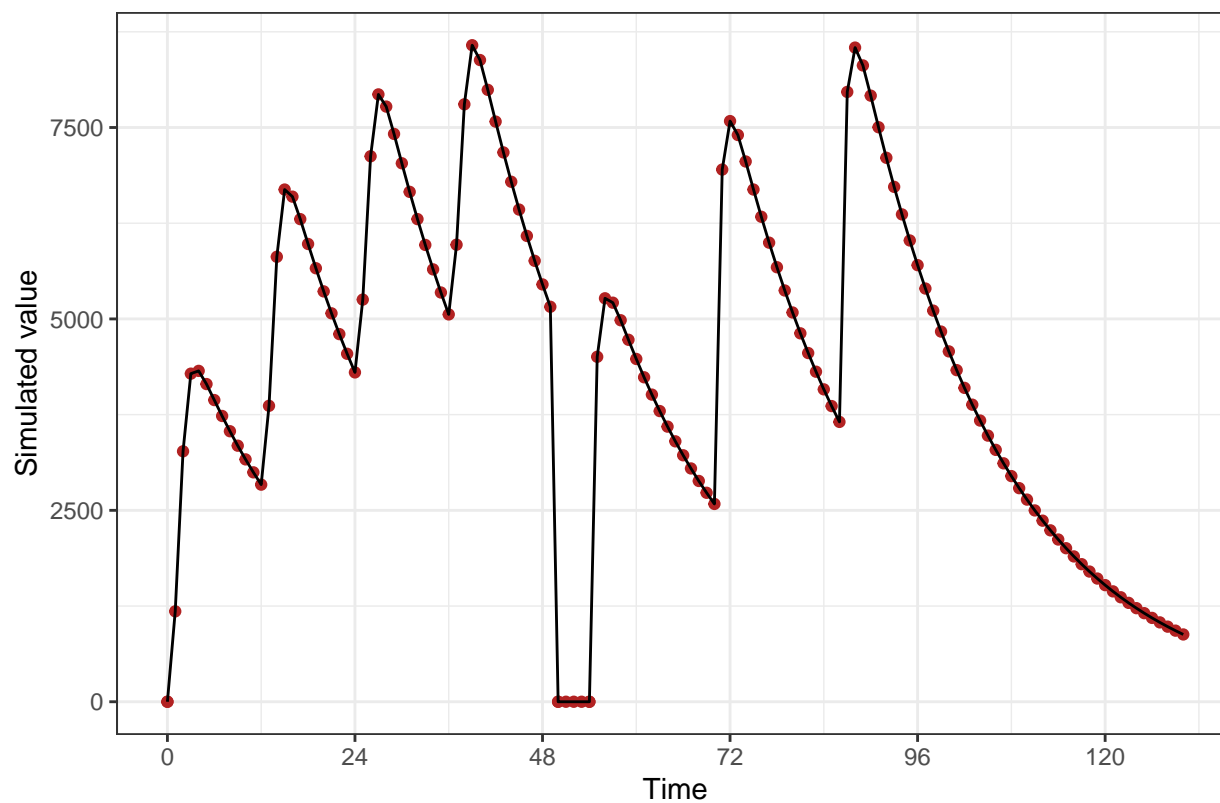

ID: 18; line: mrgsolve, point: NONMEM



8.19 19: Reset (EVID 3) with additional

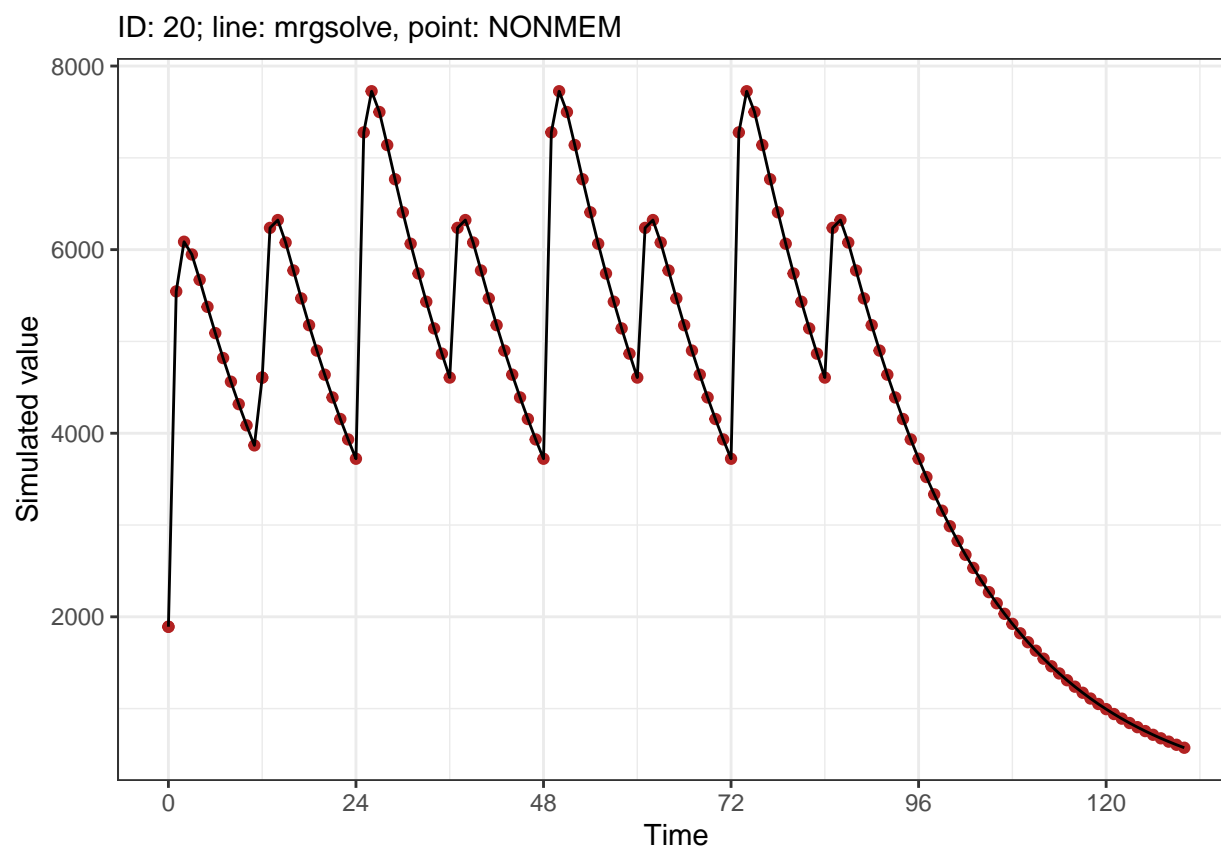
```
. $ev
. Events:
.   ID time amt rate ii addl cmt evid BIOAV
. 1 19    0 100   50 12   3   1    1 0.61
. 2 19   50   0    0  0   0   2    3 1.00
. 3 19   54 120    0 16   2   1    1 1.00
.
. $plot
```

ID: 19; line: mrgsolve, point: NONMEM



8.20 20: Steady state 1 and 2

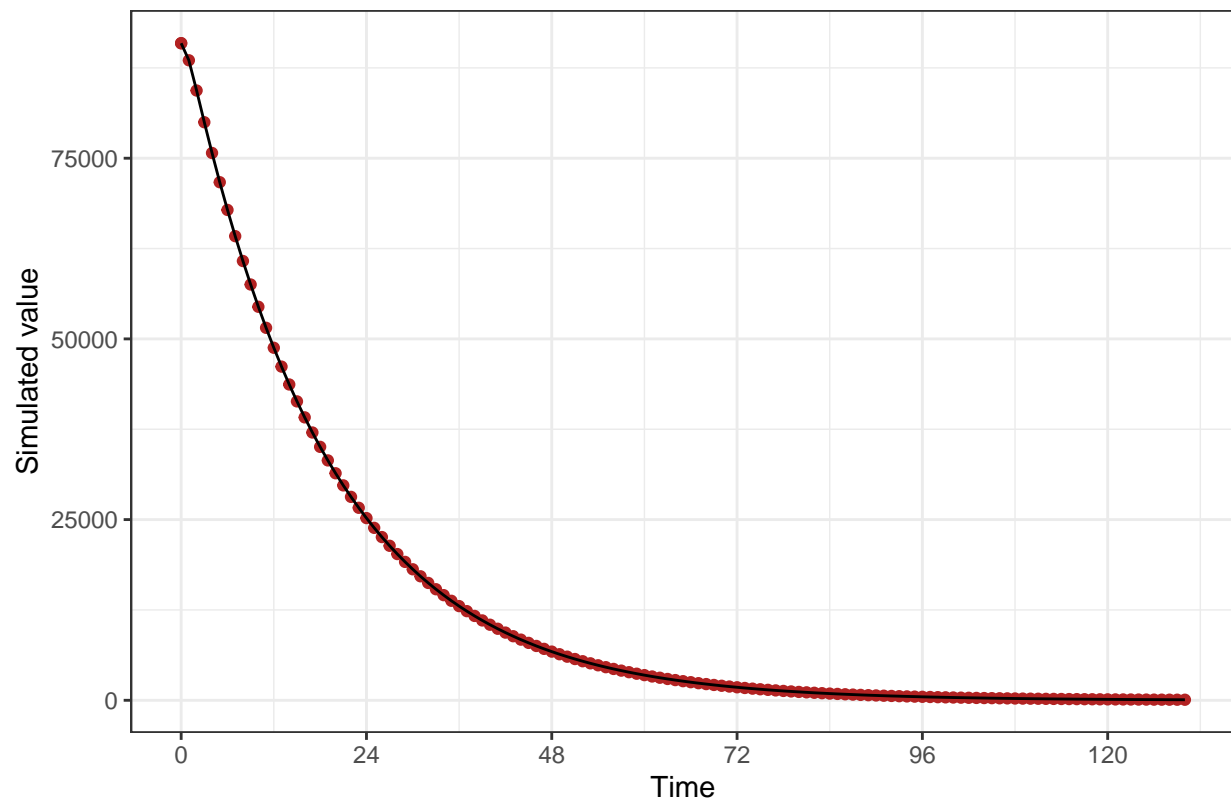
```
. $ev
. Events:
.   ID time amt ii addl cmt evid ss
. 1 20    0 100 24    3    1    1  1
. 2 20   12  50 24    3    1    1  2
.
. $plot
```



8.21 21: Steady state infusion

```
. $ev
. Events:
.   ID time amt rate cmt evid ss
. 1 21   0   0 100   1   1 1
.
. $plot
```

ID: 21; line: mrgsolve, point: NONMEM



9 Control stream

```
writeLines(readLines("model/1001/1001.lst"))
```

```
Wed Oct 9 20:12:37 UTC 2019
```

```
$PROB RUN# 101
```

```
$INPUT C ID TIME EVID AMT CMT SS II ADDL RATE LAGT MODE DUR2 RAT2 BIOAV DV
```

```
$DATA ../../data/1001.csv IGNORE=C
```

```
$SUBROUTINES ADVAN2 TRANS2
```

```
$PK
```

```
TVCL=THETA(1)
```

```
CL=TVCL*EXP(ETA(1))
```

```
TVV2=THETA(2)
```

```
V=TVV2*EXP(ETA(2))
```

```
TVKA=THETA(3)
```

```
KA=TVKA*EXP(ETA(3))
```

```
ALAG2 = LAGT
```

F2 = BIOAV

IF(MODE.EQ.1) R2 = RAT2

IF(MODE.EQ.2) D2 = DUR2

\$ERROR

IPRED=A(2)/(V/1000)

Y=IPRED*EXP(ERR(1))

CP = IPRED

\$THETA

(1.1, FIX) ;; CL

(20, FIX) ;; V

(1.5, FIX) ;; KA

\$OMEGA

0.0 FIX

0.0 FIX

0.0 FIX

\$SIGMA

0.00 FIX

\$TABLE FILE=TAB ID TIME CP NOPRINT ONEHEADER NOAPPEND

\$SIMULATION (2674474) ONLYSIMULATION

NM-TRAN MESSAGES

WARNINGS AND ERRORS (IF ANY) FOR PROBLEM 1

(WARNING 2) NM-TRAN INFERS THAT THE DATA ARE POPULATION.

License Registered to: Metrum Research Group

Expiration Date: 14 JUL 2020

Current Date: 9 OCT 2019

Days until program expires : 280

1NONLINEAR MIXED EFFECTS MODEL PROGRAM (NONMEM) VERSION 7.4.3

ORIGINALLY DEVELOPED BY STUART BEAL, LEWIS SHEINER, AND ALISON BOECKMANN

CURRENT DEVELOPERS ARE ROBERT BAUER, ICON DEVELOPMENT SOLUTIONS,

AND ALISON BOECKMANN. IMPLEMENTATION, EFFICIENCY, AND STANDARDIZATION

PERFORMED BY NOUS INFOSYSTEMS.

PROBLEM NO.: 1

RUN# 101

ODATA CHECKOUT RUN: NO

DATA SET LOCATED ON UNIT NO.: 2

THIS UNIT TO BE REWOUND: NO

NO. OF DATA RECS IN DATA SET: 2777

NO. OF DATA ITEMS IN DATA SET: 17

ID DATA ITEM IS DATA ITEM NO.: 2

DEP VARIABLE IS DATA ITEM NO.: 16

```

MDV DATA ITEM IS DATA ITEM NO.: 17
OINDICES PASSED TO SUBROUTINE PRED:
  4  3  5 10  7  8  6  0  0  0  9
OLABELS FOR DATA ITEMS:
  C ID TIME EVID AMT CMT SS II ADDL RATE LAGT MODE DUR2 RAT2 BIOAV DV MDV
O(NONBLANK) LABELS FOR PRED-DEFINED ITEMS:
  CP
OFORMAT FOR DATA:
  (E2.0,E3.0,E4.0,E2.0,E4.0,2E2.0,2E3.0,E17.0,E6.0,2E2.0,3E6.0,1F2.0)

TOT. NO. OF OBS RECS:      2751
TOT. NO. OF INDIVIDUALS:    21
OLENGTH OF THETA:      3
ODEFAULT THETA BOUNDARY TEST OMITTED:    NO
OOMEGA HAS SIMPLE DIAGONAL FORM WITH DIMENSION:    3
ODEFAULT OMEGA BOUNDARY TEST OMITTED:    NO
OSIGMA HAS SIMPLE DIAGONAL FORM WITH DIMENSION:    1
ODEFAULT SIGMA BOUNDARY TEST OMITTED:    NO
OINITIAL ESTIMATE OF THETA:
  LOWER BOUND      INITIAL EST      UPPER BOUND
    0.1100E+01      0.1100E+01      0.1100E+01
    0.2000E+02      0.2000E+02      0.2000E+02
    0.1500E+01      0.1500E+01      0.1500E+01
OINITIAL ESTIMATE OF OMEGA:
  0.0000E+00
  0.0000E+00  0.0000E+00
  0.0000E+00  0.0000E+00  0.0000E+00
OOMEGA CONSTRAINED TO BE THIS INITIAL ESTIMATE
OINITIAL ESTIMATE OF SIGMA:
  0.0000E+00
OSIGMA CONSTRAINED TO BE THIS INITIAL ESTIMATE
OSIMULATION STEP OMITTED:    NO
OBJ FUNC EVALUATED:          NO
ORIGINAL DATA USED ON EACH NEW SIMULATION:          NO
SEEDS RESET ON EACH NEW SUPERSET ITERATION:          YES
OSIMULATION RANDOM METHOD SELECTED (RANMETHOD): 4U
SEED  1 RESET TO INITIAL: YES
SOURCE  1:
  SEED1:      2674474  SEED2:          0  PSEUDO-NORMAL
OWARNING: NO. OF OBS RECS IN INDIVIDUAL REC NO.      1 (IN INDIVIDUAL
  REC ORDERING) EXCEEDS ONE WHILE INITIAL ESTIMATE OF WITHIN INDIVIDUAL VARIANCE IS ZERO
OWARNING: NO. OF OBS RECS IN INDIVIDUAL REC NO.      2 (IN INDIVIDUAL
  REC ORDERING) EXCEEDS ONE WHILE INITIAL ESTIMATE OF WITHIN INDIVIDUAL VARIANCE IS ZERO
OWARNING: NO. OF OBS RECS IN INDIVIDUAL REC NO.      3 (IN INDIVIDUAL
  REC ORDERING) EXCEEDS ONE WHILE INITIAL ESTIMATE OF WITHIN INDIVIDUAL VARIANCE IS ZERO
OWARNING: NO. OF OBS RECS IN INDIVIDUAL REC NO.      4 (IN INDIVIDUAL
  REC ORDERING) EXCEEDS ONE WHILE INITIAL ESTIMATE OF WITHIN INDIVIDUAL VARIANCE IS ZERO
OWARNING: NO. OF OBS RECS IN INDIVIDUAL REC NO.      5 (IN INDIVIDUAL
  REC ORDERING) EXCEEDS ONE WHILE INITIAL ESTIMATE OF WITHIN INDIVIDUAL VARIANCE IS ZERO
OTABLES STEP OMITTED:    NO
NO. OF TABLES:          1
SEED NUMBER (SEED):      11456
RANMETHOD:                3U
MC SAMPLES (ESAMPLE):     300

```

WRES SQUARE ROOT TYPE (WRESCHOL): EIGENVALUE

0-- TABLE 1 --

ORECORDS ONLY: ALL

04 COLUMNS APPENDED: NO

PRINTED: NO

HEADERS: ONE

FILE TO BE FORWARDED: NO

FORMAT: S1PE11.4

LFORMAT:

RFORMAT:

FIXED_EFFECT_ETAS:

0USER-CHOSEN ITEMS:

ID TIME CP

1DOUBLE PRECISION PREDPP VERSION 7.4.3

ONE COMPARTMENT MODEL WITH FIRST-ORDER ABSORPTION (ADVAN2)

OMAXIMUM NO. OF BASIC PK PARAMETERS: 3

OBASIC PK PARAMETERS (AFTER TRANSLATION):

ELIMINATION RATE (K) IS BASIC PK PARAMETER NO.: 1

ABSORPTION RATE (KA) IS BASIC PK PARAMETER NO.: 3

TRANSLATOR WILL CONVERT PARAMETERS

CLEARANCE (CL) AND VOLUME (V) TO K (TRANS2)

OCOMPARTMENT ATTRIBUTES

COMPT. NO.	FUNCTION	INITIAL STATUS	ON/OFF ALLOWED	DOSE ALLOWED	DEFAULT FOR DOSE	DEFAULT FOR OBS.
1	DEPOT	OFF	YES	YES	YES	NO
2	CENTRAL	ON	NO	YES	NO	YES
3	OUTPUT	OFF	YES	NO	NO	NO

1

ADDITIONAL PK PARAMETERS - ASSIGNMENT OF ROWS IN GG

COMPT. NO.	SCALE	BIOAVAIL. FRACTION	INDICES ZERO-ORDER RATE	ZERO-ORDER DURATION	ABSORB LAG
1	*	*	*	*	*
2	*	5	6	7	4
3	*	-	-	-	-

- PARAMETER IS NOT ALLOWED FOR THIS MODEL

* PARAMETER IS NOT SUPPLIED BY PK SUBROUTINE;
WILL DEFAULT TO ONE IF APPLICABLE

ODATA ITEM INDICES USED BY PRED ARE:

EVENT ID DATA ITEM IS DATA ITEM NO.:	4
TIME DATA ITEM IS DATA ITEM NO.:	3
DOSE AMOUNT DATA ITEM IS DATA ITEM NO.:	5
DOSE RATE DATA ITEM IS DATA ITEM NO.:	10
STEADY STATE DATA ITEM IS DATA ITEM NO.:	7
INTERVAL DATA ITEM IS DATA ITEM NO.:	8
ADDL. DOSES DATA ITEM IS DATA ITEM NO.:	9
COMPT. NO. DATA ITEM IS DATA ITEM NO.:	6

OPK SUBROUTINE CALLED WITH EVERY EVENT RECORD.

PK SUBROUTINE NOT CALLED AT NONEVENT (ADDITIONAL OR LAGGED) DOSE TIMES.

OERROR SUBROUTINE CALLED WITH EVERY EVENT RECORD.

OERROR SUBROUTINE INDICATES THAT DERIVATIVES OF COMPARTMENT AMOUNTS ARE USED.

```

1
SIMULATION STEP PERFORMED
SOURCE 1:
  SEED1:    1222495484  SEED2:          0
Elapsed simulation time in seconds:    0.01
ESTIMATION STEP OMITTED:             YES
Elapsed finaloutput time in seconds:   0.23
#CPUT: Total CPU Time in Seconds,     0.260
Stop Time:
Wed Oct  9 20:12:42 UTC 2019

```

10 Session Info

```

options(width = 120)
devtools::session_info()

```

```

. - Session info -----
. setting value
. version R version 3.5.1 (2018-07-02)
. os      Ubuntu 14.04.5 LTS
. system  x86_64, linux-gnu
. ui      X11
. language (EN)
. collate en_US.UTF-8
. ctype   en_US.UTF-8
. tz      Etc/UTC
. date    2019-10-09
.
. - Packages -----
. package      * version      date      lib source
. assertthat   0.2.1        2019-03-21 [1] CRAN (R 3.5.1)
. backports    1.1.4        2019-04-10 [1] CRAN (R 3.5.1)
. callr        3.3.1        2019-07-18 [1] CRAN (R 3.5.1)
. cli          1.1.0        2019-03-19 [1] CRAN (R 3.5.1)
. colorspace   1.4-1        2019-03-18 [1] CRAN (R 3.5.1)
. crayon       1.3.4        2017-09-16 [1] CRAN (R 3.5.1)
. desc         1.2.0        2018-05-01 [1] CRAN (R 3.5.1)
. devtools     2.1.0        2019-07-06 [1] CRAN (R 3.5.1)
. digest       0.6.20       2019-07-04 [1] CRAN (R 3.5.1)
. dplyr        * 0.8.3        2019-07-04 [1] CRAN (R 3.5.1)
. evaluate     0.14         2019-05-28 [1] CRAN (R 3.5.1)
. fork         1.2.5        2019-02-01 [1] local
. fs           1.3.1        2019-05-06 [1] CRAN (R 3.5.1)
. ggplot2      * 3.2.0        2019-06-16 [1] CRAN (R 3.5.1)
. glue         1.3.1        2019-03-12 [1] CRAN (R 3.5.1)
. gtable       0.3.0        2019-03-25 [1] CRAN (R 3.5.1)
. highr        0.8          2019-03-20 [1] CRAN (R 3.5.1)
. hms          0.5.0        2019-07-09 [1] CRAN (R 3.5.1)
. htmltools    0.3.6        2017-04-28 [1] CRAN (R 3.5.1)
. knitr        1.23         2019-05-18 [1] CRAN (R 3.5.1)
. labeling     0.3          2014-08-23 [1] CRAN (R 3.5.1)
. lattice      0.20-38      2018-11-04 [4] CRAN (R 3.5.1)

```


. lazyeval	0.2.2	2019-03-15	[1]	CRAN (R 3.5.1)
. magrittr	1.5	2014-11-22	[1]	CRAN (R 3.5.1)
. MASS	7.3-51.1	2018-11-01	[4]	CRAN (R 3.5.1)
. memoise	1.1.0	2017-04-21	[1]	CRAN (R 3.5.1)
. metrumrg	5.57	2015-10-08	[1]	R-Forge (R 3.5.1)
. mrgsolve	* 0.9.2.9002	2019-10-09	[1]	local
. munsell	0.5.0	2018-06-12	[1]	CRAN (R 3.5.1)
. pillar	1.4.2	2019-06-29	[1]	CRAN (R 3.5.1)
. pkgbuild	1.0.3	2019-03-20	[1]	CRAN (R 3.5.1)
. pkgconfig	2.0.2	2018-08-16	[1]	CRAN (R 3.5.1)
. pkgload	1.0.2	2018-10-29	[1]	CRAN (R 3.5.1)
. plyr	1.8.4	2016-06-08	[1]	CRAN (R 3.5.1)
. prettyunits	1.0.2	2015-07-13	[1]	CRAN (R 3.5.1)
. processx	3.4.1	2019-07-18	[1]	CRAN (R 3.5.1)
. ps	1.3.0	2018-12-21	[1]	CRAN (R 3.5.1)
. purrr	* 0.3.2	2019-03-15	[1]	CRAN (R 3.5.1)
. R6	2.4.0	2019-02-14	[1]	CRAN (R 3.5.1)
. Rcpp	1.0.1	2019-03-17	[1]	CRAN (R 3.5.1)
. RcppArmadillo	0.9.600.4.0	2019-07-15	[1]	CRAN (R 3.5.1)
. readr	* 1.3.1	2018-12-21	[1]	CRAN (R 3.5.1)
. remotes	2.1.0	2019-06-24	[1]	CRAN (R 3.5.1)
. reshape	0.8.8	2018-10-23	[1]	CRAN (R 3.5.1)
. rlang	0.4.0	2019-06-25	[1]	CRAN (R 3.5.1)
. rmarkdown	1.14	2019-07-12	[1]	CRAN (R 3.5.1)
. rprojroot	1.3-2	2018-01-03	[1]	CRAN (R 3.5.1)
. scales	1.0.0	2018-08-09	[1]	CRAN (R 3.5.1)
. sessioninfo	1.1.1	2018-11-05	[1]	CRAN (R 3.5.1)
. stringi	1.4.3	2019-03-12	[1]	CRAN (R 3.5.1)
. stringr	1.4.0	2019-02-10	[1]	CRAN (R 3.5.1)
. testthat	2.1.1	2019-04-23	[1]	CRAN (R 3.5.1)
. tibble	2.1.3	2019-06-06	[1]	CRAN (R 3.5.1)
. tidyr	* 0.8.3	2019-03-01	[1]	CRAN (R 3.5.1)
. tidyselect	0.2.5	2018-10-11	[1]	CRAN (R 3.5.1)
. usethis	1.5.1	2019-07-04	[1]	CRAN (R 3.5.1)
. vctrs	0.2.0	2019-07-05	[1]	CRAN (R 3.5.1)
. withr	2.1.2	2018-03-15	[1]	CRAN (R 3.5.1)
. xfun	0.8	2019-06-25	[1]	CRAN (R 3.5.1)
. XML	3.98-1.20	2019-06-06	[1]	CRAN (R 3.5.1)
. yaml	2.2.0	2018-07-25	[1]	CRAN (R 3.5.1)
. zeallot	0.1.0	2018-01-28	[1]	CRAN (R 3.5.1)
.				
. [1]	/data/Rlibs			
. [2]	/usr/local/lib/R/site-library			
. [3]	/usr/lib/R/site-library			
. [4]	/usr/lib/R/library			