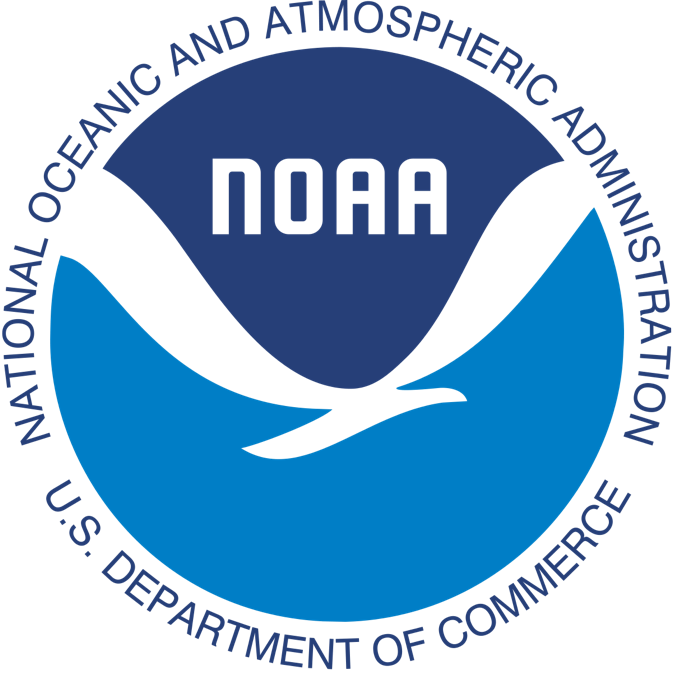
# Zymo Quick DNA Seed/Plant Miniprep Kit NOAA Protocol



* D6020

Protocol Comments

* All steps room temperature

1. Add up to 150mg of finely cut sample to a **ZR Bashing Bead Lysis Tube (2.0mm)** and add **750µl Bashing Bead Buffer**
2. Secure in bead beater and process max speed

* Fastprep – ~ 5 minutes
* Vortex attachment – 20 minutes

1. Centrifuge at >10,000g for 1 minute
2. Transfer up to 400µl supernatant to **Zymo-spin III-F Filter** in a collection tube
3. Centrifuge 8,000g for 1 minute and discard the **Zymo-spin III-F filter**
4. Add **1,200µl of** **Genomic Lysis Buffer** to the filtrate in the collection tube and mix well
5. Transfer 800µl (i.e., half) to **Zymo-spin IICR column** in a collection tube and centrifuge at 10,000g for 1 minute
   * **Zymo-spin IICR column** has max capacity 800µl
6. Empty flow through
7. Transfer **remaining 800µl** from Step 6 to the same **Zymo-spin IICR spin column** and centrifuge at 10,000g for 1 minute
8. Discard collection with flow through
9. Add **200µl DNA Pre-Wash Buffer** to the spin column with a new collection tube and centrifuge at 10,000g for 1 minute
10. Add **500µl g-DNA Wash Buffer** to the spin column and centrifuge at 10,000g for 1 minute
11. Transfer spin column to a clean 1.5ml microcentrifuge tube
12. Add **100µl DNA Elution Buffer** directly to the column matrix and centrifuge at 8,000g for 3 minutes
    * Can use water here instead of Elution Buffer
    * If using water, heat to 50-70oC and re-elute (i.e. run through same column again).
13. Place **Zymo-Spin III-HRC Filter** in a clean collection tube and add **600µl Prep Solution**. Centrifuge at 8,000g for 3 minutes
    * Matrix may seem dehydrated or powdery before adding prep solution, this is normal.
14. Transfer eluted DNA to prepared **Zymo-Spin III-HRC spin filter** in a clean 1.5ml microcentrifuge tube and centrifuge at EXACTLY 16,000g for 3 minutes.

Eluted DNA is ready for PCR and downstream applications.