



BiomedicalInformatics

3184 Graves Hall 333 W. 10th Ave. Columbus, OH 43210 614-292-4778 (phone) 614-292-7659 (fax)

Statistical Microarray Data Analysis

Review of Microarray

Elements of Gene Expression Data Analysis

- Comparative study
- Clustering

Introduction to Pathway and Gene Ontology Enrichment Analysis

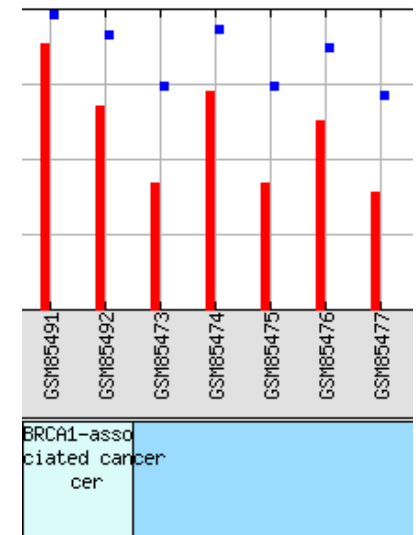
Review of Microarray

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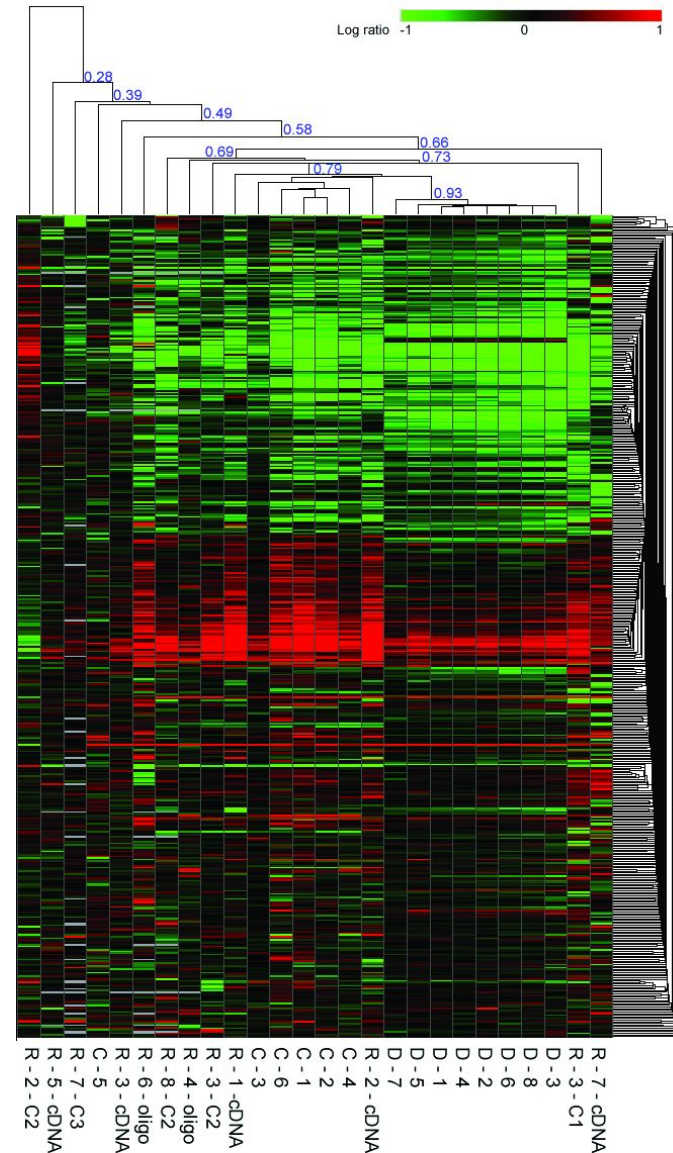
Introduction to Pathway and Gene Ontology Enrichment Analysis

Gene 1	10
Gene 2	500
Gene 3	30
Gene 4	100
Gene 5	20
Gene 6	28
Gene 7	7
...	...



How do we use microarray?

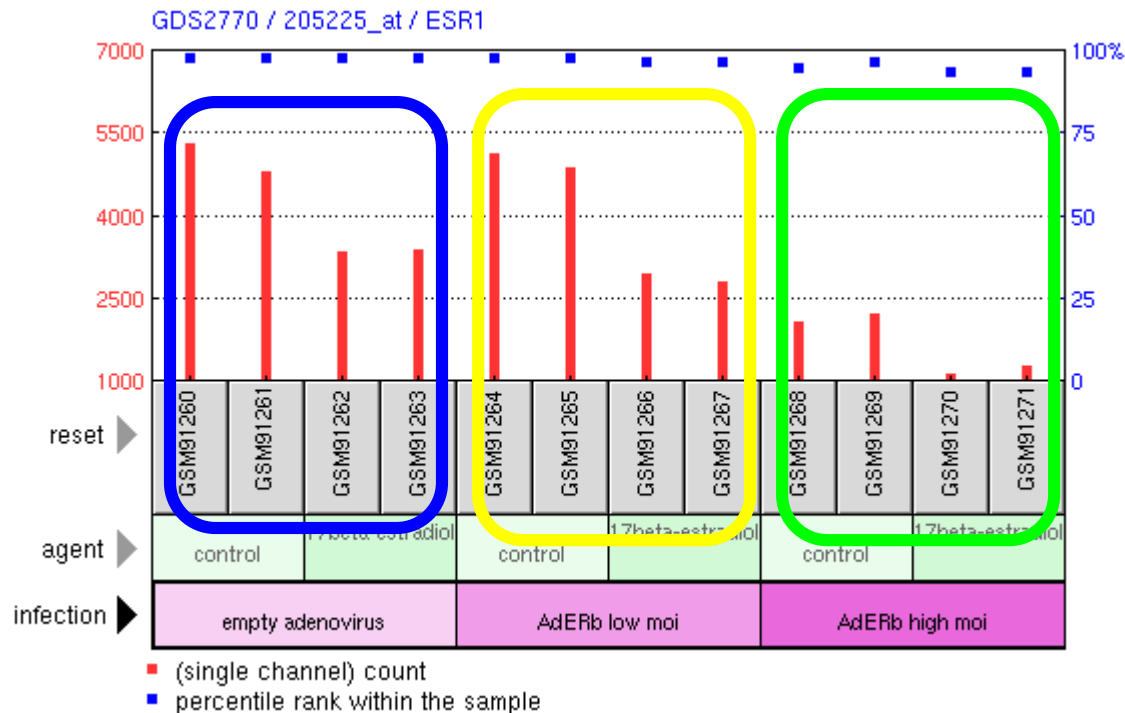
- Profiling
- Comparative study
- Clustering
- Inference



Supplementary Figure 1: Clustering of laboratory/platform combinations using log ratio values of common genes

Hypothesis Testing

- Two set of samples sampled from two distributions (N=2)



Hypothesis Testing

- Two set of samples sampled from two distributions (N=2)
- Hypothesis

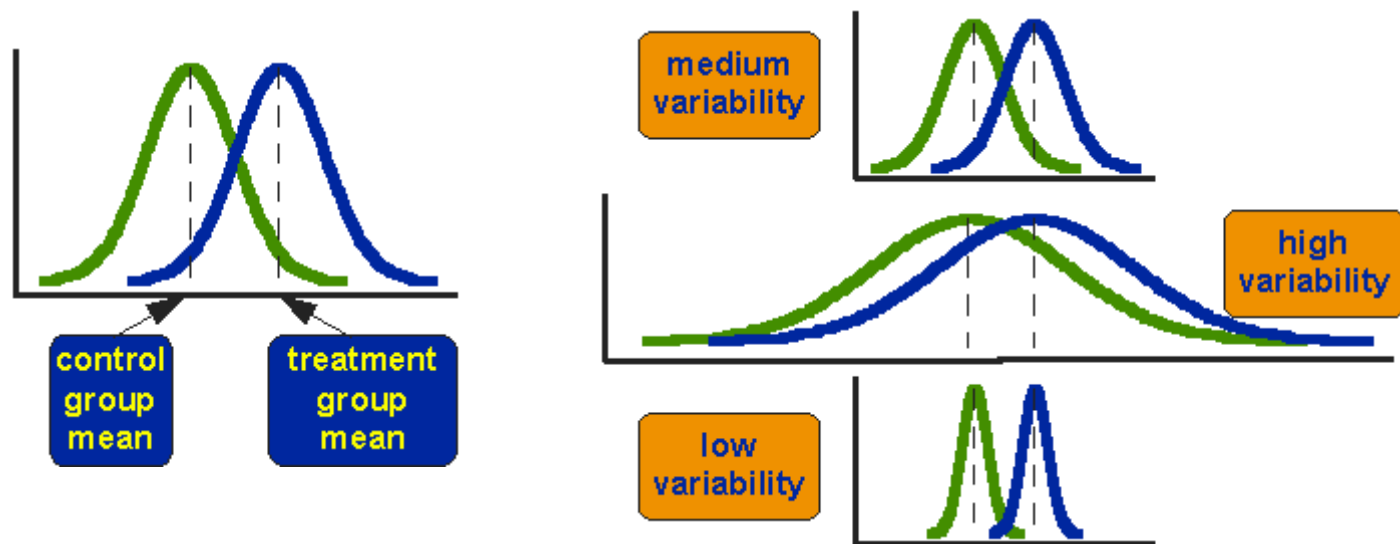
$H_0 : \mu_1 - \mu_2 = 0$ Null hypothesis

$H_1 : \mu_1 - \mu_2 \neq 0$ Alternative hypothesis

μ_1 and μ_2 are the means of the two distributions.

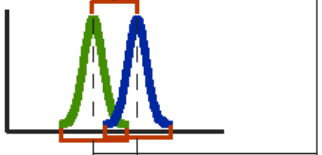
Statistical Decision	True State of the Null Hypothesis	
	H_0 True	H_0 False
Reject H_0	Type I error	Correct
Do not Reject H_0	Correct	Type II error

Student's t-test



$$\begin{aligned} \frac{\text{signal}}{\text{noise}} &= \frac{\text{difference between group means}}{\text{variability of groups}} \\ &= \frac{\bar{X}_T - \bar{X}_C}{SE(\bar{X}_T - \bar{X}_C)} \\ &= \text{t-value} \end{aligned}$$

Student's t-test

$$\begin{aligned} \frac{\text{signal}}{\text{noise}} &= \frac{\text{difference between group means}}{\text{variability of groups}} \\ &= \frac{\bar{X}_T - \bar{X}_C}{SE(\bar{X}_T - \bar{X}_C)} \\ &= \text{t-value} \end{aligned}$$


The diagram shows two overlapping normal distributions, one green and one blue, on a horizontal axis. The green distribution is shifted to the left of the blue distribution. A vertical dashed line marks the center of the blue distribution. A horizontal bracket with a vertical line at its center is positioned below the x-axis, spanning the width of the green distribution. An arrow points from the text 't-value' to the green distribution, and another arrow points from the text 'SE(\bar{X}_T - \bar{X}_C)' to the horizontal bracket.

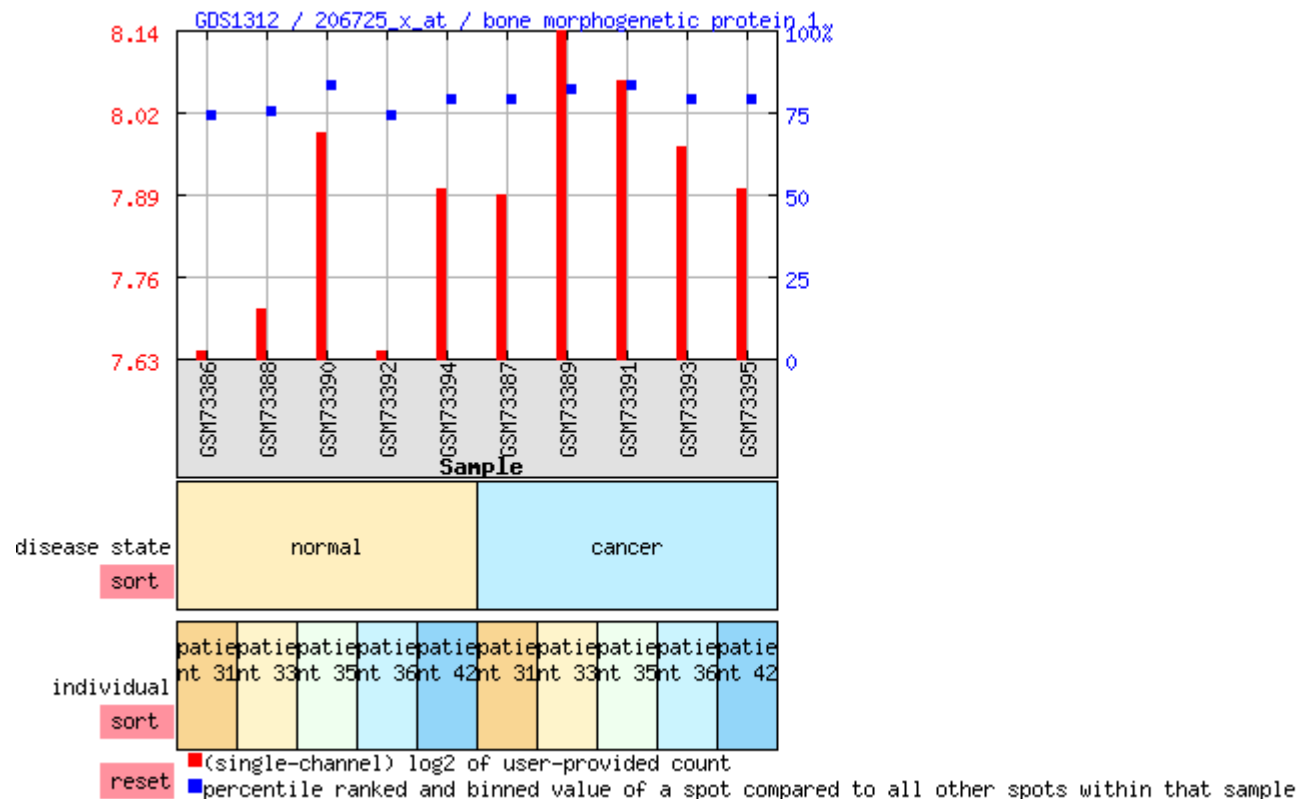
$$t = \frac{\bar{X}_T - \bar{X}_C}{\sqrt{\frac{\text{var}_T}{n_T} + \frac{\text{var}_C}{n_C}}}$$

p-value can be computed from t-value and number of freedom (related to number of samples) to give a bound on the probability for type-I error (claiming insignificant difference to be significant) assuming normal distributions.

Student's t-test

- Dependent (paired) t-test

$$t = \frac{\sum D_i}{\sqrt{\frac{n \sum D_i^2 - (\sum D_i)^2}{n-1}}} \quad D_i = X_2^i - X_1^i$$



Permutation (t-)test

T-test relies on the parametric distribution assumption (normal distribution). Permutation tests do not depend on such an assumption. Examples include the permutation t-test and Wilcoxon rank-sum test.

Perform regular t-test to obtain t-value t_0 . Then randomly permute the $N_1 + N_2$ samples and designate the first N_1 as group 1 with the rest being group 2. Perform t-test again and record the t-value t . For all possible $K = \binom{N_1 + N_2}{N_1}$ permutations, count how many t-values are larger than t_0 and write down the number K_0 .

$$p = \frac{1 + K_0}{1 + K}$$

Multiple Classes ($N > 2$)

F-test

- The null hypothesis is that the distribution of gene expression is the same for all classes.
- The alternative hypothesis is that at least one of the classes has a distribution that is different from the other classes.
- Which class is different cannot be determined in F-test (ANOVA). It can only be identified post hoc.

Example

- GEO Dataset Subgroup Effect

Gene Discovery and Multiple T-tests

Controlling False Positives

- p-value cutoff = 0.05 (probability for false positive - type-I error)
- 22,000 probesets
- False discovery $22,000 \times 0.05 = 1,100$
- Focus on the 1,100 genes in the second specimen. False discovery $1,100 \times 0.05 = 55$

Gene Discovery and Multiple T-tests

Controlling False Positives

- State the set of genes explicitly before the experiments
 - Problem: not always feasible, defeat the purpose of large scale screening, could miss important discovery
- Statistical tests to control the false positives

Gene Discovery and Multiple T-tests

Controlling False Positives

- Statistical tests to control the false positives
 - Controlling for no false positives (very stringent, e.g. Bonferroni methods)
 - Controlling the number of false positives (
 - Controlling the proportion of false positives
 - Note that in the screening stage, false positive is better than false negative as the later means missing of possibly important discovery.

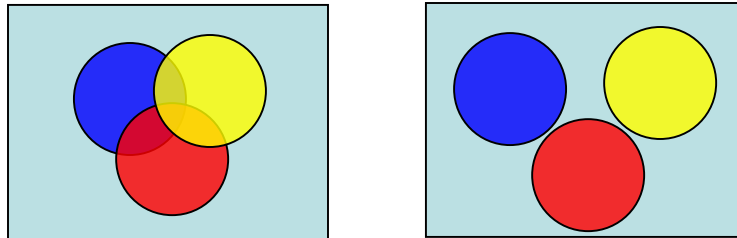
Gene Discovery and Multiple T-tests

Controlling False Positives

- Statistical tests to control the false positives
 - Controlling for no false positives (very stringent)
 - Bonferroni methods and multivariate permutation methods

Bonferroni inequality

$$Prob(E_1 \cup E_2 \cup \dots \cup E_K) \leq \sum_{i=1}^K Prob(E_i)$$



Area of union \leq Sum of areas

$$Prob(E_i) = 0.05, K = 20$$

$$Prob(E_1 \cup E_2 \cup \dots \cup E_K) \leq \sum_{i=1}^K Prob(E_i) = 1$$

Gene Discovery and Multiple T-tests

Bonferroni methods

- Bonferroni adjustment

$$Prob(E_i) = 0.05, K = 20$$

$$Prob(E_1 \cup E_2 \cup \dots \cup E_K) \leq \sum_{i=1}^K Prob(E_i) = 1$$

- If E_i is the event for false positive discovery of gene i , conservative speaking, it is almost guaranteed to have false positive for $K > 19$.
- So change the p-value cutoff line from p_0 to p_0/K . This is called **Bonferroni adjustment**.
- If $K=20$, $p_0=0.05$, we call a gene i is significantly differentially expressed if $p_i < 0.0025$.

Gene Discovery and Multiple T-tests

Bonferroni methods

- Bonferroni adjustment
- Too conservative. Excessive stringency leads to increased false negative (type II error).
- Has problem with metaanalysis.
- Variations: sequential Bonferroni test (Holm-Bonferroni test)
 - Sort the K p-values from small to large to get $p_1 \leq p_2 \leq \dots \leq p_K$.
 - So change the p-value cutoff line for the i th p-value to be $p_0/(K-i+1)$ (ie, $p_1 \leq p_0/K$, $p_2 \leq p_0/(K-1)$, ..., $p_K \leq p_0$).
 - If $p_j \leq p_0/(K-j+1)$ for all $j \leq i$ but $p_{i+1} > p_0/(K-i+1)$, reject all the alternative hypothesis from $i+1$ to K , but keep the hypothesis from 1 to i .

Gene Discovery and Multiple T-tests

Controlling False Positives

- Statistical tests to control the false positives
 - Controlling the number of false positives
 - Simple approach – choose a cutoff for p-values that are lower than the usual 0.05 but higher than that from Bonferroni adjustment
 - More sophisticated way: a version of multivariate permutation.

Gene Discovery and Multiple T-tests

Controlling False Positives

- Statistical tests to control the false positives
 - Controlling the proportion of false positives

Let γ be the portion (percentage) of false positive in the total discovered genes.

$$p_1 \leq p_2 \leq \cdots \leq p_D \leq \cdots \leq p_K$$

$$D = \arg \max (\underbrace{p_i \cdot K}_{\text{False positive}} / \underbrace{i}_{\text{Total positive}} < \gamma)$$

p_D is the choice. There are other ways for estimating false positives. Details can be found in Tusher et. al. PNAS 98:5116-5121.

Review of Microarray

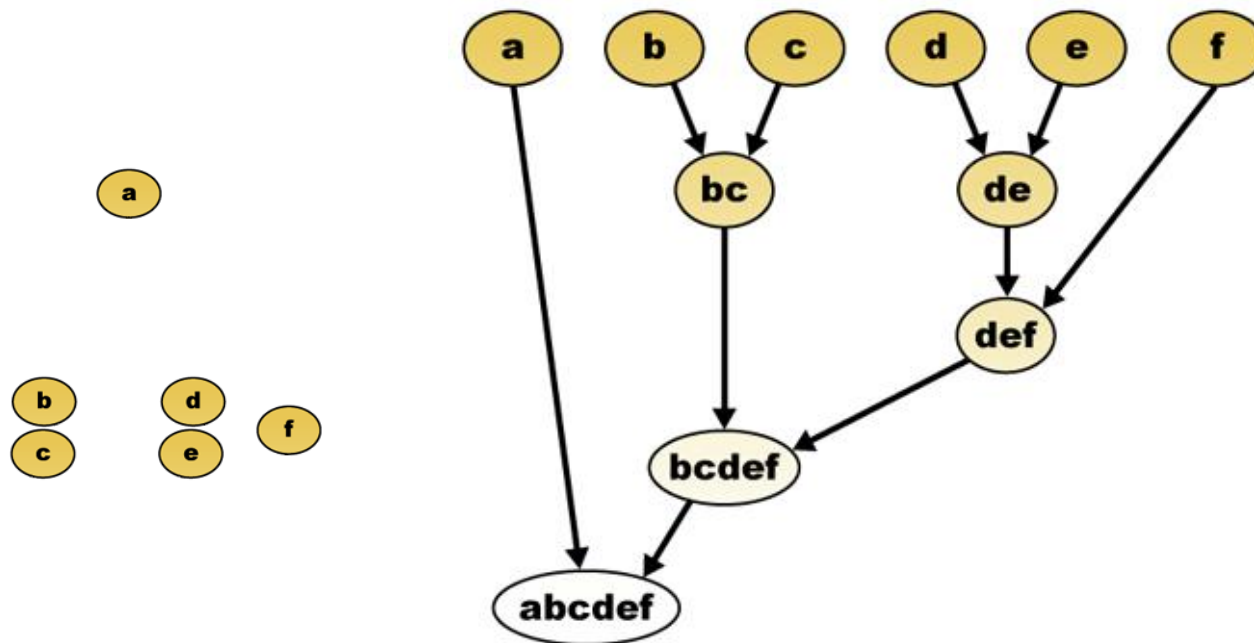
Elements of Gene Expression Data Analysis

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Introduction to Pathway and Gene Ontology Enrichment Analysis

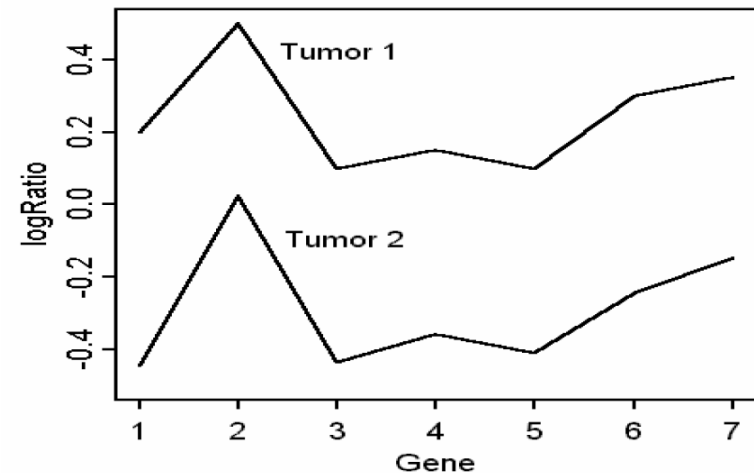
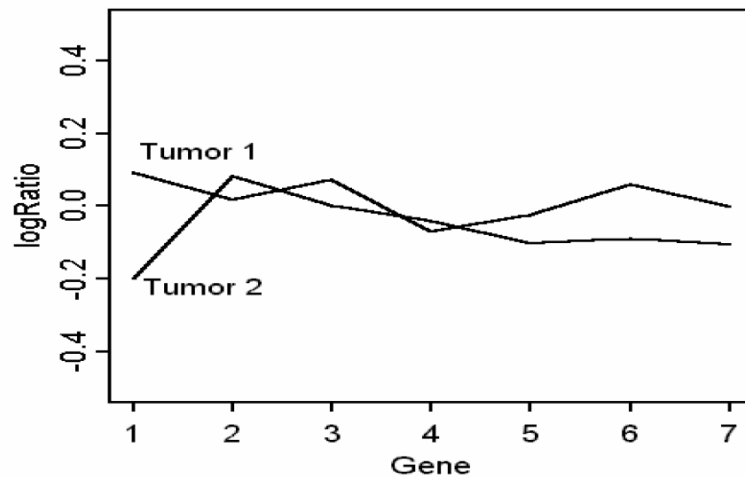
How do we process microarray data (clustering)?

– Unsupervised Learning – Hierarchical Clustering



Distance Measure (Metric?)

- What do you mean by “similar”?
- Euclidean
- Uncentered correlation
- Pearson correlation



Distance Metric

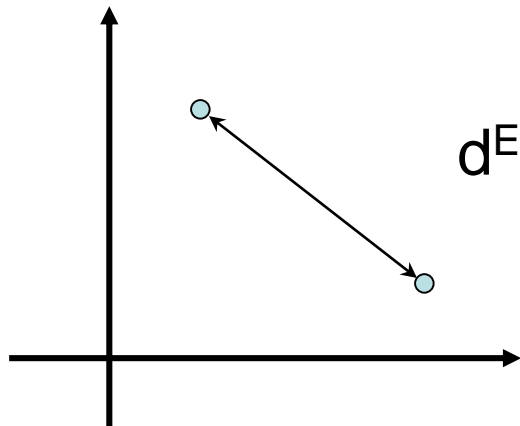
- Euclidean

$$\mathbf{x} = (x_1, x_2, \dots, x_n)^T$$

$$\mathbf{y} = (y_1, y_2, \dots, y_n)^T$$

$$d^E(\mathbf{x}, \mathbf{y}) = \sqrt{(x_1 - y_1)^2 + (x_2 - y_2)^2 + \dots + (x_n - y_n)^2}$$

102123_at	Lip1	1596.000	2040.900	1277.000	4090.500	1357.600	1039.200	1387.300
		3189.000	1321.300	2164.400	868.600	185.300	266.400	2527.800
160552_at	Ap1s1	4144.400	3986.900	3083.100	6105.900	3245.800	4468.400	7295.000
		5410.900	3162.100	4100.900	4603.200	6066.200	5505.800	5702.700



$$d^E(\text{Lip1}, \text{Ap1s1}) = 12883$$

Distance Metric

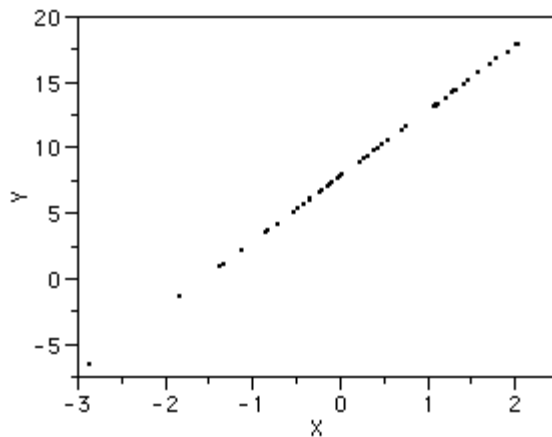
- Pearson Correlation

$$\mathbf{x} = (x_1, x_2, \dots, x_n)^T$$

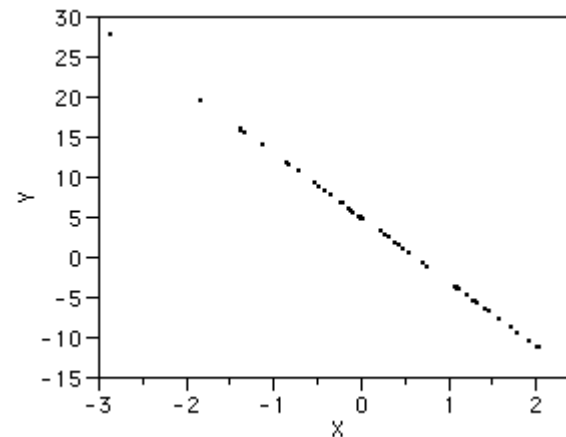
$$\mathbf{y} = (y_1, y_2, \dots, y_n)^T$$

$$r_{xy} = \frac{\sum_{i=1}^n (x_i - \bar{x})(y_i - \bar{y})}{\sqrt{\sum_{i=1}^n (x_i - \bar{x})^2} \sqrt{\sum_{i=1}^n (y_i - \bar{y})^2}}$$

Ranges from 1 to -1.



$r = 1$

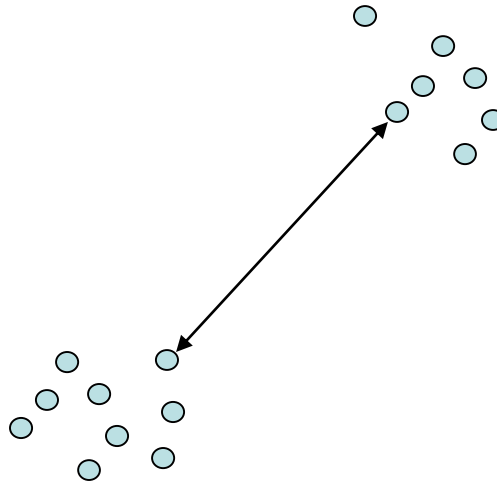


$r = -1$

How do we process microarray data (clustering)?

–Unsupervised Learning – Hierarchical Clustering

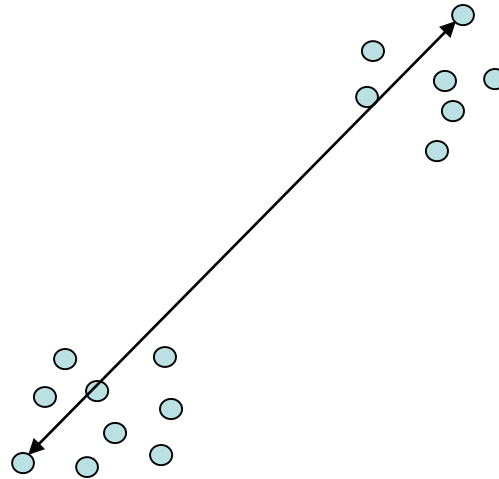
Single linkage: The linking distance is the minimum distance between two clusters.



How do we process microarray data (clustering)?

–Unsupervised Learning – Hierarchical Clustering

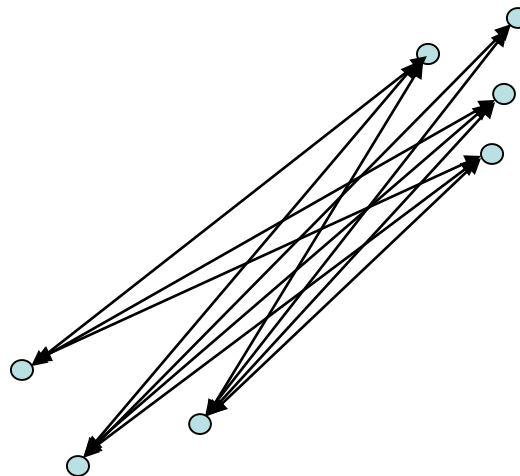
Complete linkage: The linking distance is the maximum distance between two clusters.



How do we process microarray data (clustering)?

–Unsupervised Learning – Hierarchical Clustering

Average linkage/UPGMA: The linking distance is the average of all pair-wise distances between members of the two clusters. Since all genes and samples carry equal weight, the linkage is an Unweighted Pair Group Method with Arithmetic Means (UPGMA).



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**Introduction to Pathway and Gene Ontology
Enrichment Analysis**

Where do I get the gene list?

- Comparative study
 - e.g., microarray experiments between two types of samples or two disease states (can also be from RT-PCR, proteomics, ...)
- Clustering / classification of genes
 - e.g., co-expressed genes
- Homologue analysis
 - e.g., genes from BLAST
- Other sources

What do I do with the gene list – *enrichment analysis*?

- Find commonality among the gene
 - Common **molecular functions (GO)**
 - Common **biological processes (GO)**
 - Common **cellular components (GO)**
 - Common pathways
 - Interact with common genes
 - Common sequences / molecular structures
 - Regulated by common Transcription Factors
 - Targeted by common microRNAs
 - Involved in the same disease
 - ...
- Generate new hypothesis based on the commonality

**GO
enrichment
analysis**



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The Gene Ontology project provides a controlled vocabulary to describe gene and gene product attributes in any organism. [Read more about the Gene Ontology...](#)

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The Gene Ontology Consortium is supported by a P41 grant from the National Human Genome Research Institute (NHGRI) [grant [HG002273](#)]. [See the full list of funding sources](#). The Gene Ontology Consortium would like to acknowledge the assistance of many more people than can be listed here. Please visit the [acknowledgements page](#) for the full list.

OBO

open biomedical ontologies

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<input type="checkbox"/>	PTEN -induced putative kinase 1		
<input type="checkbox"/>	Pink1_predicted		gene from <i>Rattus norvegicus</i>
	PTEN induced putative kinase 1 (predicted)		
<input type="checkbox"/>	Plip	BLAST	gene from <i>Drosophila melanogaster</i>
	PTEN -like phosphatase		
<input type="checkbox"/>	Pten	BLAST	gene from <i>Mus musculus</i>
	phosphatase and tensin homolog		
<input type="checkbox"/>	Pten		gene from <i>Rattus norvegicus</i>
	phosphatase and tensin homolog		
<input type="checkbox"/>	Pten	BLAST	gene from <i>Drosophila melanogaster</i>
<input type="checkbox"/>	pten	BLAST	gene from <i>Dictyostelium discoideum</i>
	PI3 phosphatase PTEN homolog, protein tyrosine phosphatase, 3-phosphatidylinositol 3-phosphatase		
<input type="checkbox"/>	PTEN_CANFA	BLAST	protein from <i>Canis lupus familiaris</i>
	PTEN , MMAC1 : Phosphatidylinositol-3,4,5-trisphosphate 3-phosphatase PTEN		
<input checked="" type="checkbox"/>	PTEN_HUMAN	BLAST	protein from <i>Homo sapiens</i>
	PTEN , MMAC1 , TEP1 : Phosphatidylinositol-3,4,5-trisphosphate 3-phosphatase and dual-specificity protein phosphatase PTEN		
<input type="checkbox"/>	PTEN_XENLA	BLAST	protein from <i>Xenopus laevis</i>
	pten : Phosphatidylinositol-3,4,5-trisphosphate 3-phosphatase and dual-specificity protein phosphatase PTEN		
<input type="checkbox"/>	ptena	BLAST	gene from <i>Danio rerio</i>
	phosphatase and tensin homolog A		
<input type="checkbox"/>	ptenb	BLAST	gene from <i>Danio rerio</i>
	phosphatase and tensin homolog B (mutated in multiple advanced cancers 1)		
<input type="checkbox"/>	TEP1	BLAST	gene from <i>Saccharomyces cerevisiae</i>
	Homolog of human tumor suppressor gene PTEN / MMAC1 / TEP1 that has lipid phosphatase activity and is linked to the phosphatidylinositol signaling pathway		
<input type="checkbox"/>	Tpte	BLAST	gene from <i>Mus musculus</i>
	transmembrane phosphatase with tensin homology		
	Query matches synonym Ptenj2		



Get FASTA sequences



Get annotation summary

Last updated 2008-02-19

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Search GO ☒ Terms ☐ Genes or proteins ☐ Exact Match

- Filter by ontology

All

Data source

All
CGD
dictyBase
FlyBase

Set filters

[Remove all filters](#)

all : all [477250]

 GO:0008150 : biological_process [318388]

[+](#) [i](#) [GO:0022610 : biological adhesion \[3334\]](#)

 GO:0065007 : biological regulation [44424]

GO:0033667 : negative regulation of growth or development of symbiont within host [0]

GO:0033666 : positive regulation of growth or development of symbiont within host [0]



 GO:0050789 : regulation of biological process [39400]

GO:0048519 : negative regulation of biological process [9366]

GO:0048523 : negative regulation of cellular process [8486]

GO:0043069 : negative regulation of programmed cell death [1554]

GO:0043066 : negative regulation of apoptosis [1516]

  [GO:0006916 : anti-apoptosis \[962\]](#)

 GO:0045767 : regulation of anti-apoptosis [116]



GO:001987 : negative regulation of anti-apoptosis [34]

GO:0045768 : positive regulation of anti-apoptosis [70]

 [GO:0051093](#) : negative regulation of developmental process [2670]

GO:0043069 : negative regulation of programmed cell death [1554]

GO:0043066 : negative regulation of apoptosis [1516]

  GO:0006916 : anti-apoptosis [962]

 GO:0045767 : regulation of anti-apoptosis [116]

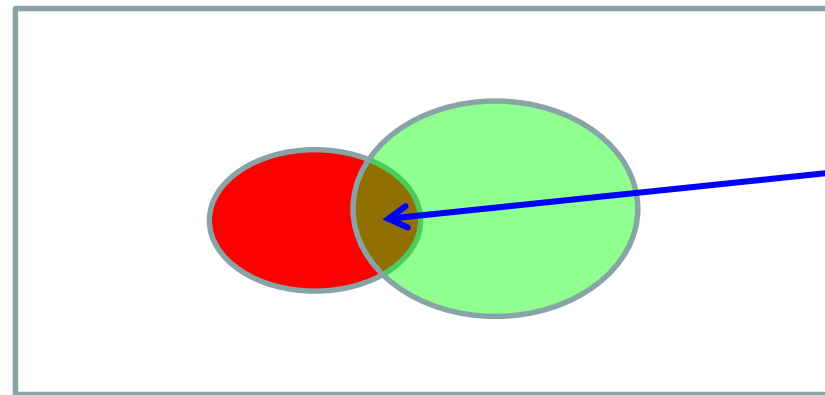
GO:0019987 : negative regulation of anti-apoptosis [34]

GO:0045768 : positive regulation of anti-apoptosis [70]

Graphical View
Permalink
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How do I find commonality from my gene list?

- Using a priori knowledge (e.g., gene ontology, pathway, annotation, etc.)
- Fisher's exact test, hypergeometric test, Bayesian-based methods, etc.

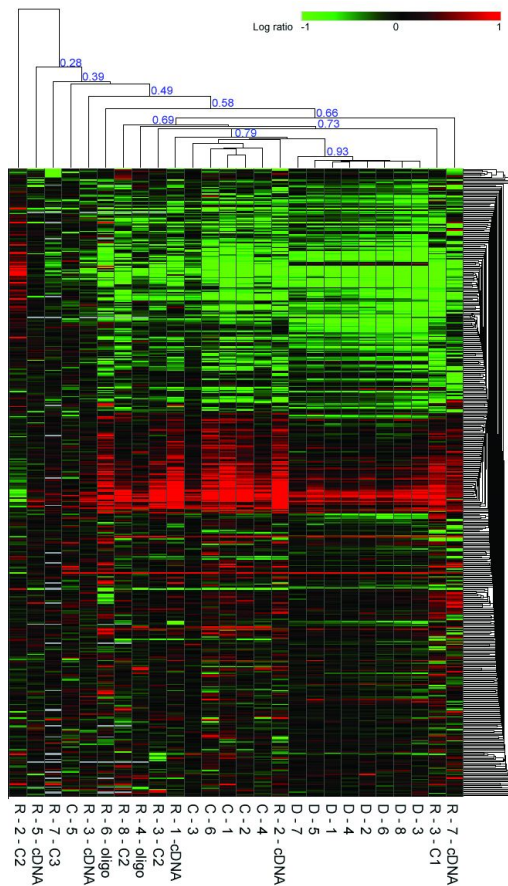


How significant is the intersection?

- Good news – most of the time you can use software to do it

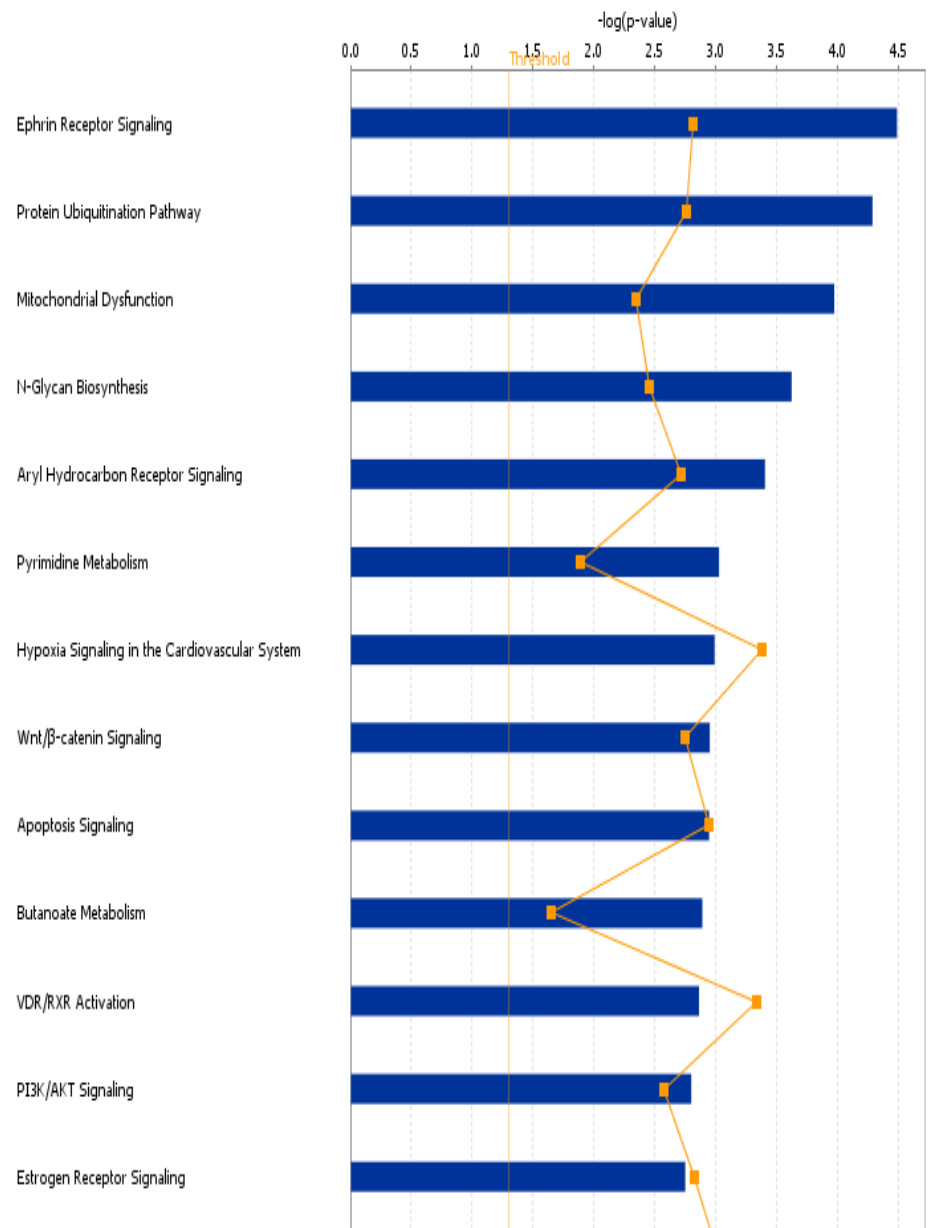
What softwares are available?

- DAVID (<http://david.abcc.ncifcrf.gov/>)
- TOPPGene
- Cytoscape
 - GOTerm
 - BiNGO
- GSEA
- GenMapp (Free)
- Pathway Architect (Commercial)
- Pathway Studio (Commercial)
- Ingenuity Pathway Analysis (Commercial)
 - Manually curated
 - On-demand computation



Supplementary Figure 1: Clustering of laboratory/platform combinations using log ratio values of common genes

Genes



Functions, pathways and networks

Pathway – What's out there?

Address  <http://www.pathguide.org/>

Google  Go   Bookmarks  4905 blocked  Check

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Pathguide» the pathway resource list

Navigation

- Protein-Protein Interactions**
- Metabolic Pathways
- Signaling Pathways
- Pathway Diagrams
- Transcription Factors / Gene Regulatory Networks
- Protein-Compound Interactions
- Genetic Interaction Networks
- Protein Sequence Focused
- Other

Search

Organisms

All 

Complete Listing of All Pathguide Resources

Pathguide contains information about ~~222~~³²⁵ biological pathway resources. Click on a link to go to the resource home page or 'Details' for a description page. Databases that are free and those supporting BioPAX, CellML, PSI-MI or SBML standards are respectively indicated.

If you know of a pathway resource that is not listed here, or have other questions or comments, please [send us an e-mail](#).

Protein-Protein Interactions

Database Name (Order: [alphabetically](#) | [by web popularity](#) )

3DID - 3D interacting domains	Details
ABCdb - Archaea and Bacteria ABC transporter database	Details
AfCS - Alliance for Cellular Signaling Molecule Pages Database	Details
AllFuse - Functional Associations of Proteins in Complete Genomes	Details
ASEdb - Alanine Scanning Energetics Database	Details

News

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are av

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statist



KEGG PATHWAY Database

Wiring diagrams of molecular interactions, reactions, and relations

KEGG2 KID PATHWAY BRITE GENES SSDB LIGAND DRUG DBGET

Pathway Maps

KEGG PATHWAY is a collection of manually drawn pathway maps representing our knowledge on the molecular interaction and reaction networks for:

1. Metabolism

Carbohydrate Energy Lipid Nucleotide Amino acid Other amino acid
Glycan PK/NRP Cofactor/vitamin Secondary metabolite Xenobiotics

2. Genetic Information Processing

3. Environmental Information Processing

4. Cellular Processes

5. Human Diseases

and also on the structure relationships (KEGG drug structure maps) in:

6. Drug Development

 Search PATHWAY for

☒ bfind mode ☐ bget mode

1. Metabolism

1.1 Carbohydrate Metabolism

Glycolysis / Gluconeogenesis
Citrate cycle (TCA cycle)
Pentose phosphate pathway
Pentose and glucuronate interconversions
Fructose and mannose metabolism
Galactose metabolism
Ascorbate and aldarate metabolism
Starch and sucrose metabolism
Aminosugars metabolism
Nucleotide sugars metabolism
Pyruvate metabolism
Glyoxylate and dicarboxylate metabolism
Propanoate metabolism
Butanoate metabolism
C5-Branched dibasic acid metabolism
Inositol metabolism
Inositol phosphate metabolism

1.2 Energy Metabolism

Oxidative phosphorylation *Revised!*
Photosynthesis *Revised!*
Photosynthesis - antenna proteins *New!*

KEGG Orthology (KO)

KEGG pathway modules
Overview of biosynthetic pathways

Enzymes (+diseases)
Compounds with biological roles

Photosynthesis proteins

File Edit View Favorites Tools Help

Back Forward Stop Home Search Favorites Reload Print Mail Print Mail Print Mail

Address http://www.kegg.com/kegg/pathway/map/map00380.html

Go Links

Google

Go

Bookmarks

4905 blocked

Check

AutoLink

AutoFill

Send to

Settings



Tryptophan metabolism - Reference pathway

[Pathway menu]

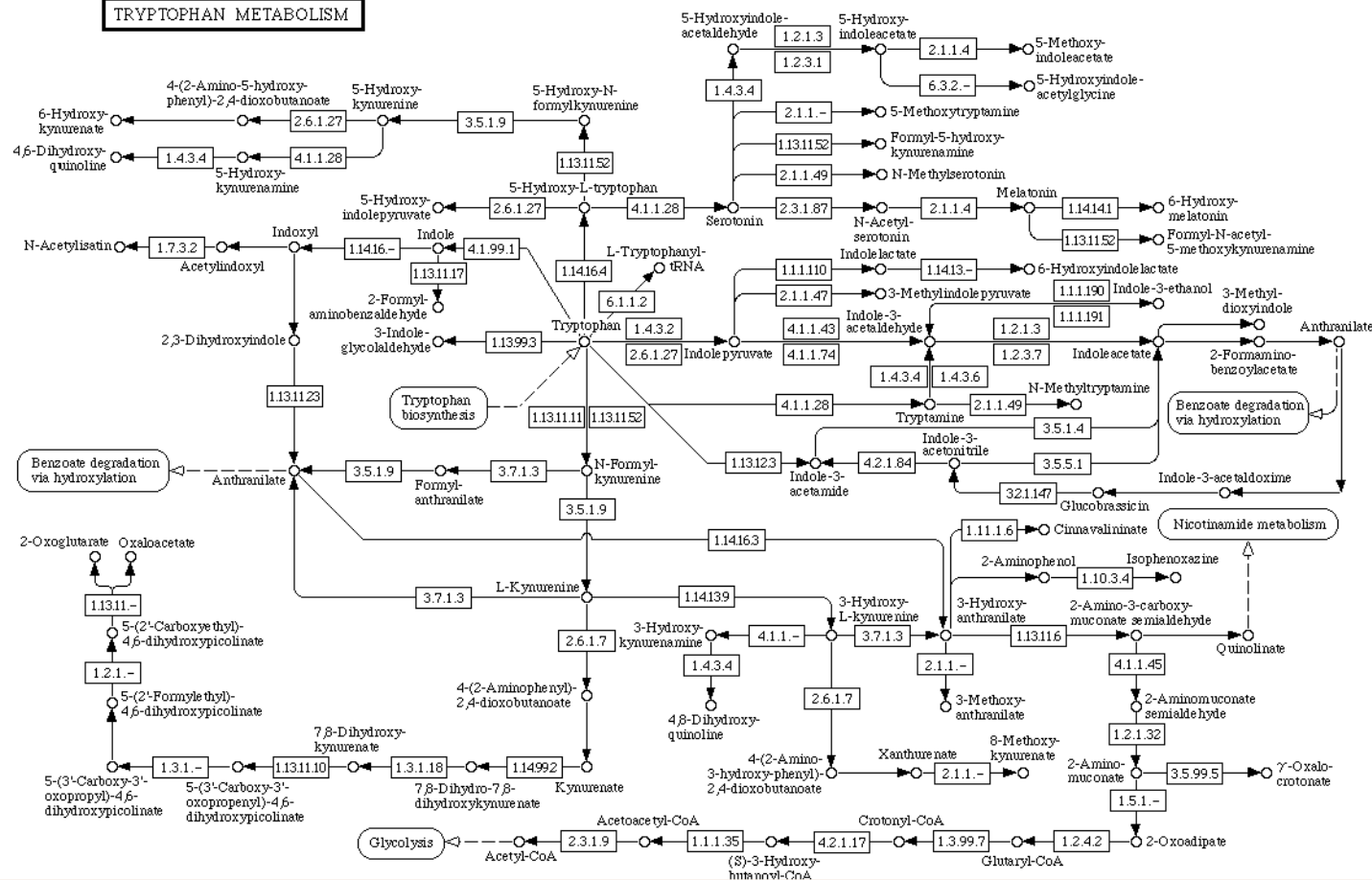
Reference pathway

Go

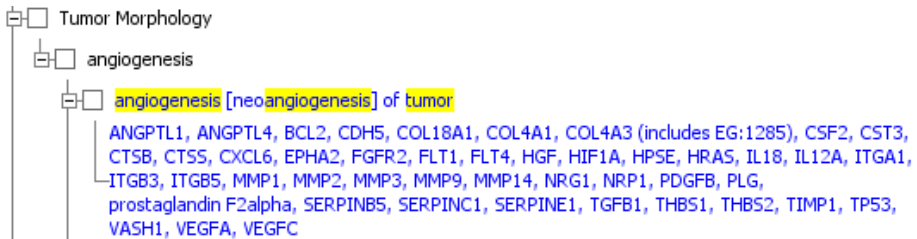
Current selection Select

Help

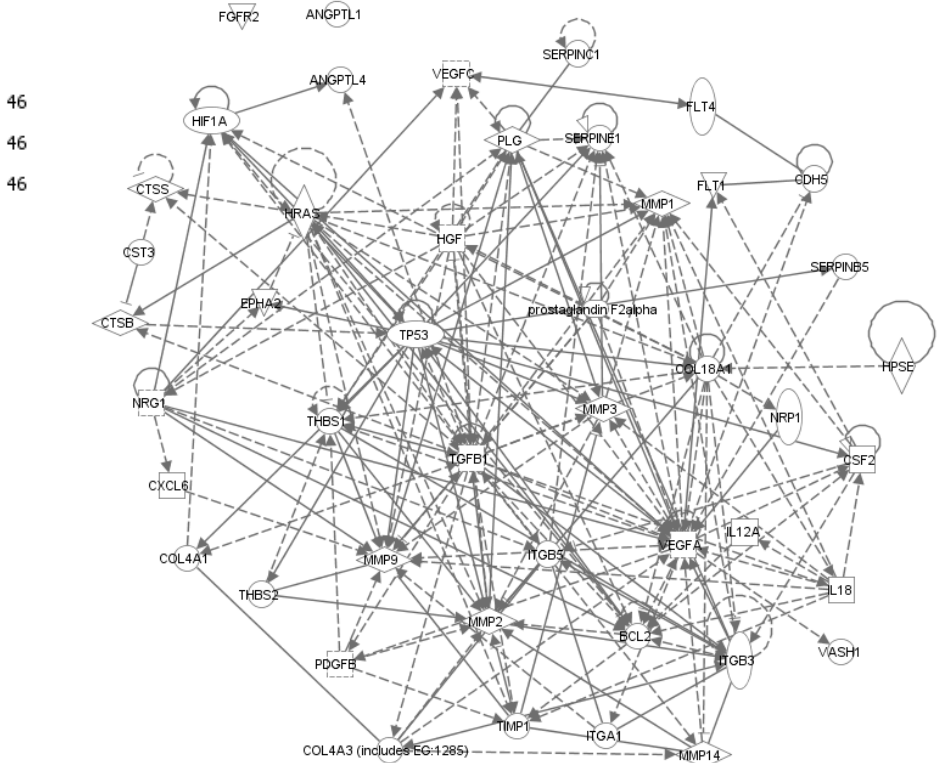
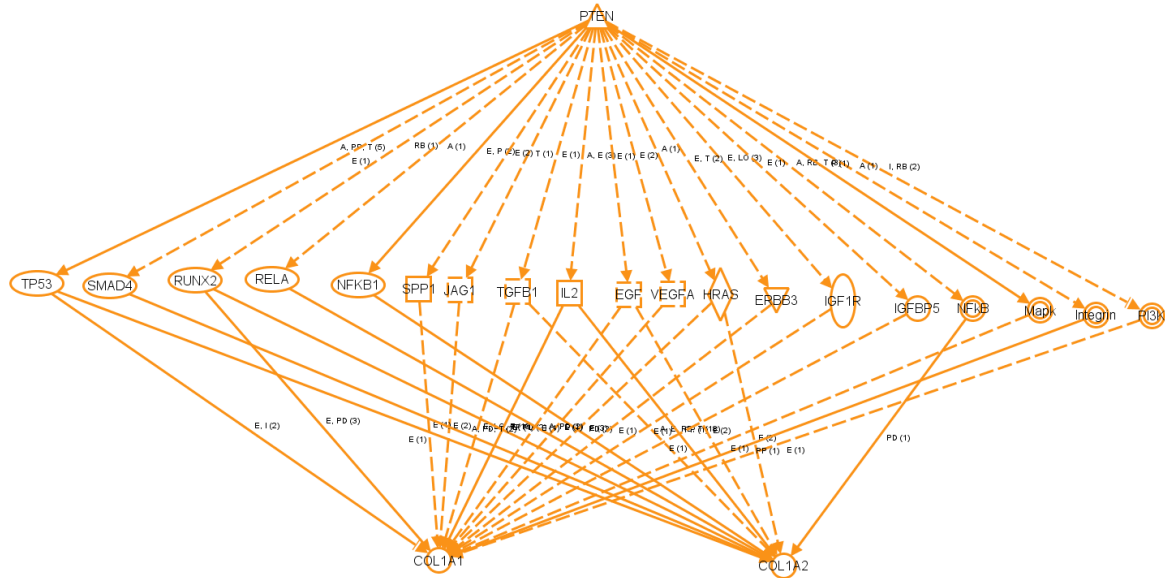
TRYPTOPHAN METABOLISM



Ingenuity Pathway Analysis (IPA)



New Pathway 6



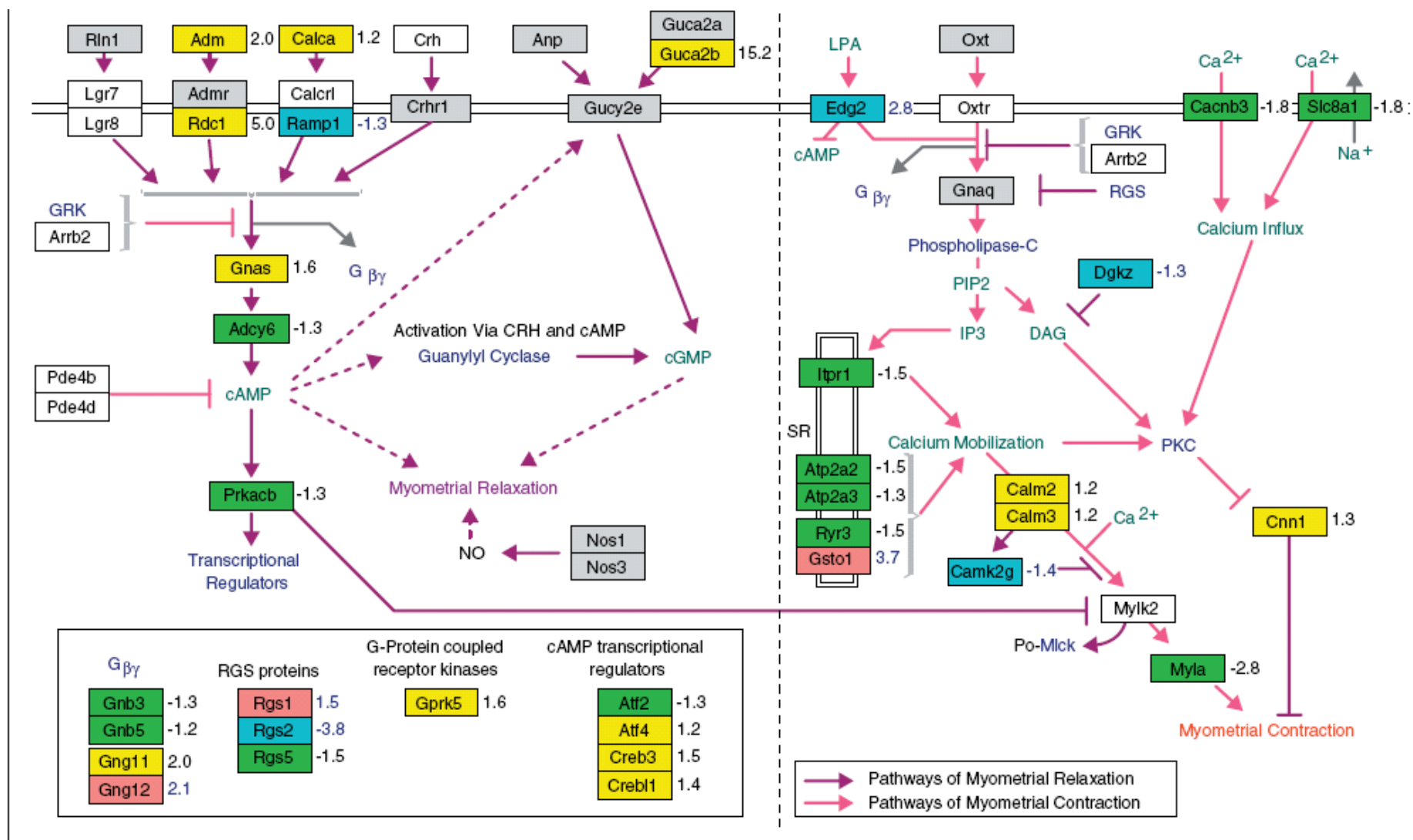


Figure 3
 Analysis of pathways of uterine smooth muscle contraction. (a) Prostaglandin synthesis and (b) G-protein signaling pathways in the myometrium are overlaid with gene-expression color criterion and fold-changes from the program GenMAPP. Interactions suggested by results of this microarray analysis are included in these figures. Detailed gene-expression data, statistics and full gene annotations are available on the GenMAPP interactive version of these pathways online [40].

Demo

- DAVID (<http://david.abcc.ncifcrf.gov/>)
- TOPPGene
- Ingenuity Pathway Analysis

Gene List1: AURKA BIRC5ASPM BUB1 CCNA2 CCNB2 CDC2
ACOT7 CDC20 CDC45L CDCA8 CENPE CENPF CEP55 CKS2
CHEK1 DKFZp762E1312 DLG7 DNA2L E2F8 EPR1
FANCI HMMR KIF4A LMNB1 MAD2L1 MELK NCAPG
RANBP1RRM2 SPAG5 STIL TACC3 TPX2 TRIP13 TTK
UBE2C UBE2S

Gene List2: AI445650 CD2 CCR5 CD247 CD27 CD38 CD3D
CD3E CD3G CD79A CD8A CRTAM CST7 CTSW
CXCR6 DENND2D FAIM3 FMNL1 GZMA GZMB GZMH
GZMK HLA-DOB IL21R IL2RB IL2RG IL7R KLRK1
LAG3 LAT LAX1 MIRN650 NKG7 NM_014792
PTPN7 RASGRP1 RUNX3 SELPLGSEPT6 SERPINB9
SH2D1ASIRPG SLAMF7SOCS1 TBX21 TRBC1 WAS XCL1
CCL4 XCL2 ZAP70