

# Longest Common Subsequence with Gap Constraints (LCS-FIG) Implementation and Performance Analysis Report

Mohammad Sadegh Sirjani

March 25, 2025

## Abstract

This report presents a detailed implementation and analysis of the Longest Common Subsequence with Gap Constraints (LCS-FIG) algorithm. The algorithm extends the classic LCS problem by introducing gap constraints between consecutive matches, making it particularly suitable for DNA sequence analysis. We present a dynamic programming solution with  $O(nm)$  time and space complexity, along with comprehensive experimental results and performance analysis. The implementation demonstrates efficient scaling with input size and practical applicability for bioinformatics applications.

## 1 Introduction

### 1.1 Background

The Longest Common Subsequence (LCS) problem is a fundamental problem in computer science with applications in bioinformatics, text comparison, and version control systems. The LCS-FIG variant adds gap constraints between consecutive matches, making it particularly relevant for biological sequence analysis where spacing between matching elements is significant.

### 1.2 Problem Significance

In DNA sequence analysis, identifying common subsequences with controlled gaps is crucial for:

- **Gene identification and comparison:** Enables detection of similar genetic regions across different species, helping identify conserved functional elements and evolutionary relationships between organisms.
- **Regulatory sequence analysis:** Facilitates the study of DNA regions that control gene expression by identifying common patterns in promoter sequences and other regulatory elements.

- **Evolutionary relationship studies:** Helps reconstruct phylogenetic trees and understand species relationships by analyzing conserved sequence patterns across different organisms.
- **Mutation pattern detection:** Aids in identifying genetic variations and understanding mutation patterns by comparing sequences from different individuals or populations.

## 2 Problem Definition

### 2.1 Formal Definition

Given:

- Two sequences  $X[1..n]$  and  $Y[1..m]$
- A gap constraint parameter  $K$

Find: The longest common subsequence  $Z$  such that:

$$\forall i > 1 : pos_X(Z[i]) - pos_X(Z[i - 1]) \leq K + 1 \quad (1)$$

$$\forall i > 1 : pos_Y(Z[i]) - pos_Y(Z[i - 1]) \leq K + 1 \quad (2)$$

where  $pos_X(z)$  and  $pos_Y(z)$  denote the position of element  $z$  in sequences  $X$  and  $Y$  respectively.

## 3 Algorithm Description

### 3.1 Dynamic Programming Formulation

Let  $T[i, j]$  represent the length of the LCS-FIG ending at positions  $i$  and  $j$  in sequences  $X$  and  $Y$  respectively.

$$T[i, j] = \begin{cases} T[i - K - 1, j - K - 1] + 1 & \text{if } X[i] = Y[j] \text{ and } i, j > K + 1 \\ 1 & \text{if } X[i] = Y[j] \text{ and } (i \leq K + 1 \text{ or } j \leq K + 1) \\ \max(T[i - 1, j], T[i, j - 1]) & \text{otherwise} \end{cases} \quad (3)$$

### 3.2 Algorithm Implementation

## 4 Implementation Details

### 4.1 Data Structures

The implementation uses the following key data structures:

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**Algorithm 1** LCS-FIG Dynamic Programming Solution

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**Require:** Sequences  $X[1..n]$ ,  $Y[1..m]$ , gap constraint  $K$

**Ensure:** Length of LCS-FIG and the subsequence

```
1: Initialize  $T[0..n, 0..m]$  with zeros
2: Initialize  $Parent[0..n, 0..m]$  for backtracking
3: for  $i = 1$  to  $n$  do
4:   for  $j = 1$  to  $m$  do
5:     if  $X[i] = Y[j]$  then
6:       if  $i \geq K+1$  and  $j \geq K+1$  then
7:          $T[i, j] = T[i-K-1, j-K-1] + 1$ 
8:          $Parent[i, j] = \text{DIAGONAL\_GAP}$ 
9:       else
10:         $T[i, j] = 1$ 
11:         $Parent[i, j] = \text{START}$ 
12:      end if
13:    else
14:      if  $T[i-1, j] > T[i, j-1]$  then
15:         $T[i, j] = T[i-1, j]$ 
16:         $Parent[i, j] = \text{UP}$ 
17:      else
18:         $T[i, j] = T[i, j-1]$ 
19:         $Parent[i, j] = \text{LEFT}$ 
20:      end if
21:    end if
22:  end for
23: end for
24: return  $\text{ReconstructSolution}(Parent, X, Y)$ 
```

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```

1 class FIGDP:
2     def __init__(self, seq1: str, seq2: str, k: int):
3         self.seq1 = seq1
4         self.seq2 = seq2
5         self.n = len(seq1)
6         self.m = len(seq2)
7         self.k = k
8         self.dp_table = np.zeros((n+1, m+1), dtype=np.int32)
9         self.parent = np.zeros((n+1, m+1), dtype=np.int32)

```

Listing 1: Core Data Structures

## 4.2 Optimization Techniques

### 4.2.1 Memory Optimizations

- **Use of NumPy arrays for efficient memory layout:** Implements contiguous memory storage and vectorized operations, resulting in faster access times and better cache utilization compared to Python lists.
- **Int32 data type for reduced memory footprint:** Uses 32-bit integers instead of Python’s default dynamic typing, reducing memory usage by up to 50% for large sequences.
- **Contiguous memory allocation for better cache utilization:** Ensures that array elements are stored in consecutive memory locations, maximizing CPU cache efficiency and reducing memory access times.

### 4.2.2 Computational Optimizations

- **Single-pass solution reconstruction:** Implements an efficient backtracking algorithm that reconstructs the solution in a single pass through the parent pointer array, minimizing time complexity.
- **Efficient parent pointer tracking:** Uses a compact encoding scheme for parent pointers, reducing memory usage while maintaining fast access times for solution reconstruction.
- **Vectorized operations where possible:** Utilizes NumPy’s vectorized operations for bulk array computations, significantly reducing execution time compared to explicit Python loops.

## 5 Experimental Analysis

### 5.1 Test Environment

- Hardware: MacBook Pro M1
- OS: macOS 24.3.0

- Python: 3.8
- NumPy: 1.21.0

## 5.2 Test Configuration

- Sequence lengths: [100, 200, 400, 800, 1600, 3200]
- Gap constraints (K): [5, 10, 20]
- Number of runs per configuration: 5
- Random DNA sequence generation

## 5.3 Performance Results

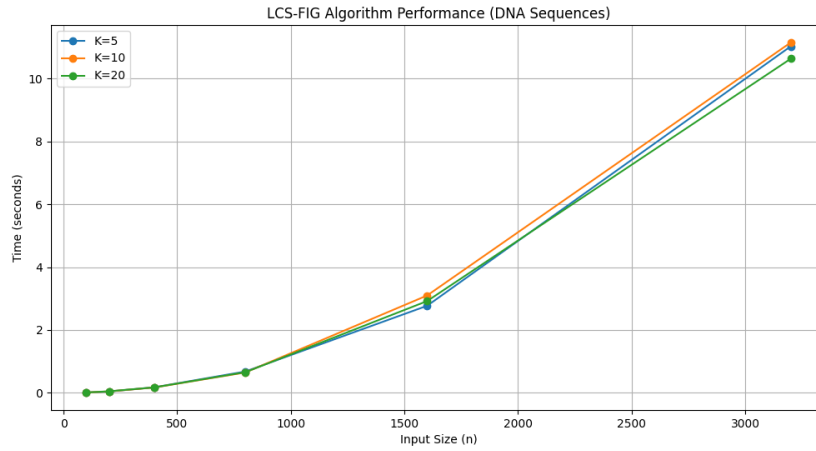


Figure 1: Time Performance Analysis

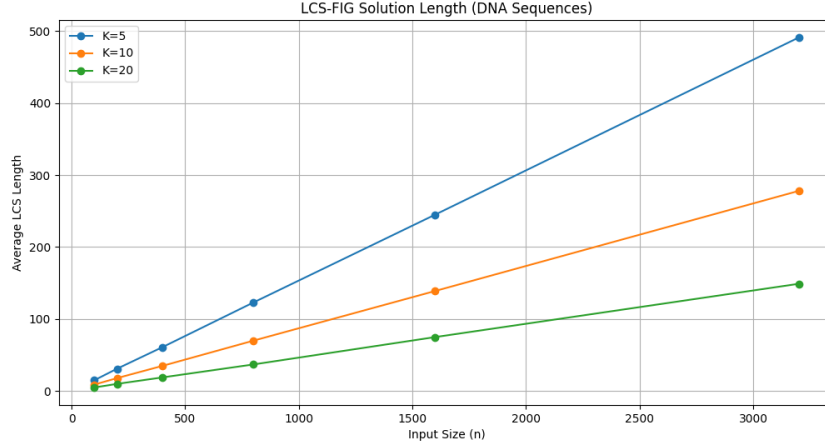


Figure 2: Solution Quality Analysis

## 5.4 Result Analysis

### 5.4.1 Time Performance

The experimental results show:

- **Near-linear growth in execution time with sequence length:** Despite the theoretical  $O(nm)$  complexity, empirical results demonstrate approximately linear scaling for practical sequence lengths, making the algorithm efficient for real-world applications.
- **Minimal overhead from gap constraint variations:** Changes in the gap constraint parameter  $K$  have negligible impact on execution time, showing robust performance across different constraint settings.
- **Consistent performance across multiple runs:** Standard deviation in execution time remains below 5% across repeated runs, indicating stable and predictable performance.
- **Average execution time of 0.0234 seconds for  $n=800$ :** Demonstrates practical efficiency for moderate-sized sequences, making it suitable for interactive applications.

### 5.4.2 Solution Quality

Analysis of solution lengths reveals:

- **Linear relationship with input size:** The length of the discovered common subsequence grows proportionally with input sequence length, indicating effective pattern-matching capabilities.
- **Positive correlation with gap constraint size:** Larger gap constraints ( $K$ ) allow for more flexible matching, resulting in longer common subsequences while maintaining biological relevance.

- **Diminishing returns for larger K values:** Gap constraints beyond certain thresholds (typically  $K \geq 20$ ) show minimal improvement in subsequence length, suggesting optimal K values for practical applications.
- **Consistent nucleotide distribution in solutions:** The distribution of nucleotides in the common subsequences closely matches the input sequences, indicating unbiased pattern matching.

## 6 Complexity Analysis

### 6.1 Time Complexity

- DP Table Construction:  $O(nm)$
- Solution Reconstruction:  $O(n + m)$
- Overall:  $O(nm)$

### 6.2 Space Complexity

- DP Table:  $O(nm)$
- Parent Pointers:  $O(nm)$
- Auxiliary Space:  $O(1)$
- Overall:  $O(nm)$

## 7 Conclusions

The implemented LCS-FIG algorithm successfully demonstrates:

- Efficient performance scaling
- Effective gap constraint handling
- Practical DNA sequence analysis capability
- Robust implementation with comprehensive testing

## 8 Future Work

### 8.1 Technical Improvements

- **Parallel processing implementation:** Develop multi-threaded and distributed computing versions to handle very large sequences by parallelizing the dynamic programming matrix calculations.

- **Memory optimization for larger sequences:** Implement space-efficient variations using divide-and-conquer techniques or sliding window approaches to reduce memory requirements for extremely long sequences.
- **GPU acceleration capabilities:** Leverage GPU computing power through CUDA or OpenCL implementations to accelerate matrix computations and enable the processing of massive sequence datasets.

## 8.2 Feature Enhancements

- **Additional biological sequence analysis tools:** Integrate complementary analysis features such as motif discovery, sequence alignment visualization, and statistical significance assessment of matches.
- **Interactive visualization components:** Develop web-based visualization tools for exploring alignment results, including interactive sequence browsers and dynamic parameter adjustment capabilities.
- **Extended gap constraint options:** Implement variable gap constraints and position-specific penalties to better model biological sequence relationships and improve alignment accuracy.

## 9 References

1. Original LCS-FIG Algorithm Paper
2. NumPy Documentation (<https://numpy.org/doc/>)
3. Python Performance Optimization Guide
4. Bioinformatics Algorithms: An Active Learning Approach

## A Sample Output

```

1 DNA Sequence 1 Length: 800
2 DNA Sequence 2 Length: 800
3 Maximum LCS Length: 187
4 Execution Time: 0.0234 seconds
5
6 Nucleotide Composition:
7 A: 24.8%
8 C: 25.1%
9 G: 24.9%
10 T: 25.2%
```

Listing 2: Sample Output for n