

# KinPFN: Bayesian Approximation of RNA Folding Kinetics using Prior-Data Fitted Networks

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## Summary

**RNA folding kinetics** describe the probabilistic dynamics of the RNA folding process.

**RNA folding times** allow to analyse the folding efficiency with applications in synthetic biology and candidate selection for drug discovery.

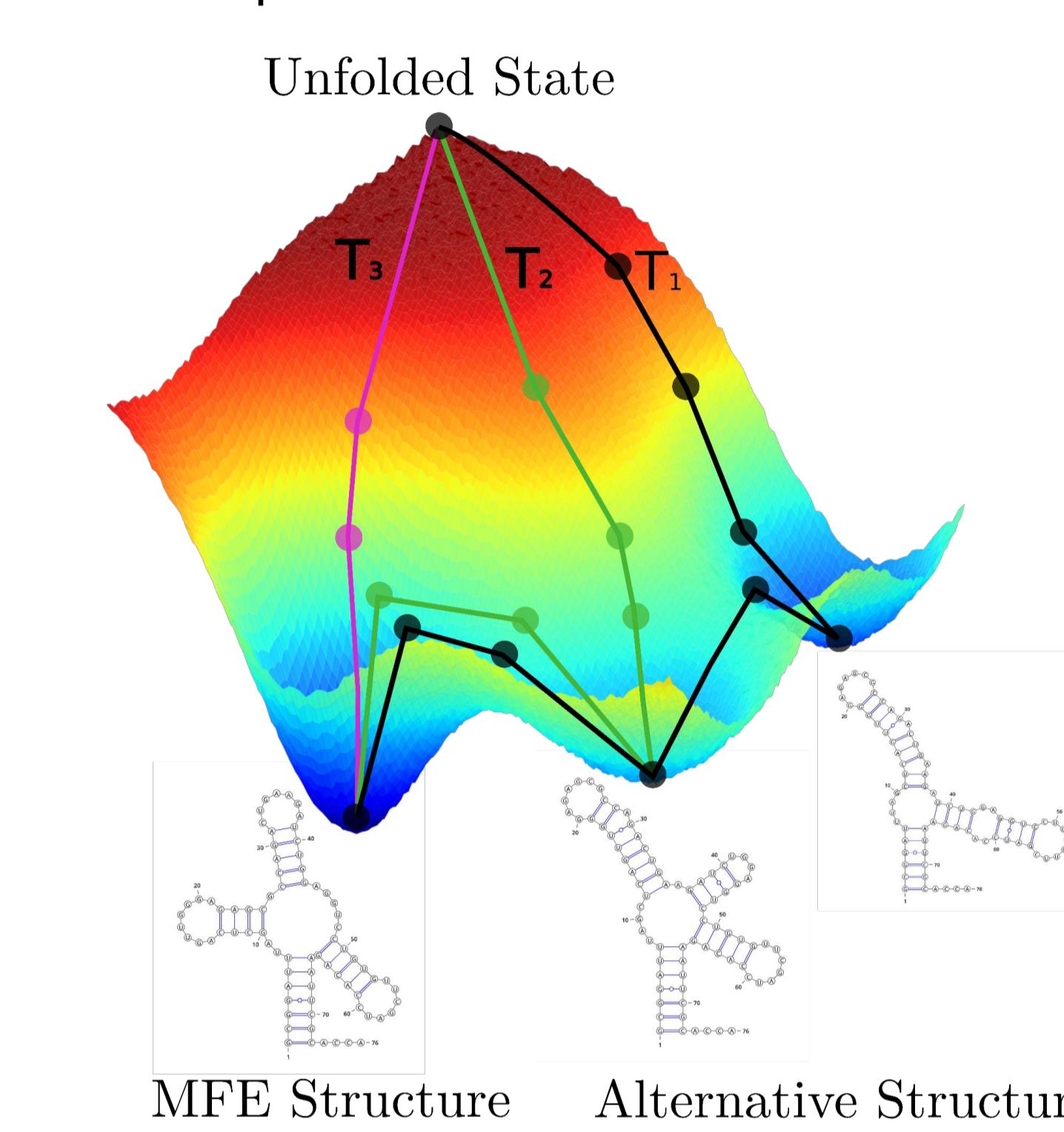
Problem: Current **RNA kinetics simulators** are costly and scale exponentially with the RNA length.

We present **KinPFN**, a novel approach for RNA folding kinetics based on prior-data fitted networks (PFNs) [1, 2].

Trained on a **synthetic prior representing RNA folding times**, KinPFN achieves comparable results while reducing simulator costs by ≥95%.

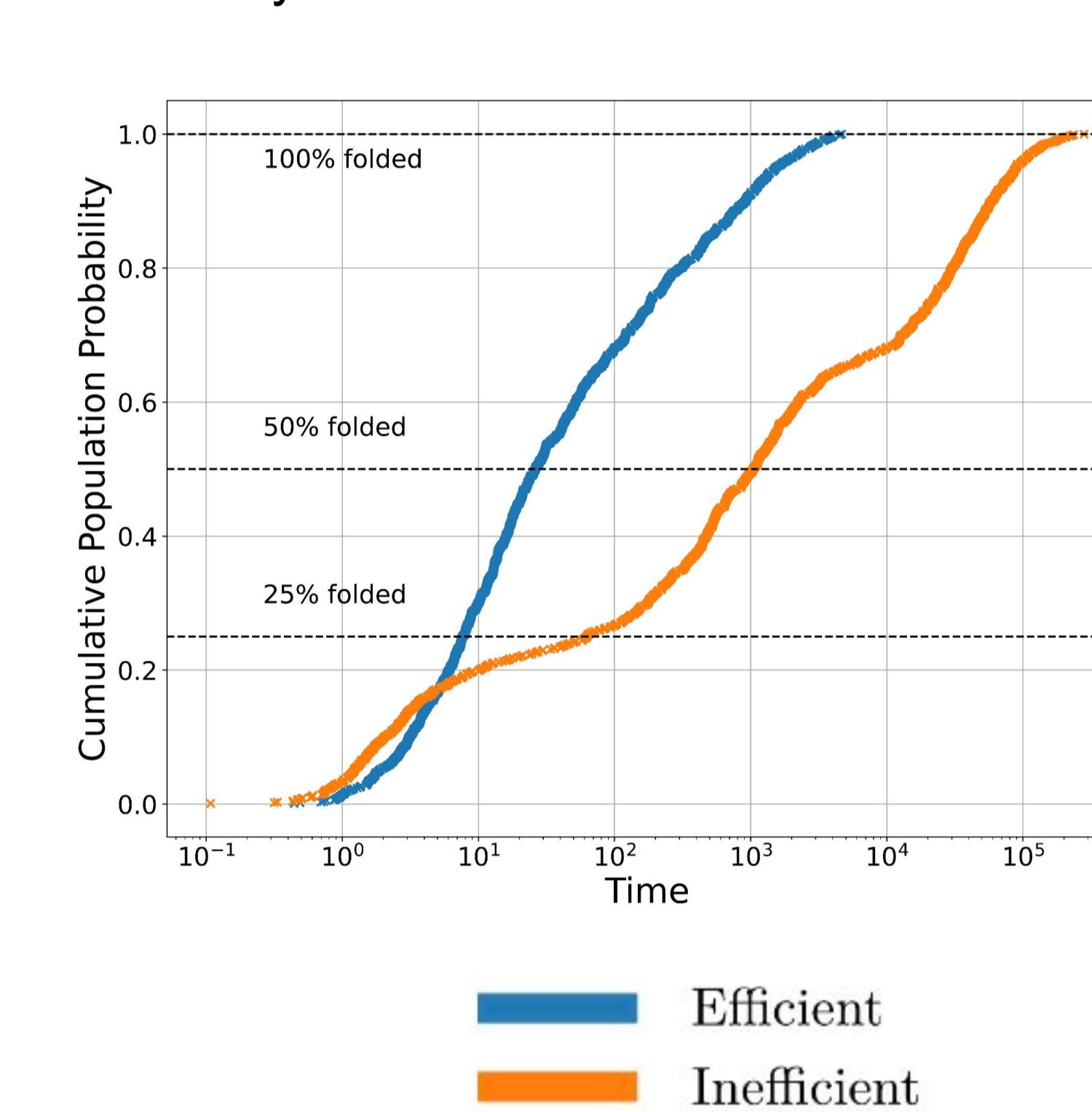
## Background: RNA Folding Kinetics

During the folding process, RNA traverses through a series of intermediate structural states, with each transition occurring at variable rates that collectively influence the time required to reach the functional form.

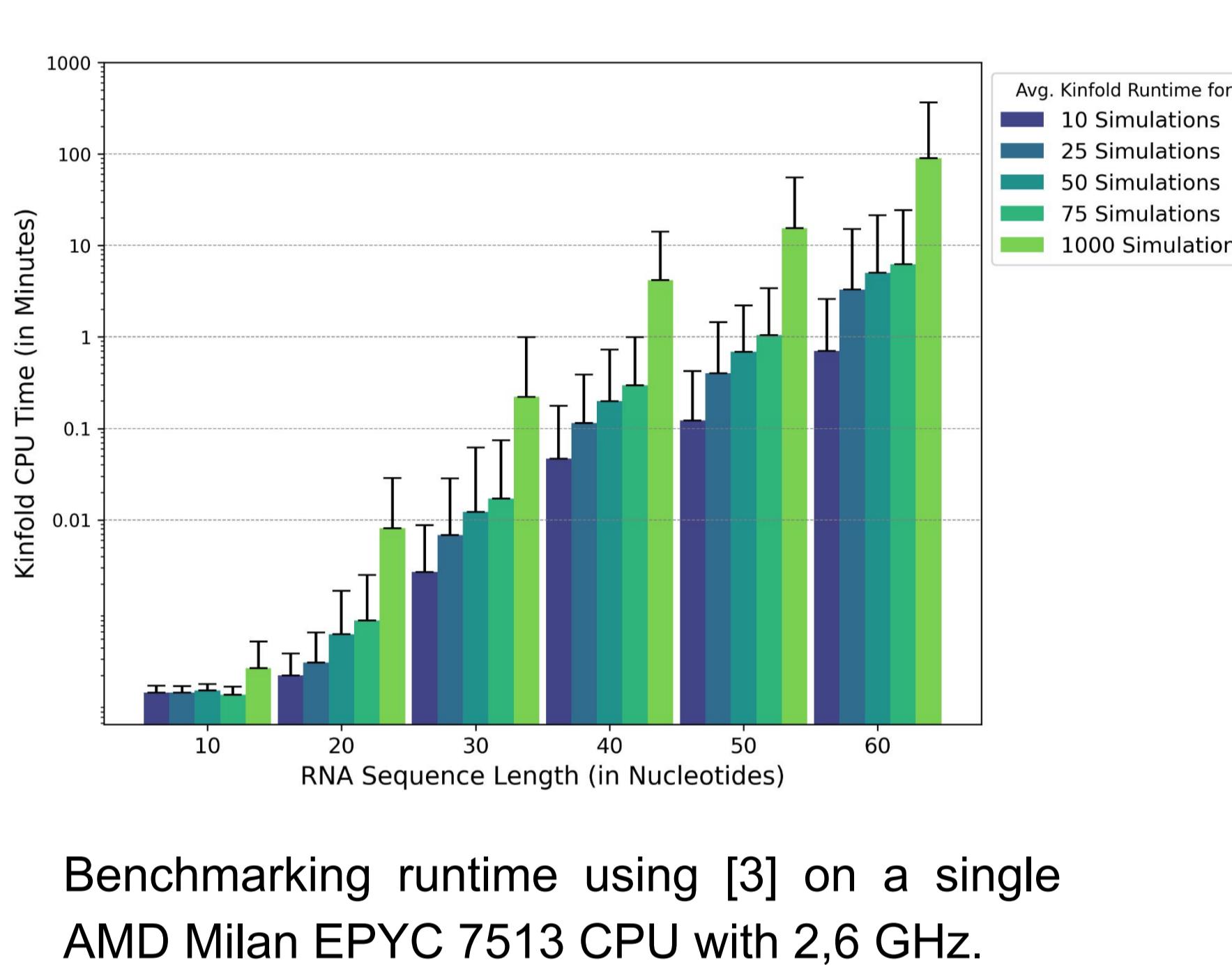


## Background: RNA Folding Kinetics

RNA Folding Times, the time required to fold into the structural form, allow to analyse the folding efficiency with applications in synthetic biology and drug discovery.

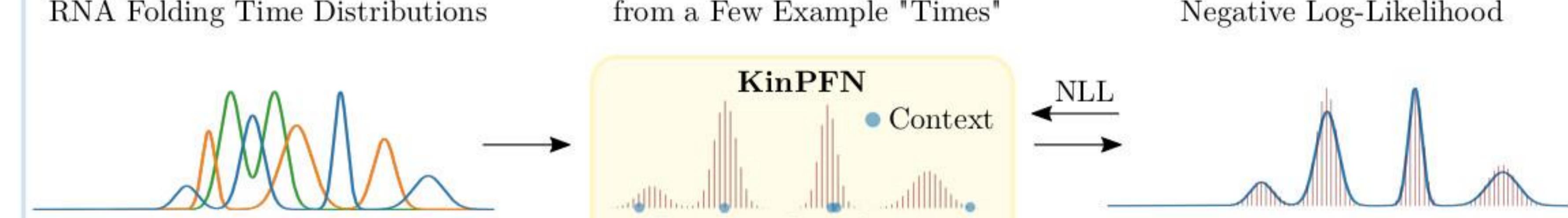


Problem: Existing RNA folding kinetics simulators are costly and scale exponentially with the RNA length, which makes them inapplicable to applications such as kinetic RNA design.



## Our Approach: KinPFN

**a Training on a Synthetic Prior**  
Synthetic Datasets are Drawn from Multi-Modal Gaussians to Mimic RNA Folding Time Distributions

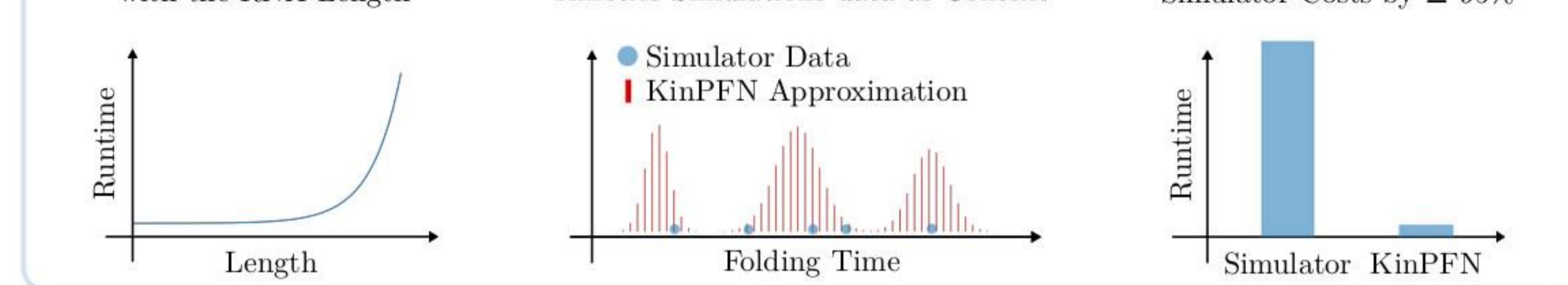


KinPFN is Trained on ~5M Distributions by Minimizing the Negative Log-Likelihood

## b Application

RNA Kinetics Simulators are Costly and Scale Exponentially with the RNA Length

KinPFN Approximates the Distribution of Folding Times Using Kinetics Simulations data as Context



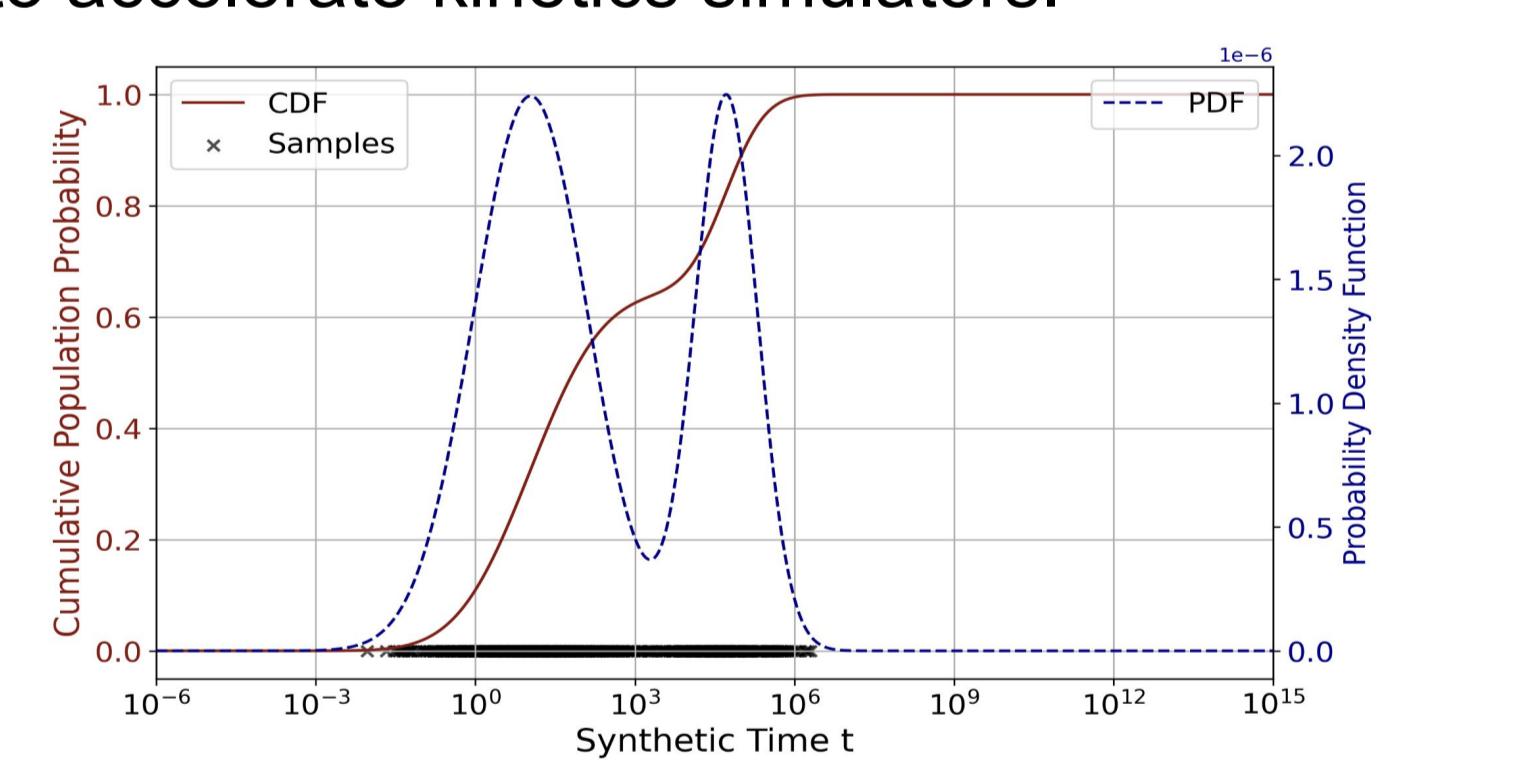
## Synthetic RNA Folding Time Prior

### Challenges:

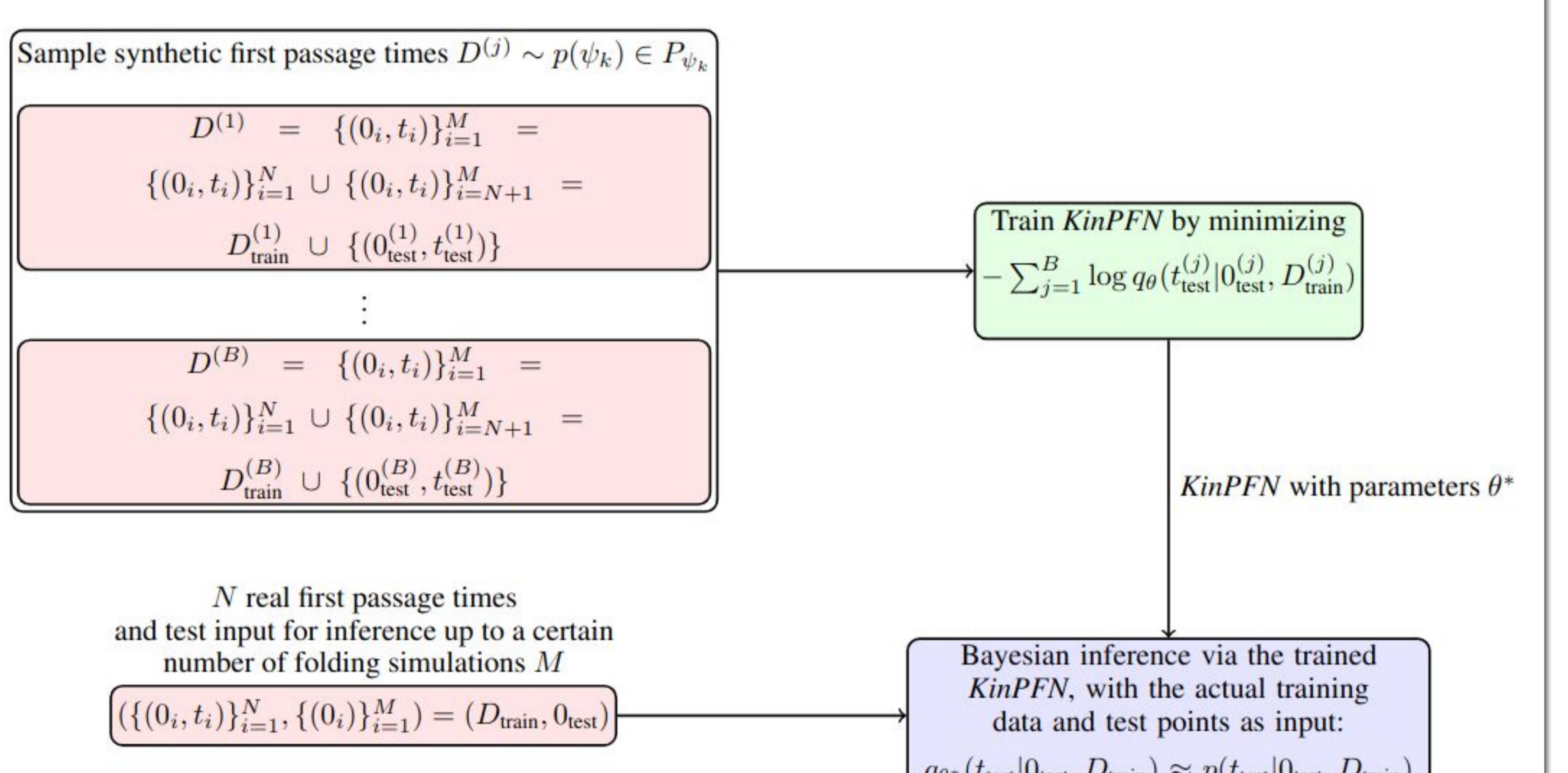
- RNA kinetics data is rare due to exponential costs of kinetic simulators.
- We have no access to the combination of RNA folding times and specific features like sequence or energy.

### Approach:

- We train on synthetic datasets drawn from parameterized multi-modal Gaussians representing RNA folding time distributions.
- We leverage in-context learning at test time to accelerate kinetics simulators.



## Training on the Synthetic Prior



## From Synthetic to Real Data

**Setup:** Use Simulator Data as Context.

**Data:** 635 randomly generated RNA sequences with 1000 Simulations from [3].

**Results:** Strong approximation performance of KinPFN across varying context sizes.

Method	First Passage Times N				
	10	25	50	75	100
KinPFN	1.3739	1.2435	1.2047	1.1916	1.1858
GMM <sub>2</sub>	2.3122	1.3612	1.2355	1.2036	1.1933
GMM <sub>3</sub>	5.2469	1.5830	1.2838	1.2132	1.1910
GMM <sub>4</sub>	13.1325	1.9922	1.3676	1.2480	1.2119
GMM <sub>5</sub>	37.5845	2.7708	1.4957	1.2953	1.2374
DP-GMM <sub>2</sub>	1.6285	1.3529	1.2618	1.2305	1.2150
DP-GMM <sub>3</sub>	1.6268	1.3549	1.2653	1.2323	1.2155
DP-GMM <sub>4</sub>	1.6294	1.3558	1.2663	1.2337	1.2169
DP-GMM <sub>5</sub>	1.6256	1.3572	1.2675	1.2337	1.2175
KDE	1.4370	1.2559	1.2133	1.2003	1.1957

## Performance is Independent of RNA Features and Kinetics Simulators

**Setup:** Use kinetics simulations from [4].

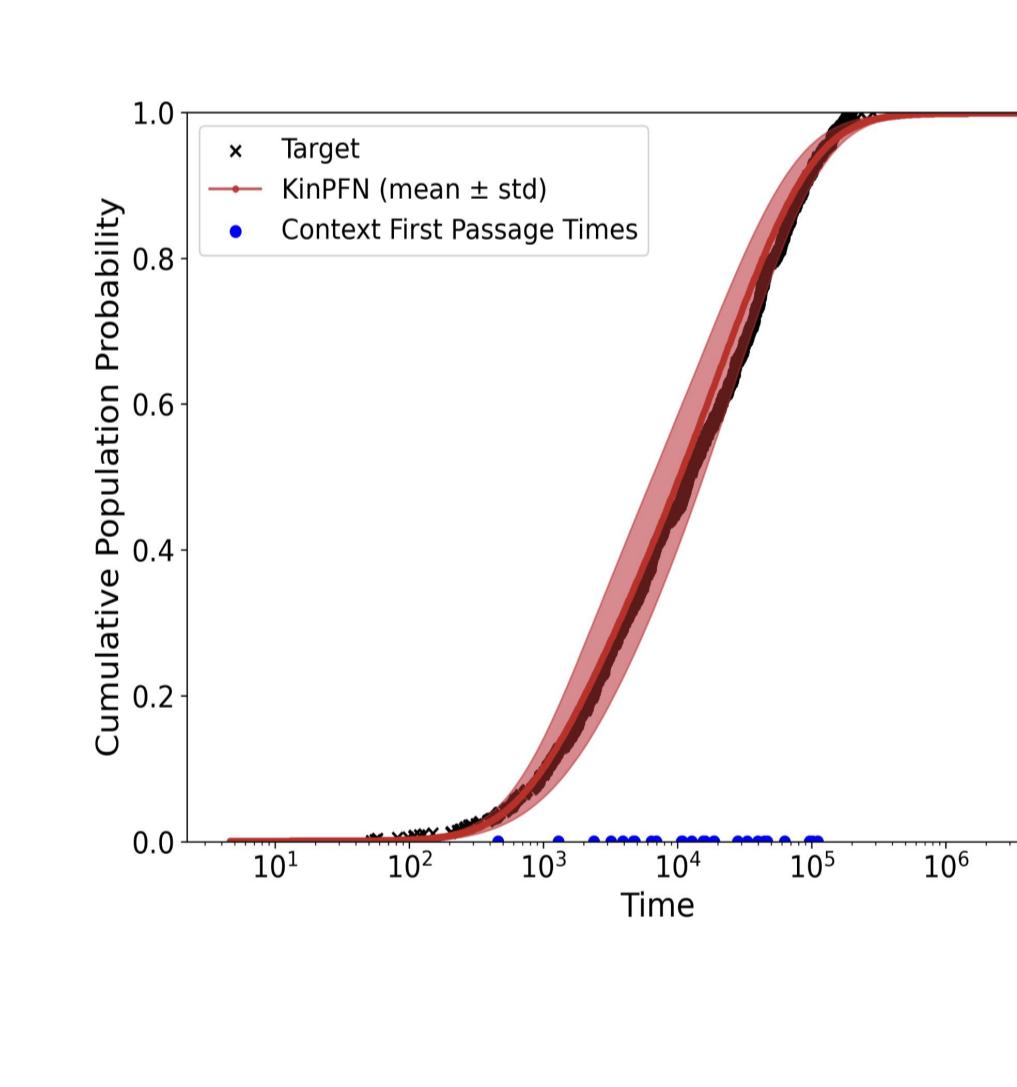
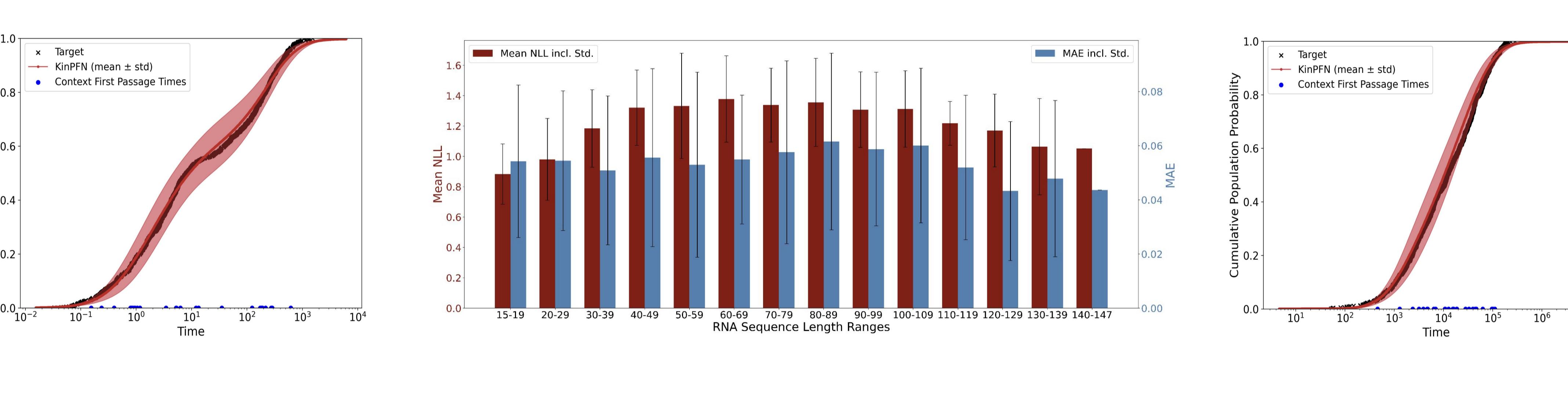
**Results:** KinPFN approximation is independent of the simulator.

**Setup:** Analyze performance across RNA sequence lengths.

**Results:** KinPFN performance is constant across sequence lengths.

**Setup:** Analyze performance with different start and stop structures.

**Results:** KinPFN is independent of the start and stop structure.



## Case Study: Kinetics of Natural RNAs

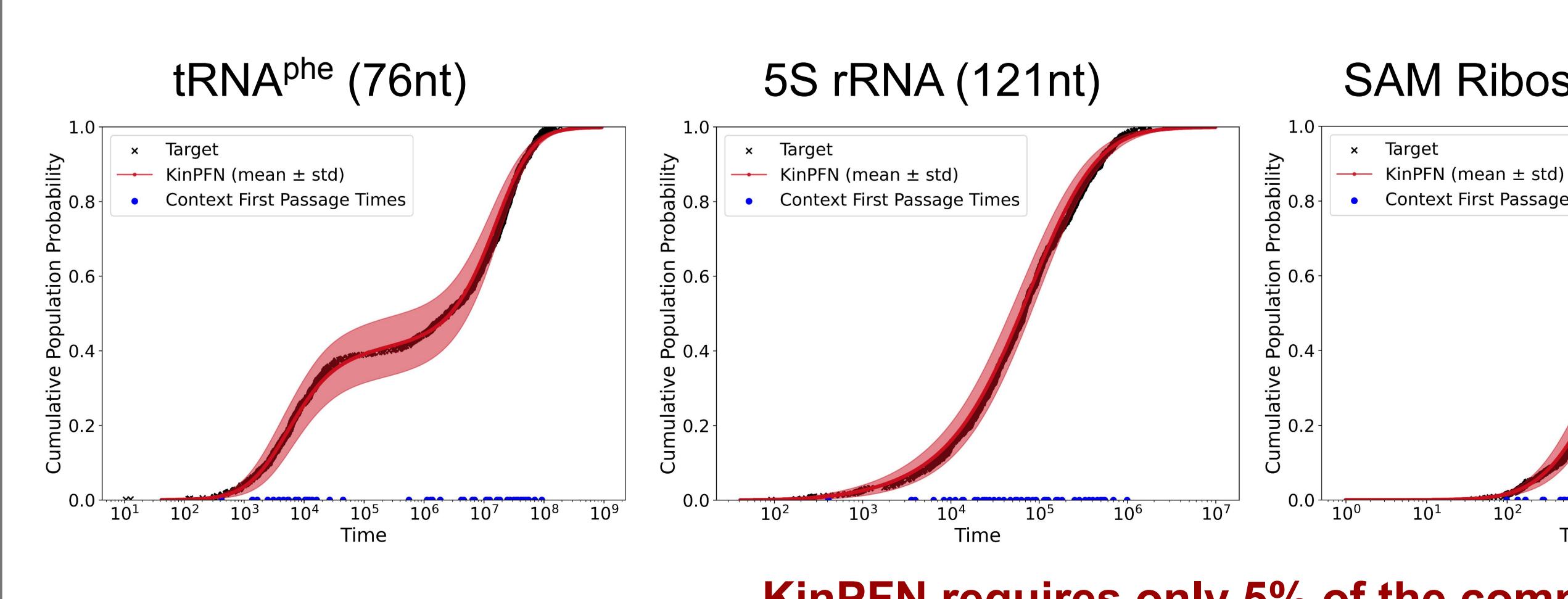
**Setup:**  
We use 50 context RNA folding times from 1,000 simulations of [3].

### Data:

Four natural RNAs: tRNA<sup>phe</sup>, 5S rRNA (both *S. cerevisiae*), SAM Riboswitch (*B. subtilis*), micro RNA (*H. sapiens*).

### Results:

95% runtime improvement (~2 days → ~3 hours) with minimal accuracy loss.

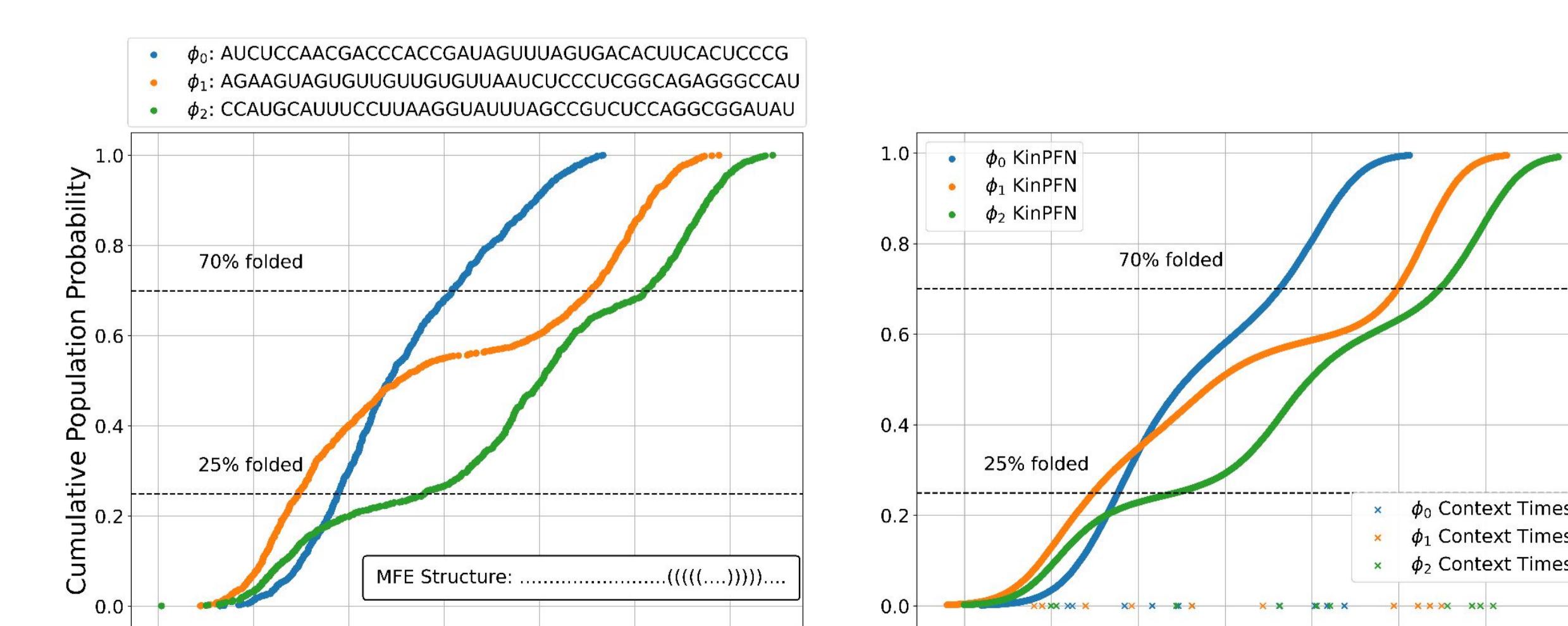


## Case Study: Folding Efficiency Analysis

**Setup:** Compare the folding efficiency of three 43nt RNAs ( $\phi_0, \phi_1, \phi_2$ ) with the same minimum free energy (MFE) structure.

**Data:** 10 context RNA folding times from 1,000 simulations of [3] for each RNA.

**Results:** 100× speed-up per RNA at comparable performance.

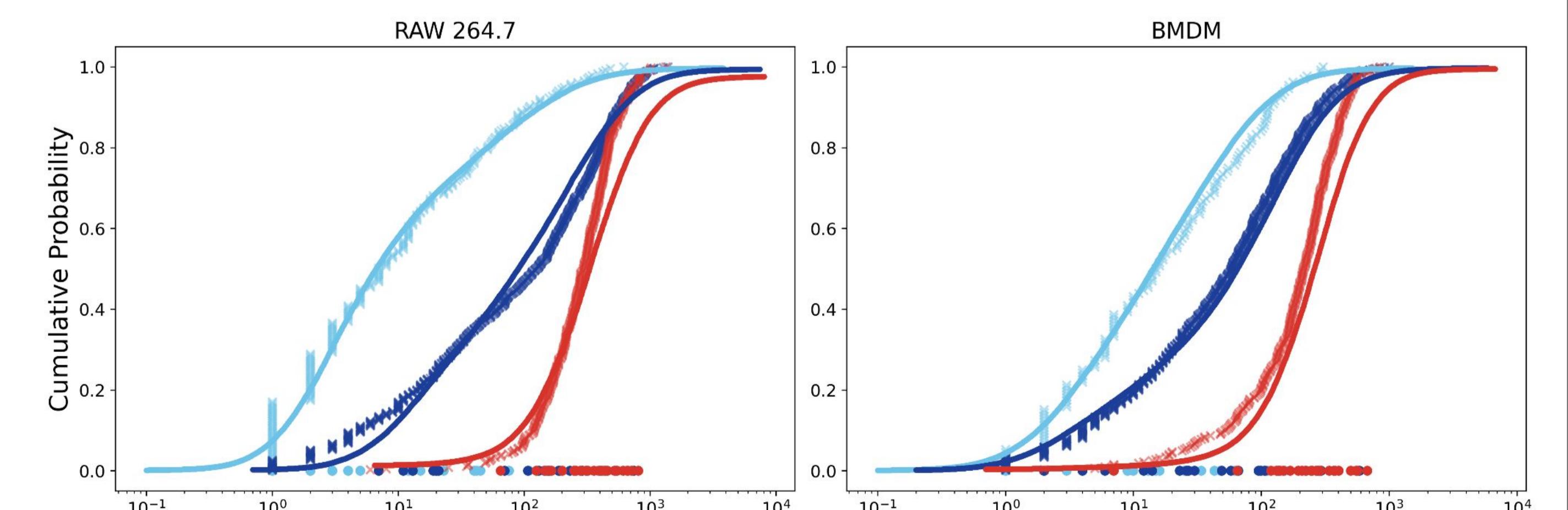


## Generalization to Gene Expression Data

**Setup:** Evaluate KinPFN generalization performance by using data from different biological contexts.

**Data:** smFISH counts from [5] for the expression of Interleukin-1 (IL-1α, IL-1β) and tumor necrosis factor alpha (TNF-α) mRNA in two immune cell lines, established RAW 264.7 macrophage cells and bone-marrow-derived macrophages (BMDM) stimulated with Lipid A.

**Results:** KinPFN achieves accurate approximations of the gene expression using only 8% of the count data.



## References

- [1] Müller, S., Hollmann, N., Arango, S. P., Grabocka, J., & Hutter, F. Transformers Can Do Bayesian Inference. In *International Conference on Learning Representations 2022*.
- [2] Adriaensen, S., Rakotomaro, H., Müller, S., & Hutter, F. (2023). Efficient bayesian learning curve extrapolation using prior-data fitted networks. *Advances in Neural Information Processing Systems*, 36, 19858-19886.
- [3] Flamm, C., Fontana, W., Hofacker, I. L., & Schuster, P. (2000). RNA folding at elementary step resolution. *Rna*, 6(3), 325-339.
- [4] Dykeman, E. C. (2015). An implementation of the Gillespie algorithm for RNA kinetics with logarithmic time update. *Nucleic acids research*, 43(12), 5708-5715.
- [5] Bagnall, J., Rowe, W., Alachkar, N., Roberts, J., England, H., Clark, C., ... & Paszek, P. (2020). Gene-specific linear trends constrain transcriptional variability of the toll-like receptor signaling. *Cell Systems*, 11(3), 300-314.

