

Class 9: Structural Bioinformatics (pt 1)

AUTHOR

Michael Romero (A18135877)

The PDB database

The main data base for structural biology is called the PDB. Lets have a look at what it contains:

Download a CSV file from the PDB site (accessible from **"Analyze" > "PDB Statistics" > "by Experimental Method and Molecular Type"**).

Move this CSV file into your RStudio project and use it to answer the following questions:

```
stats <- read.csv("Data Export Summary.csv", row.names = 1)
head(stats)
```

	X. ray	EM	NMR	Integrative
Multiple.methods				
Protein (only)	176,204	20,299	12,708	342
218				
Protein/Oligosaccharide	10,279	3,385	34	8
11				
Protein/NA	9,007	5,897	287	24
7				
Nucleic acid (only)	3,066	200	1,553	2
15				
Other	173	13	33	3
0				
Oligosaccharide (only)	11	0	6	0
1				
	Neutron	Other	Total	
Protein (only)	83	32	209,886	
Protein/Oligosaccharide	1	0	13,718	
Protein/NA	0	0	15,222	
Nucleic acid (only)	3	1	4,840	
Other	0	0	222	
Oligosaccharide (only)	0	4	22	

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy. Give your answer in 2 s.f.

```
stats$Total
```

```
[1] "209,886" "13,718" "15,222" "4,840" "222" "22"
```

Oh, these are characters not numeric...

```
sum( stats$Neutron)
```

```
[1] 87
```

```
SumTotal <- as.numeric(sub(","," ", stats$Total))
```

Long coding, inefficient

```
library("readr")
stats <- read_csv("Data Export Summary.csv")
stats
```

```
# A tibble: 6 × 9
  `Molecular Type` `X-ray`   EM   NMR Integrative
  `Multiple methods` Neutron
  <chr>           <dbl> <dbl> <dbl>      <dbl>
  <dbl> <dbl>
  1 Protein (only)     176204 20299 12708      342
  218     83
  2 Protein/Oligosacch... 10279  3385   34        8
  11     1
  3 Protein/NA         9007   5897   287      24
  7     0
  4 Nucleic acid (only) 3066    200   1553      2
  15     3
  5 Other              173     13    33        3
  0     0
  6 Oligosaccharide (o... 11      0     6        0
  1     0
# i 2 more variables: Other <dbl>, Total <dbl>
```

```
n.total <- sum(stats$Total)
```

```
stats$`X-ray`
```

```
[1] 176204 10279 9007 3066 173 11
```

```
n.xray <- sum(stats$`X-ray`)
percent.xray <- n.xray / n.total * 100
```

```
percent.xray
```

```
[1] 81.48087
```

There are 81.48 percent Xray structures in the PDB

Q2: What proportion of structures in the PDB are protein?

```
(209886 + 13718 + 15222) / (209886 + 13718 + 15222 + 4840 + 222)
```

```
[1] 0.9791562
```

97.9%

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

2406

Exploring PDB structures

Package for structural bioinformatics

```
library(bio3d)  
  
hiv <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
hiv
```

Call: read.pdb(file = "1hsg")

Total Models#: 1
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A
B)

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 172 (residues: 128)

Non-protein/nucleic resid values: [HOH (127), MK1 (1)]

Protein sequence:

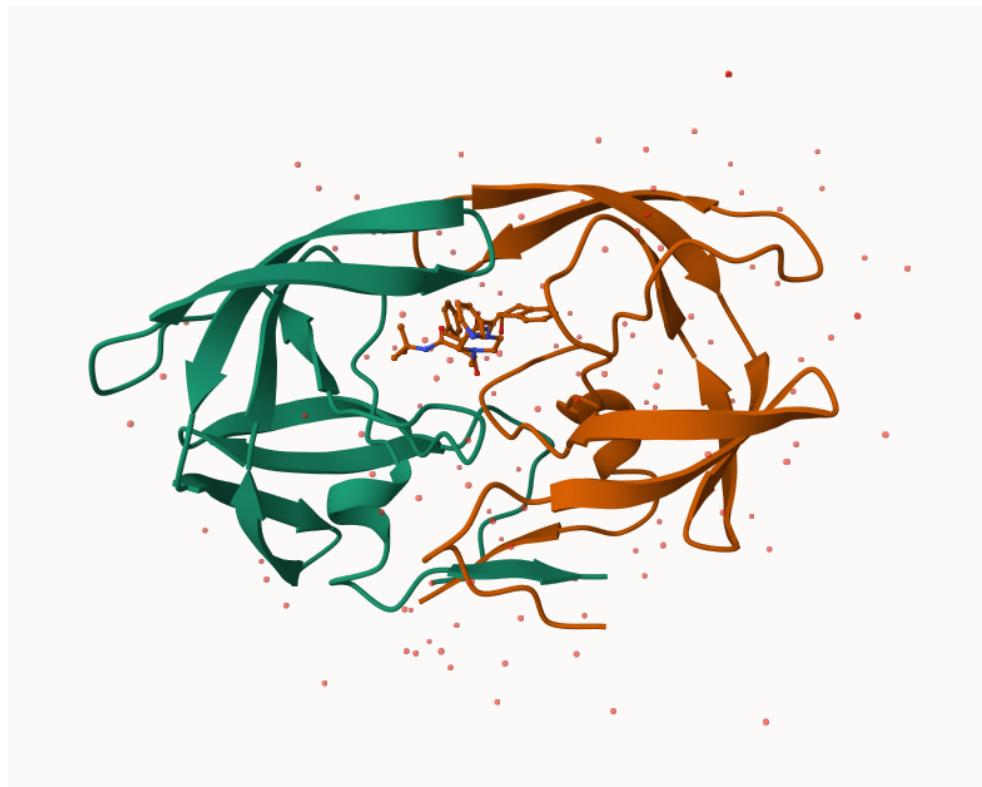
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPKMIGGIGGFIKVRQYD

QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE

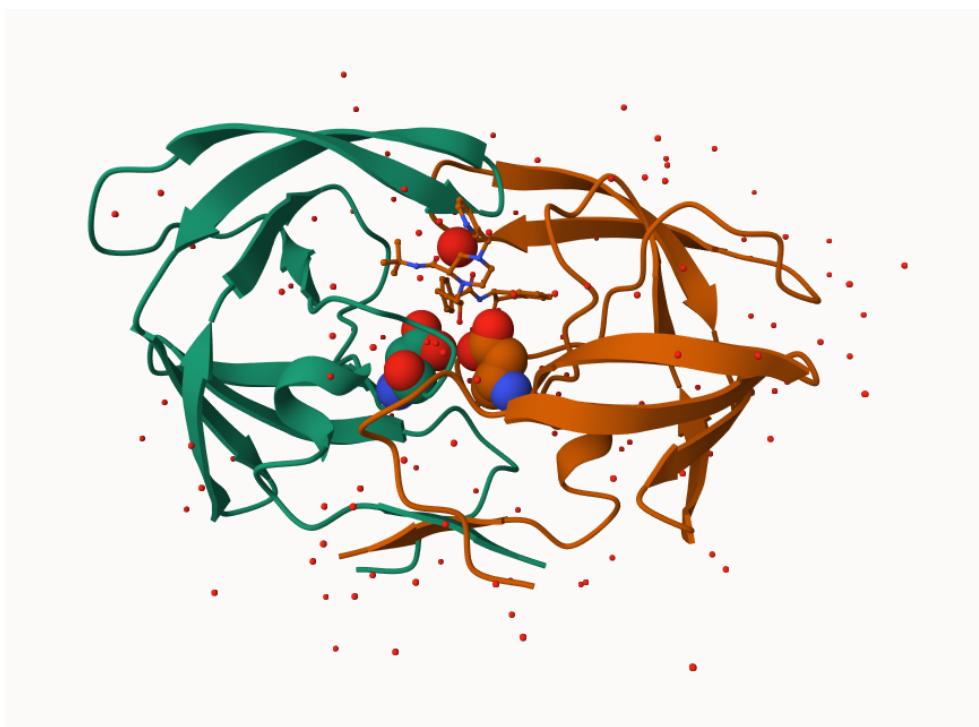
ALLDTGADDTVLEEMSLPGRWPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF

+ attr: atom, xyz, seqres, helix, sheet,
calpha, remark, call

Let's first use the Mol* viewer to explore this structure.



My first view of HIV-Pr



My first view of HIV-Pr

PDB objects in R

```
head( hiv$atom )
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y
z	o		b							
1	ATOM	1		N	<NA>	PRO	A	1	<NA>	29.361
5.862	1	38.10								
2	ATOM	2		CA	<NA>	PRO	A	1	<NA>	30.307
5.319	1	40.62								
3	ATOM	3		C	<NA>	PRO	A	1	<NA>	29.760
4.022	1	42.64								
4	ATOM	4		O	<NA>	PRO	A	1	<NA>	28.600
3.676	1	43.40								
5	ATOM	5		CB	<NA>	PRO	A	1	<NA>	30.508
6.342	1	37.87								
6	ATOM	6		CG	<NA>	PRO	A	1	<NA>	29.296
7.162	1	38.40								
	segid	elesy	charge							
1	<NA>	N	<NA>							
2	<NA>	C	<NA>							
3	<NA>	C	<NA>							
4	<NA>	O	<NA>							
5	<NA>	C	<NA>							
6	<NA>	C	<NA>							

Extract the sequence

```
pbseq(hiv)
```

```

 1  2  3  4  5  6  7  8  9  10 11 12 13 14 15
16 17 18 19 20
"P" "Q" "I" "T" "L" "W" "Q" "R" "P" "L" "V" "T" "I" "K" "I"
"G" "G" "Q" "L" "K"
 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35
36 37 38 39 40
"E" "A" "L" "L" "D" "T" "G" "A" "D" "D" "T" "V" "L" "E" "E"
"M" "S" "L" "P" "G"
 41 42 43 44 45 46 47 48 49 50 51 52 53 54 55
56 57 58 59 60
"R" "W" "K" "P" "K" "M" "I" "G" "G" "I" "G" "G" "F" "I" "K"
"V" "R" "Q" "Y" "D"
 61 62 63 64 65 66 67 68 69 70 71 72 73 74 75
76 77 78 79 80
"Q" "I" "L" "I" "E" "I" "C" "G" "H" "K" "A" "I" "G" "T" "V"
"L" "V" "G" "P" "T"
 81 82 83 84 85 86 87 88 89 90 91 92 93 94 95
96 97 98 99 1
"P" "V" "N" "I" "I" "G" "R" "N" "L" "L" "T" "Q" "I" "G" "C"
"T" "L" "N" "F" "P"
 2  3  4  5  6  7  8  9  10 11 12 13 14 15 16
17 18 19 20 21
"Q" "I" "T" "L" "W" "Q" "R" "P" "L" "V" "T" "I" "K" "I" "G"
"G" "Q" "L" "K" "E"
 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36
37 38 39 40 41
"A" "L" "L" "D" "T" "G" "A" "D" "D" "T" "V" "L" "E" "E" "M"
"S" "L" "P" "G" "R"
 42 43 44 45 46 47 48 49 50 51 52 53 54 55 56
57 58 59 60 61
"W" "K" "P" "K" "M" "I" "G" "G" "I" "G" "G" "F" "I" "K" "V"
"R" "Q" "Y" "D" "Q"
 62 63 64 65 66 67 68 69 70 71 72 73 74 75 76
77 78 79 80 81
"I" "L" "I" "E" "I" "C" "G" "H" "K" "A" "I" "G" "T" "V" "L"
"V" "G" "P" "T" "P"
 82 83 84 85 86 87 88 89 90 91 92 93 94 95 96
97 98 99
"V" "N" "I" "I" "G" "R" "N" "L" "L" "T" "Q" "I" "G" "C" "T"
"L" "N" "F"

```

```
chainA_seq <- pbseq( trim.pdb(hiv, chain = "A") )
```

I can interactively view these PDB objects in R with the new **bio3dview** package. This is not yet on CRAN.

```
pak::pak("bioboot/bio3dview")
```

! Using bundled GitHub PAT. Please add your own PAT using
`gitcreds::gitcreds_set()`.

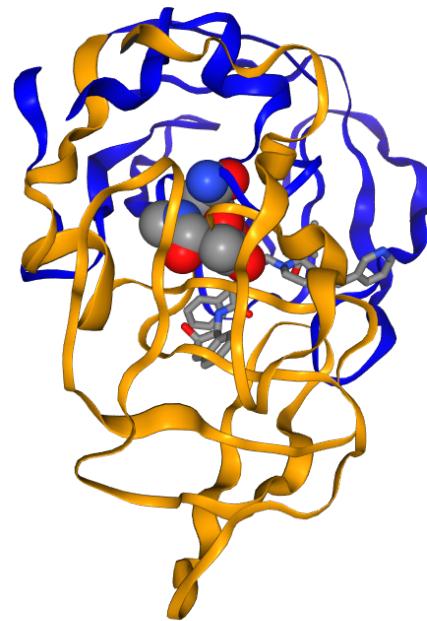
ℹ Loading metadata database
✓ Loading metadata database ... done

ℹ No downloads are needed
✓ 1 pkg + 40 deps: kept 39 [6.8s]

```
library(bio3dview)
```

```
sel <- atom.select(hiv, resno=25)

view.pdb(hiv, highlight = sel,
         highlight.style = "spacefill",
         colorScheme = "chain",
         col = c("blue", "orange"),
         backgroundColor = "pink")
```



Predict protein flexibility

We can run a bioinformatics calculation to predict protein dynamics - i.e. functional motions.

We will use the `nma()` function:

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file
PDB has ALT records, taking A only, `rm.alt=TRUE`

```
adk
```

Call: `read.pdb(file = "6s36")`

Total Models#: 1
Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)

```
Protein Atoms#: 1654 (residues/Calpha atoms#: 214)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

```
Non-protein/nucleic Atoms#: 244 (residues: 244)
Non-protein/nucleic resid values: [ CL (3), HOH (238), MG
(2), NA (1) ]
```

Protein sequence:

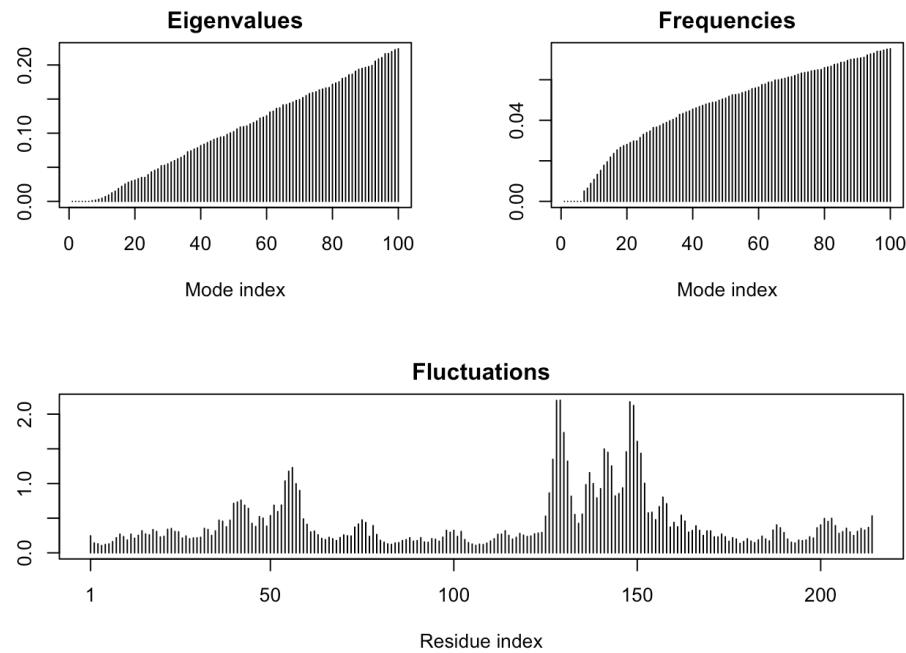
```
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGMLRAAVKSGSELGKQAKDIMALGKLVT
DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVVDYVLEFDVPDELIVDKI
VGRRVHAPSGRKYHVKFNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG
YYSKAEAGNTKYAKVDGTPVAEVRADEKILG
```

+ attr: atom, xyz, seqres, helix, sheet,
calpha, remark, call

```
m <- nma(adk)
```

```
Building Hessian... Done in 0.021 seconds.
Diagonalizing Hessian... Done in 0.454 seconds.
```

```
plot(m)
```



Generate a "trajectory" of predicted motion.

```
mktrj(m, file="ADK_nma.pdb")
```