# Practical Exercises for Exercise Collection

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## **Exercise 1**

- (a) Open R Studio
- (b) Open a new R-Script
- (c) Load data set chickwts
- (d) Do summary statistic (numerically and graphically)
- (e) For advanced R users: Try an anova (are the assumptions fulfilled?) and a Tukey-Anscombe plot. Try a histogram with a density line on top. ...

## **Exercise 2**

(a) Create a data frame with 3 columns.

## **Exercise 3**

- (a) Install package MASS.
- (b) Load data set bacteria.
- (c) Describe in your own words what the data set bacteria contains.
- (d) Do summary statistic (numerically and graphically).
- (e) Select only observations collected during the second week.

# **Exercise 4**

What is conceptionally the difference between the bracket types [...] and (...)?

```
chickwts[, 2]
summary(aov(weight ~ feed, data = chickwts))
```

- (a) How many levels has the factor variable trt from bacteria?
- (b) Define a new variable trt.new in which you combine the levels drug and drug+ into one single level and label it as treated. The new variable trt.new should in the end have two levels: placebo and treated.
- (c) Do summary statistics for placebo and treated group.

# **Exercise 6**

- (a) Load data set ToothGrowth.
- (b) Do summary statistic (numerically and graphically).
- (c) Define additional column dose.factor by converting the numeric variable dose into a factor variable.
- (d) Are the tooth length measurements normally distributed within the treatment (supp: VC or OJ) and within in the different doses (dose: 0.5, 1, 2)?

## **Exercise 7**

- (a) Import the data set perulung\_ems.csv (taken from Kirkwood and Sterne, 2nd edition) into R. Data from a study of lung function among children living in a deprived suburb of Lima, Peru. Variables:
  - fev1: in liter, "Forced Expiratory Volume in 1 second" measured by a spirometer. This is the maximum volume of air which the children could breath out in 1 second
  - age: in years
  - height: in cm
  - sex: 0 = girl, 1 = boy
  - respsymp: respiratory symptoms experienced by the child over the previous 12 months
- (b) What delimiter do you need to choose?
- (c) Do all variables have the correct data type (numeric, integer, factor)? If not, do correct and / or define them.
- (d) Check for heteroscedascity or homogeneity of variances

Apply the summary statistics to the perulung\_ems and ToothGrowth data set.

# **Exercise 9A: Plausibility Checks**

- (a) What can go wrong?
- (b) Identify different strategies for spotting these potential errors.
  - Logical errors
  - Spelling mistakes
- (c) Import the data set bacteria\_plausibility\_check.csv to R.
- (d) Detect the six errors in the imported data set bacteria\_plausibility\_check.csv in R.
- (e) Find possible solutions in R how to handle these challenges.
- (f) Do all variables have the correct data type (numeric, integer, factor)? If not, do correct / define them.

# **Exercise 9B: Missing Values**

(a) Check out the difference between the different missing values.

```
y1 <- c(2, 4, 3, NA, 6, 1)
y2 <- c("diseased", "healthy", NA, "NA")
y3 <- c(1, "NA", 0, 1, NaN)
#
is.na(y1)
which(is.na(y1))
is.na(y2)
which(is.na(y2))
is.na(y3)
which(is.na(y3))
is.na(y3)</pre>
```

- (b) Create a vector with missing values and determine the mean and median.
- (c) If x = c(22,3,7,NA,NA,67) what will be the output for the R statement length(x)?

- (d) If x = c(NA, 3, 14, NA, 33, 17, NA, 41) which line of R code removes all occurrences of NA in x.
- (e) If y = c(1, 3, 12, NA, 33, 7, NA, 21) what R statement will replace all occurrences of NA with 11?
- (f) If x = c(34, 33, 65, 37, 89, NA, 43, NA, 11, NA, 23, NA) then what will count the number of occurrences of NA in x?
- (g) Create the vector x1. Then, find again the number of missing values and their position.

```
x1 \leftarrow c(rnorm(10,5,2), NA, 5:12, NA, 6, 7.5, NA)
```

(h) Now, create the vector x2 and assess the difference to x1.

```
x2 \leftarrow c(rnorm(10,5,2), NA, 5:12, NA, 6, 7.5, NA, log(-2))
```

- (i) What is the meaning of "NA" versus "NaN"?
- (j) Replace the missing values in x1 with a 0. Check then that the NAs are no longer present. Try two different commands to coerce the NAs into 0.

## **Exercise 10**

- (a) Import the data set water\_errors.csv to R: A data frame with 61 observations on the following 6 variables.
  - location: a factor with levels North and South indicating whether the town is as north as Derby.
  - town: the name of the town.
  - mortality: averaged annual mortality per 100.000 male inhabitants.
  - hardness: calcium concentration (in parts per million).
  - smoker: If there are any smokers living in town.
  - num.of.cig: In case, smokers live in town, what number of cigarettes do they smoke per day.
- (b) Detect the errors in the imported data set water\_errors.csv in R.
- (c) Find possible solutions in R how to handle these challenges.
- (d) Do all variables have the correct data type (numeric, integer, factor)? If not, do correct / define them.

(a) Load the data set ToothGrowth within R and apply the two-sided two sample t-test to suitable variables of the data set.

```
data(ToothGrowth)
```

- (b) Interpret the results.
- (c) Read in the data set perulung\_ems and apply the two-sided t-test to suitable variables of the perulung\_ems data set and interpret the results.

#### Exercise 12

- (a) Apply the Chi-square test and the fisher exact test to the whole bacteria data set.
- (b) Apply the Chi-square test and the fisher exact test to the subset of bacteria containing only the observations taken in week 2 (cf. Exercise 3). Are there any issues?
- (c) Repeat this exercise by using the (previously defined) combined trt.new variable (cf. Exercise 5) with the two levels treated and drug.
- (d) Could you also obtain the odds ratios?
- (e) Try also a logistic regression in R. Ask Google for help!

# Exercise 13A: Outside plot frame

(a) Type demo(graphics) in your console and press enter. This command shows you a nice demonstration of possible R graphics.

```
# After the demonstration us the following commands:
dev.off()
par(mfrow=c(1,1))
```

(b) Change the x-axis and y-axis labelling of a boxplot plotting the len variable of the ToothGrowth data set.

```
data("ToothGrowth")
boxplot(ToothGrowth$len)
```

- (c) How do you set a main title for your above plot?
- (d) What does the following command do?

```
par(mfrow=c(2,2))
```

- (e) We have six different feed types in chickwts. Try to plot two separate boxplots for casein and horsebean and set the same minimum and maximum for the y-axis. Use the function subset for doing so.
- (f) How do you enlarge the font size of the axis as well as the axis labels of the following plot with the perulung data set?

```
lung <- read.csv("perulung_ems.csv", sep=";")
par(mfrow=c(1,1))
plot(lung$fev1, lung$height)</pre>
```

(g) Label the x-axis of the following plot with "Vitamin C in  $\mu$ g". Use the greek letter for  $\mu$ .

```
plot(ToothGrowth$dose, ToothGrowth$len)
```

(h) Read http://www.statmethods.net/advgraphs/parameters.html.

# Exercise 13B: Inside the square of the plot

(a) Type demo(graphics) in your console and press enter. This command shows you a nice demonstration of possible R graphics.

```
# After the demonstration us the following commands:
dev.off()
par(mfrow=c(1,1))
```

(b) Add a legend to the following barplot. Are there several different solutions for this?

(c) Add a density line to this histogram.

```
hist(ToothGrowth$len, prob = TRUE, col = "grey", ylim = c(0, 0.05))
```

(d) Add a **dotted red** linear regression line to the following plot.

```
plot(lung$height, lung$fev1)
```

(e) Color the points in the following plot according to the sex variable.

```
plot(lung$height, lung$fev1)
```

(f) Add two linear regression lines separately for female and maleto the following plot.

```
plot(lung$height, lung$fev1)
```

(g) Color the points in the following plot according to the supp variable. Use different point characters (pch) based on the supp variable.

```
plot(ToothGrowth$len, ToothGrowth$dose)
```

(h) Read http://www.statmethods.net/advgraphs/parameters.html.

## **Exercise 14**

(a) Load the below data set and for further information check the command ?water.

```
# install.packages("HSAUR3")
library("HSAUR3")
data("water")
str(water)
head(water)
summary(water)
```

- (b) Try to plot the variables mortality against hardness from the water data set.
- (c) Add a main title to the above plot (mortality against hardness).

- (d) Change the ...
  - (a) font size of the axis annotation
  - (b) font size of the x- and y-axis labels
  - (c) the point sizes within the plot
  - ... of the above plot (mortality against hardness).
- (e) Looking at the above plot: Do you think the two variables hardness and mortality correlate? What function do you use to find out the correlation coefficient? Do they have a positive or a negative correlation coefficient? How do you interpret the correlation coefficient in your own words?
- (f) In the water data set, can you graphically find out if there is a difference between the two variables hardness and mortality conditional on the location (North, South).
- (g) Add a legend to the above plot so that you can easily differentiate the locations (North or South) of the observations.
- (h) Do a barplot of the variable location from the water data set.
- (i) ADDITIONAL: Try if any of these following plotting functions can be applied to the data sets perulung or ToothGrowth.

- (a) Open the .R file ANOVA\_with\_chickwts .R from your RCourse folder and have another look on how we applied the anova to the chickwts data set. Check line for line.
- (b) Load the ToothGrowth data set into R and encode the numeric variable dose as a factor variable. Define the new factor variable as dose.factor with the three levels low, med and high and add it to the data frame of ToothGrowth.

#### data(ToothGrowth)

- (c) Visualize the variable len per dose.factor level in a boxplot.
- (d) With the help of the R-commands written in the ANOVA\_with\_chickwts.R file, apply a analysis of variance (ANOVA) to the data set ToothGrowth

- (a) Reuse the commands from the lecture slides to fit a simple as well as a multiple linear regression model to the data set of perulung\_ems. Use fev1 as your response variable y.
- (b) Check the model assumptions.
- (c) Which model is best?

# **Exercise 17**

(a) Load the ToothGrowth data set and run the following four linear regression models.

- (b) Have a look at the summary of these models.
- (c) How do you interpret the model coefficients?
- (d) Which model is best?

# **Exercise 18**

- (a) Load the water data set and fit a multiple linear regression model. Use mortality as your response variable and add hardness and location as an explanatory variable.
- (b) Check the underlying model assumptions.
- (c) Add an interaction term between hardness and location to the above estimated multiple linear regression model.
- (d) Interpret the interaction coefficient hardness:locationSouth.
- (e) Check the underlying model assumptions.
- (f) Which one is the better model? With or without the interaction term?
- (g) How to derive confidence intervals for the regression coefficient of hardness and location?

Hypothetical example - from Kirkwood and Sterne, Medical Statistics, 2nd ed., p. 177

- (a) Read in the data set lepto. This study presents a serology survey of leptospira sero-prevalence in rural and urban areas of the west indies.
- (b) Encode the numeric variable antibodies as a factor with levels 0 and 1.
- (c) Make a crosstable with the risk factor exposure and antibodies.
- (d) Run a Chi-squared test, a Fisher's exact test and a logistic regression (glm) to assess if the exposure (living in rural vs. urban areas) is a risk factor.
- (e) Create a subset for male and female based on the variable gender.
- (f) Repeat the crosstable (2-by-2 table), Chi-squared test, Fisher's exact test and a logistic regression (glm) for the subsets **separately**.
- (g) Does the conclusion of your research question change with the analysis of the subsets? (Research question: Is the exposure (rural and urban areas) a risk factor?)
- (h) Fit a logistic regression model (glm) with exposure and gender as explanatory variables.